

Original Research Article

The Distribution and Seasonal Variation of Zooplankton Species of the Great Kwa River, Calabar, Nigeria: A Reassessment Approach

ABSTRACT

Background: Human activities adversely affect the distribution and diversity of zooplankton. They are diverse group of organisms with little or no locomotive ability and quickly respond to changes in their environment. This research was aimed at providing updated information on the distribution and seasonal variation of zooplankton in Great Kwa River.

Materials and Methods: Two sampling stations (S1- ObufaEsuk and S2- EsukAtu) were mapped along the river bank, samples were collected using plankton net of 55µm mesh size and preserved in 4% formalin. Species were identified using taxonomic keys. Data were analyzed using ecological indices.

Results: The results revealed 12 taxa; belonging to 38 species. *Tintinnida*, *Protozoa*, *Cladocera*, *Copepodawere* 23.1%, 18.5%, 15.4% and 13.3% respectively. The lowest taxonomic groups were *Diptera*, *Foraminitera*, *Atenata data* and *Trichoptera* having 1.85% for each order. The highest species was recorded in S2 having 31 species. In both Stations *Ascampbelliellaacuta* was dominance over other species. Shannon-Weiner index (H) were 2.997 and 2.40 in S2 and S1 respectively. The evenness index were 0.576 and 0.547 for S1 and S2 respectively. Margalef's diversity index were 8.171 and 4.111 for Esuk Atu-S2 and Obufa Esuk-S1 respectively. Zooplankton species were abundant in wet season than dry season.

Conclusion:This present study provide updated information on the zooplankton distribution, diversity and seasonal variations of the Great Kwa River. The high dominance of *Tintinnida* in this study indicates a natural linkage between nano-planktons and macro-planktons in the food webs of the river.

Keywords: Distribution, Seasonal variation; Zooplanktonspecies; Kwa river; Calabar

1. INTRODUCTION

Human activities through industrialization, deforestation, farming, oil exploration and transportation have resulted in climate changes which adversely affect the distribution, assembly structure of aquatic communities, especially the composition, richness and species diversity [1-3]. Activities of humans also lead to discharge of untreated animal wastes like releases from sewage and septic tanks, run-off from farm lands, laundering waste into surrounding bodies of water. Continuous subjecting these water bodies with high level of pollutants will consequently affect the health status of zooplanktons negatively; therefore influencing the distribution patterns of aquatic animals in the water [3-4].

Zooplankton species are essential constituent of the food chain in aquatic ecosystem. They feed on the phytoplankton, bacteria, aggregates of detritus and microorganisms and other zooplankton species. Almost all freshwater fish feed on zooplankton at different stage in their life cycle [5-7]. Zooplankton are heterotrophic planktonic animals. These are suspended in water with limited power of movement. Like phytoplankton, they are usually defused in water. Freshwater zooplankton are dominated by four major groups of animals namely *Protozoa*, *Rotifera* and two sub-classes of *Crustacea*- *Cladocera* and *Copepoda*[4,8,9]. Zooplankton is an important plankton component that regulates phytoplankton and microbial productivity by feeding on them. Feeding on phytoplankton and bacteria serve as main food for larval fishes, juvenile fishes and other carnivorous aquatic organisms [8].The zooplankton feed on phytoplankton and facilitate the conversion of plant material into animal tissue and in turn constitute the basic food for higher animals including fishes, especially their larve [5,9], other aquatic vertebrates and man [10,11].

These fishes and some aquatic vertebrates are good source of protein, minerals, fat and oil for humans[9].

Studies have shown that phytoplankton diversity increased zooplankton productivity [10,11], while the effect of phytoplankton evenness on resource-use efficiency(RUE) switched from negative at the producer level (phytoplankton) to positive at the consumer level (zooplankton) [12] in some lakes investigated in United State of America. Zooplankton species diversity, distribution and abundance in the aquatic world has a vital influence on the healthiness of the aquatic environment [13] and differs temporally and spatially from sea to sea. It may be influence by competition, predation, physical, biological and chemical parameters [14]. Biological factors include predation, breeding, phytoplankton concentration and vertical migration. The physical factors are mixing of the oceanic water (down welling and upwelling) that influences nutrients availability; and on the other hand affect phytoplankton production and subsequently in the future affect the zooplankton speciation [9-11,15,16-18].

The water bodies play crucial roles in the economy and aquatic food supply for the inhabitants of Calabar and its neighboring villages, serving as their major source of fish and employment to the local people. Despite these vital roles played by the Great Kwa River to the good people of Cross River State, little or no measures have been put in place to conserve the endangered species due to uncontrolled fishing and other detrimental human activities in the aquatic environment. Against this back drop, there is urgent need for an updated and current information on zooplankton species in the great Kwa River due to increased human activities (uncontrolled fishing, release of untreated sewage, etc) and industrialization; geared toward species

conservation and as a tool for foretelling the impact of human activities in the river. Therefore, this study aimed at investigating on current distribution and seasonal variation of zooplankton species in the Great Kwa River, Calabar south, Nigeria.

