

## Original Research Article

# IN VITRO ANTIBACTERIAL ACTIVITY OF HYDROETHANOLIC LEAF, STEM, ROOT EXTRACT OF *ACALYPHA INDICA* - A COMPARATIVE STUDY

**Running title:** Antibacterial activity of *Acalypha indica* extract

### **ABSTRACT:**

**BACKGROUND AND AIM:** *Acalypha indica* is a weedy, short lived and monoecious plant. It has an antioxidant effect and it must be implemented to control the disease where free radicals are involved. *Acalypha* plant is useful for the bronchitis, pneumonia, pulmonary tuberculosis patients. *A.indica* plant is used in the rejuvenation in the worst conditions which may undergo infections in the microorganisms and it is the chemotherapeutic agent and distributed in the large contributions in human health and well being. The main aim of this study is to assess which part of *A. indica* has an antibacterial activity.

**MATERIALS AND METHODS:** Agar well diffusion method was used for assessing the antimicrobial activity of the plant extract. The nutrient broth is inoculated with bacterial strains *E. faecalis*. The broth was then incubated at 37°C overnight. Antibacterial activity was determined by measurement of the diameter of zones of inhibition (mm).

**RESULT:** When compared to root, stem and leaves of hydroethanolic extract of *A.indica*, leaf maximum inhibition against *E. faecalis*. As the concentration of extract increases from 25 µg/ml to 100 µg/ml, the zone of inhibition also increases. This shows the antimicrobial activity is better at higher concentration.

**CONCLUSION:** *A.indica* has effective antimicrobial activity. It reduces the growth of various human pathogenic bacteria. The root, stem and leaf of *A.indica* possess Antibacterial activity against *E. faecalis*.leaf extract has significant antibacterial activity.

**KEYWORDS:** Antibacteria, Antimicrobial, phytochemicals, inhibition, Strains, Innovative techniques.

## INTRODUCTION:

*A.indica* Linn was also called as kucinggalak. It is widely distributed in Tropical Africa, South Africa, India and Sri Lanka.(1). *A.indica* is a weed, short lived and monoecious plant. *A.indica* Linn is an annual plant and it is an erect herb. This plant is commonly called in Tamil as 'KUPPAIMENI'. It belongs to Euphorbiaceae. This plant can grow up to 1.5 to 2.5 m tall. *A.indica* plant contains the acalyphine which is used to treat deficiency of Vitamin C, Vitamin E patients. Leaf of the acalypha plant has been used for treatment of scabies and the other cutaneous diseases. It is useful to treat rheumatism and several other ailments (2). *A.indica* is most diverse within nearly 450 species. 2/3rd species found in America, 19 found in Venezuela and they are mainly used as an ornamental plant (3)(4).

It has an antioxidant effect and it must be implemented to control the disease where free radicals are involved. *Acalypha* plant is useful for the bronchitis, pneumonia, pulmonary tuberculosis patients (5). These plants are diuretic, emetic, expectorant, laxative. This plant has beneficial effects on nosocomial infections and the bacterial pathogens (6). This plant also has beneficial effects on asthma (7). It is a herbal plant and grows in the wet temperate and tropical regions. The plant can be used in medicine and in several therapeutic treatments.(6,8) *A.indica* plant is used in the rejuvenation in the worst conditions which may undergo infections in the microorganisms and it is the chemotherapeutic agent and distributed in the large contributions in the human health and the well being (9,10). Nearly 88% of the global population's derived medicines from these plants and it act as the first line of defense for maintaining health and combating the disease (11).

