

## “An Investigation of Insulinotropic Potential of Herbal Plants for Management of Diabetes”

### ABSTRACT

The Insulinotropic hormone GLP-1, which has been proposed as a modern treatment for management of diabetes, is metabolized extremely with the aid of DPP-IV. Inhibitors on DPP IV enhance the level of GLP-1, as have elevated glucose tolerance and improved insulin secretion. Recently, incretin-based treatments have end up a beneficial tool to treat diabetic patients, and distinctive research have focused on the identification of GLP-1 receptor agonists, which includes these of herbal origin.

**Study design:** *In-vitro* model

**Place and Duration of study:** Department of pharmacology, Karnataka College of pharmacy, Bangalore, India, between Jan 2022 to April 2022.

**Methodology:** Hence, after apprehend the medicine undertaking of different medicinal plants extract of management regarding diabetes right here we were the methanolic extracts of *Aegle marmelos*, *Moringa oleifera*, then *Syzygium cumini* have been tested in-vitro for DPP-IV inhibitory activity and have an impact on of incretin system like GLP-1. An in-vitro assay according to measure GLP-1 release from cultured murine EEC's under fatty acid stimulation. DPP-IV is involved in the inactivation over GLP-1, a potent Insulinotropic peptide.

**Results:** The current study underlines up to expectation the extracts inhibits the DPP-IV and enhances the GLP-1 for diabetes. Results established so the extracts on *Aegle marmelos*, *Moringa oleifera*, and *Syzygium cumini* had DPP-IV inhibitory activity on 261.3329 $\mu$ g/mL, 241.5223 $\mu$ g/mL, and then 99.1478 $\mu$ g/mL respectively. Diprotin A confirmed an IC50 virtue on 46.6277 $\mu$ g/mL, as is used as positive controls. Similarly, the study additionally demonstrates that on a cellular level of *Aegle marmelos*, *Syzygium cumini* potentially stimulate GLP-1 secretion, however *Moringa oleifera* indicate decent rises into GLP-1 secretion.

**Conclusion:** The outcomes assure the inhibitory impact about plants regarding DPP-IV, GLP-1 secretion, and the main in conformity with stand a novel, efficient and fair strategy for the management on diabetes.

**KEYWORDS:** Diabetes, Medicinal Plants, Insulinotropic Hormone, GLP-1 Secretion, DPP-IV Inhibition

## 1. INTRODUCTION:

Newer, potent, cost effective molecule is pressing want in diabetic research. More than 300 million humans worldwide are at a risk on thriving diabetes. And disease's pecuniary have an effect on within some nations ought to be higher than so about AIDS epidemic. As by the country wide health policy on Government of India, diabetes, which is associated along cardiovascular diseases, is rising so a serious health challenge in India. It is estimated to that amount in that place may also stay huge upward push in diabetes cases in India increasing from 31 million in 2005 in accordance with 46 million in 2015 and in particular ripe of civic population. Through the ages, plants have always provided tremendous possibilities toward the betterment concerning human health either through ameliorating disorder conditions or improving normal physiological activity. Medicinal plants are extensively used as an alternative cure method in the management and treatment over diabetes seeing that long. It has been estimated as about 30% on diabetics international have adopted the remedy offered via alternative and complementary medicine.

A newborn approach for therapy on diabetes mellitus is primarily based about the intestine hormone glucagon-like peptide-1 (GLP-1), which is ant-diabetic due to its combined action to stimulate insulin secretion, production of beta-cell mass, subdue glucagon secretion, decrease the rate of gastric emptying and induces satiety. The peptide is rapidly inactivated via the enzyme dipeptidyl peptidase-IV (DPP-4), ensuing between a half-life of active GLP-1 of only approximately 1-2 minutes. Inhibition of DPP-IV increases the ranges on endogenous active GLP-1 and prolongs its half-life. GLP-1 secretagogue activity on medicinal plants has less side-effects then paltry cost namely compared to GLP-1 agonists on synthetic origin.<sup>1</sup> A variety of anti-diabetic pills are currently clinically accessible who action by mimicking or improving GLP-1 action yet prohibition of DPP-IV. A new current focus of research is the identification on compounds which incite intimate secretion on GLP-1 and DPP-IV inhibition.