2. MATERIALS AND METHODS

2.1 Study location

This study was conducted in Great Kwa River, Cross River State. The area is located between latitude $8^{\circ} 15' E$ and $8^{\circ} 30' E$ and longitude $4^{\circ} 45' N$ and $5^{\circ} 15' N$. It has an estimated length of 56km and is about 2.8km wide at the mouth where it empties into the Cross River Estuary [17]. Two climatic seasons occur in the study area (wet and dry). The wet season is usually characterized by heavy rainfalls while the dry season experiences occasional downpours. The shore-lines are lined with dark plates usually exposed during low tides and the shore is brackish and rich with zooplankton. The banks are also surrounded by lush, evergreen, forest vegetation with different species of trees, shrubs and grasses.

2.2 Selection of sampling stations

Two sampling stations were mapped along the River bank. Station 1 (ObufaEsuk); designated as S1 located between Latitude $4^{\circ} 94' N$ and Longitude $8^{\circ} 35' E$, close to the Biological Sciences, University of Calabar and University of Calabar Teaching Hospital. Station 2 (EsukAtu); designated as S2 located between Latitude $4^{\circ} 95' N$ and Longitude $8^{\circ} 36' E$ close to the University of Calabar staff quarters (Figure 1) [17].

Comment [S1]: Why these two sampling stations were chosen and why only two? Explain at work

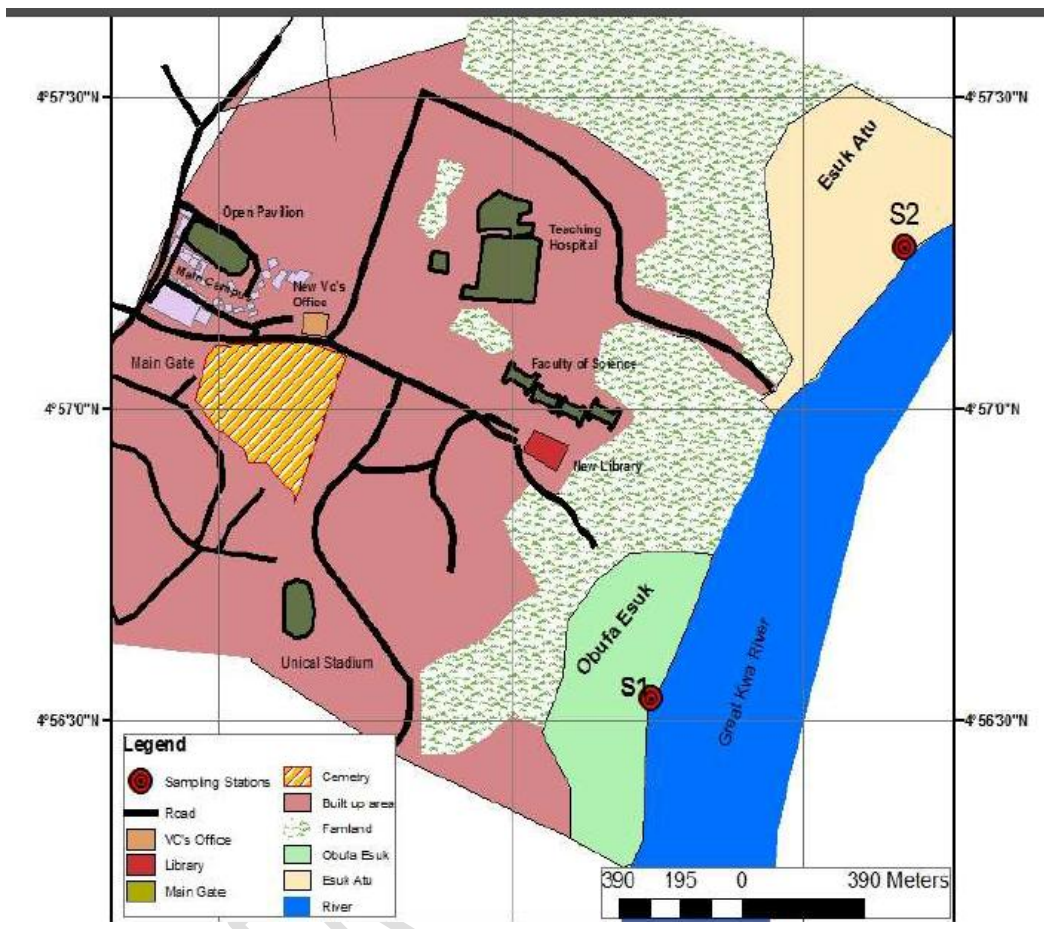


Figure 1: Map showing the study location of S1 and S2
 Source: Okorafor, K. A. *et al.*, [17].

2.3 Zooplankton sample collection

A 55µm mesh standard plankton hydrobios net was used to collect zooplankton samples by towing method. Three replicates were collected at each station. After filtration, the zooplankton was collected in sampling bottles and was preserved in 4% formalin solution before transferring to Institute of Oceanography Laboratory, University of Calabar, Calabar. Some identification of zooplankton species was also carried out in the Department of Zoology and Environmental Biology, Faculty of Biological Sciences, University of Calabar, Calabar. Samples were taken and all individual taxa present were examined and counted. Species were sorted and counted using dissecting microscope at different magnifications. The abundance of zooplankton was determined using ecological indices.

