

Evaluation of the influence of some abiotic factors on the microbiological quality of some groundwater points in Soa sub-division, Cameroon

ABSTRACT

Groundwater samples collected were collected from five hand dug wells in Soa subdivision. On the field, the hydrological and morphometric parameters of the study area were measured while the bacteriological and physicochemical parameters were determined in the laboratory. The goal was to assess the microbiological quality of the groundwater and the health risk it posed to the population of Soa. The low piezometric levels (9 – 20 m) and the little variation in curb-stone heights (0.58 – 1.05 m) suggest that the wells are vulnerable to pollution. The physicochemical parameters show that the groundwater of Soa is acidic and poorly mineralized. Most of the population that is dominated by students is not connected to the mains water making the residents recourse to groundwater for potable and general domestic use. Bacteriological analysis of the water samples focused on the search and listing of heterotrophic mesophilic aerobic bacteria (HAMB). Also, bacteria of the Enterobacteriaceae family were isolated, identified and listed (*Escherichia*, *Klebsiella*, *shigella*, *Citrobacter*, *providencia*). All the analysis was performed on standard, selective and conventional gallery culture media, using surface spreading, streaking and sub-culturing techniques. Different ions were also measured for the physicochemical component according to the usual analytical techniques. Low values of oxidability have been recorded and reflect an average organic pollution. Organic matter largely influenced the bacterial load which is high and varied. The presence of these germs degrades the quality of these waters. This varies from one site to the other with important factors being the proximity of the sources of pollution to the well and its poor maintenance. Local populations are using these waters without any prior treatment. They are exposed to health risks in the short and medium terms.

Keywords: groundwater; pathogenic bacteria; Enterobacteria; abiotic variable.

1. INTRODUCTION

In Africa and particularly in Cameroon, water supply companies are still facing major challenges in the provision of fresh potable water to the population. Access to safe drinking water therefore remains a key issue [1, 2] in most developing countries. In Cameroon, the main public water supply company (CAMWATER) has not been able to match its supply with the rapidly increasing population in the country. As a result, many have resorted to the use of alternative sources with groundwater as the most preferred. Though the trend is gradually shifting towards the use of boreholes to harness the groundwater, most of the supply is largely through unprotected hand dug wells, making the source highly vulnerable to microbial pollution. Even though the boreholes relatively more protected than the hand-dug wells, their

location is conditioned by the owner and whose decision to influence the location is also limited within the confines of his property. The result is that most boreholes are located within the contamination range (< 100 m) of pit latrines and septic tanks. Users take the clear appearance of the groundwater to imply purity in complete oblivion of its microbial load [3, 4]. Bacteria of various types make up the majority of the microflora in groundwater [4, 5]. Looking for organisms that suggest contamination or fecal pollution is one way to determine the quality of the water. Enterobacteriaceae (total and fecal coliforms) is a useful indicator whose presence in non-disinfected groundwater depicts degradation of the water quality induced by surface water inputs. Its presence in water can be an indicator of the potential presence of human enteric viruses [6, 7].

Numerous studies have been carried out on the bacteriological quality of groundwater in Yaoundé [8, 9, 10, 11]. Note should be taken that the study area in this study, Soa, is found at the peri-urban outskirts of Yaounde. The previous studies show that groundwater in Yaoundé harbors a varied microflora, made up of fecal bacteria and opportunistic pathogenic bacteria. Isolated bacteria of the Enterobacteriaceae family commonly belong to the *Enterobacter*, *Escherichia*, *Klebsiella*, *Citrobacter*, *Serratia*, *Salmonella*, *Shigella*, *Edwardsiella*, *Proteus* and *Providencia* genera. Their population dynamics is influenced by meteorological parameters and certain abiotic factors of the environment [12].

However, little is known about the groundwater quality of Soa, where pressure on groundwater is increasing rapidly as a result of increasing domestic use. This increase is driven by the inability of CAMWATER, the main public water supply company, to meet the needs of the Soa population. The population, especially that in the student residential areas of the university town of Soa, has resorted to alternative water sources with groundwater as the most favored source because of its generally good quality. That notwithstanding, groundwater remains susceptible to contamination [13-15] and this vulnerability can only be known through data. The paucity of data on this important source of potable water for the Soa community leaves a knowledge gap on its quality and health effects. It is for this reason that the aim of this work is to (i) assess the bacteriological quality (ii) determine the physicochemical composition and (iii) establish the morphometric and piezometric parameters of the sampling points in for groundwater in Soa.