It can be used for anthelmintic, antiulcer, bronchitis (12), antidiabetic, anti hyperlipidemic, anti obesity, antivenom, hepatoprotective and hypoxia (13). Extracts from the stem, root, leaf can make the drugs valuable and it has a high export potential (9). The dried leaves of the *A.indica* were made into a poultice to treat the bedsores and the wounds. (14) The leaves of *A.indica* have also been reported to possess contraceptive activity. The juice of this Euphorbiaceae plant is added to oil or lime and used to treat a variety of skin disorders. These plants possess the bronchodilation and the bronchial hyperreactivity. Essential oil which is anhydroethanolic extract has the positive outcome in the in vitro study and has the minimum inhibitory concentration methods (15). It has acaricidal effects. It is also expectorant against pneumonia

and also as an emmenagogue. This plant is also known to possess respiratory effects on experimental animals. *Acalypha* is one of the genres that show a great potential in the world of the scientific advancement due to its chemical and biological results (16). To enrich the knowledge of the antibacterial activity of *A.indica* plant extract against the gram positive and the gram negative bacteria (17).

Our team has extensive knowledge and research experience that has translated into high quality publications (18–20)(21–26),(27)(28),(29)(30),(31)(32)(33–37). So the main aim of this study is to assess which part of *A.indica* has a potential antibacterial activity.

## **MATERIALS AND METHODS:**

### **Test Organisms**

The bacterial strains such as *Enterococcus faecalis*, were obtained from the Department of Microbiology, Saveetha Dental College, Saveetha University were used for the research. They were maintained in a nutrient agar slope at 40°C.

### **Collection of Plant Powder**

#### **Leaf, Stem and Root extract of *A.indica***

The leaves stem and root of *A.indica* was obtained in powder form from a registered pharmacy in Arumbakkam, Chennai, India

### **Preparation of Extract**

The powders obtained were subjected to extraction. Soxhlet extractor was used for this purpose. After the completion of the process of extraction, the solvent is distilled and the extracts are kept in a desiccator after being concentrated to dry residue on a water bath.

### **Assessment of Antimicrobial Activity by Agar Well Diffusion Method**

Agar well diffusion method was used for assessing the antimicrobial activity of the plant extract. The nutrient broth was inoculated with *E. faecalis*. The broth was then incubated at 37°C overnight. The culture was then adjusted to 0.5 McFarland turbidity standard. Muller-Hinton agar plates [MHA-HiMedia M1084] were used for the Lawn culture of the test organism. This was done with using sterile cotton q tips. The plates were clean and dry. Then, a 6 mm diameter well was bored by a sterile cork for different concentrations of the extracts (25, 50, 75 and 100 µg/ml). The extracts were introduced into the wells using micropipettes. The culture plates were allowed to stand on the bench for 30 min for pre-diffusion and were then incubated in an upright position for 24 h at 37°C. After 24 h, antibacterial activity was determined by measurement of

the diameter of zones of inhibition (mm). To minimize the test error, all the tests are done in triplicate.

## RESULTS:

When comparing with root, stem and leaves of hydroethanolic extract of *A.indica*, leaf extract shown the maximum inhibition against *E. faecalis*. As the concentration of extract increases from 25µg/ml to 100 µg/ml, the zone of inhibition also increases. This shows the antimicrobial activity is better with increase in concentration. According to this study, hydroethanolic extract of stem, root and leaves of *A.indica* was effective against the bacterial strain *E.faecalis*. In the lower concentration 20 µg/ml of extract, the zone of inhibition of root was 15 mm, stem 16 mm, and leaves 17mm (Table 1). When compared, leaf extract had a larger zone of inhibition than root and stem extract. As the concentration of all parts of plant extract increases, the zone of inhibition gradually increases. The *E. faecalis* served as a positive control while distilled water was used as negative control. These were tested using disc diffusion methods. The results show less antibacterial activity on the root extract of *A.indica* plant (Figure 1). Culture plate showed little higher antibacterial activity on the stem extract of *A.indica* plant (Figure 2). Culture plate showed much higher antibacterial activity on the leaf than the root and stem extracts of *A.indica* plant (Figure 3).

