

RESPONSE OF PEARL MILLET (*Pennisetum typhoides* (Burm f.) TO DIFFERENT FERTILIZER APPLICATIONS UNDER FIELD CONDITIONS

ABSTRACT

Aims: To study and compare the response of Pearl millet to organic manure, NPK fertilizer, and arbuscular mycorrhizal fungi as biofertilizer.

Study design: The study employed a Complete Randomized Block Design (CRBD) with a split-split arrangement and the treatments were replicated four times each.

Place and Duration of Study: The experiment was conducted under field conditions during the rainy season of 2020 at the Agriculture Research and Demonstration Farm Land, University of Maiduguri, Nigeria.

Methodology: Pearl millet seeds were planted under three (inorganic, organic and biofertilizer) fertilizer applications and monitored for growth over a period of 60 days. Plant growth parameters were measured appropriately and statistically analyzed.

Results: Results indicated that NPK significantly ($P < 0.05$) improved the growth of millet, with mean shoot heights of 142.45 ± 10.0 cm and mean shoot diameter of $5.696-70$ cm. Leaf area was negatively affected by fertilizer application with the smallest (885.53 ± 0.49) cm^2 and largest (1070.91 ± 6.0 cm^2) areas observed in plants of the control and biofertilizer treated soil respectively. The mean root lengths ranged from 34.75 cm to 58.50 cm and dry shoot biomass ranged from 2.16 g to 15.45 g with more biomass in plants treated with NPK. Plant root colonization was however greater (40%) in plants treated with biofertilizer than in all the remaining treatments (0.71%). The shoot lengths, stem diameters, leaves leaf growth, and root biomass was also enhanced by biofertilizer application when compared to those treated with organic manure and the controls

Conclusion: A positive response was observed using arbuscular mycorrhizal fungi (AMF) as biofertilizer and its application could serve better in millet cultivation where inorganic and organic fertilizers are undesirable or inaccessible.

Keywords: Arbuscular, Biofertilizer, *Faidherbia albida*, Fungi, Millet, Mycorrhizal

1. INTRODUCTION

Pearl millet is among the leading crops produced in different parts of the world. It has been estimated that over one third of the world's Pearl millet production is in Africa [1]. In West Africa, the arable land put to Pearl millet production has been estimated to be 23.99 million hectares; in which about 5 million hectares occurs in Nigeria [2]. Millet in Nigeria is predominantly produced in Bauchi, Borno, Gombe, Kano, Katsina, Sokoto and Yobe States. Pearl millet is important in Nigeria as staple crop used in preparation of different kind of local and processed foods, feeds, pharmaceutical products and other essential products [3, 4]. It has also been fermented, processed, distilled and used in the manufacture of industrial alcohol [5].

Millet production in Nigeria is significantly affected by a myriad challenges. Millet is attacked by various diseases and pests ~~on-in the~~ field [6]. The most important of the fungal diseases of Pearl millet is downy mildew, caused by the fungus *Sclesopora, graminicola* [7]. Other diseases and pests of Pearl millet in Nigeria are smut and Ergot caused by *Moesziomyce —pennicullariae* and *Claviceps fusiformis* respectively [8, 9]. Some important invertebrate pest includes stem borer (*Coniesta ignefusalis*), head miner (*Raghuva* sp.), the head beetle (*Pocnoda* sp.) and grasshoppers, while *Quelea quelea* is the major vertebrate pest [10]. Fragility of the soils arises from various factors such as desertification, drought, security challenges, bush burning, overgrazing and limited supply of plant nutrients, thus limiting the

productivity of the soil which virtually limit agricultural and agro-forestry developments. Other predominant factors include drought, poor soil with low water holding capacity [9, 11] cultural practices such as low crop density [12], cost of fertilizers, pest invasion and disease infestations [7].