2. MATERIAL AND METHODS

2.1 Study area

Soa subdivision is located in the Mefou and Afamba Division in the Centre Region of Cameroon; some 14 km northeast of Yaoundé. It is roughly circumscribed by latitude 3°50' to 4°10' N and longitude 11°30' to 11°40' E (Fig. 1). Altitudes range from 600 to 730 m and can drop to 300 m in the valleys. It has a surface area of 326 km² [16]. The prevailing climate is humid equatorial. The vegetation consists of forest formations in the south which gradually passes to a peri-forest savannah towards the north [17]. Soa is found in the Nyong basin in which the hydrographic network that is revolves around the Afamba river and its main tributaries: Mbende and Foulou. The soils are mainly ferralitic with two main variants: red clayey and acidic ferralitic soils (pH < 5.5) and yellow clayey iron-rich ferralitic soils [18,19].

2.2 Sampling and analysis methods

Five groundwater samples were collected from five different wells in the study area. The samples were collected between April and August 2016 using protocol developed by the French Standardization Agency [AFNOR (NF-T 90-10)]. The samples were collected in 1liter polyethylene bottles. Prior to this, each bottle was washed and rinsed three times with the water to be sampled, then filled to the brim and sealed hermetically. before being carried in

coolers to the laboratory. Samples for microbiological analysis were collected using 500 mL glass bottles before autoclaving. All samples were stored at 4 °C in coolers immediately after collection and during transportation to the Laboratory of Hydrobiology and Environment and Chemical Laboratory of the Institute of Agricultural Research for Development (IRAD) and then analyzed within 24 hours. Table 1 summarizes the codes assigned to these wells, their geographical coordinate and their average altitudes. The coordinates and the elevation of the various wells were obtained on the field using a Garmin 60CSX GPS.

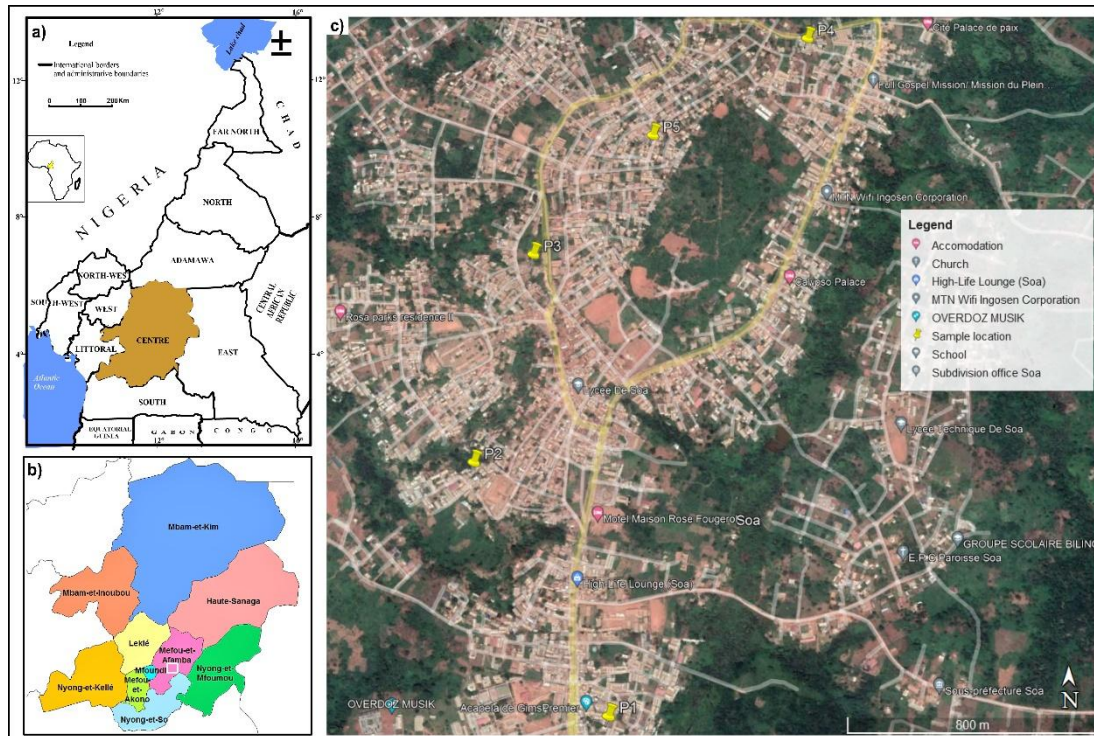


Fig. 1. Investigated area and sampling location: a) location map of the Region of the study area, b) location of Department of the study area and its surroundings, c) location of sampling area and water points in locality of Soa

2.2.1 Analysis of abiotic parameters

On the field, the height of the curb-stone and the piezometric level were measured using a graduated rope. The temperature was measured using a B11403 mercury column thermometer graduated to 1/10th of a degree and the pH was measured using a HACH HQ 30d Flexi portable multimeter. Physicochemical parameters were measured according to the recommendations of Rodier et al. [20]. The total dissolved solids (TDS) and electrical conductivity (EC) of the samples were measured using a TDS/EC portable HANNA series HT 8733 conductivity meter. The dissolved oxygen was determined using a WTW Oxy 340 brand oximeter. The values were given in percentage oxygen saturation.

