

# ANTICANCER ACTIVITY OF AEGLE MARMELOS ON HUMAN HEPG2 CELLS BY REGULATION OF MATRIX METALLOPROTEINASES EXPRESSION

Running title: Role of Aeglemarmelos against liver cancer.

## ABSTRACT:

**Background:** Aeglemarmelos belong to the family rutaceae. It is commonly known as Bael. It is used in traditional medicine, as it has antidiarrheal, antimicrobial, antiviral, radio-protective, anti cancer, chemopreventive properties which are of great medicinal use. Liver cancer is the 5th most common type of cancer, Hepatocellular carcinoma is the most common form, which originates from the liver. Aeglemarmelos is said to inhibit the proliferative action of cancer cells.

**Objective:** To investigate the role of Aeglemarmelos against human liver cancer cells (HEP G2 cell line) by inhibiting the activity of matrix metalloproteinases which is responsible for spread of cancer.

**Materials and methods:** HEP G2 cell lines were procured from NCCS (National center for cell sciences) Pune, India. It was cultured and viability of the cells before and after adding the extract was analysed using the MTT assay. mRNA amplification was done using real time PCR. Statistical analysis was done using ANOVA and dunken's multiple test. Corresponding graphs are also plotted.

**Results:** The viability of the cells decreased from 100% to 50%. The mRNA expression of MMP-2 and MMP-9 decreased after the addition of the extract.

**Conclusion:** From this study we can conclude that Aeglemarmelos, a novel innovative anticancer drug inhibited the proliferative action of liver cancer cells by reducing the expression of MMP 2 and MMP 9 which inhibits the metastasis of cancer cells.

**Key Words:** Aeglemarmelos, novel, innovative Hepatocellular carcinoma, mRNA expression, proliferative, viability.

## INTRODUCTION

Cancer has now become one of the leading causes of death globally. Various researches on inhibiting cancer cell proliferation are carried out globally(1). Cancer begins when healthy cells are mutated and grow out of control-Metastasis.(2)(3). There are various types of liver cancers, hepatocellular carcinoma is one of which that originates in the liver unlike other types where the origin is not liver.(4)(5)(6)

Primary liver cancer is of 3 types out of which hepatocellular carcinoma is the most common type (75%)(7). HEP G2 cell lines are liver cancer cell lines obtained first from an argentinian male who had hepatocellular carcinoma.(8). These cells are epithelial in morphology, they are suitable in vitro polarised human hepatocytes for study purposes. These cells have high degrees of morphological and functional differentiation. (9)(10)

Aeglemarmelos inhibit the in vitro proliferation of tumor cell lines like leukemia K562, T Lymphoid Jurkat, Melanoma col 038, this is compared to the anti tumor agent 5-fluorouracil.(11,12) Aeglemarmelos also possess chemopreventive potential and has antitumor effects.(13). (5) Methanolic extracts of Aeglemarmelos when evaluated against cisplatin which might induce renal toxicity in rats shows that this plant extract possesses nephroprotective and antioxidant property(6)(14). It also reduces murrine ascites which reduce tumor volume and a viable tumor is prophylactically.(15)(16,17)

A Bael compound named skimmianine extracted from leaf and immature barks has anticancer property, analgesic property and even anti diuretic property.(18)(19) The fruits also have antiviral properties with essential vitamins like vitamin A, vitamin C, riboflavin niacin.(20). In the view of the wide ranging pharmacological activity this study was conducted to investigate the role of Aeglemarmelos in prevention of liver carcinogenesis.(21)(16)

## **MATERIALS AND METHODS:**

In order to test the anticancer property of aeglemarmelos a cancer cell line is procured and it is treated with varied doses of the leaf extract of aeglemarmelos, then the viability of cancer cells is analysed using MTT assay. A decrease in viability would indicate the effectiveness of the drug. The detailed explanation of the tests performed and chemicals used is described below.

### **Cell lines and cell culture:**

The Human Liver cell line (Hep G2) was purchased from the National Centre for Cell Sciences (NCCS), Pune, India. Cells were cultured in DMEM medium (Thermo Fisher Scientific, CA, USA) containing 10% fetal bovine serum (Thermo Fisher Scientific, CA, USA), 100 U/ml penicillin and 100 µg/ml streptomycin (Thermo Fisher Scientific, CA, USA) at 37°C with 5% CO<sub>2</sub>.

