

ANTIMICROBIAL ACTIVITY OF SELENIUM NANOPARTICLES AND *PTEROCARPUS SANTALINUS* BASED MOUTHWASH

RUNNING TITLE - Antimicrobial activity of Selenium nanoparticles and *Pterocarpus santalinus* based mouthwash

ABSTRACT

Introduction - Their unique size-dependent properties make these materials superior and indispensable in many areas of human activity. Selenium possesses excellent photo electrical and semiconductor properties which make it extensively used in duplicate, photography, cells and rectifiers. Selenium is also one of essential trace elements in the human body and has great importance in nourishment and medicine. In India, medicinal plants are widely used by all sections of people either directly as folk remedies or in different indigenous systems of medicine or indirectly in the pharmaceutical preparations of modern medicines. The aim of the present study is to determine antimicrobial activity of selenium nanoparticles and *Pterocarpus santalinus* based mouthwash.

Materials and methods - Antimicrobial activity of respective nanoparticles against the strain staphylococcus aureus, Bacillus and E Coli. MH Aagar was utilized for this activity to determine the zone of inhibition. Muller hinton agar was prepared and sterilized for 45 minutes at 120 lbs. Media poured into the sterilized plates and let them stabilize for solidification. The wells were cut using the well cutter and the test organisms were swabbed. The nanoparticles with different concentrations were loaded and the plates were incubated for 24 hours at 37 degree celsius. After the incubation the zones of inhibition were measured.

Result - The highest zones of inhibition were exhibited against two bacterial strains Staphylococcus aureus and Staphylococcus mutans proving that it acts as a good antibacterial against S.aureus and S.mutans infections.

KEYWORDS: *Pterocarpus santalinus*, selenium, nanoparticle, microbial activity

INTRODUCTION

In India, medicinal plants are widely used by all sections of people either directly as folk remedies or in different indigenous systems of medicine or indirectly in the pharmaceutical preparations of modern medicines (1), (2,3). A survey by UNCTAD has shown that 33% of total drugs produced by the industrialized nations are plant derived and microbes are considered, 60% of medicinal products are of natural origin. (4), (5)(6). Rig Veda mentions 67 plants having therapeutic effects, Yajurveda lists 81 plants and Atharveda 290 plants (7)(7,8). The World Health Organization recently compiled a list of over 20 000 medicinal plant species. Medicinal plants and their products from India are used to treat a variety of ailments including catarrh, bronchitis, pneumonia, ulcers, and diarrhoea (9), (10), (10,11).

Researchers are increasingly turning their attention to folk medicine looking for new leads to develop better drugs against cancer, as well as viral and microbial infections (12),(13),(14). Although hundreds of plant species have been tested for antimicrobial properties, the vast majority have not yet been adequately evaluated. In India alone, 2000 different plants are used for medicinal preparations for both internal and external use, according to National Health Experts (15), (16). Among them only 200 are of animal origin, and 300 of mineral origin, while 1500 drugs are extracted from various plants. The development of techniques for the controlled synthesis of metal nanoparticles of well-defined scale, shape, and composition is a major challenge in nanotechnology. Electronic, magnetic, catalytic, and optical properties of metal nanoparticles and nanocomposites vary from those of bulk metals (17),(18),(19). Nanomaterials are at the forefront of nanotechnology's rapidly evolving sector.

Their unique size-dependent properties make these materials superior and indispensable in many areas of human activity (20),(21),(22). Selenium possesses excellent photo electrical and semiconductor properties which make it extensively used in duplicate, photography, cells and rectifiers (23). Selenium is also one of essential trace elements in the human body and has great

importance in nourishment and medicine (24). It has been reported that the redness of selenium nanoparticles has high biological activities and low toxicity (25–27). Medical diagnostic field also developed to use the selenium nanoparticle and also studies on the increase efficiency of glutathione peroxidase and thioredoxin reductase. Elemental selenium is one of those materials. Nowadays, selenium is well-known as an essential micronutrient of fundamental importance to human and animal health (28). An especially interesting field of research regarding the antimicrobial activity of SeNPs, which recently gained attention, is the inhibition of biofilm formation and activity against resistant microbial strains (29), (30), (31), (32), (33,34). Previously our team has published extensive research on various aspects (35–54), this vast research experience has inspired us to research about the present study. The aim of the present study is to determine antimicrobial activity of selenium nanoparticles and *Pterocarpus santalinus* based mouthwash.

MATERIALS AND METHOD-

Plant collection -

The leaf, stem, and bark of *P. santalinus* Linn. f. were collected. They were thoroughly washed with tap water and dried under shade. The dried plant parts were homogenized to a fine powder and stored in airtight bottles and later used for extraction.

Plant extract preparation -

Commercially available dry powder of *Pterocarpus santalinus* was used for this experiment. This experiment was conducted in Saveetha Dental College, Chennai, Tamilnadu. This experiment was carried out by dissolving 1g of *Pterocarpus santalinus* in 100ml of water. This mixture was then boiled in a heating mantle at 70 degrees Celsius for up to 10 minutes. The boiled mixture was then filtered using Whatman number 1 filter paper to obtain the plant extract. Then 40ml of plant extract was measured using a measuring cylinder and the mixture was added to 60ml of 1mM selenium dissolved in 60 ml distilled water.

Preparation of mouthwash -

To an eppendorf tube, 10 ml of distilled water was taken then 0.3g of sucrose was measured using an electrical weighing scale and was added to the tube containing the water. The mixture was mixed well and then 0.01g of sodium lauryl sulphate and 0.001g of sodium benzoate were

added and mixed well. Then 12 drops of the plant pellets were added to the above mixture followed by adding 2 drops of peppermint oil.