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2.4 Procedure for zooplankton analysis

In the laboratory, zooplankton samples were allowed to settle for two days, thereafter some liquid were decanted to get concentrated samples which were stored in 50ml bottles. A homogenate of the sample fixed with 4% formalin solution was put in a 1ml plankton chamber (AJAH Model 001) and allowed to settle after covering it with a glass slide. Examination was carried out at 100x magnification; using the x10 magnification lens. They were identified to species levels using identification key guide or taxonomic keys [19-20]. Also relevant materials were utilized for species identification [21-26].

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2.5 Ecological indices and statistical analysis

Analysis was carried out using software packages. Software package XLSTAT and JMP were used to analyze biological parameters in order to determine the zooplankton abundance and

diversity of the Great Kwa River. Ecological indices of zooplankton was estimated using margalef's index (D), Shannon-weiner (H), Pielou's evenness (E) and Simpson's dominance indices as documented [27]. Simple percentage was also used.

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3.RESULTS

3.1 Zooplankton taxa and species distribution

The results of this study revealed a total of 12 taxa belonging to 38 species. The order *Tintinnida* was represented by three species (*Ascampbelliella acuta*, *Coxiella longa* and *Coxiella mariana*) consisting of 23.1%, followed by *Protozoa*; which was represented by 10 species (like *Blephanirisma* species, *Loxodes* species, *Nebella caudate*, *Trigonopyxis arcula*, *Plagiopyxis callids*, *Arcelladiscoides*, etc) with 18.5%, while *Cladocera* was 15.4% of the total abundance of zooplankton species. The lowest taxonomic groups were *Diptera*, *Foraminifera*, *Atenadata* and *Trichoptera* having only one representative with 1.85% for each order as displayed on Table 1 and figure 2. The highest number of species were recorded in Esukatu (S2) station with 31 species represented during the research period. In both Stations (Obufa Esuk and Esukatu), *Ascampbelliella acuta* displayed high dominance over other species. The following taxonomic groups namely: *Diptera*, *Foraminifera*, *Atenadata* and *Trichoptera* where only found in station two (Esukatu) (Table 1).

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3.2 Zooplankton diversity

The trend of Shannon-Weiner index for both stations in the Great Kwa River can be illustrated as $S2 > S1$. The highest value of 2.997 (Shannon-Weiner index) was recorded in S2 (Esukatu) whereas the minimum of 2.40 was detected in S1 (Obufa Esuk) station. Margalef's

diversity index were 8.171 and 4.111 for Esuk Atu-S2 and Obufa Esuk-S1 respectively depicting richness in diversity of zooplankton species in the study area. The Equitability or evenness index was slightly high in S1 (0.576) than in S2 (0.547) as shown on Table 2. In Obufa Esuk (S1), 8 individuals belonging to 3 taxa were recorded in February; while 21 individuals from 5 taxa were recorded in July. In S2 (EsukAtu), 20 individuals belonging to 6 taxa were recorded in February, while 57 individuals belonging to 5 taxa were recorded in July. In summary, zooplankton species were relatively abundant in wet season (June and July) than during dry season (February and March) (Table 3).

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Table 1: Composition and Distribution of zooplankton species in the Great Kwa River during the study period

| Taxonomic group | Species | No. of species | | Individual | |
|---------------------|------------------------------|----------------|-----------|------------|----|
| | | S1 | S2 | S1 | S2 |
| <i>Copepoda</i> | <i>Copepodnauplii</i> | 2 (1.03) | | 2 (1.03) | |
| | <i>Tropocyclopsprasinus</i> | 3(1.54) | | 7(3.59) | |
| | <i>Eurytemora</i> | 2(1.03) | | 2(1.03) | |
| | <i>Bryocaptusbeisteinii</i> | 1(0.51) | | 5(2.56) | |
| | <i>Paracyclopsfimbriata</i> | 1(0.51) | | 1(0.51) | |
| <i>Nemata</i> | <i>Ethmolaimusamericanus</i> | 1(0.51) | | 2(1.03) | |
| | <i>Anonchusmonhystera</i> | 5(2.56) | | 7(3.59) | |
| | <i>Bastianaexilis</i> | 4(2.05) | | 4(2.05) | |
| <i>Ostracoda</i> | <i>Ostracod spp.</i> | 1(0.51) | | 8(4.10) | |
| | <i>Limnocythereverrucosa</i> | 2(1.03) | | 3(1.54) | |
| <i>Rotifera</i> | <i>Notholcaacuminata</i> | 2(1.03) | | 5(2.56) | |
| | <i>Aspladinaprodonta</i> | 1(0.51) | | 1(0.51) | |
| | <i>Keratellalongispina</i> | | | 2(1.03) | |
| | <i>Keratellaquadrata</i> | | | 1(0.51) | |
| | <i>Platytiasquadriformis</i> | | | 1(0.51) | |
| <i>Oligochaeta</i> | <i>Pentanueraspp</i> | 1(0.51) | | 2(1.03) | |
| <i>Protozoa</i> | <i>Blephanirisma spp.</i> | 1(0.51) | | 3(1.54) | |
| | <i>Protozoa spp.</i> | 4(2.05) | | 6(3.08) | |
| | <i>Arthrodesmum incus</i> | 2(1.03) | | 5(2.56) | |
| | <i>Centropyxis spp.</i> | 1(0.51) | | 2(1.03) | |
| | <i>Loxodes spp.</i> | 1(0.51) | | 4(2.05) | |
| | <i>Arcella spp.</i> | | | 1(0.51) | |
| | <i>Nebella caudate</i> | | | 2(1.03) | |
| | <i>Trigonopyxisarcula</i> | | | 1(0.51) | |
| | <i>Plagiopyxiscallids</i> | | | 2(1.03) | |
| | <i>Arcelladiscoides</i> | | | 1(0.51) | |
| <i>Cladocera</i> | <i>Daphnia magna</i> | | | 3(1.54) | |
| | <i>D. rosa</i> | 3(1.54) | | 6(3.08) | |
| | <i>D. pulex</i> | 1(0.51) | | 3(1.54) | |
| | <i>Conchoeciaelegans</i> | 2(1.03) | | 3(1.54) | |
| | <i>Gyrinus spp</i> | 2(1.03) | | 7(3.59) | |
| <i>Tintinnida</i> | <i>Ascampbelliellaacuta</i> | 8(4.10) | | 29(14.87) | |
| | <i>Coxiella longa</i> | | | 4(2.05) | |
| | <i>Coxiella mariana</i> | | | 4(2.05) | |
| <i>Diptera</i> | <i>Chironomus spp.</i> | | | 1(0.51) | |
| <i>Foraminifera</i> | <i>Globigerina bulloides</i> | | | 1(0.51) | |
| <i>Atentadata</i> | <i>Bolinopsiscumis</i> | | | 1(0.51) | |
| <i>Trichoptera</i> | <i>Glossosoma spp.</i> | | | 1(0.51) | |
| TOTAL | 12 | 38 | 51 | 144 | |