The color of the water was obtained using a HACH DR/2000 spectrophotometer and the reading was taken at the wavelength of 455 nm. The nitrogen forms in mg/L (nitrate and ammoniacal nitrogen) were determined by colorimetry using a spectrophotometer (HACH DR/2000) and the Nessler reagents were used for the determination of ammoniacal nitrogen and nitrate IV for nitrates. Oxidability, alkalinity and dissolved carbon dioxide were determined by volumetric assay using the appropriate reagents. The major cations, magnesium (Mg^{2+}), calcium (Ca^{2+}), sodium (Na^{+}) and potassium (K^{+}) were determined by

conductimetry using a cleonex ICS 90 and ICS 1100 liquid phase ion chromatograph and the results were expressed in mg /L.

Table 1. GPS coordinates of the wells selected at Soa

Wells	Sites	GPS coordinates		
		Altitudes (m)	Latitudes	Longitudes
P1	In front of the University	663	03°57'57.3	11°36'40.0
P2	Behind the public primary school	659	03°58'23.1	11°35'26.2
P3	Entered general high school	684	03°58'45.2	11°35'32.5
P4	Police station	684	03°59'09.6	11°36'02.8
P5	Château (Foe Road)	700	03°58'58.4	11°36'45.5

2.2.2 Bacteriological analysis

Bacteriological analysis consisted of searching, isolating and counting HMAB and Enterobacteriaceae; as well as the identification of strains of Enterobacteriaceae isolated from well water samples. These microorganisms were identified and enumerated by filtering homogeneous aliquots of 100 mL through a membrane with a pore diameter of 0.45 µm. The membranes were then placed on selective media for 24 hours at 37 °C in a thermostatic oven. The following media were used: Plate Count Agar (PCA) was used for the isolation of HMAB and Eosin Methylene Blue (EMB) lactose agar was used for the isolation of Enterobacteriaceae [21].

The identification of the bacterial colonies isolated on the EMB medium was carried out first by macroscopic observation of the colonies, including the shape, size, outline and color of the colonies. Subsequently, biochemical tests were carried out using the classical gallery [21]. The colony count was performed with the naked eye using a colony counter. The results were expressed in log (CFU / 100 mL).

2.2.3 Statistical analysis

The correlation coefficients between bacterial densities and physicochemical variables were calculated using Spearman's "r" correlation test. The comparisons of the mean of the bacterial densities were carried out by means of the H tests of Kruskal-Wallis and U of Mann-Whitney. These tests were performed using SPSS 16.0 software.

3. RESULTS AND DISCUSSION

3.1 Hydrological and morphometric parameters of wells

The hydrological parameters vary from one well to the other (Table 2). The piezometric levels vary between 9 and 20 m while the heights of the curb-stone range between 0.58 and 1.05 m. The highest curb-stone height was recorded at well P3 and the lowest at well P2. The measured parameters (Table 2) indicate that wells P1 and P2 are not protected, while wells P3, P4, and P5 are protected. the P1 well was not covered while the P2 well had a rusted and perforated metal cover. On the other hand, wells P3, P4 and P5 were properly covered and relatively well equipped (Table 2).

Table 2. Morphometric and hydrological parameters of the wells studied

Wells	P1	P2	P3	P4	P5
Piezometric level (m)	9	19.5	16	18	20

Height of curb-stone (m)	0.9	0.58	1.05	0.85	0.80
Well protection	absent	absent	present	present	present

3.2 Physicochemical parameters

Table 3 shows the mean values and standard deviations of the physicochemical variables for each well. The temperature of the groundwater samples that ranges from ($22.80 \pm 0.2 - 23.60 \pm 0.4^\circ\text{C}$) shows very little variation. This low thermal variation could be explained by the low conductivity of the soil (Ait, 2007). This is in agreement with the report of Asi and Ajeegah [22] in the same area ($23.5^\circ\text{C} - 24^\circ\text{C}$). The pH, that ranges from 5.3 to 6.3 ascribes an acidic character to the groundwater of Soa. Within this range however, well P1 is more acidic than the other four wells (P2, P3, P4 and P5). These results are similar to those obtained by Moussina Yaka et al. [10], in the groundwater of Yaoundé. The acidity of the groundwater is believed to be due to the acidic nature of the soils in the region [23]. It could also be linked to the production of CO_2 in the surface layers of the soil under the action of biological activities [24, 25, 26]. In addition, the pH values obtained suggests a decrease in the dissolution of CO_2 in the groundwater. This result is similar to that obtained in the groundwater of Cross River Basin in Nigeria [27].