Antimicrobial activity -

Antimicrobial activity of respective nanoparticles against the strain staphylococcus aureus, Bacillus and E Coli. MHAagar was utilized for this activity to determine the zone of inhibition. Muller hinton agar was prepared and sterilized for 45 minutes at 120 lbs. Media poured into the sterilized plates and let them stabilize for solidification. The wells were cut using the well cutter and the test organisms were swabbed. The nanoparticles with different concentrations were loaded and the plates were incubated for 24 hours at 37 degree celsius. After the incubation the zones of inhibition were measured.

RESULT -



FIGURE 1 - GREEN SYNTHESIS OF SELENIUM NANOPARTICLES AND *PTEROCARPUS SANTALINUS* BASED MOUTHWASH

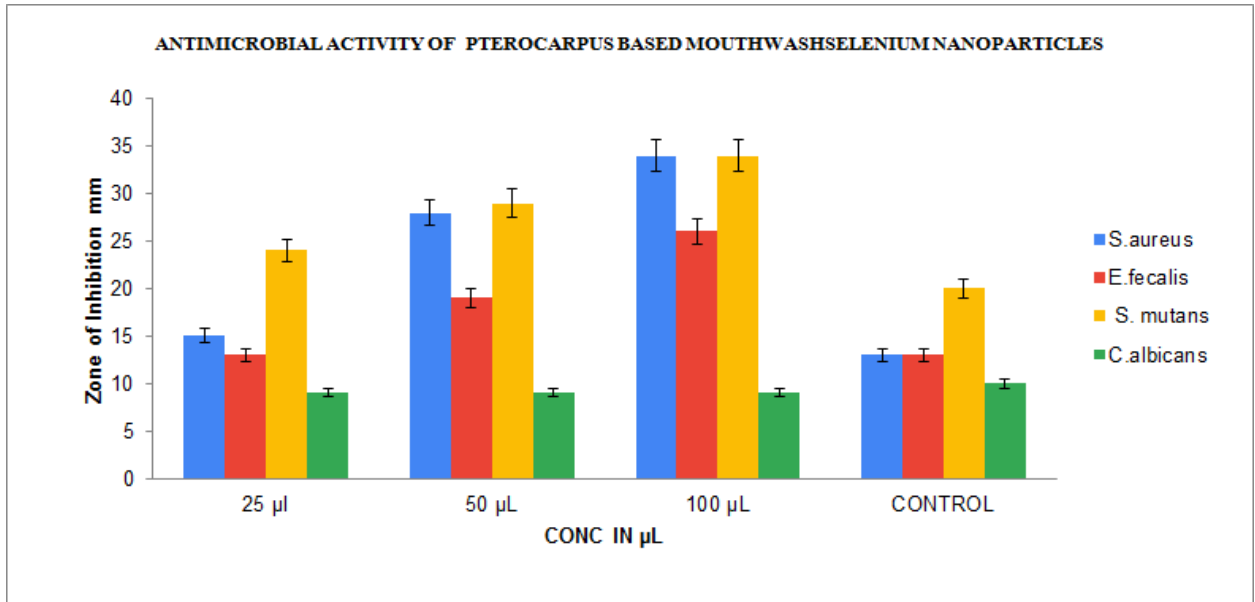


FIGURE 2 - Antimicrobial activity of P.santa based mouthwash selenium nanoparticles, data implies as mean±SEM .

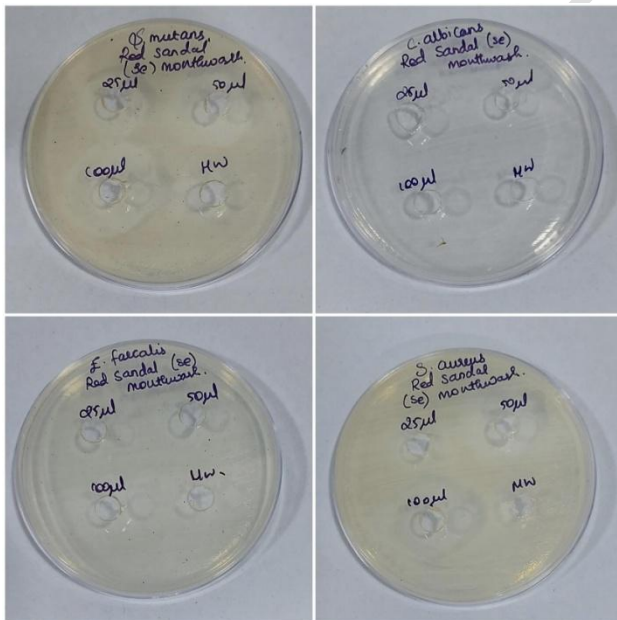


FIGURE3- Zone Of Inhibition Against S.Mutans, C.Albicans,E.Faecalis,S.Aureus

In the present study, the MIC of bacterial and fungal growth at varying concentrations of biosynthesized selenium nanoparticles are analyzed. The antibacterial activity of selenium nanoparticles was evaluated based on their zone of inhibitions and the results were compared

with the standard antibacterial agent. The antimicrobial activity of selenium nanoparticles against *S. aureus* showed a zone of inhibition of 15 mm at the concentration of 25 μ l, 28mm at the concentration of 50 and 35mm at 100 μ l concentration which is comparatively more than the used standard antibiotic which showed zone of inhibition of 13mm.

The antimicrobial activity of selenium nanoparticles against *E.faecalis* showed a zone of inhibition of 13 mm at the concentration of 25 μ l, which is comparatively less than the used standard antibiotic which showed zone of inhibition of 15mm, 19mm at the concentration of 50 and 26mm at 100 μ l concentration which is comparatively more than the used standard antibiotic.

