

Original Research Article

Phytochemical analysis of *Fagonia schweinfurthii* Hadidi

ABSTRACT

Fagonia schweinfurthii Hadidi (family *zygophyllaceae*) is plant of desert region .In Gujarat it is found on the coastal area of Dwarka, Bet Dwarka and Rann of Kachcha. It is locally called dhamasia or dhamaso. Plants parts including leaves, roots, rhizomes, stems, barks, flowers, fruits, grains or seeds which can be employed in the control or treatment of infectious disease which causes damage to the respiratory system, urinary tract, gastrointestinal and biliary systems and on the skin. The plant parts contain chemical components that are medically active. Tribal people living in desert region use this plant to cure number of ailments such as skin eruptions, skin diseases, anti- pyretic, in pain relief, in heal sores, ear infection, venereal diseases etc. In the new era of science and technology there is increasing need to validate the claims of traditional knowledge database for safe, easily available, cheap, side- effect free healthcare provisions. In the present study preliminary phytochemical analysis of plant parts of *Fagonia schweinfurthii* Hadidi was done by the standard method of Sofowora, 1982; Trease & Evans, 1989 method, which confirms the presence of alkaloids, tannin, saponin, terpenoid and steroids. High Performance Thin Layer Chromatography method was developed using mobile phase containing Chloroform: methanol: acetic acid in ratio 70:30:0.2 for detecting phyto-active constituents from the methanolic extracts of the leaf, stem and root of *Fagonia schweinfurthii* Hadidi. Methanolic extract of *Fagonia schweinfurthii* Hadidi of leaf, stem, fruit and root detected 8, 8, 7 and 4 numbers of peaks at 254 nm and at 366 nm 10, 12, 13 and 12 numbers of peaks from leaf, stem, fruit and root extract of *Fagonia schweinfurthii* Hadidi were detected.

Keywords: {*Fagonia schweinfurthii*, phytochemical, secondary metabolites}

1. INTRODUCTION

Fagonia schweinfurthii (Hadidi) belongs to family *Zygophyllaceae* and found as a shrub in Western India, upper Gangetic plains and Peninsular India. *Fagonia arabica* Linn. is synonym of *F. schweinfurthii* Hadidi. In Ayurveda it is called as Dhanvayaasa, Dhanvayavaasa, Dhanvayaasaka, Duraalabhaa, Samudraantaa, Gaandhaari, Kachhuraa, Anantaa, Duhsparshaa and it is locally known as Dhramau, Dhamaso, Kandhera, dhamasa and dhamasia (1).The plant parts contain chemical components that are medically active. Tribal people living in desert region use this plant to cure number of ailments such as skin eruptions, skin diseases and anti- pyretic, heal sores, ear infection, venereal diseases etc (1). In the new era of science and technology there is increasing need to validate the claims of traditional knowledge database for safe, easily available, cheap, side- effect free healthcare provisions. Plant produces a wide varieties of phytochemicals which are responsible for protecting the plant against environmental hazards such as pollution, stress, drought, UV exposure, pathogenic attack, microbial infections (2). The phytochemicals produced by plant are categorized in to:1) primary metabolites producing food for plants and 2)Secondary metabolites for growth and protection of plants.

Standardization of plant materials is a need now days. Several pharmacopoeia containing monographs of the plant materials describe only the physicochemical parameters. Hence the modern methods describing the identification and quantification of active constituents in the plant material may be useful for proper standardization of herbals and its formulations. The studies on phytochemical analysis have attracted the plant scientists due to its potential to cure diseases and development of new biotechnological tools and techniques. This biotechnological tool played a major role in giving the explanation to efficient problems on the one hand and provides the supplementary resources of raw material for pharmaceutical industry on the other hand. The potential of chemotaxonomy is now become increasingly obvious because of improvement of natural product chemistry. The application of chemical data to methodical has received attention of a large number of biochemists and botanists during the last three decades (3). Chromatographic fingerprinting techniques are most significant methods which can be used for the routine herbal drug analysis and for quality assurance. HPTLC offers better resolution and estimation of active constituents with reasonable accuracy in a shorter time (4). Pharmaceutical industry is developing very fast by analyzing phytochemicals from medicinal plants. Farmers remove some plants for their cash crops yield, although these plants have medicinal properties. The plants regularly removed from field. In the present study attempts were made to study of Phytochemistry of various phyto constituents of leaf, stem, fruit and root of *Fagonia schweinfurthii* (Hadidi) using Wagner's test and HPTLC.