Bael, *Aegle marmelos*, a plant of Indian origin grudging massive medicine potential, it is belong in conformity with family Rutaceae, it is acknowledged through the several mean names between the specific parts over the united states and also backyard of the country.<sup>2</sup> The utility of bael is point out in the Indian historic system of medicine, each portion of the bael plant such as root, bark, leaf, flower, fruits, seed or even its latex are also important between several traditional system of medicine, that's in what it is certain concerning the almost essential plant of the India. The bael fruit is grudging lots of pharmacological activity; fruit on such possesses anti-dyspepsia, anti diarrhoea and anti-dysentery. The crop is additionally used as a dietary supplements, such is additionally chronic in accordance with cure intermittent fever, mental disease, hypoglycemic effect, anti-fungal effect, anti-microbial, analgesic, anti-inflammatory, antipyretic, anti

dyslipidemic activity, Immunomodulatory activity, anti-proliferative activity, wound-healing activity, anti-fertility, insecticidal endeavor and many more.<sup>3,4</sup>

*Moringa oleifera* is certain on the vegetables of the Brassica order and belongs to the family Moringaceae. The Moringaceae is a single genus family including 13 known species (Khawaja et al., 2010). *Moringa oleifera* is a small provincial plant on the sub-Himalayan areas of North West India. Moringa is also broadly acknowledged and used for its health benefits. Among commoners, such has earned its name as like 'the miracle tree' due after its wondrous healing capabilities for a range of illnesses and also some persistent diseases. Several investigations have been conveyed out to isolate bioactive compounds from more than a few components on the sow appropriate to it's a variety of applications (Guevara et al., 1999). The *Moringa's* extremely good medicinal usage which is claimed via dense cultures and communities primarily based about Real-life experiences is nowadays slowly existence proven through science. Through research, the Moringa was observed to incorporate deep indispensable nutrients, for instance, vitamins, minerals, amino acids, beta-carotene, antioxidants, anti-inflammatory nutrients and omega 3 or 6 fatty acids (Fahey, 2005; Hsu et al., 2006; Kasolo et al., 2010).<sup>5</sup>

*Syzygium cumini* (Family Myrtaceae) is additionally recognized so *Syzygium jamunum* and *Eugenia cumini*. Other frequent names are Jambul, Black Plum, Java Plum, Indian Blackberry, Jamblang, Jamun etc. is a vital medicinal sow chronic within a variety of common systems on medicine. It is high quality into the therapy of diabetes mellitus, inflammation, ulcers and diarrhea then preclinical research hold also proven such in imitation of possesses chemopreventive, radioprotective and antineoplastic properties. The plant is prosperous of compounds containing anthocyanins, glucoside, ellagic acid, isoquercetin, kaemferol and myrecetin. The seeds are claimed in conformity with contain alkaloid, jambosine, then glycoside jambolin and antimellin, which halts the diastatic conversion of starch into sugar.<sup>6</sup>

The targets on this study have been in imitation of clarify methanolic extract over plants inhibits DPP-IV to decide inhibitory activities yet to compare it things to do among Diprotin, a reference standard. The current study additionally explains the influences about GLP-1 secretions on STC-1 cell line which is certain of the objectives for diabetes and such helps in accelerated glucose tolerance and including provocation of insulin biosynthesis and secretion.

## **2. MATERIALS AND MEHTODS:**

### **2.1 Collection of Plant Materials:**

The plants were brought from Bangalore, Karnataka, India. The plant specimen has been identified and authenticated by department of botany, University of Rajasthan, Jaipur and specimens were kept for the reference. And reference numbers were *Aegle marmelos* fruits (RUBL 211761), *Moringa oleifera* fruits (RUBL 211760), and *Syzygium cumini* seeds (RUBL 211759).

### **2.2 Extraction of Plants**<sup>7-11</sup>

Preparation of Extract: The Plants were chopped into small pieces and dried under shade at room temperature for seven days. The dried fruits were powdered and passed through the sieve (Coarse 10/40). The powder was used for the preparation of methanolic extract.

Method of extraction: Each 100gm powder was subjected to extraction with 1000ml methanol in a reflux condenser for 3 cycles of 7hrs each till the volume reduced to half. Extract was filtered through Whattman filter paper No.1 and evaporated to dryness to get constant weight.