Efforts to address the current challenges of millet production have been targeted towards improving soil fertility and controlling pests and diseases. Soils in semi-arid zones have moderate to poor fertility due to low content of nitrogen (N), available phosphorus (P) and sometimes potassium (K) [10]. This has made soil amendments with fertilizers necessary to enable adequate food and feed supply. However, most soils in dry land areas are very low in plant available water which makes artificial fertilizers rarely used by farmers [13]. The use of organic manure such as cow dung on farmlands is also essential for enhancing soil fertility and protecting agricultural soils from wind and water erosion, leaching and volatilization [14]. The effect of organic manure on soil is also noticed through improvement of soil physicochemical and biological properties [15]. In the recent past, the availability of organic manure remained a major challenge to the farmers especially that the rate of livestock production has significantly decreased. As a result, the search for a better alternative have has remained imperative.

The use of fungi in agriculture as an alternative biofertilizer play a significant role of providing an economically viable level for achieving ultimate goal to enhance plant growth and developments. Arbuscular mycorrhizal fungi (AMF) are important components of agricultural ecosystems, and can directly influence the productivity of crops. Mycorrhiza is the symbiotic associations between fungi and the living root cells of plants. In this association, the fungus obtains its carbohydrate requirements from the plants, and in return it absorbs mineral elements particularly, phosphorus, zinc, copper [16,] and potassium [16,17] and potassium [17] and transport them to the plants through its ramifying hyphae. The hyphae of AMF extend beyond the root hair uptake zone into un-depleted soil zones and this increases water and nutrients movement to the plant roots [18]. Mycorrhizal fungi as a biofertilizer, form a bridge between the roots and the soil and also gather nutrients from the soil and give it to the roots [19]. In our previous study Grema *et al.* [20], the role of AMF in enhancing millet growth have been established under greenhouse conditions. However, crop growth and productivity in the arid zone depends on photo-respiration length and level of the plant that is affected by moisture [21]. It is against this backdrop, the present study investigated the effect of AMF on millet growth under field conditions as compared to organic and inorganic fertilizer application.

2. MATERIALS AND METHODS

2.1 STUDY AREA

The experiment was conducted during the rainy season of 2020 at the Agriculture Research and Demonstration Farm Land in the Faculty of Agriculture, University of Maiduguri, Maiduguri, Borno State, North Eastern Nigeria (Latitude: 11.48° 16.29' North, Longitude: 13° 11' 55.11", East and on an altitude 300 meters above ~~the~~ sea level and covering an area of 54.2 m²).

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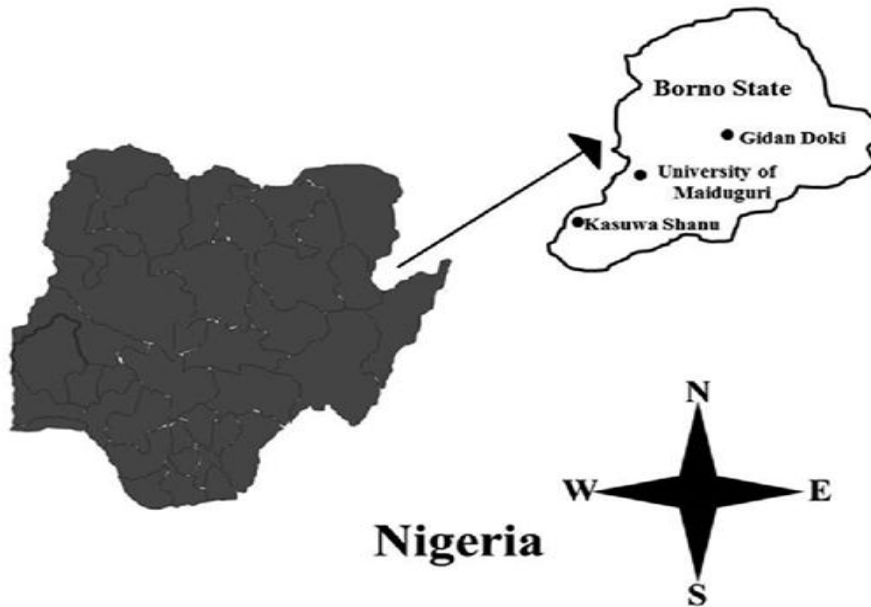


Figure 1: Map of Nigeria Showing the Study Area Borno State

2.2 Sample collection

Pearl millet and fertilizers were obtained from the Lake Chad Research Institute Maiduguri, and the seeds were tested for viability by simple floatation method. One hundred percent (100%) viable seeds tested was surface sterilized with 5% Sodium hypochloride to eliminate surface contaminants and rinsed 3 to 4 times with distilled water [22]. While decomposed cow dung (organic manure) procured from University of Maiduguri Livestock Farm, Dalori, Maiduguri was used for the experiment. Biofertilizers in the form of AMF spores consisting of *Glomus mosseae* and *Glomus intraradices* was extracted as described by Grema *et al.* [20].