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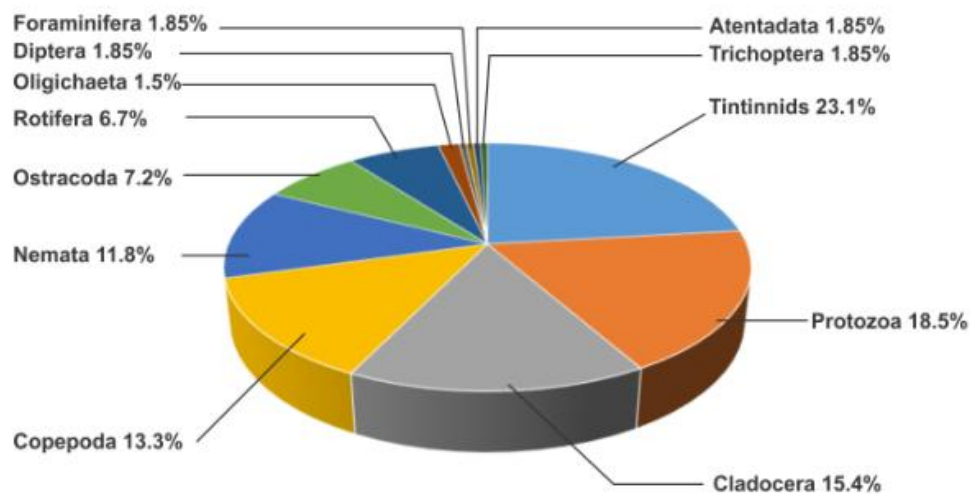


Fig. 2: A chart showing the relative abundance of zooplankton during the period of study

Table 2: Species diversity indices of zooplankton taxonomic group in the Great Kwa River in relation to sampling stations

| Parameters | Sampling stations | Total | |
|--------------------------------------|--------------------------|---------------------|---------------|
| | ObufaEsuk (S1) | EsukAtu (S2) | |
| No of taxonomic group | 12 | 8 | |
| No of individual | 51 | 144 | |
| Margalef's Index (d) | 4.111 | 8.171 | 12.282 |
| Shannon-wiener Index (H) | 2.40 | 2.997 | 5.397 |
| Pielou's evenness Index (E) | 0.576 | 0.547 | 1.123 |
| Simpson's Dominance Index (D) | 0.403 | 1.553 | 1.956 |

Table 3: Summary of Seasonal distribution of zooplankton in the Great Kwa River for S1 and S2 stations

| Month of sampling | Number of individual | | Relative Abundance (%) | |
|-------------------|----------------------|------------|------------------------|------------|
| | S1 | S2 | S1 | S2 |
| Feb | 8 | 20 | 15.7 | 13.9 |
| March | 6 | 18 | 11.8 | 12.5 |
| June | 16 | 49 | 31.4 | 34.0 |
| July | 21 | 57 | 41.2 | 39.6 |
| Total | 51 | 144 | 100 | 100 |

S1- ObufaEsuk, S2- EsukAtu, February and March represent dry season, June and July represent wet season