### **2.3 Experimental design:**

#### **2.3.1 In vitro DPP-IV inhibition assay**

##### **Reagents:**

- DPP-IV from porcine kidney, Gly-pro-p-nitroanilide (GPPN), Diprotin-A, Tris-HCl Buffer was purchased from Sigma–Aldrich (St. Louis, MO, USA).
- HPLC-grade methanol for extraction of plants
- Water was purified by 0.22µm membrane filtration and deionization (Milli-Q Plus system from Millipore, Billerica, MA, USA).

**Procedure:**<sup>12,13</sup>

The assay was performed as described by Al-Masri et al. (2009). Briefly, experiments were done in triplicate in a 96- well microplate with total volume of 100µL. Plant extracts were dissolved in water and used in various concentrations (0, 2.5, 5, 10, 20, 30, 40 and 80 µg/mL). The DMSO concentration was less than 1.0% in all experiments. A mixture containing 22.5µL of Tris-HCl buffer, different concentration of µg/mL of plant extracts, and 7.5µL of DPP-IV enzyme solution (0.05 U/mL) was pre-incubated for 10 min at 37°C, and subsequently 50µL of Gly-pro-p-nitroanilide (GPPN 0.2mM in Tris-HCl) was added to the mixture. Final incubation was done at 37°C for 30min. The absorbance was measured at 405 nm by using a plate reader. Diprotin A was used as positive reference inhibitor. Percent enzyme inhibition was calculated using the following formula:

% Inhibition = Slope (Control) – Slope (Sample) / Slope (Control) x 100%

**2.3.2 To study the influence of herbal plants in incretin system like GLP-1**<sup>14-</sup>

<sup>16</sup>

**Requirements:**

- Entero-endocrine cells (EECs), STC-1 cells (From NCCS, Pune, India), Docosahexaenoic acid (DHA), Glucogan like peptide (GLP-1) ELISA Kit was purchased from Sigma–Aldrich (St. Louis, MO, USA).

**1. Prepare 10mg/ml DHA stock solution**

- a. Dilute DHA solution with pure ethanol at 10mg/ml.
- b. Aliquot DHA stock solution into small glass bottles, 10ml each.
- c. Ambient air above the solution should be displaced to avoid oxidation of DHA by priming pure nitrogen gas on the surface of the solution prior to closing the container.
- d. Store at -20°C.

**2. DHA Stimulation assay**

1. Seed 2×10<sup>6</sup> STC-1 cells in 6cm dishes two days prior to experiment in triplicates. Culture the cells in tissue culture incubator at 37°C, 5% CO<sub>2</sub>, using Dulbecco's Modified Eagle Medium (DMEM) containing high glucose, 4.5 g/L, with L-glutamine, and without sodium pyruvate with 10% FBS and Antibiotics (100 U/mL penicillin and 100 mg/L streptomycin).
2. On experimental day cells should be 80% confluent.
3. Aspirate media and wash the cells twice with 3ml of HEPES buffer.
4. Add 3ml of HEPES buffer into each dish.
5. Put cells back to tissue incubator for 30min. the lack of nutrients in the buffer is meant to serve as a starvation period to nutrient stimulation.
6. Prepare 100µM DHA stimulation buffer.
  - A. To prepare 20ml of 100µM DHA stimulation buffer, add 66µl of 10mg/ml DHA (30.4mM) stock solution into a glass beaker using capillary micropipettes. Then add 20ml of HEPES buffer.
  - B. Sonicate the solution immediately at output level 5 and constant duty cycle for 3 min prior to stimulation to prevent DHA precipitation. If using a different sonicator, set the amplitude as 40%. After sonication, the solution should be clear, with minimal bubbling.
7. Aspirate HEPES buffer gently. Then add 1 ml of HEPES buffer into each control dish and 1ml of DHA stimulation buffer into each stimulation dish. Put all dishes back to tissue culture incubator for 15 min.
8. Collect 600µl of media from each dish and put into a fresh 1.5ml micro-centrifuge tube, then place them on ice.
9. Add 0.6µl of 100 mM PMSF in to 600µl of media at final concentration of 100mM.
10. Spin the micro-centrifuge tubes at 850 x g for 5min, at 4°C.