### **Cell viability by MTT assay:**

Cell viability was analysed using a modified colorimetric technique that is based on the ability of living cells to convert MTT, a tetrazolium compound into purple formazan crystals by mitochondrial reductases which is present in every living cell (Mosmann, 1983). Briefly, the cells (1 ×10<sup>4</sup>/well) were exposed to different concentrations of Aeglemarmelos extract (100-500µg/ml) with HepG2 cells for 48 h. At the end of the treatment, 100 µl of 0.5 mg/ml MTT solution was added to each well and incubated at 37 °C for an hour. Then the formazan crystals formed were dissolved in dimethyl sulfoxide (100 µl ) and incubated in the dark for an hour. Then the intensity of the colour developed was assayed using a Micro ELISA plate reader at 570 nm. The number of cells that were viable was expressed as the percentage of control cells cultured in serum-free medium. Cell viability of the cells in control medium without any treatment was represented as 100%. The cell viability can be calculated using the following formula: % cell viability = [A<sub>570 nm</sub> of treated cells/A<sub>570 nm</sub> of control cells] × 100.

### **Gene expression analysis by Real Time-PCR:**

Samples obtained from each group were submerged in 2 ml Trizol (Invitrogen, Carlsbad, CA, USA) for the purpose of RNA

extraction and stored at  $-80^{\circ}\text{C}$  until further processed. cDNA synthesis was performed on 2  $\mu\text{g}$  RNA in a 10  $\mu\text{l}$  sample volume using Superscript II reverse transcriptase (Invitrogen) as recommended by the manufacturer. Real-time PCR array analysis was performed in a total volume of 20  $\mu\text{l}$  including 1  $\mu\text{l}$  cDNA, 10  $\mu\text{l}$  qPCR Master Mix 2x (Takara, USA) and 9  $\mu\text{l}$  ddH<sub>2</sub>O. Reactions were run on an CFX96 Touch Real-Time PCR Detection System (Bio-Rad, USA) using universal thermal cycling parameters (95 $^{\circ}\text{C}$  for 5 min, 40 cycles of 15 sec at 95 $^{\circ}\text{C}$ , 15 sec at 60 $^{\circ}\text{C}$  and 20 sec at 72 $^{\circ}\text{C}$ ; followed by a melting curve: 5 sec at 95 $^{\circ}\text{C}$ , 60 sec at 60 $^{\circ}\text{C}$  and continued melting). For checking quality control, melting curves were obtained for all samples. The specificity of the amplification product was determined by analysing the melting curve for each primer pair. The data were analyzed by comparative CT method and the fold change is calculated by  $2^{-\Delta\Delta\text{CT}}$  method described by Schmittgen and Livak (2008) using CFX Manager Version 2.1 (Bio Rad, USA).

The following chemicals Dimethyl sulfoxide (DMSO), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were purchased from Sigma Chemical Pvt Ltd, USA. Trypsin-EDTA, fetal bovine serum (FBS), antibiotics-antimycotics, RPMI 1640 medium and phosphate buffered saline (PBS) were purchased from Gibco, Canada. (5,5,6,6-tetrachloro-1,1,3,3-tetraethyl benzimidazolecarbocyanine iodide) and Real Time PCR kit was purchased TAKARA (Meadowvale Blvd, Mississauga, ON L5N 5S2, Canada).

**Table 1: Primer Sequence**

S.No	Gene	Primer sequence
------	------	-----------------

2	Human MMP-2	Forward: 5'-ACC TAC ACC AAG AAC TTC CG-3' Reverse: 5'-TTG GTT CTC CAG CTT CAG GT-3'
3	Human MMP-9	Forward:5'-TCC CTG GAG ACC TGA GAA CC-3' Reverse: 5'-TCC CTG GAG ACC TGA GAA CC-3'
4	Human $\beta$ -actin	Forward:5'-CTACAATGAGCTGCGTGTGG -3' Reverse: 5'TAGCTCTTCTCCAGGGAGGA-3'

### Statistical analysis

The obtained data were analyzed statistically by one-way analysis of variance (ANOVA) and Duncan's multiple range test with a computer-based software (Graph Pad Prism version 5) to analyze the significance of individual variations among the control and experimental groups. The significance was considered at  $p < 0.05$  level in Duncan's test.