**TABLE 1:** Zone of inhibition(diameter mm). Effect of antibacterial activity of leaf, stem and root extract of *A.indica* against *E.faecalis*

Extract	25 µg/ml	50 µg/ml	75 µg/ml	100 µg/ml
<i>A.indica root</i>	15	16	18	21
<i>A.indica stem</i>	16	18	18	20
<i>A.indica leaf</i>	17	19	19	21

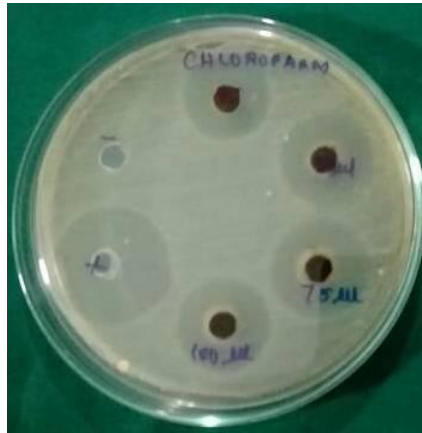


Figure 1: Culture plate showing efficacy of antibacterial activity of *A.indica* root extract was tested with four different concentrations which were 25  $\mu\text{g/ml}$ , 50  $\mu\text{g/ml}$ , 75  $\mu\text{g/ml}$ , 100  $\mu\text{g/ml}$ .

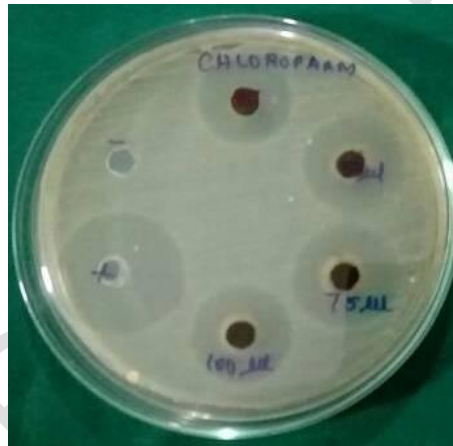


Figure 2: Culture plate showing efficacy of antibacterial activity of *A.indica* stem extract was tested with four different concentrations which were 25  $\mu\text{g/ml}$ , 50  $\mu\text{g/ml}$ , 75  $\mu\text{g/ml}$ , 100  $\mu\text{g/ml}$ .

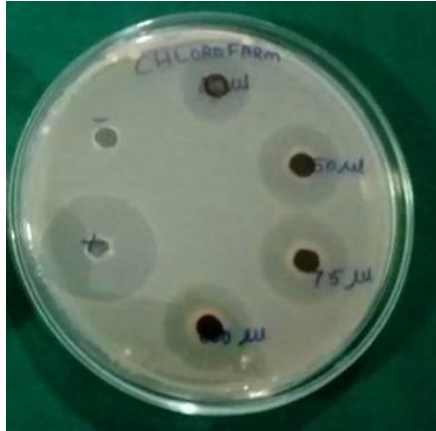


Figure 3: Culture plate showing efficacy of antibacterial activity of *A.indica* leaf extract was tested with four different concentrations which were 25 µg/ml, 50 µg/ml, 75 µg/ml, 100 µg/ml.

#### **DISCUSSION:**

*A.indica* species destroys the growth of all gram positive bacteria but it didn't act on gram negative bacteria (38). Due to its antioxidant activity, it fights against many diseases (39). When compared to ethanol, methanol and acetone extract of root, stem and leaf extract of *A.indica*, ethanol leaf extract has potential antimicrobial activity with the zone of inhibition 20cm against *K.pneumoniae*(40). Antibacterial activity of the hydroethanolic root extract of the *A.indica* carried out by a maceration method (41). Phytochemical quantitative analysis of the total flavonoid content and the alkaloid content (42,43). Aqueous and ethanolic extract of *A.indica* manifested moderately antibacterial activity against gut pathogens (44). *A.indica* has effective antimicrobial activity (45). Antimicrobial activity is due to the presence of phytochemicals like tannins, flavonoids and phenolics(46). The presence of bioactive compounds such as alkaloids, tannins, steroids, saponins, flavonoids, glycosides and the phenolic compounds was also detected during the phytochemical testing (47). *A.indica* needs more extensive laboratory and clinical work in order to know preferable antibacterial principles (41).