The mean values of EC and TDS varied depending on the water sampling points. They were high for well P1 ($289.4 \mu\text{S} / \text{cm}$ and $145 \text{ mg} / \text{L}$ respectively) and relatively low for wells P2 ($35.7 \mu\text{S} / \text{cm}$ and $17 \text{ mg} / \text{L}$), P3 ($47 \mu\text{S} / \text{cm}$ and $23 \text{ mg} / \text{L}$), P4 ($33.2 \mu\text{S} / \text{cm}$ and $16.4 \text{ mg} / \text{L}$) and P5 ($47.5 \mu\text{S} / \text{cm}$ and $24 \text{ mg} / \text{L}$). The major cation (Na^+ , K^+ , Mg^{2+} and Ca^{2+}) and nitrate ($\leq 2.5 \text{ mg} / \text{L}$ of NO_3^-) contents remain generally low. These values show that these waters are soft and lowly mineralized. However, the high values observed in well P1 ($27.80 \text{ mg} / \text{L}$ of Na^+ , $4.97 \text{ mg} / \text{L}$ of K^+ , $1.10 \text{ mg} / \text{L}$ of Mg^{2+} , $8.30 \text{ mg} / \text{L}$ of Ca^{2+} and $1.60 \text{ mg} / \text{L}$ of NO_3^-) for these different variables could be due to the low level of protection of the well (absence of well cover and presence of grasses along the internal wall of the well) and also to the shallow depth of the well. This is in conformity with the assertion that groundwater that is near to the surface of earth is more susceptible to pollution, Mbawala et al. [28]. The mineralogical composition of the groundwater could also be influenced by the mineralogy of the host rock, the contact time with the minerals, the water circulation speed, the aquifer water renewal time and the rainfall [29].

The mean water color values fluctuated from 11.4 to 56.6 Pt. Co. This high color value was largely influenced by the drop in the water column in each well in July. The oxidability values ranged from 3.3 to 5.3 mg /L. They showed that the water is moderately loaded with organic matter. The WHO sets a threshold of 5 mg /L of oxidability in drinking water.

The oxygen saturation percentage (Sat %) varied from 64.8% to 66.5% and reflected a fairly good oxygenation of the water. These results differed from those obtained by Nougang et al. [30] where a low to medium saturation rate in dissolved oxygen was recorded. The relatively high value (66.50 %) recorded at the P2 well may be attributed to the fact that open the well is open. This increases the contact and exchanges between water and air in the ambient environment.

The dissolved carbon dioxide contents fluctuated between 2.8 and 4.2 mg /L. The NO_3^- contents were relatively low for wells P2 (0.4 mg /L), P3 (0.8 mg /L), P4 (0.5 mg /L) and P5 (0.9 mg/L) as compared to well P1 (1.6 mg/L). However, they remain below the standards recommended by the WHO in drinking water (50 mg /L).

Table 3. Average values and standard deviations of the physicochemical parameters measured by station

Parameters	Well 1	Well 2	Well 3	Well 4	Well 5
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T (°C)	23.20 ± 0.2	22.80 ± 0.2	23.60 ± 0.3	23.10 ± 0.3	23.60 ± 0.4
pH	6.30 ± 0.4	5.60 ± 0.3	5.40 ± 0.6	5.30 ± 0.4	5.50 ± 0.6
TDS (mg/L)	145 ± 27.1	17 ± 4.4	23 ± 4.5	16.40 ± 6.1	24 ± 3.7
EC (µS/cm)	289.40 ± 50.5	35.70 ± 9.5	47 ± 10.1	33.20 ± 13.8	47.50 ± 5.8
Color (Pt.Co)	20.20 ± 19.4	56.60 ± 59.3	45.40 ± 56.8	11.40 ± 16.8	23 ± 74
Alcalinity (mg/L)	1.80 ± 0.4	1.80 ± 0.4	1.80 ± 0.4	1.80 ± 0.4	2.20 ± 1
CO ₂ (mg/L)	4.20 ± 1.5	2.80 ± 0.9	2.80 ± 1.5	3.50 ± 1.2	2.80 ± 0.9
O ₂ (%)	64.80 ± 2.3	66.50 ± 6.1	65.20 ± 2.9	65.40 ± 3.7	65.50 ± 3.2
Oxydability (mg/L)	3.40 ± 1.4	3.30 ± 2.3	5.10 ± 0.3	4.80 ± 1	5.30 ± 0.6
NO ₃ ⁻ (mg/L)	1.60 ± 0.4	0.40 ± 0.3	0.80 ± 0.5	0.50 ± 0.2	0.90 ± 0.7
Mg ²⁺ (mg/L)	1.10 ± 0.5	0.40 ± 0.1	0.60 ± 0.2	0.50 ± 0.3	0.70 ± 0.4
Ca ²⁺ (mg/L)	8.30 ± 4.5	3.50 ± 0.6	4.80 ± 2.2	3.10 ± 0.9	5.30 ± 2.5
K ⁺ (mg/L)	4.97 ± 2.6	0.93 ± 0.2	1.38 ± 1.1	1.12 ± 0.8	0.5 ± 0.1
Na ⁺ (mg/L)	27.80 ± 16.9	1.02 ± 0.6	1.83 ± 1.6	1.42 ± 0.8	2.05 ± 2.4