### RESULTS:

From the study we can infer that the viability of cancer cells which were 100% viable, after addition of Aeglemarmelos extract decreased in viability based on the dosage. It almost reached 50% when the concentration of extract was 300 to 500 micrograms.(figure 1).The fold changes over control of the mRNA expression of MMP-2 decreases significantly on the addition of Aeglemarmelos extract.(Figure 2). The fold changes over control of the mRNA expression of MMP-9, decreases significantly on the addition of Aeglemarmelos extract.(figure 3)

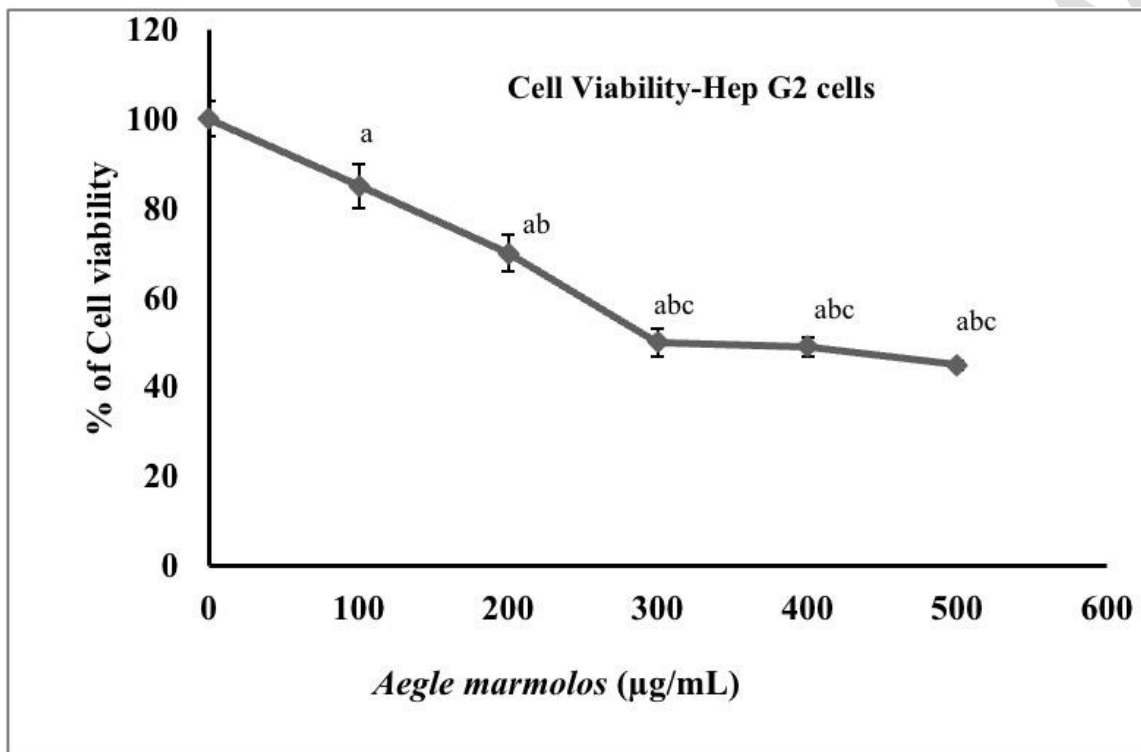


Figure 1 represents the effect of *Aegle marmelos* leaf extraction cell viability in HepG2 cells. Each bar represents a mean  $\pm$  SEM of 6 observations. Significance at  $p < 0.05$ , a-compared with untreated control cells, b-compared with 100µg treated HepG2 cells. X axis represents the concentration of extract and Y axis represents the percentage of cell viability. There is a significant decrease in the cell viability based on the dosage.