2.3 Experimental Design and Treatments

Randomized Complete Block Design (RCBD) was used to assess the effects of Arbuscular mycorrhizal fungi (AMF), organic manure (cow dung) and inorganic fertilizer (NPK) on the growth and root colonization of Pearl millet crop. The gross land area was disc-harrowed, leveled and marked out into plots. The apportioned area was divided into four groups each plot measured 19 m x 4.5 m, with spacing of 1.0 m between groups. Each of the plots was divided into four replicates in which the four treatments (AMF, cow dung, NPK and control) were apportioned. Each plot measuring 4 m x 4.5 m was fertilized with either AVM, cow dung or NPK. A set of control plots remained without applying any treatment (zero) with spacing of 1.0 m between them [23].

2.3.1 Fertilizer application

To one group of the plots, 100 g (containing about 1000 spores) of soil was added on each replicate as biofertilizer-inoculum. Similarly, 67.5 g /18 m² cow dung (equivalent to 7.5 tonnes / ha⁻¹, field rate for Pearl millet) was added to the second group of the replicate plots [24]. For the inorganic fertilizer, 0.117 Kg N, 0.117 Kg P₂O₅, 0.117 Kg K₂O/18 m² combined with 46 kg/18 m² of urea to give one treatment

combinations plus 60 Kg ha^{-1} N, 30 Kg ha^{-1} P $_2$ O $_5$, 30 Kg ha^{-1} K $_2$ O which is the recommended rate for Pearl millet crop was added to the replicate plots. Soil without AMF spores, cow dung and NPK was considered as control treatment [22].

2.3.2 Sowing and Experimental Management Practices

The seed of Pearl millet was surface sterilized with 5 % Sodium hypo-chloride and about 10 seeds of millet was sown in the center of each of the respective field plots. The millet seedling was thinned to one per plot under rain fed to encourage root system formation [24].

2.4 Data Collection

2.4.1 Percentage of Root Colonization by AMF

This was carried out by collecting, staining and microscopic observations as described by Grema *et al.* [20]. The percentage (%) Arbuscular mycorrhizal fungi root colonization was calculated using the formula (1) below [25]:

$$\% \text{ AMF colonization} = \frac{\text{Number of inter-section with AMF structures}}{\text{Total Number of intersections viewed}} \times 100 \quad (1)$$

2.4.2 Shoot Heights

Shoot heights ~~was-were~~ measured in centimeters from cotyledonary scar to shoot tips of all the crops using a Laboratory wooden meter ruler from seedling plants in each replicate for the cereal (millet) and the mean was computed [26].

2.4.3 Stem Diameter

Stem girths of the plants ~~was-were~~ measured using a macro Vernier caliper for the field experiment and was expressed in cm [26].

2.4.4 Number of Leaves

Number of leaves on each plant of Pearl millet ~~leaf~~ was counted on the seedling plants per replicate for each plant per field trial and the means was computed using [26].

2.4.5 Leaf Area

The leaf area of the field crops was measured by using laboratory wooden meter ruler. The diameter of the leaf and the length of the leaf was taken and then ~~multiply-multiplied~~ by the leaf weight of the seedling plants under field study and expressed in cm 2 [12].

2.4.6 Root Length

A root length of the plants was also measured using meter ruler after harvest. The length was determined by placing the ruler at the cotyledonary scar (base) of the shoot to the tip of the longest root [27].

2.4.7 Shoot and Roots Dry Biomass

The fresh weight of the shoot was taken as well as the shoots and roots dry biomass of each field trial millet plants harvested, oven dried at 70 $^{\circ}$ C for 24 hours to constant weight and their dry weights was determined using Digital electric balance with capacity of 5000 g and expressed in grams [15].