3.3 Bacteriological parameters

Qualitatively, whitish to yellowish colonies of various diameters were obtained color on PCA medium. Those obtained on EMB medium and identified belong to 5 bacterial genera within the Enterobacteriaceae family in all the water samples. These include the genera *Escherichia*, *Klebsiella*, *Shigella*, *Citrobacter* and *Providencia*.

Quantitatively, the means of bacterial abundances shown in (Table 4) varied from one bacteria to another. The abundances of heterotrophic aerobic mesophilic bacteria (HAMB) ranged from 5.45 to 5.95 log CFU / 100 mL in different wells. These recorded abundances are above the WHO standard [31] which sets a threshold of 2 log CFU / 100 mL and significantly higher ($P < 0.05$) than those of the identified genera of Enterobacteria (Table 4). Bacteriological analysis carried out on water samples taken from the wells during the study period show that, these waters contain bacteria resulting from fecal contamination. Among the main groups of bacteria indicating fecal contamination, two have been represented. These are total coliforms, represented by bacteria of the genera *Klebsiella* and *Citrobacter*, and fecal coliforms represented mainly by the genera *Escherichia*. The abundances of these two groups of bacteria have varied from one well to another over time. The abundances of bacteria of the genus *Klebsiella* ranged from 3.15 to 3.76 log CFU / 100 mL and those of the genera *Citrobacter* fluctuated from 3.45 to 4.03 log CFU / 100 mL. The mean values of the *Klebsiella* and *Citrobacter* abundances recorded were 1.56 and 1.85 log CFU / 100 mL respectively, while the abundances of *Escherichia* ranged from 0 to 3.08 log CFU / 100 mL (Table 4). These values were all higher than the standards set by the WHO for drinking water. These standards set the levels of fecal coliforms and total coliforms at 1.3 and 1.69 log CFU / 100 mL respectively. These results are similar to those obtained by Buguioh et al. [4], in spring water in the West region of Cameroon.

Table 4. Averages of bacterial abundances according to wells

Bacteria (Log UFC/ 100 ml)	Wells				
	P1	P2	P3	P4	P5
HAMB	5.95	5.69	5.78	5.79	5.45
<i>Escherichia</i>	3.08	2.90	2.30	0	0
<i>Klebsiella</i>	3.45	3.38	3.15	3.76	3.34
<i>Shigella</i>	3.66	3.25	4.17	4.07	3.80
<i>Citrobacter</i>	3.60	3.88	3.45	3.5	4.03
<i>Providencia</i>	3.41	3.08	3.73	2.90	3.25