2. MATERIAL AND METHODS / PRELIMINARY OBSERVATION

Chemical tests were carried out using aqueous extracts of plant parts of *Fagonia schweinfurthii* Hadidi to identify various phyto-constitutes using standard methods.(5)(6)(7).

Sample preparation for phytochemical screening: 50 gm powdered sample was weighed and taken separately. The powder was moistened with ammonia and evaporated to dryness. Dried sample was extracted with chloroform and filtered. After filtration, 10% sulfuric acid was added to the filtrate using separating funnel and aqueous layer was separated by adjusting pH 8 with ammonia. After adjusting pH of extract the solution with chloroform and the organic extract which obtained is evaporated to concentrate by keeping in open at room temperature. However, aqueous extraction was evaporated to dryness by heating in water-bath to obtain semi solid mass. Dried extract was stored in refrigerator for their future use in phytochemical analysis. Dried extract was re- dissolved by adding 5ml of distilled water.

Test for Alkaloid: 3 ml aqueous extract was stirred with 3 ml of 1% HCl on steam bath. Wagner's reagent was then added to mixture. Turbidity of the resulting precipitate was taken as an evidence for the presence of alkaloid

Test for Flavonoids: To 1 ml of aqueous extract, 1 ml of 10% lead acetate solution was added. The formation of a yellow precipitate was taken as a positive test for flavonoids.

Tests for steroids: A red color produced in the lower chloroform layer when 2 ml of **organic extract** was dissolved in 2 ml of chloroform and 2 ml concentrated sulphuric acid was added in it. This indicates the presence of steroids. Development of a greenish color when 2 ml of the organic extract was dissolved in 2 ml of chloroform and treated with sulphuric and acetic acid indicates the presence of steroids.

Preparation of extracts for HPTLC

1 g of plant parts of leaf, stem, fruit and root of *Fagonia schweinfurthii* Hadidi was taken and reflux with 250 ml of methanol using a soxhlet apparatus on a water bath for 60 minutes for two cycles. Filter the extract and concentrate to 5 ml then the sample extract obtained so far is used for further analysis. Then the extract was filtered with Whatman filter paper No 1. The filtrates were stored in a glass bottle in the freeze at 8°C temperature.(8)

3. RESULTS AND DISCUSSION

PRELIMINARY SCREENING

The presence of alkaloids was confirmed in all parts of *Fagonia schweinfurthii* Hadidi by using Wagner's test. Flavonoids and Steroids were present in all parts of *Fagonia schweinfurthii*. Preliminary screening of plant parts of *Fagonia schweinfurthii* reveals the presence of phyto-constituents' in it but cannot predict the maximum no of compounds in the plant parts which influence the plant medicinal properties. Hence, to determine the number of compounds HPTLC technique was used.

Chart 1. Preliminary observation of *Fagonia schweinfurthii* Hadidi.

Plant parts of <i>Fagonia schweinfurthii</i> Hadidi.	Alkaloids (Wagner's test)	Flavonoids	Steroids
Leaf	+	+	+
Stem	+	+	+
Fruit	+	+	+
Root	+	+	+

(+) = present;(-)= absent

HPTLC (High Performance Thin Layer Chromatography)

Determination of Phyto-constitutes by HPTLC: For HPTLC aluminium plate support silica gel60F254, 10X100 cm (Merck) plates were cut with ordinary household scissors and markings were made with soft pencil. Silica gel plates were impregnated by dipping to in methanol followed by drying at room temperature for 1 hr. Sample was applied using Band wise with Linomat 5 (camag, muttez; Switzerland) spray on automated instrument for HPTLC. Applied sample band length 6.9 mm 8 tracks, track distance 10.7mm, distance from lower edge 15mm; application volume 10 and 20 μ l of sample at 8 tracks. Camag twin through chamber with Chloroform: methanol: acetic acid (70:30:0.2, v/v/v) after 20 min pre-saturation with mobile phase for development were used for plant parts of *Fagonia schweinfurthii* Hadidi (API vol 5). The eight developments over 75 mm with intermediate drying after the run plate were dried with the help of Air Dryer for detection of active compound. The camag TLC Scanner 3 controlled by win CATS software was used for densitometry analysis. For this densitometry analysis observed Absorption measurement at 254 and 366 with TLC Scanner 3 controlled by win CATS software. Retention factors are calculated by the densitometer. (9)

The R_f value indicated the position at which a substance is located in the chromatogram. R_f values were calculated using the following formula: The R_f values of respective compounds for all the chromatograms are depicted in Table 1 and Table 2 which showed number of compounds present in that particular class of phytoconstituents.

Retention time (R_f) = Distance travelled by the solute/distance travelled by the solvent.