2.5 Statistical Analysis

The data collected was subjected to two-way Analysis of Variance (ANOVA) based on Randomized Complete Block Design (RCBD) with Split - Split arrangement using the Analytical Statistical Software, Statistix Version 8.0 (SX). Differences between means ~~was-were~~ determined using the Duncan's Multiple Range Test (DMRT) at 5% probability level.

3. RESULTS

The response of millet to ~~the~~ application of organic, inorganic and biofertilizer was investigated. Results in Table 1 showed that the shoot height of Pearl millet was significantly taller ($p < 0.05$) in the treatments that received NPK fertilizer. It was followed by millet in the control (138.78 cm) and biofertilizer (120.75 cm) treated millet. Shoots of the plants that received AMF as fertilizer were significantly longer ($p < 0.05$) than the plants treated with organic manure. The results of millet stem diameter showed that plants treated with biofertilizer, NPK and the control had significantly greater ($p < 0.05$) stem diameter than plant treated with organic manure (cow dung). However, statistical analysis showed no significant difference between the biofertilizer, NPK fertilized plant and that of the control plants respectively (Table 1). The millet leaves observed in this study ranged between 50 - 73 leaves, where plant treated with NPK fertilizer exceeded (73.04) the other treatments and the least (50.83) number of leaves occurred in millet grown without fertilizer. Analysis of variance indicated that millet that received NPK has significantly ($P_{<0.05}$) more leaves than organic manure and also those of the plant inoculated with AMF was significantly more ($P_{<0.05}$) than ~~the~~ control plants.

The result showed that mean root length of the field planted millet under the four treatments ranged from 34.75 - 58.50 cm, where NPK fertilizer treated plants had the longest (58.50 cm) root length and the shorter ~~one~~ was observed in the control plants (Table 1). The outcome of the study also shows that plants treated with organic manure had significantly ($P_{<0.05}$) taller roots than that of the plant treated with biofertilizer ~~whereas,whereas~~; millet treated with biofertilizer had significantly longer roots than that of the control.

Results in Table 1 showed that fertilizer application reduced the ~~leaves-leaf~~ area of millet crop with ~~the~~ lowest area observed in the plants treated with biofertilizer (885.53 cm²). ~~The Highest-highest leaves-leaf~~ area (1070.90 ± 0.68 cm²) were observed in the control plants which were significantly ($P_{<0.05}$) larger than plants in the remaining treatments.

For the plants dry matter, the results showed that plants treated with NPK fertilizer had the highest (15.45 g) shoot dry weight and the lowest (2.16 g) was observed in the plants inoculated with biofertilizer which were significantly lighter than those in the control. Similarly, the plant root dry weight followed the same ~~trend,trend~~; except that biofertilizer treated millet was significantly heavier than that of the control plants. The mean values ranged between 8.64 g - 1.96 g, in which plants treated with inorganic fertilizer had the greater (8.64 g) dry root biomass. The percentage (%) root colonization observed in Pearl millet showed that biofertilizer had the highest percentage (40%) of root colonization and was significantly ($P_{<0.05}$) different from all other treatments.

Table 1: The Pearl millet growth parameters under field conditions

Treatment	Mean* shoot height(cm)	Mean* stem diameter (cm)	Mean* number of leaves	Mean* root length (cm)	Mean* root area(cm ²)	leaf biomass (g)	Mean* dry biomass (g) Shoot	Mean* dry biomass (g) root
AMF	120.75 ± 0.25	5.17 ± 0.50	56.41 ± 0.67	35.00 ± 0.50	885.53 ± 0.49	2.16 ± 0.25	2.10 ± 0.50	
OM	107.67 ± 0.50	4.98 ± 0.75	64.95 ± 0.83	39.25 ± 0.66	945.39 ± 6.0	5.84 ± 0.46	3.62 ± 0.23	
NPK	142.45 ± 10.0	5.69 ± 0.58	73.04 ± 1.67	58.50 ± 2.20	951.53 ± 10.17	15.45 ± 1.20	8.64 ± 1.40	
Control	138.78 ± 3.33	5.58 ± 0.75	50.83 ± 0.33	34.75 ± 2.40	1070.90 ± 6.80	4.00 ± 0.12	1.96 ± 0.20	
Mean*	127.41	5.36	61.31	41.88	963.34	6.86	4.08	
SE ±	9.144480	0.930	9.2369924	3.93	101.35447	1.58	0.89	
LSD _{0.05}	0.689	0.502	0.153	0.086	0.098	0.002	0.007	