11. Transfer 500µl of media into a fresh tube.
12. Measure GLP1 concentration in samples by ELISA kit.

### **Lyse the cells**

For STC-1 cells, gently wash the cells with moderate amount of pre-cooled PBS and dissociate the cells using trypsin. Collect the cell suspension into a centrifuge tube and centrifuge for 5 min at 1000×g. Discard the medium and wash the cells 3 times with pre-cooled PBS. For each  $2 \times 10^6$  cells, add 150-250µL of pre-cooled PBS to keep the cells suspended. Repeat the freeze-thaw process several times until the cells are fully lysed. Centrifuge for 10min at 1500×g at 2-8°C. Remove the cell fragments; collect the supernatant to carry out the assay. Avoid repeated freeze-thaw cycles.

Bring all reagents to room temperature (18~25°C) before use. Follow the Microplate reader manual for set-up and preheat it for 15 min before OD measurement.

### **Assay procedure**

1. Add the Standard working solution to the first two columns: Each concentration of the solution is added in duplicate, to one well each, side by side (100µL for each well). Add the samples to the other wells (100uL for each well). Cover the plate with the sealer provided in the kit. Incubate for 90 min at 37°C. Note: solutions should be added to the bottom of the micro ELISA plate well, avoid touching the inside wall and causing foaming as much as possible.
2. Remove the liquid out of each well, do not wash. Immediately add 100µL of Biotinylated Detection Ab working solution to each well. Cover with the Plate sealer. Gently mix up. Incubate for 1 hour at 37°C.
3. Aspirate or decant the solution from each well, add 350µL of wash buffer to each well. Soak for 1~2 min and aspirate or decant the solution from each well and pat it dry against clean absorbent paper. Repeat this wash step 3 times. Note: a microplate washer can be used in this step and other wash steps.
4. Add 100µL of HRP Conjugate working solution to each well. Cover with the Plate sealer. Incubate for 30 min at 37°C.
5. Aspirate or decant the solution from each well, repeat the wash process for five times as conducted in step 3.
6. Add 90µL of Substrate Reagent to each well. Cover with a new plate sealer. Incubate for about 15 min at 37°C. Protect the plate from light. Note: the reaction time can be shortened or extended according to the actual color change, but not more than 30min.
7. Add 50µL of Stop Solution to each well. Note: Adding the stop solution should be done in the same order as the substrate solution.
8. Determine the optical density (OD value) of each well at once with a micro-plate reader set to 450 nm.

### **3. STATISTICAL ANALYSIS:**

The results are expressed as Mean  $\pm$  SEM (N=3) in each group. Data were analysed using statistical software Microsoft Excel worksheet. The significance of difference among the groups was assessed using Student t-test compared between Normal control (Untreated) vs. all groups  $p < 0.05$  were considered significant.

#### 4. RESULTS:

##### 4.1 In vitro DPP-IV inhibition assay

Table 1: DPP-IV inhibitory activity of different plants & Diprotin A

S. No.	Tested Material	Concentration (µg/mL)	% Inhibition ± S.E.M.	IC <sub>50</sub> µg/mL
1.	<i>Aegle marmelos</i> (n = 3)	0	1.21 ± 0.64	261.3329 µg/mL
		2.5	10.74 ± 0.46	
		5	14.75 ± 0.58	
		10	19.15 ± 0.64	
		20	22.65 ± 0.54	
		30	35.8 ± 0.69	
		40	52.11 ± 0.57	
		80	81.64 ± 0.48	
2.	<i>Moringa oleifera</i> (n = 3)	0	1.01 ± 0.67	241.5223 µg/mL
		2.5	8.44 ± 0.59	
		5	12.64 ± 0.47	
		10	14.52 ± 0.64	
		20	19.64 ± 0.49	
		30	25.19 ± 0.67	
		40	42.21 ± 0.58	
		80	64.44 ± 0.69	
3.	<i>Syzygium cumini</i> (n = 3)	0	1.19 ± 0.66	99.1478 µg/mL
		2.5	9.34 ± 0.67	
		5	13.646 ± 0.59	
		10	16.27 ± 0.56	
		20	21.45 ± 0.76	
		30	31.27 ± 0.94	
		40	52.01 ± 0.46	
		80	69.48 ± 1.35	
4.	Diprotin –A (n = 3) Reference Inhibitor	0	1.28 ± 0.44	46.6277 µg/mL
		2.5	18.13 ± 0.56	
		5	27.61 ± 0.87	
		10	32.27 ± 0.64	