Ayurvedic Pharmacopeia of India volume five reported the Thin Layer Chromatography of the whole plant of *Fagonia cretica* Linn. was performed by using the mobile phase of chloroform: methanol: acetic acid in the ratio of 70:30:0.2,v/v/v under UV at 254 nm which elutes four spots, from those spots three spots were appeared in violet colored, their R_f values are 0.14, 0.32, 0.46 and a yellowish green colored spot at R_f 0.72. Under 366nm six fluorescent spots appeared. Brown colour spots were observed at R_f . 0.14, 0.32 while pink colored spots were observed at R_f 0.39, 0.51, 0.61 and 0.72. On exposure to iodine vapour eight spots were appeared at R_f . 0.14, 0.19, 0.28, 0.35 0.51, 0.61 and 0.72 all were in yellow color; at R_f 0.46 faint orange colored was observed. On spraying with vanillin sulphuric acid reagent and heating the plate at 110°C for 10 min. ten spots were appeared as follows (10)

Sr. no.	Rf value	Color
1	0.06	Bluish grey
2	0.14	Violet
3	0.19	Brown
4	0.2	Violet
5	0.35	Brown
6	0.39	Violet
7	0.46	Brown
8	0.51	Violet
9	0.61	Brown
10	0.72	Violet

The mobile phase of API volume 5 was applied on *Fagonia schweinfurthii*. Methanolic extract of *Fagonia schweinfurthii* of leaf, stem, fruit and root extracts of 20 µl were taken. *Fagonia schweinfurthii* leaf, stem, fruit and root the ratio of chloroform: methanol: acetic acid (70:30:0.2) was used as mobile phase.

Fagonia schweinfurthii leaf methanolic extract of volume 20µl at 254nm detected 8 peaks with R_f values 0.01, 0.04, 0.08, 0.18, 0.36, 0.42, 0.58 and 0.71 were calculated. Stem methanol extract detected 8 peaks with R_f values 0.01, 0.07, 0.11, 0.15, 0.34, 0.40, 0.51 and 0.72. Fruit methanol extract detected 7 peaks with R_f values are 0.03, 0.09, 0.16, 0.35, 0.45, 0.56 and 0.72. Root methanol extract detected 4 peaks with R_f values are 0.03, 0.15, 0.38 and 0.40 respectively. (Table 1, Figure 1) The present study of HPTLC reveals that methanolic leaf extract and stem extract of *Fagonia schweinfurthii* at 254nm separates 8 compounds, respectively. Fruit methanolic extract separated 7 compounds, while root methanol extract separated 4 compounds. All the components separated from the plant parts are unknown except from fruit extract at peak number 2 at R_f 0.09.

Fagonia schweinfurthii leaf methanolic extract of 20µl at 366nm detected 10 peaks with R_f values 0.02, 0.05, 0.08, 0.12, 0.18, 0.36, 0.42, 0.46, 0.59 and 0.86 were calculated. Stem methanol extract detected 12 peaks with R_f values 0.03, 0.07, 0.15, 0.25, 0.34, 0.40, 0.48, 0.51, 0.46, 0.64, 0.67 and 0.86. Fruit methanol extract detected 13 peaks with R_f values 0.01, 0.06, 0.16, 0.35, 0.41, 0.44, 0.61, 0.64, 0.72, 0.80, 0.83, 0.86 and 0.98. Root methanol extract detected 12 peaks their R_f values are 0.16, 0.23, 0.26, 0.36, 0.38, 0.52, 0.55, 0.69, 0.72, 0.83, 0.85 and 0.89 respectively. (Table 2, Figure 2) Hence, methanolic extracts of leaf, fruit, stem and root of *Fagonia schweinfurthii* at 366nm separates maximum number of components.

Table 2. Rf values of *Fagonia schweinfurthii* Hadidi leaf, stem, fruit and root extract in Volume 20µl at 366 nm