*Mean obtained from 4 replications

Table 2: Rate of millet crop root colonization

Treatment	AMF	NPK	Cowdung	Control	Mean	SE±	LSD _{0.05}
Colonization (%)	40 ± 0.50	0.71 ± 0.02	0.71 ± 0.10	0.71 ± 0.2	10.53	19.64	41.84

*Mean obtained from 4 replications

4. DISCUSSION

Arbuscular mycorrhizal fungi could be exploited to source for nutrients to enhance crop performance as biofertilizer. Thus, in the present study, the inorganic fertilizer (NPK) was found to enhance millet shoot length, stem diameter, root length, leaf production, shoot and root biomass which the results indicated that treatment in this group had significantly ($P < 0.05$) improved growth. However, the root colonization was very much (40%) enhanced in plants treated with biofertilizer than the remaining treatments. A positive response from such investigation has demonstrated a higher percentage of root colonization in plants treated with arbuscular mycorrhizal fungi spores than other treatments which were negative in the other experimental sites.

-The lower response of millet to biofertilizer in the field experiment could be due to the routine tillage of the field soils, which could have resulted in poor root system development compared to the plants grown under the influence of inorganic fertilizer. The higher rate of growth observed in the latter could be attributed to the higher N and P contents in the field as reported by Kwari *et al.*, [28], which resulted in similar crop growth. Adequate supply of N and P induce faster root development which would lead to better nutrients absorption and improved plant growth. According to Shinde [29], plant growth may be attributed to the production of more roots which increases absorption of water and minerals (N, P and K⁺). Thus, the increases may be attributed to the improved mineral nutrition of the inoculated plants which evidently resulted in increased physiological process such as photosynthesis and photorespiration [30].

Organic fertilizer (cow dung) was less effective in field conditions as observed in this study compared to inorganic fertilizer. This agrees with the observations of ~~the~~ [31] who showed that organic manure application to soils have longer residual effect in terms of nutrient availability on soil fertility. Thus, the slow release of nutrients by organic manure as a fertilizer affects their capacity to elicit significant growth and yield responses.

The result of the present study revealed that lower and shorter roots were produced by crops treated with biofertilizer than non-biofertilized ones except the controls. Previously, arbuscular mycorrhizal fungi has been reported to have negative effect on root length and root biomass of plants [15]. According to [27], most of the carbohydrate (about 1 - 17 %)

released by the plants for the purpose of root formation (root lengths and roots biomass), is taken up by the mycorrhizal fungi in the associations. As a result, root development is grossly jeopardized. In addition, mycorrhizal fungi may take up the function of root hairs as absorbing-agents which could further decrease the rate of root elongation. As such, it could be expected that mycorrhizal plants should contain less and shorter roots length than their non-mycorrhizal counterparts. Bonnie *et al* [32] have indicated that the production of root extensions, branching and root hairs are strongly inhibited in the presence of arbuscular mycorrhizal fungi mycelia.

There was a higher percentage (40%) of Pearl millet roots colonization in plants treated with biofertilizer than in all other treatments (0.71% each). This could be attributed to environmental factors and soil nutrients, such as the high quantity of phosphorus, since some researchers have reported the negative effect of high P levels on arbuscular mycorrhizal fungi (AMF) colonization [33]. The variation in the level of root colonization of crops in the field conditions could also be due to uneven distributions of arbuscular mycorrhizal fungi (AMF) spores in the soil or due to the heavy rain. Comparatively, the millet plants treated with biofertilizer had longer shoots and stem diameters compared to those treated with organic manure, which contrastingly had better leaves-leaf properties and dry biomass. Similarly, biofertilizer application promoted the plants' shoot development, leaves area and shoots dry biomass as compared to the plants used as control. This indicated that, biofertilizer application could enhance food and feed production.