		20	49.29 ± 0.25
		30	67.74 ± 1.27
		40	73.18 ± 0.21
		80	89.17 ± 1.24

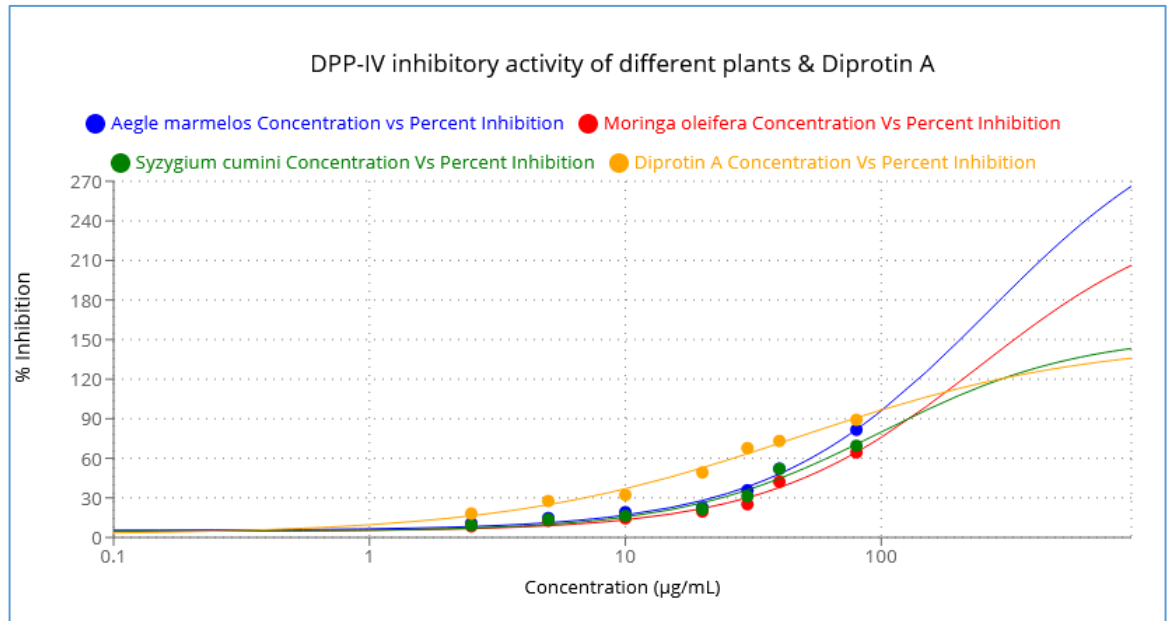


Figure 1: DPP-IV inhibitory activity of different plants & Diprotin A

Table 2: IC<sub>50</sub> Regression Results of *Aegle marmelos* concentration Vs. % Inhibition

IC <sub>50</sub> Regression Results [ <i>Aegle marmelos</i> Concentration vs Percent Inhibition]	
Parameter	Value
IC <sub>50</sub>	261.3329
Equation	$Y = 5.3571 + \frac{336.5031 - 5.3571}{1 + \left(\frac{X}{261.3329}\right)^{-1.0163}}$
Equation Form	$Y = \text{Min} + \frac{\text{Max} - \text{Min}}{1 + \left(\frac{X}{\text{IC}_{50}}\right)^{\text{Hill coefficient}}}$

Table 3: IC<sub>50</sub> Regression Results of *Moringa oleifera* concentration Vs. % Inhibition

**IC<sub>50</sub> Regression Results [Moringa oleifera Concentration Vs Percent Inhibition ]**

Parameter	Value
IC <sub>50</sub>	241.5223
Equation	$Y = 4.6346 + \frac{254.5655 - 4.6346}{1 + \left( \frac{X}{241.5223} \right)^{-1.0433}}$
Equation Form	$Y = \text{Min} + \frac{\text{Max} - \text{Min}}{1 + \left( \frac{X}{\text{IC}_{50}} \right)^{\text{Hill coefficient}}}$