Volume	Plant part Extracts	No. of peaks detected	Peak	Max Rf	Assigned Substance			
20µl	Leaf	8	1	0.02	Autogenerated 4			
			2	0.05	Autogenerated 6			
			3	0.08	Autogenerated 9			
			4	0.12	Autogenerated 10			
			5	0.18	Autogenerated 7			
			6	0.36	Autogenerated 1			
			7	0.42	Autogenerated 2			
			8	0.47	Unknown*			
			9	0.58	Autogenerated 25			
			10	0.86	Unknown*			
			20µl	Stem	12	1	0.03	Autogenerated 4
						2	0.07	Autogenerated 9
						3	0.15	Autogenerated 13
4	0.25	Autogenerated 10						
5	0.34	Autogenerated 15						
6	0.40	Autogenerated 1						
7	0.48	Autogenerated 19						
8	0.51	Autogenerated 3						
9	0.46	Autogenerated 16						
10	0.64	Autogenerated 33						
11	0.67	Autogenerated 32						
12	0.86	Unknown*						
20µl	Fruit	13	1	0.01	Autogenerated 4			
			6	0.40	Unknown*			
			2	0.06	Autogenerated 6			
			3	0.56	Autogenerated 10			
			4	0.35	Autogenerated 1			
			5	0.41	Autogenerated 19			
			6	0.44	Autogenerated 3			
			7	0.61	Autogenerated 25			
			2	0.09	Autogenerated 17			
			8	0.64	Autogenerated 33			
			9	0.72	Autogenerated 12			
			10	0.80	Unknown*			
			11	0.83	Unknown*			
12	0.86	Unknown*						
20µl	Root	12	1	0.16	Autogenerated 10			
			2	0.23	Autogenerated 7			
			3	0.26	Autogenerated 15			
			4	0.36	Autogenerated 1			
			2	0.15	Unknown*			
			5	0.38	Autogenerated 17			
			6	0.58	Autogenerated 16			
			7	0.55	Autogenerated 30			
			8	0.69	Autogenerated 32			
			9	0.72	Autogenerated 12			

10	0.83	Unknown*
11	0.85	Unknown*
12	0.89	Unknown*

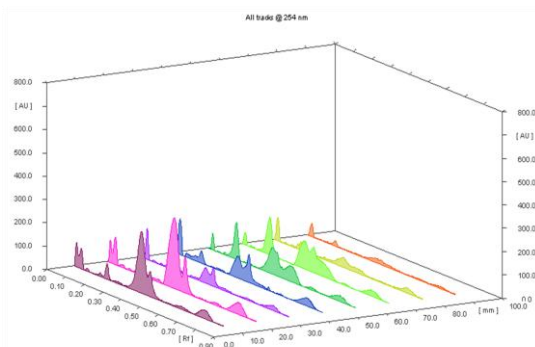


Figure 1

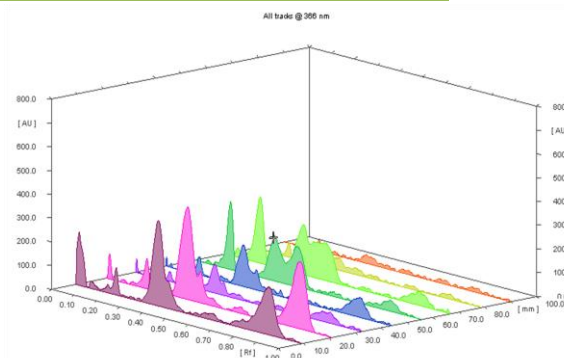


Figure 2

Babu and Radhaswamy separated compounds from the ethanolic extract of leaf, stem and root parts of *G. pentaphylla*. The chromatograms under UV 254nm 9 peaks were separated from the leaf extract indicating the occurrence of 9 different components. The stem and root showed 4 and 5 peaks, respectively. From the HPTLC analysis of plant parts of *Fagonia schweinfurthii* at 254 nm all peaks were detected are unknown except one and at 366nm all the peaks which are detected from the leaf and stem methanolic extracts are known except one. In leaf 10th numbered peak and in stem 12th numbered peak at Rf value 0.86 is unknown component, while in fruit 4 peaks at Rf values 0.80, 0.83, 0.86 and 0.98 are unknown and in root extract 3 peaks at Rf values 0.83, 0.85 and 0.89 are unknown.(11)

Gupta *et al* (2019) isolated 13 main secondary compounds namely alkaloids, anthracene derivatives, arbutin derivatives, bitter compounds, cardiac glycosides, coumarin derivatives, essential oils, flavonoids, lignans, pungent-tasting principles, saponins, triterpenes and valepotriates from the bark, leaves and seeds of *Careya arborea*. Bark extracts showed the presence of 1alkaloid, 3 anthracene derivatives, 1 arbutin derivative, 7 bitter compounds, 3 cardiac glycosides, 2 coumarin derivatives, 7 essential oils, 5 flavonoids, 5 lignans, 4 pungent-tasting principles, 4 saponins, 4 triterpenes and 6 valepotriates. Leaf extracts showed the presence of 6 anthracene derivatives, 1 arbutin derivative, 8 bitter compounds, 8 cardiac glycosides, 4 coumarin derivatives, 7 essential oils, 9 flavonoids, 7 lignans, 6 pungent-tasting principles, 5 saponins, 7 triterpenes and 6 valepotriates. Seeds showed the presence of 1 anthracene derivative, 6 bitter compounds, 6 cardiac glycosides, 3 coumarin derivatives, 4 essential oils, 3 flavonoids, 6 lignans, 5 pungent-tasting principles, 5 saponins, 5 triterpenes, and 6 valepotriates.(12)