As it stated earlier, the antibacterial activity of leaf stem and root extract is worthy of investigation. But, in the *A.indica* plant for further laboratory and clinical studies of this plant

was required in order to understand better in antimicrobial principles which will allow the scientific community to recommend their uses.

## CONCLUSION:

*A.indica* has effective antibacterial activity. It reduces the growth of various human pathogenic bacteria. The root, stem and leaf of *A.indica* possess Antibacterial activity against *E.faecalis*. Leaf has significant antibacterial activity. Even though there are many drugs, still there is no complete cure. No side effects as they are natural.

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## REFERENCES:

1. Sridharan G, Ramani P, Patankar S, Vijayaraghavan R. Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma [Internet]. Vol. 48, Journal of Oral Pathology & Medicine. 2019. p. 299–306. Available from: <http://dx.doi.org/10.1111/jop.12835>
2. Mothana RAA, Abdo SAA, Hasson S, Althawab FMN, Alaghbari SAZ, Lindequist U. Antimicrobial, antioxidant and cytotoxic activities and phytochemical screening of some yemeni medicinal plants. Evid Based Complement Alternat Med. 2010 Sep;7(3):323–30.
3. PerumalSamy R, Ignacimuthu S, Raja DP. Preliminary screening of ethnomedicinal plants from India. J Ethnopharmacol. 1999 Aug;66(2):235–40.
4. Saraswathi I, Saikarthik J, Senthil Kumar K, Srinivasan KM, Ardhanaari M, Gunapriya R. Impact of COVID-19 outbreak on the mental health status of undergraduate medical students in a COVID-19 treating medical college: a prospective longitudinal study [Internet]. Vol. 8, PeerJ. 2020. p. e10164. Available from: <http://dx.doi.org/10.7717/peerj.10164>
5. Gangadev V. The antibacterial activity of *Acalypha indica* L [Internet]. Vol. 1, Indian Journal of Science and Technology. 2008. p. 1–5. Available from: <http://dx.doi.org/10.17485/ijst/2008/v1i6.2>
6. Egbuna C, Mishra AP, Goyal MR. Preparation of Phytopharmaceuticals for the

Management of Disorders: The Development of Nutraceuticals and Traditional Medicine. Academic Press; 2020.570 p.