The antimicrobial activity of selenium nanoparticles against *S.mutans* showed a zone of inhibition of 24 mm at the concentration of 25 μ l, 29mm at the concentration of 50 and 34mm at 100 μ l concentration which is comparatively more than the used standard antibiotic which showed zone of inhibition of 20mm. The antifungal activity of selenium nanoparticles against *C.albicans* showed a zone of inhibition of 9 mm at the concentration of 25 μ l, 9mm at the concentration of 50 and 9mm at 100 μ l concentration which is comparatively more than the used standard antibiotic which showed zone of inhibition of 10mm.

DISCUSSION

Analyzing the results of the present study, it can be found that the highest zones of inhibition were exhibited against two bacterial strains *Staphylococcus aureus* and *Staphylococcus mutans* proving that it acts as a good antibacterial against *S.aureus* and *S.mutans* infections. Selenium has been described as a potential cancer treatment and prevention candidate, as well as an anti-inflammatory agent, in the treatment of cardiovascular diseases and thyroid disorders, and as a key component of bone and muscle metabolism (65), (66), (67), (68). SeNPs have been shown in many studies to have a lower risk of selenium toxicity while having the same bioavailability and efficacy in increasing the activities of selenoenzymes as Se-Met and selenite (69–71).

Many researchers are now focusing on the anticancer activity of SeNPs, either alone or in combination with other anticancer agents, as a result of these findings (72–74). One of the pioneering work that reported the antimicrobial activity of SeNPs was done by (75) The authors show that SeNPs with a diameter of about 100 nm inhibit the growth of *Staphylococcus aureus* at concentrations as low as 7.8 g/mL in this paper. Inhibition of biofilm formation and activity

against resistant microbial strains is a particularly important area of research on SeNPs' antimicrobial activity that has recently gained attention (29),(30),(31),(32),(33,34). Bacterial biofilm is thought to be the highest degree of bacterial defence against the immune system and antibiotics.

Due to microbial synergy, polymicrobial biofilms strengthened this tolerance, raising the risk of serious health complications (76). Polymicrobial infections are often caused by a combination of bacteria and fungi. Many of these infections, such as those caused by *Candida albicans* and *Staphylococcus* species, are associated with high mortality rates (77) (78) (79) (80) (81) (82) (83) (84) (85) (86) (87) (88) (89) (90) (91). The morphology of nanoparticles also has an effect on their ability to destroy microbial organisms (92,93). The ability of nanoparticles to operate simultaneously through these multiple mechanisms is their key advantage as antimicrobial agents. As a result, unlike commercial antibiotics, microbes are unable to establish resistance to these articulated mechanisms of action. Positively charged NPs, according to some authors, have a greater capacity for inhibiting bacterial growth due to their stronger attachment to bacteria (67,94).

CONCLUSION

The present study is a preliminary evaluation of antimicrobial activity of plants. It indicates that several plants have the potential to generate novel metabolites(55-64). The crude extract demonstrating anticandidal activity could result in the discovery of novel anticandidal agents, The plants demonstrating broad spectra of activities may help to discover new chemical classes of antibiotics that could serve as selective agents for the maintenance of animal or human health and provide biochemical tools for the study of infectious disease.

NOTE:

The study highlights the efficacy of "FOLK MEDICINE" which is an ancient tradition, used in some parts of India. This ancient concept should be carefully evaluated in the light of modern medical science and can be utilized partially if found suitable.

REFERENCE:

1. Shankar SB, Barani Shankar S, Arivarasu L, Rajeshkumar S. Biosynthesis of Hydroxy Citric Acid Mediated Zinc Nanoparticles and Its Antioxidant and Cytotoxic Activity [Internet]. *Journal of Pharmaceutical Research International*. 2020. p. 108–12. Available from: <http://dx.doi.org/10.9734/jpri/2020/v32i2630845>
2. Karthik V, Arivarasu L, Rajeshkumar S. Hyaluronic Acid Mediated Zinc Nanoparticles against Oral Pathogens and Its Cytotoxic Potential [Internet]. *Journal of Pharmaceutical Research International*. 2020. p. 113–7. Available from: <http://dx.doi.org/10.9734/jpri/2020/v32i1930716>
3. Shree MK, Kavya Shree M, Arivarasu L, Rajeshkumar S. Cytotoxicity and Antimicrobial Activity of Chromium Picolinate Mediated Zinc Oxide Nanoparticle [Internet]. *Journal of Pharmaceutical Research International*. 2020. p. 28–32. Available from: <http://dx.doi.org/10.9734/jpri/2020/v32i2030726>
4. International Trade Centre UNCTAD/GATT. *Markets for Selected Medicinal Plants and Their Derivatives*. 1982. 206 p.
5. Jaisankar AI, Arivarasu L. Free Radical Scavenging and Anti-Inflammatory Activity of Chlorogenic Acid Mediated Silver Nanoparticle [Internet]. *Journal of Pharmaceutical Research International*. 2020. p. 106–12. Available from: <http://dx.doi.org/10.9734/jpri/2020/v32i1930715>
6. *Herbal Sources Used by The Public Against Infections* [Internet]. Vol. 12, *International Journal of Pharmaceutical Research*. 2020. Available from: <http://dx.doi.org/10.31838/ijpr/2020.sp1.015>
7. Niveditha AS, Sankari Niveditha A, Geetha RV, Arivarasu L. Will Alternative Medicine Help Us to Fight Against COVID-19 [Internet]. *International Journal of Current Research and Review*. 2020. p. 112–6. Available from: <http://dx.doi.org/10.31782/ijcrr.2020.sp47>
8. Devaraj E, Roy A, Veeraragavan GR, Magesh A, Sreeba AV, Arivarasu L, et al. β -Sitosterol attenuates carbon tetrachloride–induced oxidative stress and chronic liver injury in rats [Internet]. Vol. 393, *Naunyn-Schmiedeberg's Archives of Pharmacology*. 2020. p. 1067–75. Available from: <http://dx.doi.org/10.1007/s00210-020-01810-8>
9. Nasim I, Kamath K, Rajeshkumar S. Evaluation of the re-mineralization capacity of a gold nanoparticle-based dental varnish: An in vitro study [Internet]. Vol. 23, *Journal of Conservative Dentistry*. 2020. p. 390. Available from: http://dx.doi.org/10.4103/jcd.jcd_315_20
10. S SK, Satheesha KS. In-Vitro Antibacterial Activity of Black Tea (*Camellia sinensis*) Mediated Zinc Oxide Nanoparticles Against Oral Pathogens [Internet]. Vol. 13, *Bioscience Biotechnology Research Communications*. 2020. p. 2077–80. Available from: <http://dx.doi.org/10.21786/bbrc/13.4/66>