4. DISCUSSION

Urban development, industrialization around coastal areas and high levels of anthropogenic contaminants from catchment area affect water quality globally [28] and in-turn influences zooplankton availability and diversity. The Great Kwa River displays a typical tropical zooplankton species composition [4,18, 32]. Overall zooplankton species distribution, diversity and abundance in the study sites are influenced by various environmental and biological

factors such as water colour, surface area, depth and trophic level, predation, breeding and vertical migration[9-11,15,16-18]. The results of this present study revealed that some common types of freshwater zooplankton are *Cladocera*, *Copepoda*, *Protozoa*, *Tintinnida*, *Nemata*,*Rotifera*;which forms the bulk of zooplankton groups in previous studies [2-4,13-17,29-35] and are in agreement with our present findings. The submissions of other researchers suggested that the availability of these zooplankton species could be as a result of genetic adaptation; coupled with other relevant environmental factors [3,4,36].The zooplankton composition(abundance and diversity)in this studyrevealed that the various zooplankton species varied seasonally andspatially. The highest number of zooplankton species were recorded during the wet season (June and July), while the low zooplankton species were recorded in the dry season(February and March)in both study locations.Thisresult is in harmony withother findings in River Ossiamo [10], Opobo river, Rivers State [11], Okhuo River, Edo state [37] and Ehoma lake [38]. The seasonal difference in zooplankton species abundance as observed in this study maybe due to the chemical composition of the water; although not examined in this research. Also, flooding during wet season (June and July) because of high rainfall maycontribute positively by recruiting zooplankton from other water bodies;thereby increasing the zooplankton community during wet season.Mixing of high water masses causes high turbidity, this may affect theavailability and distribution of zooplankton species [33-35].The water level in Great Kwa River can be categorized as low to moderate during the study period. This same level of waterwas documented in Ehomalake, Nigeria [38].The condition may be due to thedischarge and draining of water from the mainland of Parliamentary extension (the central dump site in Calabar metropolis), finally ending-up in Great Kwa River. This conditionmay be related to relatively low diversity of zooplankton species. Reports from different studies supported that low

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zooplankton species availability was affected by discharge of water from mainland, and therefore collaborates with our study [40-45].

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The results of this study showed that *Tintinnids* are the most abundance zooplankton (23.1%), followed by other protozoans (18.5%). *Tintinnids* is a type of protozoa found in marine environments belonging to the order Tintinnida [46]. *Tintinnids* represent a group of ciliates belonging to the subclass *Choreotrichida*. *Tintinnids* are essential link between nano-planktons and macro-planktons in the food webs of the marine environment. It is also known that *Tintinnids* are one of the major components of marine planktonic ciliates and has a cosmopolitan character [46]. In Western Philippines, South China Sea, the taxonomic group named *Tintinnids* was reported; although was relatively rare and contributed to only 1% of the total zooplankton identified [31], but in our study high occurrences of *Tintinnids* (23.1%) was documented. Also in Manila-bay, Philippines the taxonomic groups of *Tintinnids* and *Trichoptera* were documented [30], which is in tandem with our present finding in the Great Kwa River. Therefore, *Tintinnids* distribution in our study area formed an important linkage between nano-planktons and macro-planktons in the food webs of the marine environment in Great Kwa River, Calabar. Rotifers species like *Keratella* species (typical of oligotrophic to mesotrophic in nature) were also observed in the study, collaborating with the documented research conducted in Ikwori Lake, central Cross River State, Nigeria. The occurrences of *Protozoa*, *Cladocera*, *Copepoda* and *Rotifera* were observed, which is similar to different documented results [11,47-48]. Specifically, *Copepod nauplii* in the *Copepoda* taxonomic group was identified in this study, agreeing to previous reports [31] and they are among the free-living filter feeder zooplankton utilized as in bio-monitors for pollutants in the water bodies [39]. The variations in the number of individual

species recorded in these findings may be due to the sampling method and sampling time which are supported by other investigations from different aquatic populations [10,11,18].

The ecological diversity indices in this present study indicated richness and abundance in the distribution of zooplankton (Margalef's index and Shannon-Wiener index) in both stations (ObufaEsuk and EsukAtu), with relatively low evenness or equality in both station; especially in station one from the Pielous evenness index values. The relatively low zooplankton species evenness maybe attributed to high abundance and dominance of some species like *Copepods*, etc [49]. Evenness indices of the zooplankton species in this study was lower than 0.99 – 0.993 values documented by [51] in a Nigerian forest river. The values of the diversity indices in our study showed that zooplankton at station 2 (EsukAtu) were more diverse than station one (ObufaEsuk). The diversity shown by EsukAtu may be due to the presence of phytoplankton at that station; constituting rich food web in the location. This study is similar to other documented researches [32,50] but lower than the value obtained by Ekwu and colleagues [51] in lower Cross River estuaries. Although, the zooplankton community showed diversity; the different species of zooplankton that make up the community were relatively unevenly distributed. This finding is similar with the findings in Opobo River (precisely Opobo town station); that displayed diversity of zooplankton species, but evenness index was 0.825 relatively higher than the values in our study.

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Numerous mechanisms are associated with zooplankton occurrence, distribution and diversity in an aquatic ecosystem. These mechanisms include intense predation [52], anti-predator refuge

effect [53], increased niche availability, food sources [48], and the physical complexity habitat provided by banks of macrophytes; which are positively associated with zooplankton diversity [53]. These mechanisms may have contributed to the distributions and diversity of these zooplankton species identified in this study during the duration of the study.