4. CONCLUSION

Fagonia schweinfurthii (Hadidi) belongs to family *Zygophyllaceae*. There are no reports on phytochemical analysis of *Fagonia schweinfurthii*. Author investigated and collected *Fagonia schweinfurthii* from Bet Dwarka and Dwarka, Sea Coast of Gujarat. Preliminary screening method of phyto-constitute was followed by Sofowara1982; Trease & Evans, 1989 and Harbone, 1998. Preliminary investigation reveals the presence of alkaloids, flavonoids and steroids in the aqueous extract of plant parts. Phytochemical screening along with these HPTLC profiles is thus needed in setting up the

standard of this plant. The present study provides evidence that *Fagonia schweinfurthii* (Hadidi) solvent extract of contains medicinally important bioactive compounds and this justifies the use of plant species as traditional medicine for treatment of various diseases.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

REFERENCES

1. Rathore. M. K., Sharma. M. C., Goyal. M., Singh. G. K. and Nagori. B. P. Pharmacognostical studies on root of *Fagonia scheiwnfurthii* Hadidi. International Journal of Pharmaceutical and Biological Archieve. 2011; 2 (5): 514 – 1517.
2. Doughari, Hamuel James. Phytochemicals: Extraction Methods, Basic Structures and Mode of Action as Potential Chemotherapeutic Agents. Phytochemicals - A Global Perspective of Their Role in Nutrition and Health, Dr Venketeshwer Rao (Ed.), ISBN: 978-953-51-0296-0.InTech; 2011.
3. Santosh M. K., Sharanabasappa G. K., D. Shaila, Y. N. Seetharam and I. Sanjeevarao, Shaila D. Phytochemical studies on *Bauhinia racemosa* Lam., *Bauhinia purpurea* Linn. and *Hardwickia binata* Roxb. E-Journal of Chemistry. 2007; 4 (1): 21-31.
4. Rashmi Tambe, Maushumi Kulkarni, Kiran Bhise *et al.* Preliminary Phytochemical Screening and HPTLC Fingerprinting of Bark Extracts of *Symplocos racemosa*. *Journal of pharmacognosy and Phytochemistry*. 2013; 2 (3): 45-49.
5. Sofowora, E.A. Medicinal Plants and Traditional Medicine in Africa. John Wiley and Sons Ltd., Hoboken;1982.
6. Trease G.E. and Evans W.C. Pharmacognosy. 11th Edition, Bailliere Tindall, London; 1989.
7. Harborne, J.B. Textbook of Phytochemical Methods. A Guide to Modern Techniques of Plant Analysis. 5th Edition, Chapman and Hall Ltd, London;1998.
8. Mulla, Khaja Peer, Chaithra Dandu, Vasavi Thirumalanadhuni, Shaheen Shaik, Iliyaz Ali Shaik, and Yasodamma Nimmanapalli *et al.* Phytochemical, elemental, physico-chemical, HPTLC and anticancer investigations of *Ceropegia spiralis* Wight. Tuber extracts. The Pharma Innovation Journal. 2021; 10(3): 95-105. DOI: <https://doi.org/10.22271/tpi.2021.v10.i3b.5918>
9. Mehta, K., Patel, B. N., Jain, B.K. Phytochemical analysis of leaf extract of *Phyllanthus fraternus*. Research Journal of Recent Science. 2013; 2:12-15.
10. The Ayurvedic Pharmacopoeia of India. Createsspace Independent publishing platform. GOVERNMENT OF INDIA, MINISTRY OF HEALTH AND FAMILY WELFARE, DEPARTMENT OF AYUSH.2015;1(5):1-246.
11. Babu V. S and Radhamany P. M. Preliminary Phytochemistry, HPTLC and FTIR analysis in *Glycosmis pentaphylla* (Retz.)Dc.- an ethnobotanically important medicinal plant. International Journal of Pharmaceutical Sciences and Research.2021;12(2), 1135-114.
12. Gupta, P., Patil, D., & Patil, A. Qualitative HPTLC phytochemical profiling of *Careya arborea* Roxb. bark, leaves and seeds. 3 Biotech. 2019; 9(8),1-8. <https://doi.org/10.1007/s13205-019-1846-x>