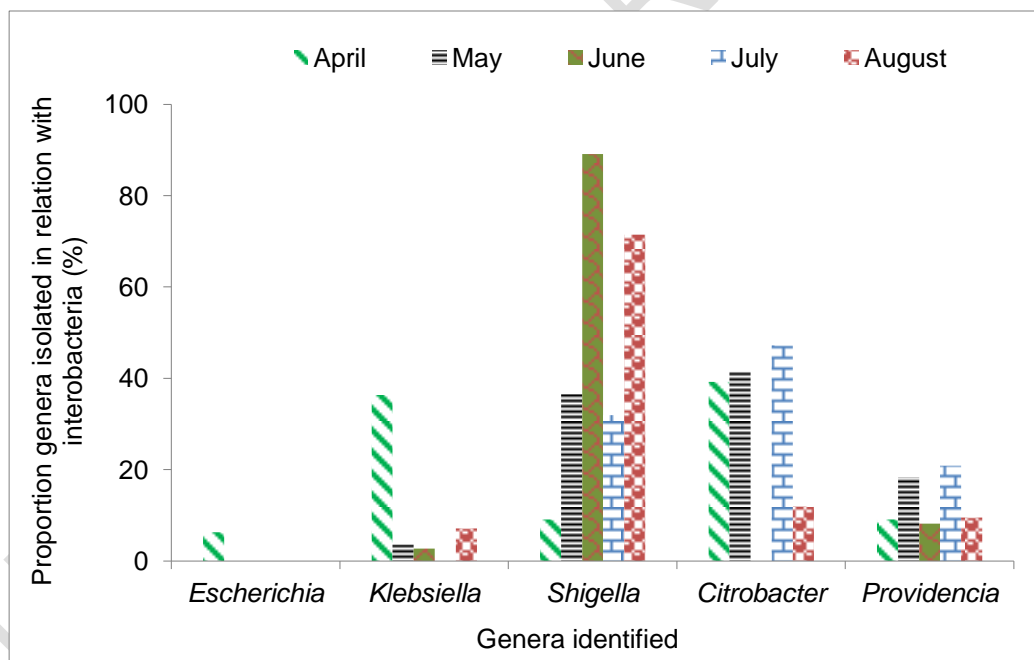
Apart from these two bacterial groups, other pathogenic and opportunistic bacteria identified are bacteria of the genera *Shigella* and *Providencia*. The abundances of bacteria of the genera *Shigella* oscillated between 3.25 and 4.17log CFU / 100 mL. Those of the genera *Providencia* fluctuated between 2.9 and 3.73 log CFU / 100 mL (Table 4). The permanent presence of these pathogenic bacteria and their high abundance 3.79 log CFU / 100 mL (*Shigella*) reflects the degree of pollution in these water samples and the high health risk. Similar results have been obtained by Mpakam et al. [32] in Bafoussam and Moussina Yaka et al. [10], in Yaoundé, where many pathogens such as *Escherichia coli*, *Salmonella*, *Shigella* and many other bacteria witnessing fecal contamination have been observed in the groundwaters.

The major differences in bacterial abundance observed in the five analyzed wells are influenced by the structural character (opened or closed) of the well. A greater bacterial density in closed wells (P3, P4 and P5) than in open wells (P1 and P2) was noted. This shows that closed wells are more polluted than open wells. In fact, the closed wells considered in this study, although protected on the outside by adequate copings, covers and pulley systems, contain other sources of pollution in their environment. These are most likely the toilets in the case of well P4 located upstream of sloping ground and about 2 m from the well. In addition, these wells we observed to be close the agricultural fields and gardens. All these parameters account for the high abundance of the observed bacterial populations as well as their great diversity. Socioeconomic characteristics of households influences the type of toilet and the mode of waste management, Wright et al. [33]. This implies that high income households will have better sanitary facilities and manage their wastes better. The low income status of most of the households in the study area explains the poor sanitary facilities and low waste management capabilities which are directly impinging on the quality of groundwater in the Soa.

3.4 Relative frequencies of bacterial abundance

The ratios (monthly relative frequencies) of the average abundances of each bacterial genus identified to those of HAMB on one hand and to the average sum of Enterobacteriaceae identified on the other hand were calculated and are represented respectively in Figures 2 and 3. Figure 2 shows that the abundances of the different identified genera vary from one month of sampling to the other, appearing high in some months and dropping to rarity in some other months. This is the case with bacteria of the genus *Escherichia*, which turned out to be very rare during the months of May, June, July and August. In addition, the abundances of the bacterial genera identified represent only 4% of the HAMB counted.

Fig. 2. Variations in the proportions of each bacterial genus identified compared to HAMBs