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5 CONCLUSION

This present study provides updated information on the zooplankton distribution, diversity and seasonal variations of the Great Kwa River. The results revealed 12 major taxonomic groups including *Tintinnida* consisting of 23.1%, followed by other *protozoa* (18.5%), *Cladocera* (15.4%), *Copepoda* (13.3%) and the lowest taxonomic groups were *Diptera*, *Foraminifera*, *Atenatada* and *Trichoptera* having only one representative with 1.85% for each order of total abundance of zooplankton species. The highest number of species was recorded in Esukatu (S2) station with 31 species represented during the research period. In both Stations, *Ascampbelliella acuta* displayed high dominance over other species. The abundance of Zooplankton species in the river varies seasonally, with more species recruited during the wet season. The high dominance of the *Tintinnida* (*Ascampbelliella acuta*) in our current study indicates that the river is a natural important linkage between nano-planktons and macro-planktons in the food webs of the marine environment.

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6. RECOMMENDATIONS

The following recommendations are suggested:

- a. The government of Cross River State should establish a waste treatment facility to control and reduce the level of waste disposed in open surface areas (especially in the

Calabar central dump site located at Parliamentary Extension); which finally ends-up in the aquatic ecosystem causing contaminations.

- b. Proper environmental monitoring should be put in place to check sporadic behavior of direct dumping of untreated waste into water bodies; to reduction aquatic/water pollution in the Great Kwa River.

Limitations of the study

Genetic and molecular characterization of zooplanktons were not carried out in this study.

REFERENCES

1. Frisch D, Cottenie K, Badosa A, Green AJ. Strong spatial influence on colonization rates in a pioneer zooplankton metacommunity. *PLoS One*, 2012; 7(7):e40205.
2. Eskinazi-Sant'Anna E.M, Santos G.de-S, Alves N. J. da-S, Brito L. A.F, Leite M. G. P. (2020) The relative importance of regional and local factors in shaping zooplankton diversity in high-altitude tropical shallow lakes. *Journal of Freshwater Ecology*, 2020; 35(1): 203-221. DOI:10.1080/02705060.2020.1770874.
3. Oku E. E, Andem A. B, Arong, G. A, Odjadjare E. Effect of Water Quality on the Distribution of Aquatic Entomofauna of Great Kwa River, Southern Nigeria. *American Journal of Engineering Research*, 2014; 3(04): 265-270.
4. Eyo, V. O, Andem, A.B, Ekpo, P.B. Ecology and diversity of zooplankton in the Great Kwa River, Cross River State, Nigeria. *International Journal of Science and Research*; 2013; 2310-7064.
5. Alfred J.R.B. Faunal diversity in India: An overview. In: Alfred JRB, Das AK, *et al.* editors. *Faunal diversity in India*: Zoological Society of India Kolkatta, 1998:1–9.
6. Otero J, Álvarez-Salgado X.A, Bode A. Phytoplankton Diversity Effect on Ecosystem Functioning in a Coastal Upwelling System. *Front. Mar. Sci*, 2020; 7: 1-15. doi: 10.3389/fmars.2020.592255
7. Dhargalkar V. K, Verlecar X. N. *Zooplankton Methodology Collection and Identification: A Field Manual*, National Institute of Oceanography, Dona Paula, Goa, 2004; 6:403–404.

8. Fielding, S., Ward, P., Poulton, A. J., Pollard, R. T., Seeyave, S., Read, J. F., Hughes, J. A., Smith, T, Castellani, C. Community Structure and Grazing Impact Of Mesozooplankton During Late Spring/Early Summer 2004/2005 in the vicinity of the Crozet Islands (Southern Ocean). *Deep Sea Res. Part II*, 2007; 54:2106-2125.
9. Striebel, M., Singer, G., Stibor, H., Andersen, T. Trophic overyielding: Phytoplankton diversity promotes zooplankton productivity. *Ecology*, 2012; 93: 2719–2727. doi: 10.1890/12-0003.1
10. Ikhuoriah S. O., Oronsanya G. C, Adebajo I. A. Zooplankton Community of the River Ossiomo, Ologbo, Niger Delta. *Animal Research International*, 2015; 12 (3):2249-2259.
11. Shayebi E. M, Patricia U. A, Moslen M. Abundance and Diversity of Zooplankton in the Lower Reach of the Opobo River, Rivers State Nigeria. *African Journal of Environment and Natural Science*, 2020; 3(2): 49-59.
12. Filstrup, C. T., Hillebrand, H., Heathcote, A. J., Harpole, W. S., and Downing J. A. Cyanobacteria dominance influences resource use efficiency and community turnover in phytoplankton and zooplankton communities. *Ecol. Lett*, 2014; 17: 464–474. doi: 10.1111/ele.12246
13. Jafari, N., Nabavi, S. M, Akhavan, M. Ecological Investigation of Zooplankton Abundance in the River Haraz, Northeast Iran: Impact of Environmental Variables. *Arch. Biol. Sci. Belgrade*, 2011; 63(3): 785-798.
14. Emmanuel U, Jude O, George I. U, Raymond A, Cletus I, Isaac, O. A. The Taxa Structure and Composition of Zooplankton Communities of Bonny Estuary: A Bio-indication of Anthropogenic Activities. *The Pacific Journal of Science and Technology*, 2013; 14(2).
15. Kennie A. M, Akinade G.T, Ogialekhe P, Mohammed N. Zooplanktons Assemblage along Jebba Upper Basin, Nigeria. *International Journal of Pure and Applied Zoology*, 2017 5(3): 100-103.
16. Sharma R. C. Habitat ecology and diversity of freshwater zooplankton of Uttarakhand Himalaya, India. *Biodiversity Int Journal*, 2020; 4(5):188–196. DOI: 10.15406/bij.2020.04.00184.
17. Okorafor K. A, Andem A. B, Okete J. A, Ettah S. E. The Composition, Distribution and Abundance of Macroinvertebrates in the Shores of the Great Kwa River, Cross River State, South east, Nigeria. *European Journal of Zoological Research*, 2012, 1 (2):31-36.