7. U A, Ashwini U, Asha S. IN VITRO ANTIBACTERIAL ACTIVITY OF ACALYPHA INDICA LINN LEAVES EXTRACT AGAINST GRAM NEGATIVE BACTERIA [Internet]. Vol. 8, International Journal of Research in Ayurveda & Pharmacy. 2017. p. 195–8. Available from: <http://dx.doi.org/10.7897/2277-4343.083198>
8. Ezhilarasan D. Critical role of estrogen in the progression of chronic liver diseases [Internet]. Vol. 19, Hepatobiliary & Pancreatic Diseases International. 2020. p. 429–34. Available from: <http://dx.doi.org/10.1016/j.hbpd.2020.03.011>
9. Mutiarawati DT. In Vitro Anthelmintic Activity of Acalypha Indica Leaves Extracts [Internet]. Vol. 4, Health Notions. 2020. p. 94–9. Available from: <http://dx.doi.org/10.33846/hn40305>
10. Adnan M, Ali S, Sheikh K, Amber R. Review on antibacterial activity of Himalayan medicinal plants traditionally used to treat pneumonia and tuberculosis. J Pharm Pharmacol. 2019 Nov;71(11):1599–625.
11. Santiago C, Pang EL, Lim K-H, Loh H-S, Ting KN. Reversal of ampicillin resistance in MRSA via inhibition of penicillin-binding protein 2a by Acalyphawilkesiana. Biomed Res Int. 2014 Jun 30;2014:965348.
12. Gangadevi V. Screening endophytic fungi isolated from a medicinal plant, Acalypha indica L. for antibacterial activity [Internet]. Vol. 1, Indian Journal of Science and Technology. 2008. p. 1–6. Available from: <http://dx.doi.org/10.17485/ijst/2008/v1i5.11>
13. Sivaraj R, Rahman PKSM, Rajiv P, Narendhran S, Venckatesh R. Biosynthesis and characterization of A.indica mediated copper oxide nanoparticles and evaluation of its antimicrobial and anticancer activity. SpectrochimActa A MolBiomolSpectrosc. 2014 Aug 14;129:255–8.
14. Shabgah AG, Ezzatifar F, Aravindhana S, Zekiy AO, Ahmadi M, Gheibihayat SM, et al. Shedding more light on the role of Midkine in hepatocellular carcinoma: New perspectives on diagnosis and therapy [Internet]. Vol. 73, IUBMB Life. 2021. p. 659–69. Available from: <http://dx.doi.org/10.1002/iub.2458>
15. Navarro MC, Montilla MP, Cabo MM, Galisteo M, Cáceres A, Morales C, et al. Antibacterial, antiprotozoal and antioxidant activity of five plants used in Izabal for infectious diseases. Phytother Res. 2003 Apr;17(4):325–9.
16. Govindarajan M, Jebanesan A, Reetha D, Amsath R, Pushpanathan T, Samidurai K. Antibacterial activity of A.indica L. Eur Rev Med Pharmacol Sci. 2008 Sep;12(5):299–302.
17. Mudigonda SK, Murugan S, Velavan K, Thulasiraman S, Krishna Kumar Raja V. Non-suturing microvascular anastomosis in maxillofacial reconstruction- a comparative study [Internet]. Vol. 48, Journal of Cranio-Maxillofacial Surgery. 2020. p. 599–606. Available

from: <http://dx.doi.org/10.1016/j.jcms.2020.04.005>

18. Saraswathi I, Saikarthik J, Senthil Kumar K, MadhanSrinivasan K, Ardhanaari M, Gunapriya R. Impact of COVID-19 outbreak on the mental health status of undergraduate medical students in a COVID-19 treating medical college: a prospective longitudinal study. *PeerJ*. 2020 Oct 16;8:e10164.
19. Santhakumar P, Roy A, Mohanraj KG, Jayaraman S, Durairaj R. Ethanolic Extract of *Capparis decidua* Fruit Ameliorates Methotrexate-Induced Hepatotoxicity by Activating Nrf2/HO-1 and PPAR $\gamma$  Mediated Pathways. *Ind J Pharm Educ*. 2021 Mar 19;55(1s):s265–74.
20. Nambi G, Kamal W, Es S, Joshi S, Trivedi P. Spinal manipulation plus laser therapy versus laser therapy alone in the treatment of chronic non-specific low back pain: a randomized controlled study. *Eur J PhysRehabil Med*. 2018 Dec;54(6):880–9.
21. Rajakumari R, Volova T, Oluwafemi OS, Rajesh Kumar S, Thomas S, Kalarikkal N. Grape seed extract-soluplus dispersion and its antioxidant activity. *Drug DevInd Pharm*. 2020 Aug;46(8):1219–29.
22. Clarizia G, Bernardo P. Diverse Applications of Organic-Inorganic Nanocomposites: Emerging Research and Opportunities: Emerging Research and Opportunities. IGI Global; 2019.237 p.
23. Prakash AKS, Devaraj E. Cytotoxic potentials of *S. cumini* methanolic seed kernel extract in human hepatoma HepG2 cells [Internet]. Vol. 34, *Environmental Toxicology*. 2019. p. 1313–9. Available from: <http://dx.doi.org/10.1002/tox.22832>
24. Tahmasebi S, Qasim MT, Krivenkova MV, Zekiy AO, Thangavelu L, Aravindhan S, et al. The effects of oxygen-ozone therapy on regulatory T-cell responses in multiple sclerosis patients. *Cell Biol Int*. 2021 Jul;45(7):1498–509.
25. Wadhwa R, Paudel KR, Chin LH, Hon CM, Madheswaran T, Gupta G, et al. Anti-inflammatory and anticancer activities of Naringenin-loaded liquid crystalline nanoparticles in vitro. *J Food Biochem*. 2021 Jan;45(1):e13572.
26. Vivekanandhan K, Shanmugam P, Barabadi H, Arumugam V, Daniel Raj Daniel Paul Raj D, Sivasubramanian M, et al. Emerging Therapeutic Approaches to Combat COVID-19: Present Status and Future Perspectives. *Front MolBiosci*. 2021 Mar 8;8:604447.
27. Ezhilarasan D. Critical role of estrogen in the progression of chronic liver diseases. *HepatobiliaryPancreat Dis Int*. 2020 Oct;19(5):429–34.
28. Egbuna C, Mishra AP, Goyal MR. Preparation of Phytopharmaceuticals for the Management of Disorders: The Development of Nutraceuticals and Traditional Medicine. Academic Press; 2020.574 p.
29. Kamath SM, Manjunath Kamath S, Jaison D, Rao SK, Sridhar K, Kasthuri N, et al. In vitro