11. Rajeshkumar S, Malarkodi C, Al Farraj DA, Elshikh MS, Roopan SM. Employing sulphated polysaccharide (fucoidan) as medium for gold nanoparticles preparation and its anticancer study against HepG2 cell lines [Internet]. Vol. 26, *Materials Today Communications*. 2021. p. 101975. Available from: <http://dx.doi.org/10.1016/j.mtcomm.2020.101975>
12. Galal M, Bashir AK, Salih AM, Adam SEI. Activity of water extracts of *Albizia anthelmintica* and *A. lebbek* barks against experimental *Hymenolepis diminuta* infection in rats [Internet]. Vol. 31, *Journal of Ethnopharmacology*. 1991. p. 333–7. Available from: [http://dx.doi.org/10.1016/0378-8741\(91\)90019-a](http://dx.doi.org/10.1016/0378-8741(91)90019-a)
13. Hoffmann JJ, Timmermann BN, McLaughlin SP, Punnapayak H. Potential Antimicrobial Activity of Plants from the Southwestern United States [Internet]. Vol. 31, *International Journal of Pharmacognosy*. 1993. p. 101–15. Available from: <http://dx.doi.org/10.3109/13880209309082926>
14. Shunmugam R, Balusamy SR, Kumar V, Menon S, Lakshmi T, Perumalsamy H. Biosynthesis of gold nanoparticles using marine microbe (*Vibrio alginolyticus*) and its anticancer and antioxidant analysis [Internet]. Vol. 33, *Journal of King Saud University - Science*. 2021. p. 101260. Available from: <http://dx.doi.org/10.1016/j.jksus.2020.101260>
15. Balandrin MF, Klocke JA, Wurtele ES, Bollinger WH. Natural plant chemicals: sources of industrial and medicinal materials. *Science*. 1985 Jun 7;228(4704):1154–60.
16. Rajeshkumar S, Sherif MH, Malarkodi C, Ponnaniakajamideen M, Arasu MV, Al-Dhabi NA, et al. Cytotoxicity behaviour of response surface model optimized gold nanoparticles by utilizing fucoidan extracted from *padina tetrastratica* [Internet]. Vol. 1228, *Journal of Molecular Structure*. 2021. p. 129440. Available from: <http://dx.doi.org/10.1016/j.molstruc.2020.129440>
17. Klaus-Joerger T, Joerger R, Olsson E, Granqvist C-G. Bacteria as workers in the living factory: metal-accumulating bacteria and their potential for materials science [Internet]. Vol. 19, *Trends in Biotechnology*. 2001. p. 15–20. Available from: [http://dx.doi.org/10.1016/s0167-7799\(00\)01514-6](http://dx.doi.org/10.1016/s0167-7799(00)01514-6)
18. Barma MD, Kannan SD, Indiran MA, Rajeshkumar S, Pradeep Kumar R. Antibacterial Activity of Mouthwash Incorporated with Silica Nanoparticles against *S. aureus*, *S. mutans*, *E. faecalis*: An in-vitro Study [Internet]. *Journal of Pharmaceutical Research International*. 2020. p. 25–33. Available from: <http://dx.doi.org/10.9734/jpri/2020/v32i1630646>
19. Vikneshan M, Saravanakumar R, Mangaiyarkarasi R, Rajeshkumar S, Samuel SR, Suganya M, et al. Algal biomass as a source for novel oral nano-antimicrobial agent [Internet]. Vol. 27, *Saudi Journal of Biological Sciences*. 2020. p. 3753–8. Available from: <http://dx.doi.org/10.1016/j.sjbs.2020.08.022>
20. Salata OV. *Journal of Nanobiotechnology* [Internet]. Vol. 2. 2004. p. 3. Available from: <http://dx.doi.org/10.1186/1477-3155-2-3>