5. CONCLUSION

Fertilizer application have-has become a cornerstone for sustainable crop production. Due to the obvious challenges associated with inorganic fertilizers, alternative sources are-have been sought for. It has been established in this study that inorganic fertilizer promote millet growth better than other sources of fertilizer used in the study. Biofertilizer application enhanced shoots lengths and stem diameters when compared to those treated with organic manure. In addition to shoot diameter, biofertilizer application supported better leaves-leaf growth and root biomass. The present study has therefore highlighted the fact that Arbuscular mycorrhizal fungi (AMF) could serve better in millet cultivation than cow dung under field conditions and thus, its suitability for use as alternative for inorganic fertilizer for improved millet production.

REFERENCES

1. Food and Agriculture Organization (2010). Global Forest Resources assessment 2010-main report. FAO forestry paper no 163 Rome. Available at www.fao.org/docrep/013/i1757e/i1757e00.htm. Retrieved date: 6/7/2018.
2. Food and Agriculture OrganisationOrganization. (2011). The State of Food and Agriculture 2010/2011: Women in Agriculture – closing the gender gap for development. Rome. Available at www.fao.org/docrep/013/i2050e.pdf. Retrieved date: 6/7/2018.
3. Nkama I, Abbo ES, Igene JO (1994). Traditional Production and Chemical composition of "ndaleyi," a Nigeria millet food. Plant Foods for Human Nutrition 46: 109-116.
4. Anon S (1999). Borno State Agricultural Development Programme Diary, BOSADP, 19 pp.

5. Hamilton R. (2009). Agriculture sustainable future: Breeding better crops. *Science of American* 19: 16-17.
6. Ascon-Aguilar C, Barea JM (1996). Arbuscular Mycorrhizas and Biological control of soil bore plant pathogens-an overview of the mechanisms involved. *Mycorrhiza* 6:457-464.
7. Anon M (2007). Johannas [announcesannounces](http://www.ssda.gov/portal/ut/p/-s.7-010B/sd.sr/sa) nitrogen fertilizer tool as part of energy strategy. Retrieved from [http:// www.ssda.gov/portal/ut/p/-s.7-010B/sd.sr/sa](http://www.ssda.gov/portal/ut/p/-s.7-010B/sd.sr/sa), (Accessed on September, 14-2007.Retrieved date: 6/7/2018.
8. Abebe F, Tefera T, Mugo S, Beyene Y, Vidal S (2009). Resistance of maize varieties to the maize weevil *Sitophilus zeamais* (Motsch) (Coleoptera: Curculionidae). *African Journal of Biotechnology* 8: 5937 – 5943.
9. Sharma YP, Watpade S, Thakur JS (2014). Role of mycorrhizae: A component of integrated disease management strategies. *Journal of Mycology of Plant Pathology*. 44 (1): 12-20.
10. Food and Agriculture Organization. (2017). Legume web. Retried 20th January, 2017.
11. Udaiyan K, Muthukumar T (2006). Growth improvement of *Acacias* by indigenous arbuscular mycorrhizal fungi rhizobium and phosphate solubilizing bacteria in [non-sterile](#) field soil. In: *Mycorrhiza*: Ed. Anil prakash, V.S. Mehrotra. Scientific Publishers, Jodhpur. 67-74.
12. Ajayi O, Owonubi JJ, Uyovbisere EO, Zarafi AB (1998). Climatic, edaphic and biological factors limiting pearl millet yield in Nigeria. Pp 9-36. In: *Pearl millet in Nigerian Agriculture. Production, Utilization and Research Priorities. Proceedings of the pre-season national co-ordination and planning meeting of the Nationally Coordinated Research Programme on Pearl millet, Maiduguri, 21-24 April, 1998.*
13. Helen T, Ardo J (2009). Relating soil properties to biomass consumption and land management in semiarid Sudan. A minor Field Study in North Kordofan, M. Sc Thesis, Lund University, Geobiosphere Science Centre Physical Geography and Ecosystems Analysis, p 45.
14. Ewulo BS, Hassan KO, Ojeniyi SO (2007). Comparative effect of cow dung manure on soil and leaf nutrient and yield of pepper. *International Journal of Agricultural Research* (2):1043-1048.
15. Akande MO, Makinde EA, Otuwe MO (2011). Dry matter partitioning of Sesame and nutrient dynamics with Organic and inorganic fertilizers. *Tropical and subtropical agroecosystems*, 14:1063-1069.
16. Babajide PA, Fagbola O (2014). Growth yield and nutrient uptake of Sesame (*Sesamum indicum* Linn.) as influenced by biofertilizer inoculants. *International Journal of Current Microbiology and Applied Sciences* 3 (8): 859-879.
17. Bhale UN (2013). Occurrence of vesicular arbuscular mycorrhizas (VAM) in medicinal plants of Marathwada region of Maharashtra, India. *Journal of Chemical and Biophysical. Sciences* 3(3):1912-1919.
18. Dalpe Y, Seguire SY (2013). Microwave-assisted technology for the clearing and staining of Arbuscular mycorrhizal fungi in roots. *Mycorrhiza*, 23(4):333-340.
19. Peters S (2002). *Mycorrhiza 101*. Reforestation Technologies International, Salinas, CA. Reforestation Technologies International 2003 Mycopak Product Information, pp. 1-7.
20. Grema MN, Ismail HY, Muhammad S (2022). Effect of arbuscular mycorrhizal fungi, organic and inorganic fertilizers on the growth parameters and root colonization of pennisetum typhoides (burm f.) under greenhouse conditions. *Arid Zone Journal of Basic and Applied Science Research*, In press.
21. Havargi RS (2007). Mitigation of drought stress through plant growth regulators and vesicular arbuscular mycorrhizae (VAM) in cotton. M.Sc. Thesis: University of Agricultural Sciences, Dharwad, Biological Sciences Department, Pp 1-20.