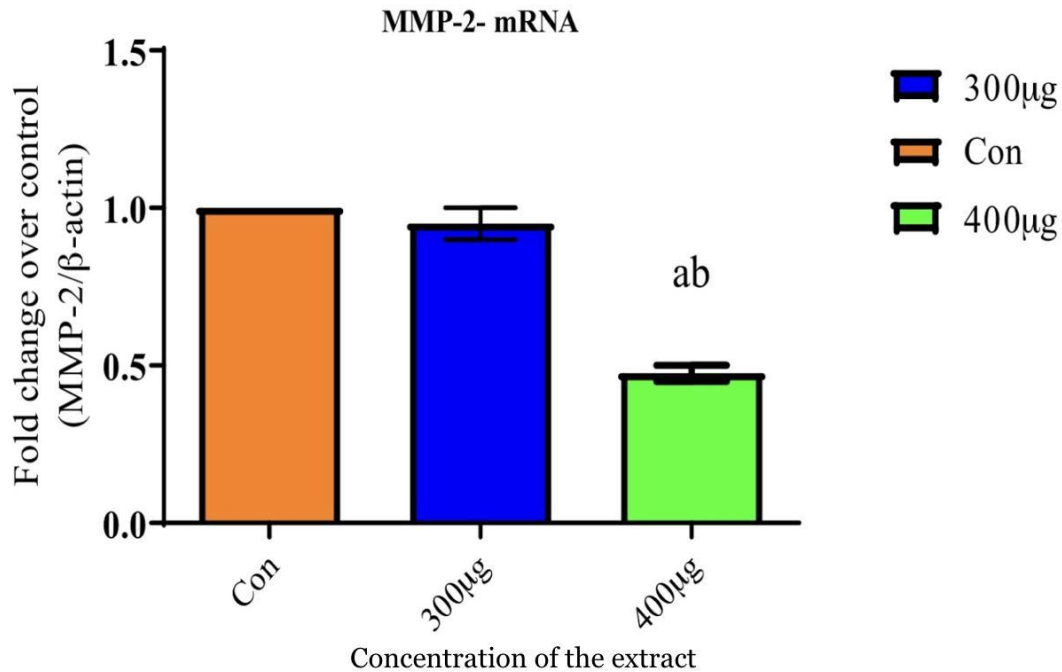
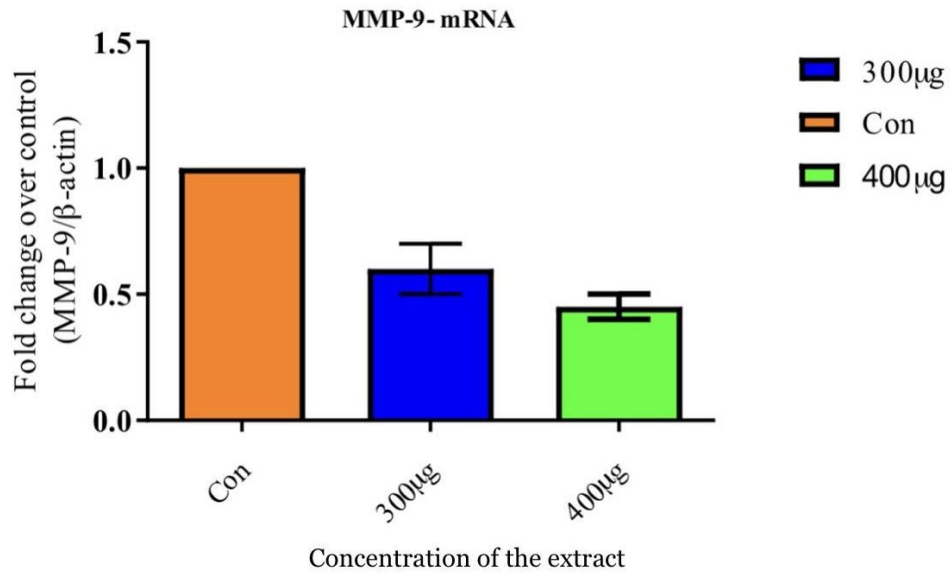


Figure 2 shows the Effect of Aeglemarmelos leaf extract on MMP-2 mRNA expression in HepG2 cells. Each bar represents a mean  $\pm$  SEM of 6 observations. Significance at  $p < 0.05$ , a-compared with untreated control cells, b-compared with 300 $\mu$ g treated cells. X axis represents the concentration of extract added and Y axis represents the fold change over control of the mRNA expression of MMP-2. There is a significant reduction in the mRNA expression of the MMP 2 based on the dosage.