21. Barma MD. Synthesis of Triphala Incorporated Zinc Oxide Nanoparticles and Assessment of its Antimicrobial Activity Against Oral Pathogens : An In-Vitro Study [Internet]. Vol. 13, Bioscience Biotechnology Research Communications. 2020. p. 74–8. Available from: <http://dx.doi.org/10.21786/bbrc/13.7/14>
22. Syed MH, Gnanakkan A, Pitchiah S. Exploration of acute toxicity, analgesic, anti-inflammatory, and anti-pyretic activities of the black tunicate, *Phallusia nigra* (Savigny, 1816) using mice model [Internet]. Vol. 28, Environmental Science and Pollution Research. 2021. p. 5809–21. Available from: <http://dx.doi.org/10.1007/s11356-020-10938-2>
23. Berger LI. Semiconductor Materials [Internet]. 2020. Available from: <http://dx.doi.org/10.1201/9780138739966>
24. Liu H, Xu H, Huang K. Selenium in the prevention of atherosclerosis and its underlying mechanisms [Internet]. Vol. 9, Metallomics. 2017. p. 21–37. Available from: <http://dx.doi.org/10.1039/c6mt00195e>
25. Zhang Y, Huang P, Guo J, Shi R, Huang W, Shi Z, et al. Photodetectors: Graphdiyne-Based Flexible Photodetectors with High Responsivity and Detectivity (Adv. Mater. 23/2020) [Internet]. Vol. 32, Advanced Materials. 2020. p. 2070175. Available from: <http://dx.doi.org/10.1002/adma.202070175>
26. Bhattacharya D, Gupta RK. Nanotechnology and Potential of Microorganisms [Internet]. Vol. 25, Critical Reviews in Biotechnology. 2005. p. 199–204. Available from: <http://dx.doi.org/10.1080/07388550500361994>
27. Mohanpuria P, Rana NK, Yadav SK. Biosynthesis of nanoparticles: technological concepts and future applications [Internet]. Vol. 10, Journal of Nanoparticle Research. 2008. p. 507–17. Available from: <http://dx.doi.org/10.1007/s11051-007-9275-x>
28. Rayman MP. The importance of selenium to human health [Internet]. Vol. 356, The Lancet. 2000. p. 233–41. Available from: [http://dx.doi.org/10.1016/s0140-6736\(00\)02490-9](http://dx.doi.org/10.1016/s0140-6736(00)02490-9)
29. Webster TJ, Wang. Short communication: inhibiting biofilm formation on paper towels through the use of selenium nanoparticles coatings [Internet]. International Journal of Nanomedicine. 2013. p. 407. Available from: <http://dx.doi.org/10.2147/ijn.s38777>
30. Shakibaie M, Mohazab NS, Mousavi SAA. Antifungal Activity of Selenium Nanoparticles Synthesized by Bacillus species Msh-1 Against *Aspergillus fumigatus* and *Candida albicans* [Internet]. Vol. 8, Jundishapur Journal of Microbiology. 2015. Available from: <http://dx.doi.org/10.5812/jjm.26381>
31. Guisbiers G, Lara HH, Mendoza-Cruz R, Naranjo G, Vincent BA, Peralta XG, et al. Inhibition of *Candida albicans* biofilm by pure selenium nanoparticles synthesized by pulsed laser ablation in liquids [Internet]. Vol. 13, Nanomedicine: Nanotechnology, Biology and Medicine. 2017. p. 1095–103. Available from: <http://dx.doi.org/10.1016/j.nano.2016.10.011>

32. Prateeksha, Prateeksha, Singh BR, Shoeb M, Sharma S, Naqvi AH, et al. Scaffold of Selenium Nanovectors and Honey Phytochemicals for Inhibition of *Pseudomonas aeruginosa* Quorum Sensing and Biofilm Formation [Internet]. Vol. 7, *Frontiers in Cellular and Infection Microbiology*. 2017. Available from: <http://dx.doi.org/10.3389/fcimb.2017.00093>
33. Cremonini E, Boaretti M, Vandecandelaere I, Zonaro E, Coenye T, Lleo MM, et al. Biogenic selenium nanoparticles synthesized by *Stenotrophomonas maltophilia* SeITE02 loose antibacterial and antibiofilm efficacy as a result of the progressive alteration of their organic coating layer. *Microb Biotechnol*. 2018 Nov;11(6):1037–47.
34. Tran PA, O'Brien-Simpson N, Palmer JA, Bock N, Reynolds EC, Webster TJ, et al. Selenium nanoparticles as anti-infective implant coatings for trauma orthopedics against methicillin-resistant *Staphylococcus aureus* and *epidermidis*: in vitro and in vivo assessment [Internet]. Vol. 14, *International Journal of Nanomedicine*. 2019. p. 4613–24. Available from: <http://dx.doi.org/10.2147/ijn.s197737>
35. Rajeshkumar S, Venkat Kumar S, Ramaiah A, Agarwal H, Lakshmi T, Roopan SM. Biosynthesis of zinc oxide nanoparticles using *Mangifera indica* leaves and evaluation of their antioxidant and cytotoxic properties in lung cancer (A549) cells [Internet]. Vol. 117, *Enzyme and Microbial Technology*. 2018. p. 91–5. Available from: <http://dx.doi.org/10.1016/j.enzmictec.2018.06.009>
36. Nandhini NT, Rajeshkumar S, Mythili S. The possible mechanism of eco-friendly synthesized nanoparticles on hazardous dyes degradation [Internet]. Vol. 19, *Biocatalysis and Agricultural Biotechnology*. 2019. p. 101138. Available from: <http://dx.doi.org/10.1016/j.bcab.2019.101138>
37. Vairavel M, Devaraj E, Shanmugam R. An eco-friendly synthesis of *Enterococcus* sp.–mediated gold nanoparticle induces cytotoxicity in human colorectal cancer cells [Internet]. Vol. 27, *Environmental Science and Pollution Research*. 2020. p. 8166–75. Available from: <http://dx.doi.org/10.1007/s11356-019-07511-x>
38. Gomathi M, Prakasam A, Rajkumar PV, Rajeshkumar S, Chandrasekaran R, Anbarasan PM. Green synthesis of silver nanoparticles using *Gymnema sylvestre* leaf extract and evaluation of its antibacterial activity [Internet]. Vol. 32, *South African Journal of Chemical Engineering*. 2020. p. 1–4. Available from: <http://dx.doi.org/10.1016/j.sajce.2019.11.005>
39. Rajasekaran S, Damodharan D, Gopal K, Rajesh Kumar B, De Poures MV. Collective influence of 1-decanol addition, injection pressure and EGR on diesel engine characteristics fueled with diesel/LDPE oil blends [Internet]. Vol. 277, *Fuel*. 2020. p. 118166. Available from: <http://dx.doi.org/10.1016/j.fuel.2020.118166>
40. Santhoshkumar J, Sowmya B, Venkat Kumar S, Rajeshkumar S. Toxicology evaluation and antidermatophytic activity of silver nanoparticles synthesized using leaf extract of *Passiflora caerulea* [Internet]. Vol. 29, *South African Journal of Chemical Engineering*. 2019. p. 17–23. Available from: <http://dx.doi.org/10.1016/j.sajce.2019.04.001>