18. Offem B. O, Ayotunde E. O, Ikpi G. U, Ochang S. N, Ada F. B. Influence of Seasons on Water Quality, Abundance of Fish and Plankton Species of Ikwori Lake, South-Eastern Nigeria. *Fisheries and Aquaculture Journal*, 2011; 13:1-18.
19. Newell G. E, Newell, R.C. *Marine plankton: a practical guide* (5th edition). Hutchinson and Co. Publishers limited, 1977.
20. Ward H.B, Whipple G.C. *Freshwater Biology*. Edited by I.V. T. Edmondson (2nd Edition). John Wiley and Sons Inc, 1959.
21. Alfred R. B, Bricice S, Isaac M. L, Michael R. G, Rajendran M, Royan J. P, Sumitra V, Wycliffe J. A guide to the study of freshwater organisms. *Journal of Madras University, Supplementary 1*; 1973: 103-151.
22. Adoni D. G, Joshi K, Gosh S. K, Chourasia A. K, Vaishya M. Y, Verma H. G. A work book on limnology, (Pratibha Publisher) Sagar, 1985.
23. Newell G. E, Newell R. C. *Marine Plankton: A practical guide*. Revised Edition. Hutchinson, London, 1966: 225.
24. Imoobe T. O. T. Crustacean of Jamieson River, Nigeria. Phd thesis, University of Benin, Benin city, Nigeria, 1997: 155.
25. Korinek 1997. A Guide to Limnetic Species of *Cladocera* of African Inland Waters (*Crustacea, Branchiopoda*). Occasional Publication No 1. The International Association of Theoretical and Applied Limnology; BTL, Geneva, 1999.
26. Smirnov N. N. Fauna of the USSR, *Crustacea, Chydoridae*. Academy of Sciences of the USSR (English translation). Israel Program of Scientific Translation, Jerusalem, 1974; 1(2):644.
27. Ogbeigbu E. *Biostatistics: A practical approach to Research and data handling*. Minex publishing Company Limited, Benin City Nigeria, 2005: 153-155.
28. Islam M S, Tanaka M. Impacts of pollution on coastal and marine ecosystems including coastal and marine fisheries and approach for management: a review and synthesis. *Mar. Pollut. Bull*, 2004;48: 624-649.

29. Negi R. K. and Mamgain S. Zooplankton Diversity of Tons River of Utarakhand State India. *International Journal of Zoology and Research*, 2013; 3(2): 1-8.
30. Jose EC, Furio EF, Borja VM, Gatdula NC, Santos MD. Zooplankton Composition and Abundance and its Relationship with Physicochemical Parameters in Manila Bay. *Journal of Oceanography and Marine Res*, 2015; 3: 136. doi:[10.4172/2332-2632.1000136](https://doi.org/10.4172/2332-2632.1000136)
31. Relox J. R, Furio Jr-E. F, Borja V. M. Abundance and Distribution of Zooplankton in the South China Sea, Area III: Western Philippines. *Proceedings of the SEAFDEC Seminar on Fishery Resources in the South China Sea, Area III: Western Philippines*, 164-176.
32. Eyo, V. O, Ekpo P. B, Andem A. B, Okorafor, K. A. Ecology and Diversity of Phytoplankton in the GreatKwa River, Cross River State, Nigeria. *International Journal of Fisheries and Aquatic Studies*, 2013; 1(2):1-7.
33. Izadi A, Dobaradaran S, Nabipour I, Karbasdehi VN, Abedi E, Darabi H, Ansarizadeh M, Ramavandi B. Data on diversity and abundance of zooplanktons along the northern part of the Persian Gulf, Iran. *Data in Brief*, 2018; 19: 1418-1422. <https://doi.org/10.1016/j.dib.2018.06.012>. <https://doi.org/10.1016/j.ejar.2015.03.002>.
34. Koszarowska M.M, Gowacka L.D, Weydmann A. Influence of environmental factors on the population dynamics of key zooplankton species in the Gulf of Gdańsk (southern Baltic Sea). *Oceanologia*, 2019 61(1): 17-25. <https://doi.org/10.1016/j.oceano.2018.06.001>.
35. Berraho A, Abdelouahab H, Baibai T, Charib S, Larissi J, Agouzouk A, Makaoui A. Short-term variation of zooplankton community in Cintra Bay (Northwest Africa). *Oceanologia*, 2019: 207. <https://doi.org/10.1016/j.oceano.2019.02.001>.
36. Kremp A K, Reusch T. B. H, Wood A. M. Genetic diversity and evolution in eukaryotic phytoplankton: revelations from population genetic studies *Journal of Plankton Research*, 2017; 39(2): 165-179. <https://doi.org/10.1093/plankt/fbw098>
37. Imoobe, T.O.T. Diversity and Seasonal Variation of Zooplankton in Okhuo River, a Tropical Forest River in Edo State, Nigeria. 2011.
38. Okogwu, O. I. Seasonal variations of species composition and abundance of Zooplankton in Ehoma Lake, a floodplain Lake in Nigeria. *Revista de Biologia Tropical*, 2010; 58(1): 171-182.
39. Banerjee A, Chakrabarty M, Rakshit N, Bhowmick AR, Ray S. Environmental factors as indicators of dissolved oxygen concentration and zooplankton abundance: Deep learning