- augmentation of chondrogenesis by Epigallocatechin gallate in primary Human chondrocytes - Sustained release model for cartilage regeneration [Internet]. Vol. 60, Journal of Drug Delivery Science and Technology. 2020. p. 101992. Available from: <http://dx.doi.org/10.1016/j.jddst.2020.101992>
30. Barabadi H, Mojab F, Vahidi H, Marashi B, Talank N, Hosseini O, et al. Green synthesis, characterization, antibacterial and biofilm inhibitory activity of silver nanoparticles compared to commercial silver nanoparticles [Internet]. Vol. 129, Inorganic Chemistry Communications. 2021. p. 108647. Available from: <http://dx.doi.org/10.1016/j.inoche.2021.108647>
  31. Bharath B, Perinbam K, Devanesan S, AlSalhi MS, Saravanan M. Evaluation of the anticancer potential of Hexadecanoic acid from brown algae *Turbinaria ornata* on HT-29 colon cancer cells [Internet]. Vol. 1235, Journal of Molecular Structure. 2021. p. 130229. Available from: <http://dx.doi.org/10.1016/j.molstruc.2021.130229>
  32. GowhariShabgah A, Ezzatifar F, Aravindhan S, OlegovnaZekiy A, Ahmadi M, Gheibihayat SM, et al. Shedding more light on the role of Midkine in hepatocellular carcinoma: New perspectives on diagnosis and therapy. *IUBMB Life*. 2021 Apr;73(4):659–69.
  33. Sridharan G, Ramani P, Patankar S, Vijayaraghavan R. Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma. *J Oral Pathol Med*. 2019 Apr;48(4):299–306.
  34. R H, Hannah R, Ramani P, Ramanathan A, Jancy MR, Gheena S, et al. CYP2 C9 polymorphism among patients with oral squamous cell carcinoma and its role in altering the metabolism of benzo[a]pyrene [Internet]. Vol. 130, Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology. 2020. p. 306–12. Available from: <http://dx.doi.org/10.1016/j.oooo.2020.06.021>
  35. J PC, Pradeep CJ, Marimuthu T, Krithika C, Devadoss P, Kumar SM. Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study [Internet]. Vol. 20, Clinical Implant Dentistry and Related Research. 2018. p. 531–4. Available from: <http://dx.doi.org/10.1111/cid.12609>
  36. Wahab PUA, Madhulaxmi M, Senthilnathan P, Muthusekhar MR, Vohra Y, Abhinav RP. Scalpel Versus Diathermy in Wound Healing After Mucosal Incisions: A Split-Mouth Study. *J Oral Maxillofac Surg*. 2018 Jun;76(6):1160–4.
  37. Mudigonda SK, Murugan S, Velavan K, Thulasiraman S, Krishna Kumar Raja VB. Non-suturing microvascular anastomosis in maxillofacial reconstruction- a comparative study. *Journal of Cranio-Maxillofacial Surgery*. 2020 Jun 1;48(6):599–606.
  38. Govindarajan M, Jebanesan A, Pushpanathan T, Samidurai K. Studies on effect of *Acalypha indica* L. (Euphorbiaceae) leaf extracts on the malarial vector, *Anopheles stephensi* Liston (Diptera: Culicidae) [Internet]. Vol. 103, Parasitology Research. 2008. p. 691–5. Available from: <http://dx.doi.org/10.1007/s00436-008-1032-2>