41. R KR, Kathiswar RR, Ezhilarasan D, Rajeshkumar S. β -Sitosterol-assisted silver nanoparticles activates Nrf2 and triggers mitochondrial apoptosis via oxidative stress in human hepatocellular cancer cell line [Internet]. Vol. 108, Journal of Biomedical Materials Research Part A. 2020. p. 1899–908. Available from: <http://dx.doi.org/10.1002/jbm.a.36953>
42. Saravanan M, Arokiyaraj S, Lakshmi T, Pugazhendhi A. Synthesis of silver nanoparticles from *Phenerochaete chrysosporium* (MTCC-787) and their antibacterial activity against human pathogenic bacteria [Internet]. Vol. 117, Microbial Pathogenesis. 2018. p. 68–72. Available from: <http://dx.doi.org/10.1016/j.micpath.2018.02.008>
43. Gheena S, Ezhilarasan D. Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells [Internet]. Vol. 38, Human & Experimental Toxicology. 2019. p. 694–702. Available from: <http://dx.doi.org/10.1177/0960327119839173>
44. Ezhilarasan D, Sokal E, Najimi M. Hepatic fibrosis: It is time to go with hepatic stellate cell-specific therapeutic targets [Internet]. Vol. 17, Hepatobiliary & Pancreatic Diseases International. 2018. p. 192–7. Available from: <http://dx.doi.org/10.1016/j.hbpd.2018.04.003>
45. Ezhilarasan D. Oxidative stress is bane in chronic liver diseases: Clinical and experimental perspective [Internet]. Vol. 19, Arab Journal of Gastroenterology. 2018. p. 56–64. Available from: <http://dx.doi.org/10.1016/j.ajg.2018.03.002>
46. Gomathi AC, Xavier Rajarathinam SR, Mohammed Sadiq A, Rajeshkumar S. Anticancer activity of silver nanoparticles synthesized using aqueous fruit shell extract of *Tamarindus indica* on MCF-7 human breast cancer cell line [Internet]. Vol. 55, Journal of Drug Delivery Science and Technology. 2020. p. 101376. Available from: <http://dx.doi.org/10.1016/j.jddst.2019.101376>
47. Dua K, Wadhwa R, Singhvi G, Rapalli V, Shukla SD, Shastri MD, et al. The potential of siRNA based drug delivery in respiratory disorders: Recent advances and progress [Internet]. Vol. 80, Drug Development Research. 2019. p. 714–30. Available from: <http://dx.doi.org/10.1002/ddr.21571>
48. Ramesh A, Varghese S, Jayakumar ND, Malaiappan S. Comparative estimation of sulfiredoxin levels between chronic periodontitis and healthy patients - A case-control study [Internet]. Vol. 89, Journal of Periodontology. 2018. p. 1241–8. Available from: <http://dx.doi.org/10.1002/jper.17-0445>
49. Arumugam P, George R, Jayaseelan VP. Aberrations of m6A regulators are associated with tumorigenesis and metastasis in head and neck squamous cell carcinoma. Arch Oral Biol. 2021 Feb;122:105030.
50. Joseph B, Prasanth CS. Is photodynamic therapy a viable antiviral weapon against COVID-19 in dentistry? [Internet]. Vol. 132, Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology. 2021. p. 118–9. Available from: <http://dx.doi.org/10.1016/j.oooo.2021.01.025>
51. Ezhilarasan D, Apoorva VS, Vardhan NA. *Syzygium cumini* extract induced reactive oxygen species-mediated apoptosis in human oral squamous carcinoma cells [Internet].

Journal of Oral Pathology & Medicine. 2018. Available from:
<http://dx.doi.org/10.1111/jop.12806>