**Table 4: IC<sub>50</sub> Regression Results of *Syzygium cumini* concentration Vs. % Inhibition**

**IC<sub>50</sub> Regression Results [Syzygium cumini Concentration Vs Percent Inhibition ]**

Parameter	Value
IC <sub>50</sub>	99.1478
Equation	$Y = 4.6495 + \frac{154.4454 - 4.6495}{1 + \left( \frac{X}{99.1478} \right)^{-1.1125}}$
Equation Form	$Y = \text{Min} + \frac{\text{Max} - \text{Min}}{1 + \left( \frac{X}{\text{IC}_{50}} \right)^{\text{Hill coefficient}}}$

**Table 5: IC<sub>50</sub> Regression Results of *Diprotin A* concentration Vs. % Inhibition**

**IC<sub>50</sub> Regression Results [Diprotin A Concentration Vs Percent Inhibition ]**

Parameter	Value
IC <sub>50</sub>	46.6277
Equation	$Y = 2.1214 + \frac{149.3319 - 2.1214}{1 + \left( \frac{X}{46.6277} \right)^{-0.7618}}$
Equation Form	$Y = \text{Min} + \frac{\text{Max} - \text{Min}}{1 + \left( \frac{X}{\text{IC}_{50}} \right)^{\text{Hill coefficient}}}$

**4.2 An *in-vitro* assay to measure GLP1 release from cultured murine EEC's under fatty acid stimulation**

The IC<sub>50</sub> values of plants were done on the basis of MTT assay and result found to be as follows:

SI. No.	Sample	IC <sub>50</sub>
1	<i>Aegle marmelos</i>	50µg/ml
2	<i>Moringa oleifera</i>	

3	<i>Syzygium cumini</i>	
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A number of anti-diabetic drugs are now clinically available which act by mimicking or enhancing GLP-1 action, and a new recent focus of research is the identification of compounds which stimulate intestinal secretion of GLP-1.

This study investigated the *in vitro* effects of some natural sources of GLP-1-releasing activity.

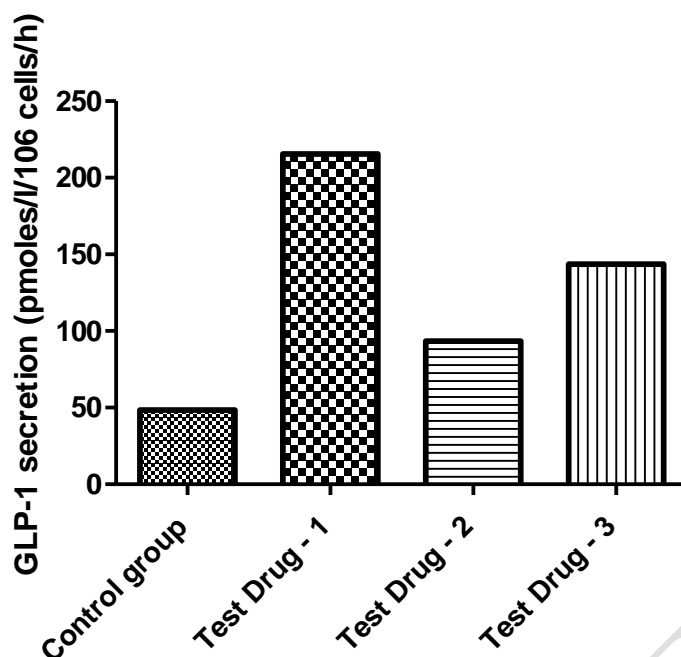
Name of Groups	GLP-1 secretion (pmoles/l/10 <sup>6</sup> cells/h)
Control Group –Media (DMEM containing high glucose, 4.5 g/L) only (Without test drug)	48.38± 0.55
Test Drug – 1 (DMEM containing high glucose, 4.5 g/L + <i>Aegle marmelos</i> 50 µg/ml)	215.5± 0.64
Test Drug – 2 (DMEM containing high glucose, 4.5 g/L + <i>Moringa oleifera</i> 50 µg/ml)	93.46± 0.89
Test Drug – 3 (DMEM containing high glucose, 4.5 g/L + <i>Syzygium cumini</i> 50 µg/ml)	143.7± 0.81

Values are expressed as Mean ± S.E.M (n=3).  
STC-1 cells were incubated for 3h with *Aegle marmelos*, *Moringa oleifera*, *Syzygium cumini* before determination of GLP-1 secretion.