39. Nambi G, Kamal W, Es S, Joshi S, Trivedi P. Spinal manipulation plus laser therapy versus laser therapy alone in the treatment of chronic non-specific low back pain: a randomized controlled study [Internet]. Vol. 54, European Journal of Physical and Rehabilitation Medicine. 2019. Available from: <http://dx.doi.org/10.23736/s1973-9087.18.05005-0>
40. Odeja O, Ogwuche CE, Elemike EE, Obi G. Phytochemical screening, antioxidant and antimicrobial activities of *Acalypha acida* plant. Clinical Phytoscience. 2016 Jun 21;2(1):1–6.
41. Rajakumari R, Volova T, Oluwafemi OS, Rajesh Kumar S, Thomas S, Kalarikkal N. Grape seed extract-soluplus dispersion and its antioxidant activity [Internet]. Vol. 46, Drug Development and Industrial Pharmacy. 2020. p. 1219–29. Available from: <http://dx.doi.org/10.1080/03639045.2020.1788059>
42. G. GJ, Goptep, G. J, Agada, G. OA, Gbise, et al. Antibacterial activity of ethanolic extract of *Acalypha wilkesiana* leaves growing in Jos, Plateau State, Nigeria [Internet]. Malaysian Journal of Microbiology. 2010. Available from: <http://dx.doi.org/10.21161/mjm.21309>
43. Ishak FD. In Vitro Study of Antimicrobial Activity of *Acalypha Indica* Linn. Extract [Internet]. Vol. 4, The Open Conference Proceedings Journal. 2013. p. 57–60. Available from: <http://dx.doi.org/10.2174/2210289201304020057>
44. Priya VT, Thamil Priya V, Department of Chemistry, VHNSN College, - V, Nadu T, et al. Phytochemical Properties of *Acalypha indica* (L), and its Antimicrobial Potential against Human Pathogens [Internet]. Vol. 14, Journal of Pure and Applied Microbiology. 2020. p. 319–26. Available from: <http://dx.doi.org/10.22207/jpam.14.1.33>
45. Santhakumar P, Roy A, Mohanraj KG, Jayaraman S, Durairaj R. Ethanolic Extract of *Capparis decidua* Fruit Ameliorates Methotrexate-Induced Hepatotoxicity by Activating Nrf2/HO-1 and PPAR $\gamma$  Mediated Pathways [Internet]. Vol. 55, Indian Journal of Pharmaceutical Education and Research. 2021. p. s265–74. Available from: <http://dx.doi.org/10.5530/ijper.55.1s.59>
46. Basha SK, Khaleel Basha S, Sudarsanam G, Silar Mohammad M, Parveen N. Investigations on anti-diabetic medicinal plants used by Sugali tribal inhabitants of Yerramalais of Kurnool district, Andhra Pradesh, India [Internet]. Vol. 4, Stamford Journal of Pharmaceutical Sciences. 2012. p. 19–24. Available from: <http://dx.doi.org/10.3329/sjps.v4i2.10435>
47. Fitri K, Khairani TN, Sinaga FA, Talunohi AG. Burn Wound Healing Activity of Ethanolic Extract of *Acalypha indica* in Ointment Formulated against Rabbits (*Oryctolagus cuniculus*) [Internet]. Vol. 8, Asian Journal of Pharmaceutical Research and Development. 2020. p. 18–20. Available from: <http://dx.doi.org/10.22270/ajprd.v8i3.740>