52. Duraisamy R, Krishnan CS, Ramasubramanian H, Sampathkumar J, Mariappan S, Sivaprakasam AN. Compatibility of Nonoriginal Abutments With Implants [Internet]. Vol. 28, *Implant Dentistry*. 2019. p. 289–95. Available from:
<http://dx.doi.org/10.1097/id.0000000000000885>
53. Gnanavel V, Roopan SM, Rajeshkumar S. Aquaculture: An overview of chemical ecology of seaweeds (food species) in natural products [Internet]. Vol. 507, *Aquaculture*. 2019. p. 1–6. Available from: <http://dx.doi.org/10.1016/j.aquaculture.2019.04.004>
54. Markov A, Thangavelu L, Aravindhana S, Zekiy AO, Jarahian M, Chartrand MS, et al. Mesenchymal stem/stromal cells as a valuable source for the treatment of immune-mediated disorders [Internet]. Vol. 12, *Stem Cell Research & Therapy*. 2021. Available from:
<http://dx.doi.org/10.1186/s13287-021-02265-1>
55. Rajendran R, Kunjusankaran RN, Sandhya R, Anilkumar A, Santhosh R, Patil SR. Comparative Evaluation of Remineralizing Potential of a Paste Containing Bioactive Glass and a Topical Cream Containing Casein Phosphopeptide-Amorphous Calcium Phosphate: An in Vitro Study. *Pesqui Bras Odontopediatria Clin Integr*. 2019 Mar 12;19(0):4668.
56. Ashok BS, Ajith TA, Sivanesan S. Hypoxia-inducible factors as neuroprotective agent in Alzheimer's disease. *Clin Exp Pharmacol Physiol* [Internet]. 2017 Mar [cited 2021 Sep 15];44(3). Available from: <https://pubmed.ncbi.nlm.nih.gov/28004401/>
57. Malli SN, Selvarasu K, Jk V, Nandakumar M, Selvam D. Concentrated Growth Factors as an Ingenious Biomaterial in Regeneration of Bony Defects after Periapical Surgery: A Report of Two Cases. *Case Rep Dent* [Internet]. 2019 Jan 22 [cited 2021 Sep 15];2019. Available from: <https://pubmed.ncbi.nlm.nih.gov/30805222/>
58. Mohan M, Jagannathan N. Oral field cancerization: an update on current concepts. *Oncol Rev* [Internet]. 2014 Jun 30 [cited 2021 Sep 15];8(1). Available from:
<https://pubmed.ncbi.nlm.nih.gov/25992232/>
59. Menon S, Ks SD, R S, S R, Vk S. Selenium nanoparticles: A potent chemotherapeutic agent and an elucidation of its mechanism. *Colloids Surf B Biointerfaces* [Internet]. 2018 Oct 1 [cited 2021 Sep 15];170. Available from: <https://pubmed.ncbi.nlm.nih.gov/29936381/>
60. Samuel SR, Acharya S, Rao JC. School Interventions-based Prevention of Early-Childhood Caries among 3-5-year-old children from very low socioeconomic status: Two-year randomized trial. *J Public Health Dent* [Internet]. 2020 Jan [cited 2021 Sep 15];80(1). Available from: <https://pubmed.ncbi.nlm.nih.gov/31710096/>
61. Praveen K, Narayanan V, Muthusekhar MR, Baig MF. Hypotensive anaesthesia and blood loss in orthognathic surgery: a clinical study. *Br J Oral Maxillofac Surg* [Internet]. 2001 Apr [cited 2021 Sep 15];39(2). Available from: <https://pubmed.ncbi.nlm.nih.gov/11286449/>

62. Neelakantan P, Subbarao C, Subbarao CV, De-Deus G, Zehnder M. The impact of root dentine conditioning on sealing ability and push-out bond strength of an epoxy resin root canal sealer. *Int Endod J* [Internet]. 2011 Jun [cited 2021 Sep 15];44(6). Available from: <https://pubmed.ncbi.nlm.nih.gov/21255047/>
63. Oligonucleotide therapy: An emerging focus area for drug delivery in chronic inflammatory respiratory diseases. *Chem Biol Interact*. 2019 Aug 1;308:206–15.
64. Kumar MS, Vamsi G, Sripriya R, Sehgal PK. Expression of matrix metalloproteinases (MMP-8 and -9) in chronic periodontitis patients with and without diabetes mellitus. *J Periodontol*. 2006 Nov;77(11):1803–8.
65. Clark LC, Combs GF Jr, Turnbull BW, Slate EH, Chalker DK, Chow J, et al. Effects of selenium supplementation for cancer prevention in patients with carcinoma of the skin. A randomized controlled trial. Nutritional Prevention of Cancer Study Group. *JAMA*. 1996 Dec 25;276(24):1957–63.
66. Zeng H, Combs GF Jr. Selenium as an anticancer nutrient: roles in cell proliferation and tumor cell invasion. *J Nutr Biochem*. 2008 Jan;19(1):1–7.
67. Fang B, Jiang Y, Nüsslein K, Rotello VM, Santore MM. Antimicrobial surfaces containing cationic nanoparticles: how immobilized, clustered, and protruding cationic charge presentation affects killing activity and kinetics. *Colloids Surf B Biointerfaces*. 2015 Jan 1;125:255–63.
68. Moreno-Reyes R, Egrise D, Nève J, Pasteels JL, Schoutens A. Selenium deficiency-induced growth retardation is associated with an impaired bone metabolism and osteopenia. *J Bone Miner Res*. 2001 Aug;16(8):1556–63.
69. Zhang J, Wang X, Xu T. Elemental Selenium at Nano Size (Nano-Se) as a Potential Chemopreventive Agent with Reduced Risk of Selenium Toxicity: Comparison with Se-Methylselenocysteine in Mice [Internet]. Vol. 101, *Toxicological Sciences*. 2008. p. 22–31. Available from: <http://dx.doi.org/10.1093/toxsci/kfm221>
70. Zhang JS, Gao XY, Zhang LD, Bao YP. Biological effects of a nano red elemental selenium. *Biofactors*. 2001;15(1):27–38.
71. Wang H, Zhang J, Yu H. Elemental selenium at nano size possesses lower toxicity without compromising the fundamental effect on selenoenzymes: comparison with selenomethionine in mice. *Free Radic Biol Med*. 2007 May 15;42(10):1524–33.
72. Zhang Y, Li X, Huang Z, Zheng W, Fan C, Chen T. Enhancement of cell permeabilization apoptosis-inducing activity of selenium nanoparticles by ATP surface decoration [Internet]. Vol. 9, *Nanomedicine: Nanotechnology, Biology and Medicine*. 2013. p. 74–84. Available from: <http://dx.doi.org/10.1016/j.nano.2012.04.002>
73. Xia Y, You P, Xu F, Liu J, Xing F. Novel Functionalized Selenium Nanoparticles for Enhanced Anti-Hepatocarcinoma Activity In vitro [Internet]. *nano Online*. Available from:

http://dx.doi.org/10.1515/nano.11671_2015.49

74. Maiyo F, Singh M. Selenium nanoparticles: potential in cancer gene and drug delivery. *Nanomedicine* . 2017 May;12(9):1075–89.
75. Tran PA, Webster TJ. Selenium nanoparticles inhibit *Staphylococcus aureus* growth. *Int J Nanomedicine*. 2011 Jul 29;6:1553–8.
76. Carolus H, Van Dyck K, Van Dijck P. *Candida albicans* and *Staphylococcus* Species: A Threatening Twosome [Internet]. Vol. 10, *Frontiers in Microbiology*. 2019. Available from: <http://dx.doi.org/10.3389/fmicb.2019.02162>
77. Feng Y, Su J, Zhao Z, Zheng W, Wu H, Zhang Y, et al. Differential effects of amino acid surface decoration on the anticancer efficacy of selenium nanoparticles [Internet]. Vol. 43, *Dalton Trans*. 2014. p. 1854–61. Available from: <http://dx.doi.org/10.1039/c3dt52468j>
78. Pushpaanjali G, Geetha RV, Lakshmi T. Knowledge and Awareness about Antibiotic Usage and Emerging Drug Resistance Bacteria among Dental Students. *Journal of Pharmaceutical Research International*. 2020 Aug 24;34–42.
79. Aathira CM, Geetha RV, Lakshmi T. Knowledge and Awareness about the Mode of Transmission of Vector Borne Diseases among General Public. *Journal of Pharmaceutical Research International*. 2020 Aug 24;87–96.
80. Baskar K, Lakshmi T. Knowledge, Attitude and Practices Regarding HPV Vaccination among Undergraduate and Postgraduate Dental Students in Chennai. *Journal of Pharmaceutical Research International*. 2020 Aug 25;95–100.
81. Manya Suresh LT. Wound Healing Properties of *Aloe Barbadensis* Miller-In Vitro Assay. *Journal of Complementary Medicine Research*. 2020;11(5):30–4.
82. First Report on Marine Actinobacterial Diversity around Madras Atomic Power Station (MAPS), India [Internet]. [cited 2021 Aug 31]. Available from: <http://alinteridergisi.com/article/first-report-on-marine-actinobacterial-diversity-around-madras-atomic-power-station-maps-india/>
83. Physicochemical Profile of *Acacia Catechu* Bark Extract – An in Vitro Stud - *International Journal of Pharmaceutical and Phytopharmacological Research* [Internet]. [cited 2021 Aug 31]. Available from: <https://eijppr.com/article/physicochemical-profile-of-acacia-catechu-bark-extract-an-in-vitro-stud>
84. Lakshmi T. Antifungal Activity of *Ficus racemosa* Ethanolic Extract against Dermatophytes-An in vitro Study. *Journal of Research in Medical and Dental Science*. 2021;9(2):191–3.
85. Awareness of Drug Abuse among Teenagers - *International Journal of Pharmaceutical and Phytopharmacological Research* [Internet]. [cited 2021 Aug 31]. Available from: <https://eijppr.com/article/awareness-of-drug-abuse-among-teenagers>

86. Mangal CSK, Anitha R, Lakshmi T. Inhibition of Nitric oxide Production and Nitric oxide Synthase Gene Expression in LPS Activated RAW 264 .7 Macrophages by Thyme oleoresin from *Thymus vulgaris*. *J Young Pharm*. 2018;10(4):481.
87. COX2 Inhibitory Activity of *Abutilon Indicum* - *Pharmaceutical Research and Allied Sciences* [Internet]. [cited 2021 Aug 31]. Available from: <https://ijpras.com/article/cox2-inhibitory-activity-of-abutilon-indicum>
88. Jibu RM, Geetha RV, Lakshmi T. Isolation, Detection and Molecular Characterization of *Staphylococcus aureus* from Postoperative Infections. *Journal of Pharmaceutical Research International*. 2020 Aug 24;63–7.
89. Sindhu PK, Thangavelu L, Geetha RV, Rajeshkumar S, Raghunandhakumar S, Roy A. Anorectic drugs: an experimental and clinical perspective – A Review. *Journal of Complementary Medicine Research*. 2020;11(5):106–12.
90. Nivethitha R, Thangavelu L, Geetha RV, Anitha R, RajeshKumar S, Raghunandhakumar S. In Vitro Anticancer Effect of *Sesamum Indicum* Extract -. *Journal of Complementary Medicine Research*. 2020;11(5):99–105.
91. Mariona P, Roy A, Lakshmi T. Survey on lifestyle and food habits of patients with PCOS and obesity. *Journal of Complementary Medicine Research*. 2020;11(5):93–8.
92. Hong X, Wen J, Xiong X, Hu Y. Shape effect on the antibacterial activity of silver nanoparticles synthesized via a microwave-assisted method [Internet]. Vol. 23, *Environmental Science and Pollution Research*. 2016. p. 4489–97. Available from: <http://dx.doi.org/10.1007/s11356-015-5668-z>
93. Cheon JY, Kim SJ, Rhee YH, Kwon OH, Park WH. Shape-dependent antimicrobial activities of silver nanoparticles [Internet]. Vol. 14, *International Journal of Nanomedicine*. 2019. p. 2773–80. Available from: <http://dx.doi.org/10.2147/ijn.s196472>
94. Pan X, Wang Y, Chen Z, Pan D, Cheng Y, Liu Z, et al. Investigation of Antibacterial Activity and Related Mechanism of a Series of Nano-Mg(OH)₂ [Internet]. Vol. 5, *ACS Applied Materials & Interfaces*. 2013. p. 1137–42. Available from: <http://dx.doi.org/10.1021/am302910q>