**Figure 3 represents the effect of Aeglemarmelos leaf extract on MMP-9 mRNA expression in HEPG2 cells. Each bar represents a mean  $\pm$  SEM of 6 observations. Significance at  $p < 0.05$ , a-compared with untreated control cells, b-compared with 300 $\mu$ g treated cells. X axis represents the concentration of extract added and Y axis represents the fold change over control of the mRNA expression of MMP-9. There is a significant reduction in the mRNA expression of the MMP 9 based on the dosage.**

## **DISCUSSION**

Hepatocellular carcinoma is placed fifth in overall worldwide cancer rates and third cause of death in the world(22).Anticancer substances present in the diet is an attractive strategy to inhibit various cancers including liver cancer.(23).Aeglemarmelos fruit, stem and leaf extracts in diet can also have similar effects.(24)

Plant phenols may be considered as potential compounds for selective blocking signal transduction pathways, this is similar to this study where the aeglemarmelos extract blocks the MAPK pathway which inhibits the maturation of the MMPs.(25)

Several medicinal plants prescribed as a constituent of liver protective herbal drugs have been shown to inhibit chemically induced hepatic carcinogenicity in experimental animals.(26). Aeglemarmelos inhibit the in vitro proliferation of tumor cell lines like leukemia K562, (27,28)breast cancer cells, and jurkat cells similar to the findings of this study where it inhibits the human HEPG2 cell lines(29).

Aeglemarmelos also have chemopreventive potential, antitumor , reducing tumour volume and size.They also possess nephroprotective and antioxidant activity.(30)The phytochemical profile of Aeglemarmelos show pharmacological activity and hepatoprotective activity.(31)

In this study Aeglemarmelos plant extract inhibit the MMPs which causes degradation of proteins in the extracellular matrix, (27)degrades outer membrane of cells, degrades basement membrane so that cancer cells easily enter lymphatic and blood vessels , release growth factors for easy proliferation of cancer cells. (32)This effect is similar to the findings of (33) which discusses prevention of cancer by Aeglemarmelos.(34)

Limitation of this study is, only when done invivo it will give accurate results but before that advanced research needs to be done, various steps of drug testing needs to be approved. In vitro results have proven to be positive and the future scope of this study is of high significance against cancer treatment.

## **CONCLUSION;**

From this study we can conclude that Aeglemarmelos help in inhibiting the proliferation of HEP G2 cell line of hepatocellular carcinoma. It prevents the maturation of MMPs which reduces the viability of cancer cells. The mRNA expression of MMPs has decreased in folds after being treated with the plant extract. Liver cancer can be treated using Aeglemarmelos extract.

❖ **COMPETING INTERESTS DISCLAIMER:**

❖ Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

**REFERENCES:**

1. Barabadi H, Mojab F, Vahidi H, Marashi B, Talank N, Hosseini O, et al. Green synthesis, characterization, antibacterial and biofilm inhibitory activity of silver nanoparticles compared to commercial silver nanoparticles [Internet]. Vol. 129, Inorganic Chemistry Communications. 2021. p. 108647. Available from: <http://dx.doi.org/10.1016/j.inoche.2021.108647>
2. Goldfeder A, Baserga R. Cell Proliferation, Cancer, and Cancer Therapy: A Conference in Honor of Anna Goldfeder. 1982. 328 p.
3. Bharath B, Perinbam K, Devanesan S, AlSalhi MS, Saravanan M. Evaluation of the anticancer potential of Hexadecanoic acid from brown algae *Turbinaria ornata* on HT-29 colon cancer cells [Internet]. Vol. 1235, Journal of Molecular Structure. 2021. p. 130229. Available from: <http://dx.doi.org/10.1016/j.molstruc.2021.130229>
4. &na;, &NA; EVIDENCED-BASED LIVER ALLOCATION FOR HEPATOCELLULAR CARCINOMA [Internet]. Vol. 82, Transplantation. 2006. p. 627. Available from: <http://dx.doi.org/10.1097/00007890-200607152-01681>
5. Sridharan G, Ramani P, Patankar S, Vijayaraghavan R. Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma [Internet]. Vol. 48, Journal of Oral Pathology & Medicine. 2019. p. 299–306. Available from: <http://dx.doi.org/10.1111/jop.12835>
6. Shabgah AG, Ezzatifar F, Aravindhyan S, Zekiy AO, Ahmadi M, Gheibihayat SM, et al. Shedding more light on the role of Midkine in hepatocellular carcinoma: New perspectives on diagnosis and therapy [Internet]. Vol. 73, IUBMB Life. 2021. p. 659–69. Available from: <http://dx.doi.org/10.1002/iub.2458>