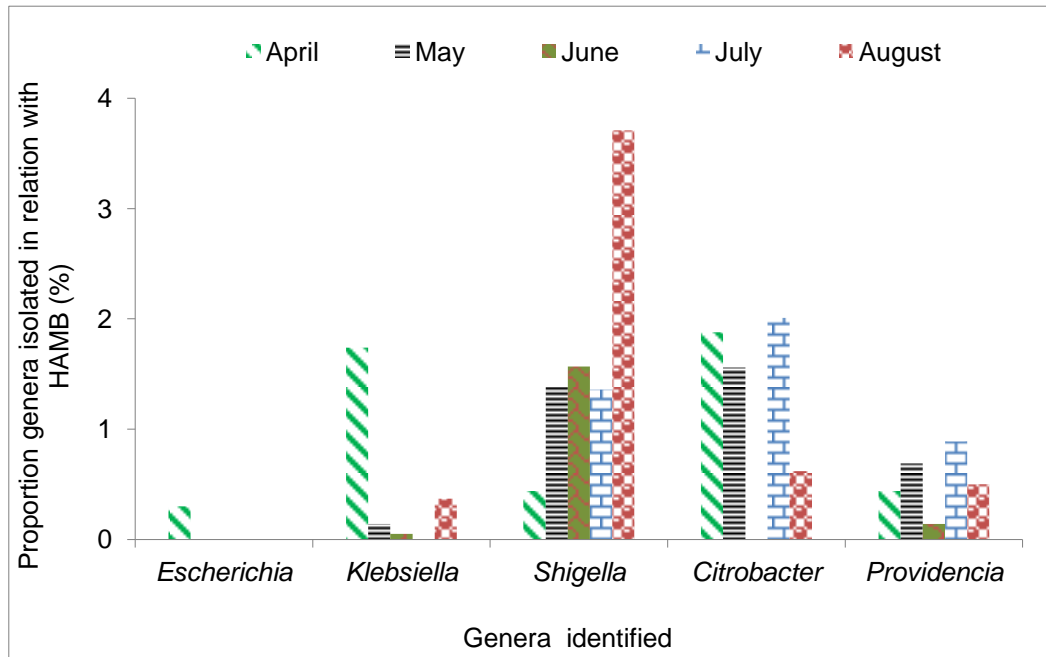


Fig. 3. Variations in the proportions of each bacterial genus identified compared to all Enterobacteria

In all the bacteria identified, those of the genus *Shigella* showed the greatest abundance (Fig. 3). Eighty percent of the enterobacteria identified in June were of the genus *Shigella* while less than 20% was of the genera *Escherichia* and *Providencia*. Similar results have been obtained by Nanga et al. [12] in Yaoundé and Ndjama et al. [34] in Douala.

3.5 Statistical analysis

3.5.1 Spearman's correlation

Spearman's "r" correlation tests were carried out among the mean bacterial abundances of the various wells (Table 5) and between the bacterial mean abundances and physicochemical parameters (Table 6). The correlations made between the mean abundances of different isolated bacterial strains indicate a very significant and positive correlations between the abundances of the genera *Escherichia* and those of the genera *Klebsiella* (0.549) and *Citrobacter* (0.512). A significant and positive correlation was also noted between *Citrobacter* and *Klebsiella* (0.397) (Table 5).

Table 5. Correlations between the different bacterial strains isolated

	<i>Escherichia</i>	<i>Klebsiella</i>	<i>Shigella</i>	<i>Citrobacter</i>	<i>Providencia</i>	HAMB
<i>Escherichia</i>	1	0.549**	-0.335	0.512**	-0.098	0.264
<i>Klebsiella</i>		1	0.06	0.397*	-0.303	0.086
<i>Shigella</i>			1	0	0.084	0.162

<i>Citrobacter</i>	1	0.173	0.147
<i>Providencia</i>		1	0.316
HAMB			1

*: $P < 0.05$ **: $P < 0.01$

From the correlations made between the physicochemical parameters and the mean abundances of isolated bacterial, it is seen that, very significant ($P < 0.01$) and positive correlations exist between the abundances of *Klebsiella* and parameters such as electrical conductivity (0.742), TDS (0.840), NO_3^- (0.614) and Mg^{2+} (0.713), Ca^{2+} (0.572) and K^+ (0.589) (Table 6). The same observation was made between *Escherichia* and Mg^{2+} (0.549), Ca^{2+} (0.565), Na^+ (0.563) and K^+ (0.565) and between *Citrobacter* and K^+ (0.549) (Table 6). Significant ($P < 0.05$) and positive correlations were recorded on the one hand between *Escherichia* and parameters such as pH (0.447), electrical conductivity (0.491), TDS (0.476) and NO_3^- (0.487) and on the other hand between *Citrobacter* and pH (0.485) and NO_3^- (0.397) (Table 6). A significant and negative correlation was noted between *Citrobacter* and oxidability (-0.430) (Table 6).

The results of the correlations between biological and physicochemical variables show that 9 variables significantly influenced the population and the distribution of bacteria throughout the study. Increase in pH of water significantly increases the abundance of bacteria of the genera *Escherichia*, *Klebsiella* and *Citrobacter*. This is congruent with Nola et al's [35] observation that in a given medium, increases in pH sometimes favor the development of *Pseudomonas aeruginosa*, *Aeromonas hydrophila*, as well as the abundance of faecal coliforms and faecal streptococci. The levels of nitrates and oxidability, which represent the organic matter in the medium are significantly positively correlated with the genera *Escherichia*, *Klebsiella* and *Citrobacter*, for nitrates, and negatively with the genera *Citrobacter*, for oxidability. This difference results from the fact that, bacteria react differently with organic matter depending on the composition of the latter. Organic matter influences the availability of nutrients while serving at the same time as a source of energy and carbon for certain microorganisms [36]. Major cations (Mg^{2+} , Ca^{2+} , K^+ and Na^+ ions) and other mineral elements represented by the electrical conductivity of water and TDS are very significantly correlated with bacteria of the genera *Escherichia*, *Klebsiella* and *Citrobacter*. Same negative correlations between these cations (Mg^{2+} , Ca^{2+} , K^+ and Na^+) and the other genera (*Shigella* and *Providencia*) are noted. The influence of major cations on microflora of the soil and the subsoil varies depending on the ability of the bacterial species to degrade these cations (either to neutralize its toxicity or to make nutrients available) and the energy source which is necessary for their biosynthesis. In addition, the activities of certain microorganisms lead to the release and migration of certain ions such as Mg^{2+} , Ca^{2+} , SO_4^{2-} and many others [24].