22. Ndirmbula GM (1998). Growth performance of pearl millet, maize, cow pea and groundnut under the influences of arbuscular mycorrhizal fungi and inorganic fertilizers. Dissertation, University of Maiduguri, Department of Biological Sciences.
23. Ajayi CA, Awodun MA, Ojeniyi SO (2007). Comparative effect of Cocoa husk ash and NPK fertilizer on the soil and root nutrient contents and growth of kola seedlings. *International Journal of Soil Science*. 2 (2): 148-153.
24. Odo PE, Gwary DM (1994). Land use and cropping within Jere Fadama in the Nigerian Sudan Savanna In: John SE, Ed. Strategies for the sustainable use of Fadama lands in Northern Nigeria. Ibadan, University Press: 56-70.
25. McGonigle TP, Miller MH (2000). Inconsistent effect of soil disturbance on colonization of roots by arbuscular mycorrhizal fungi .A test of the inoculum density hypothesis. *Applied Soil Ecology* 14:147-153.
26. Statistical Analysis System (SAS) Institute (2008). Carry, NC, USA.
27. Food and Agriculture Organisation. (2016). The state of food and agriculture. Rome. Doi: [10:/007/511027-012-9374-6](https://doi.org/10.1007/511027-012-9374-6). Retrieved 6th July, 2018.
28. Kwari JD, Shukla UC, Nwaka GIC (1995). Integrated soil fertility/plant nutrition studies in Northeast Arid Zone Development Programme (NEAZDP) Area. Pp 120-125. Paper presented at the First International Conference on Research for Development held at the University of Maiduguri, Nigeria, 19th - 25th, June, 1995.
29. Shinde SK (2014). The role of arbuscular mycorrhizal fungi on growth and productivity of onion (*Allium cepa* L.). Ph.D. Thesis, University of Pune, Plant Science Department.
30. Auge RM (2001). The effects of different species of arbuscular mycorrhiza fungus (AMF) on the water relations photosynthesis, and some growth parameters of maize (*Zea mays* L.) during drought and recovery from drought. *Aspects of Applied Biology*, 63: 1-2.
31. Bababe B, Sandabe MK, Ibrahim A (1998). Paper presented at workshop on soil fertility management and production/ utilization of organic manures in Borno State. Maiduguri on 14th April, 1998. 3-13pp.
32. Bonnie LA, Wrigths SF, Watson CA (2001). From city trees. *The Journal of the Society of Municipal Arborist* 37 (6): 1-2.
33. Blanke V, Renke C, Wagner M, Fuller K, Held M, Kuhn AJ, Bruscot F (2005). Nitrogen supply affects arbuscular mycorrhizal colonization of *Artemisia vulgaris* in a phosphate polluted field sites, *New Phytol*. 166: 981- 992.