7. Clarizia G, Bernardo P. Diverse Applications of Organic-Inorganic Nanocomposites: Emerging Research and Opportunities: Emerging Research and Opportunities. IGI Global; 2019. 237 p.
8. Liao S-H, Chen C-L, Hsu C-Y, Chien K-L, Kao J-H, Chen P-J, et al. Long-term Effectiveness of Population-wide Multifaceted Interventions for Hepatocellular Carcinoma in Taiwan. *J Hepatol* [Internet]. 2021 Mar 6; Available from: <http://dx.doi.org/10.1016/j.jhep.2021.02.029>
9. Tahervand A, Mahmoudi M, Roushandeh AM. Digoxin Effectively Decreased Proliferation of Liver Cancer Cell Line [Internet]. Vol. 2, Focus on Sciences. 2016. p. 1–10. Available from: <http://dx.doi.org/10.20286/focsci-020113>
10. Saraswathi I, Saikarthik J, Senthil Kumar K, Srinivasan KM, Ardhanaari M, Gunapriya R. Impact of COVID-19 outbreak on the mental health status of undergraduate medical students in a COVID-19 treating medical college: a prospective longitudinal study [Internet]. Vol. 8, PeerJ. 2020. p. e10164. Available from: <http://dx.doi.org/10.7717/peerj.10164>
11. Egbuna C, Mishra AP, Goyal MR. Preparation of Phytopharmaceuticals for the Management of Disorders: The Development of Nutraceuticals and Traditional Medicine. Academic Press; 2020. 570 p.
12. Ezhilarasan D. Critical role of estrogen in the progression of chronic liver diseases [Internet]. Vol. 19, Hepatobiliary & Pancreatic Diseases International. 2020. p. 429–34. Available from: <http://dx.doi.org/10.1016/j.hbpd.2020.03.011>
13. Priyadarshini P, Raj A, Warriar RR. Original article Phytochemical and antimicrobial efficacy of in vivo and in vitro tissues of *Aegle marmelos* (L.) Corrêa [Internet]. Vol. 8, Annals of Phytomedicine: An International Journal. 2019. p. 140–7. Available from: <http://dx.doi.org/10.21276/ap.2019.8.1.18>
14. Wadhwa R, Paudel KR, Chin LH, Hon CM, Madheswaran T, Gupta G, et al. Anti-inflammatory and anticancer activities of Naringenin-loaded liquid crystalline nanoparticles in vitro. *J Food Biochem*. 2021 Jan;45(1):e13572.
15. Bhatti R, Singh J, Saxena AK, Suri N, Ishar MPS. Pharmacognostic standardisation and antiproliferative activity of *Aegle marmelos* (L.) Correa leaves in various human cancer cell lines. *Indian J Pharm Sci*. 2013 Nov;75(6):628–34.
16. Vivekanandhan K, Shanmugam P, Barabadi H, Arumugam V, Daniel Raj Daniel Paul Raj D, Sivasubramanian M, et al. Emerging Therapeutic Approaches to Combat COVID-19: Present Status and Future Perspectives. *Front Mol Biosci*. 2021 Mar 8;8:604447.
17. Wahab PUA, Madhulaxmi M, Senthilnathan P, Muthusekhar MR, Vohra Y, Abhinav RP. Scalpel Versus Diathermy in Wound Healing After Mucosal Incisions: A Split-Mouth Study. *J Oral Maxillofac Surg*. 2018 Jun;76(6):1160–4.