Table 6. Correlations between the bacteria isolated and the physicochemical parameters measured

Physicochemical variables	Microbiological variables					
	HAMB	<i>Escherichia</i>	<i>Klebsiella</i>	<i>Shigella</i>	<i>Citrobacter</i>	<i>Providencia</i>
Temperature	-0.033	0.094	0.124	-0.163	-0.045	0.294
pH	0.222	0.447*	0.424*	-0.209	0.485*	0.176

EC	0.35	0.491*	0.742**	-0.045	0.263	-0.18
Color	-0.13	-0.295	0.119	0.361	-0.167	0.123
O ₂	-0.034	-0.132	0.05	0.32	0.23	0.14
CO ₂	-0.031	0.392	0.212	-0.14	0.29	0.164
TDS	0.24	0.476*	0.840**	0.03	0.2	-0.312
Nitrates	0.256	0.487*	0.614**	-0.041	0.397*	-0.047
Oxidability	-0.094	-0.137	-0.259	-0.144	-0.430*	0.113
NH ₄ ⁺	-0.129	-0.123	0.255	0.319	0.135	0.143
Mg ²⁺	-0.079	0.549**	0.713**	-0.197	0.278	-0.374
Ca ²⁺	0.17	0.565**	0.572**	0.018	0.268	-0.251
Na ⁺	0.322	0.563**	0.267	-0.373	0.052	-0.064
K ⁺	0.233	0.565**	0.589**	-0.018	0.549**	-0.015

*: $P < 0.05$ **: $P < 0.01$

3.5.2 Kruskal Wallis comparison

Several comparison tests (Kruskal Wallis H test) between the mean abundances of HAMB and the different bacterial genera and between the abundances of bacterial and the months of sampling were carried out. The tests reveal that, a significant difference ($p < 0.05$) exists between the mean abundances of HAMB and those of Enterobacteriaceae (Table 7). The abundance of HAMB and those of bacteria of the genera *Escherichia*, *Klebsiella* and *Citrobacter* varied significantly ($p < 0.05$) between months of sampling. Only the abundances of bacteria of the genera *Shigella* remained constant over time (Table 8).

Table 7. P-values indicating the significance thresholds relative to the comparison of the mean abundances of the genera of Enterobacteriaceae and of HAMB

	Bacterial genera				
	<i>Escherichia</i>	<i>Klebsiella</i>	<i>Shigella</i>	<i>Citrobacter</i>	<i>Providencia</i>
HAMB	0.001*	0.001*	0.001*	0.001*	0.001*

*: $P < 0.05$

Table 8. P-values indicating the significance thresholds relating to the Kruskal-Wallis H test between the genera of Enterobacteriaceae and the months of sampling

	<i>Escherichia</i>	<i>Klebsiella</i>	<i>Shigella</i>	<i>Citrobacter</i>	<i>Providencia</i>	HAMB
Month	0.011*	0.003*	0.908	0.028*	0.544	0.024*

*: $P < 0.05$

4. CONCLUSION

Groundwater samples taken from certain wells in the Soa region were analyzed to deduce their bacteriological and physicochemical contents. Prior to that, the morphometric and hydrological characteristics of the sampling points were determined since they are known to influence the bacteriological and physicochemical characteristics of the water.

Piezometric and morphometric evaluation reveals that the groundwater of Soa especially that accessed through hand dug wells has a low piezometric level and is not protected. This exposes the water to several sources of pollution.

Physicochemical assessment points towards an acidic, poorly mineralized, groundwater that is susceptible to pollution and degradation of its physicochemical quality.

Bacteriological analysis reveals water that is not recommended for drinking because of the presence of pathogenic germs that are indicative of high levels of fecal pollution. These bacteria are responsible for gastroenteritis, hepatovesicular, vaginal and urinary tract infections in humans.

Physicochemical parameters such as pH, electrical conductivity, TDS, oxidability, nitrates and major cations (Ca^{2+} , Na^+ , K^+ and Mg^{2+} ions) are clearly seen to influence the load and distribution of bacteria in the water. The bacterial load is also influenced by the proximity of the wells to sources of pollution and the maintenance of wells.

In general, this study can be considered as a baseline study that underscores the risk of groundwater pollution. It should therefore serve as a reference for future studies on the risk of pollution and contamination of groundwater in the Soa subdivision of Cameroon.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

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