#### 4.2.1 Statistical Analysis:

**Table 6: Comparison between the groups**

Tukey's Multiple Comparison Test	Mean Diff.	Significant? P < 0.05?	Summary
Control group vs Test Drug - 1	-167.1	Yes	***
Control group vs Test Drug - 2	-45.08	Yes	***
Control group vs Test Drug - 3	-95.30	Yes	***
Test Drug - 1 vs Test Drug - 2	122.0	Yes	***
Test Drug - 1 vs Test Drug - 3	71.82	Yes	***
Test Drug - 2 vs Test Drug - 3	-50.22	Yes	***



Values are expressed as Mean  $\pm$  S.E.M (n=3).

**Figure 2: GLP-1 Secretions between the groups**

## 5. DISCUSSION:

The wide variety of medical trials focusing on the outcomes of medicinal plant extracts yet herbal products is quite scarce, and established limited effects.  $\beta$ -cell destruction within DM is prominent up to expectation a conduct to insulin need and it has to stay managed for the duration of the life. If Phytoconstituent(s) concerning the flora discovered hopeful among ameliorating the severity of diabetes, will be a solution role to play in the production on insulin and their anointment or protection.

GLP-1 is a substrate for the enzyme DPP-IV, a serine protease as degrades GLP-1 within its indolent form. Exogenous GLP-1 administration has been shown to be beneficial of the remedy on kind of diabetes. However, the short half-life makes GLP-1 unattractive for continual therapy on diabetes. DPP-IV inhibition is method in conformity with prolong the circulating half-life of GLP-1, thus making DPP-IV inhibitors a promising goal for the remedy on diabetes. DPP-IV is involved within the inactivation on GLP-1, a potent Insulinotropic peptide. Thus, DPP-IV inhibition do stay an tremendous approach in accordance with treat diabetes mellitus by means of potentiating insulin secretion (Umezawa, *et al.*, 1984 & Aderibigbe, *et al.*, 2001).<sup>17,18</sup> Our result explains inhibitory things to do over DPP-IV yet can also bear therapeutic potential of diabetes. GLP-1, an incretin hormone, acts over pancreatic  $\beta$  cells by way of enhancing insulin secretion from the cells.<sup>19,20</sup> In summation to directly stimulating insulin secretion, GLP-1 also helps in imitation of recommend glucose sensitivity in accordance with  $\beta$  cells via stimulating glucose transporters and glucokinase.<sup>21,22</sup> Thus, GLP-1 is important for insulin-mediated glucose homeostasis. Indeed, pharmacological GLP-1 analogues and inhibitors on dipeptidyl peptidase-4, as inactivates GLP-1, have been chosen for the remedy on diabetes.<sup>23</sup> In the existing study, we examined the influences about *Aegle marmelos*, *Moringa oleifera* and then *Syzygium cumini* concerning GLP-1 secretion by way of using STC-1 cell line, yet it has observed probably excite GLP-1 secretion among STC-1 cells. This should stand the phytochemicals can also spark off GLP-1 receptor on the enteroendocrine cells of gut, resulting in activation of a sequence of sign transducers such as G protein-gustducin, phospholipase C beta 2, inositol 1,4,5-trisphosphate receptor type 3, and transient receptor potent channels. These methods subsequently

effects into depolarization of the enetroendocrine cell membrane via elevation on intracellular Ca<sup>2+</sup> concentration and releases GLP-1.

## 6. CONCLUSION:

In summary, Incretions GLP-1 secretions and DPP-IV inhibitors are altogether advantageous drugs for management on Diabetes Mellitus. Different in vivo and in vitro research confirmed that partial concerning the medicinal plants are rich into Insulinotropic compound. Here our present study underlines as medicinal plants i.e. *Aegle marmelos*, *Moringa oleifera*, and *Syzygium cumini* were inhibits the DPP-IV and enhances the GLP-1 secretions among STC-1 cells of diabetes. Thus, study demonstrates up to expectation it medicinal plants over methanolic extracts are potential stimulators yet may want to be a helpful conduct for further development as a recent anti-diabetic agent.

## 7. CONSENT:

Not Applicable

## 8. ETHICAL APPROVAL:

Not Needed

## 9. CONFLICT OF INTEREST STATEMENT:

We declare that we have no conflict of interest.

## 10. ACKNOWLEDEMENT:

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