18. J PC, Pradeep CJ, Marimuthu T, Krithika C, Devadoss P, Kumar SM. Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study [Internet]. Vol. 20, *Clinical Implant Dentistry and Related Research*. 2018. p. 531–4. Available from: <http://dx.doi.org/10.1111/cid.12609>
19. Tahmasebi S, Qasim MT, Krivenkova MV, Zekiy AO, Thangavelu L, Aravindhyan S, et al. The effects of oxygen-ozone therapy on regulatory T-cell responses in multiple sclerosis patients. *Cell Biol Int*. 2021 Jul;45(7):1498–509.
20. Dixit A, Bharati SK, Singh S, Kumar S. THERAPEUTIC SIGNALS OF BILVA (AEGLE MARMELOS CORR.) IN BRIHATTRAYI: A REVIEW [Internet]. Vol. 8, *International Journal of Research in Ayurveda & Pharmacy*. 2017. p. 110–6. Available from: <http://dx.doi.org/10.7897/2277-4343.085256>
21. Kamath SM, Manjunath Kamath S, Jaison D, Rao SK, Sridhar K, Kasthuri N, et al. In vitro augmentation of chondrogenesis by Epigallocatechin gallate in primary Human chondrocytes - Sustained release model for cartilage regeneration [Internet]. Vol. 60, *Journal of Drug Delivery Science and Technology*. 2020. p. 101992. Available from: <http://dx.doi.org/10.1016/j.jddst.2020.101992>
22. Darvesh AS, Aggarwal BB, Bishayee A. Curcumin and Liver Cancer: A Review [Internet]. Vol. 13, *Current Pharmaceutical Biotechnology*. 2012. p. 218–28. Available from: <http://dx.doi.org/10.2174/138920112798868791>
23. Potter JD. Diet and cancer [Internet]. *EPIDEMIOLOGY OF DIET AND CANCER*. p. 65–94. Available from: [http://dx.doi.org/10.4324/9780203168929\\_chapter\\_3](http://dx.doi.org/10.4324/9780203168929_chapter_3)
24. Mudigonda SK, Murugan S, Velavan K, Thulasiraman S, Krishna Kumar Raja VB. Non-suturing microvascular anastomosis in maxillofacial reconstruction- a comparative study. *J Craniomaxillofac Surg*. 2020 Jun;48(6):599–606.
25. Hathway DE. PLANT PHENOLS AND TANNINS [Internet]. *Chromatography*. 1969. p. 390–436. Available from: <http://dx.doi.org/10.1016/b978-0-433-30503-3.50024-9>
26. Girish C, Pradhan SC. Herbal Drugs on the Liver [Internet]. *Liver Pathophysiology*. 2017. p. 605–20. Available from: <http://dx.doi.org/10.1016/b978-0-12-804274-8.00044-8>
27. R H, Hannah R, Ramani P, Ramanathan A, Jancy MR, Gheena S, et al. CYP2 C9 polymorphism among patients with oral squamous cell carcinoma and its role in altering the metabolism of benzo[a]pyrene [Internet]. Vol. 130, *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. 2020. p. 306–12. Available from: <http://dx.doi.org/10.1016/j.oooo.2020.06.021>
28. Santhakumar P, Roy A, Mohanraj KG, Jayaraman S, Durairaj R. Ethanolic Extract of Capparis decidua Fruit Ameliorates Methotrexate-Induced Hepatotoxicity by Activating Nrf2/HO-1 and PPAR $\gamma$  Mediated Pathways [Internet]. Vol. 55, *Indian Journal of Pharmaceutical Education and Research*. 2021. p. s265–74. Available from: <http://dx.doi.org/10.5530/ijper.55.1s.59>

29. Akhouri V, Kumari M, Kumar A. Therapeutic effect of Aeglemarmelos fruit extract against DMBA induced breast cancer in rats. *Sci Rep.* 2020 Oct 22;10(1):18016.
30. Nambi G, Kamal W, Es S, Joshi S, Trivedi P. Spinal manipulation plus laser therapy versus laser therapy alone in the treatment of chronic non-specific low back pain: a randomized controlled study. *Eur J PhysRehabil Med.* 2018 Dec;54(6):880–9.
31. Rathee D, Kamboj A, Sidhu S. Augmentation of hepatoprotective potential of Aeglemarmelos in combination with piperine in carbon tetrachloride model in wistar rats. *Chem Cent J.* 2018 Aug 20;12(1):94.
32. Solai Prakash AK, Devaraj E. Cytotoxic potentials of *S. cuminimethanolic* seed kernel extract in human hepatoma HepG2 cells. *Environ Toxicol.* 2019 Dec;34(12):1313–9.
33. Baliga MS, Thilakchand KR, Rai MP, Rao S, Venkatesh P. Aeglemarmelos (*L. Correa* (Bael) and its phytochemicals in the treatment and prevention of cancer. *Integr Cancer Ther.* 2013 May;12(3):187–96.
34. Rajakumari R, Volova T, Oluwafemi OS, Rajesh Kumar S, Thomas S, Kalarikkal N. Grape seed extract-soluplus dispersion and its antioxidant activity. *Drug DevInd Pharm.* 2020 Aug;46(8):1219–29.