versus traditional regression approach. *Ecological Indicators*, 2019; 100: 99-117. <https://doi.org/10.1016/j.ecolind.2018.09.051>.

40. Afonina EY, Tashlykova N. A. Plankton community and the relationship with the environment in saline lakes of Onon-Torey plain, Northeastern Mongolia. *Saudi Journal of Biological Sciences*, 2018; 25(2): 399-408. <https://doi.org/10.1016/j.sjbs.2017.01.003>.
41. El-Naggar HA, Khalaf Allah HMM, Masood MF, Shaban WM, Bashar M.A.E. Food and feeding habits of some Nile River fish and their relationship to the availability of natural food resources. *The Egyptian Journal of Aquatic Research*, 2019; 45(3): 273-280. <https://doi.org/10.1016/j.ejar.2019.08.004>.
42. Rozirwan R, Iskandar I, Hendri M, Apri R, Azhar N, Mardiansyah W. Distribution of phytoplankton diversity and abundance in Maspari island waters, South Sumatera, Indonesia. *Journal of Physics: Conf. Series*, 2019; 1282. <https://doi.org/10.1088/1742-6596/1282/1/0121052>
43. Zaghoul FAR, Khairy HM, Hussein NR. Assessment of phytoplankton community structure and water quality in the Eastern Harbor of Alexandria, Egypt. *The Egyptian Journal of Aquatic Research*, 2020. <https://doi.org/10.1016/j.ejar.2019.11.008>.
44. Shalloof KAS, El-Far AM, Aly W. Feeding habits and trophic levels of cichlid species in tropical reservoir, Lake Nasser, Egypt. *The Egyptian Journal of Aquatic Research*, 2020. <https://doi.org/10.1016/j.ejar.2020.04.001>.
45. Wu P, Kainz MJ, Bravo AG, Åkerblom S, Sonesten L, Bishop K. The importance of bioconcentration into the pelagic food web base for methylmercury biomagnification: A meta-analysis. *Science of the total Environment*, 2019; 646: 357-367. <https://doi.org/10.1016/j.scitotenv.2018.07.328>
46. Dolan JR, Ritchie ME, Ras J. The “neutral” community structure of planktonic herbivores, tintinnid ciliates of the microzooplankton, across the SE Tropical Pacific Ocean. *Biogeosciences Discuss*, 2007; 4: 561–593. <https://doi.org/10.5194/bg-4-297-2007>
47. Yakubu F, Sikoki F. D, Abowei J. F. N, Hart S. A. A. Comparative study of phytoplankton communities of some rivers creeks and borrow pits in the Niger Delta Area. *Journal of Applied Science, Environment and Management*, 2000; 4(2): 41-46.
48. Waterkeyn A, Grillas P, Vanschoenwinkel B, Brendonck L. Invertebrate community patterns inMediterranean temporary wetlands along hydroperiod and salinity gradients. *Freshwater Biol*, 2008; 53(9): 1808–1822.

49. Kadiri, O. M. Phytoplankton survey in the Western Niger Delta, Nigeria. *African Journal of Environmental Pollution and Health*, 2006;5(1): 48-58.
50. Ekwu O, Sikoki F. D. Species composition and distribution of zooplankton in the Lower Cross River Estuary. *African Journal of Applied Zoology and Environmental Biology*, 2005; 7: 5-10.
51. Imoobe T. O. T, Adeyinka M. L. Zooplankton-based assessment of the trophic state of a tropical forest river. *International Journal of Fisheries and Aquaculture*, 2010; 2(2): 064-070.
52. Rennie MD, Jackson L.J. The influence of habitat complexity on littoral invertebrate distributions: patterns differ in shallow prairie lakes with and without fish. *Can J Fish AquatSci*, 2005; 62(9):2088–2099.
53. Meerhoff M, Iglesias C, DE Mello FT, Clemente JM, Jensen E, Lauridsen TL, Jeppesen E. Effects of habitat complexity on community structure and predator avoidance behavior of littoral zooplankton in temperate versus subtropical shallow lakes. *Freshwater Biol*, 2007; 52(6):1009–1021.
54. Bolduc P, Bertolo A, Pinel-Alloul B. Does submerged aquatic vegetation shape zooplankton community structure and functional diversity? A test with a shallow fluvial lake system. *Hydrobiologia*, 2016; 778(1): 151–165.