

CONTROLLING WOUND PATHOGEN USING STEVIA AND PEPPERMINT BASED MOUTH GEL

Running Title: Antifungal activity of grape seed mediated titanium dioxide nanoparticles

Abstract:

Aim:

To evaluate the antimicrobial activity of Stevia and peppermint based gel on streptococcus aureus, E. coli and pseudomonas.

Background:

In recent years, the research concerning the use of herbal products has been vastly analysed to assess their antimicrobial properties. It has been found that these herbs have certain phytochemicals which contributes to its antimicrobial property. The common oral wound pathogens are *Staphylococcus aureus*, *E. coli* and *Pseudomonas* hence our sample, Stevia and peppermint based moth gel was formulated to check its antimicrobial property on *S. aureus*, *E. coli* and *pseudomonas*.

Materials and methods:

The plants were collected, stevia and peppermint and an extract was prepared. 0.5g of carbopol was added to the extract to make it in a gel consistency. This gel was concentrated against the three wound pathogens, *S. aureus*, *pseudomonas*, *E.coli*. Through agar well diffusion method, the antimicrobial property of various concentrations of the gel was tested and compared with known antibiotic amoxicillin.

Results:

The current study revealed that stevia and peppermint based mouth gel had antimicrobial activity at all concentrations but was significantly less when compared with the standard and there was no significant difference in the antimicrobial activity between different concentrations of the gel used.

Conclusion:

From the present study, it may be concluded that stevia and peppermint based mouth gel have antimicrobial properties but are comparatively lesser when compared with the antibiotic.

Keywords: antimicrobial; Stevia; peppermint; Streptococcus aureus; E.coli; pseudomonas

INTRODUCTION:

Wound infections are a significant health problem after trauma or surgery as it increases the mortality. Wounds are common in the oral cavity caused by either trauma, or surgery; these wounds heal by the same principle as the external wounds heal. Healing always starts with blood clotting that initially seals the wound(1–3). Platelet activation during the primary haemostasis releases a number of important cytokines that start the healing process via chemotactic signals to inflammatory and resident cells(4).If bleeding persists, wound healing is delayed as there is disturbance in the formation of granulation tissue. Wound healing is also impaired when the site is infected with bacteria(5,6).

The most common bacteria found in an infection are *Staphylococcus aureus*, *pseudomonas* and *E. coli*. Most of these bacteria have drug resistance, which has resulted in difficulties in curing the related infectious diseases(7). This brings about the screening of plant extracts of great interest to scientists in the search for new drugs for greater effective treatment of several diseases(8). Therefore, plant extracts and phytochemicals with known antimicrobial properties can be of great significance in therapeutic treatments (9,10). The antimicrobial action of the plant phytochemicals, the same "natural antibiotics" used in ecology, medicine and agriculture(11).

The greater part of the world population relies on traditional medicine for healthcare(12,13). This is also the case in the treatment of wound infections. In developing countries, formulation preparation from plants has been widely used for the treatment of soft tissue wounds and burns by medical personnel trained in western medicine as well as by traditional practitioners(14).

Stevia rebaudiana (commonly referred to as Honey leaf, Candy leaf and Sweet leaf), which is popular in the world not only as a low-calorie, medicinal, natural sweetener, also known for its antioxidant activity of leaf extract (15,16) It is rich in terpenes, flavonoids, phenols and flavonoids, which cause the antimicrobial properties of the plant (17) . The phytochemicals present in *Stevia rebaudiana* are austroinullin, β carotene, dulcoside, niacin, rebaudi oxides, riboflavin, steviol, stevioside and tiamin (18). *Stevia* is also proved to inhibit the growth of certain bacteria and other infectious organisms hence used against wounds sores and gum disease.

Our team has extensive knowledge and research experience that has translated into high quality publications.(19–31),(32–36) (37) (38). This study examined the effects of *Stevia* and peppermint based mouth gel on various pathogens. Since, this is a herbal formulation, side

effects of this formulation are low and hence large quantities can be applied. This may help in the future to be used as an attachment to other antimicrobial agents. Hence, through this in vitro study, we want to evaluate the potency of herbal plants of stevia and peppermint gel against the wound pathogen.

MATERIAL AND METHODS:

COLLECTION OF PLANT MATERIALS AND PREPARATION OF THE EXTRACT:

Stevia and peppermint powder were obtained readymade. The powdered stevia and peppermint powder was measured to 10g and was taken in a beaker. 100ml of distilled water was added into the beaker. The beaker was mixed, stirred well and was subjected to boiling and 90°C until the aqueous mixture was well concentrated. The concentrated mixture was then subjected to filtration. The obtained filtrate was subjected to more heating, till the volume of the filtrate was decreased to half.

PREPARATION OF THE GEL:

0.5 g of carbopol was taken in a sterile glass beaker to this beaker 50 ML of distilled water was added in small concentration and turned well until the solute was completely dissolved. This solution, 1ml of the extract (Stevia and peppermint) was added and stirred well.

Evaluation of antibacterial activity:

Three petri plates with Muller Hinton agar were taken. four wells on each plate were formed using a gel puncher. To three wells, Stevia and peppermint gel was loaded in the concentration range from 25µl, 50µl and 100µl. The fourth well was loaded with positive control, a known antibiotic (amoxicillin) . The plates were incubated at 37°C for 24 hours. After incubation, the Zone of Inhibition was measured in mm.

STATISTICAL ANALYSIS:

All the data was analyzed statistically using SPSS version 23 (IBM). One way Anova test and post hoc test using Tukey's HSD were done to compare the standard and stevia and peppermint based mouth gel at different concentrations. Confidence limit was set at 95% and $p < 0.05$ was considered as statistically significant

RESULTS:

The results showed that the prepared gel had anti microbial effect but the zone of inhibition for all three tested organisms (S.aureus, Pseudomonas, E.coli) at all three tested concentrations was found to be significantly lesser than that of the standard antibiotic used($p < 0.05$). It was also found that there was no significant difference between the Zone of inhibition at different concentrations ($p > 0.05$) (figure 1,table 2).

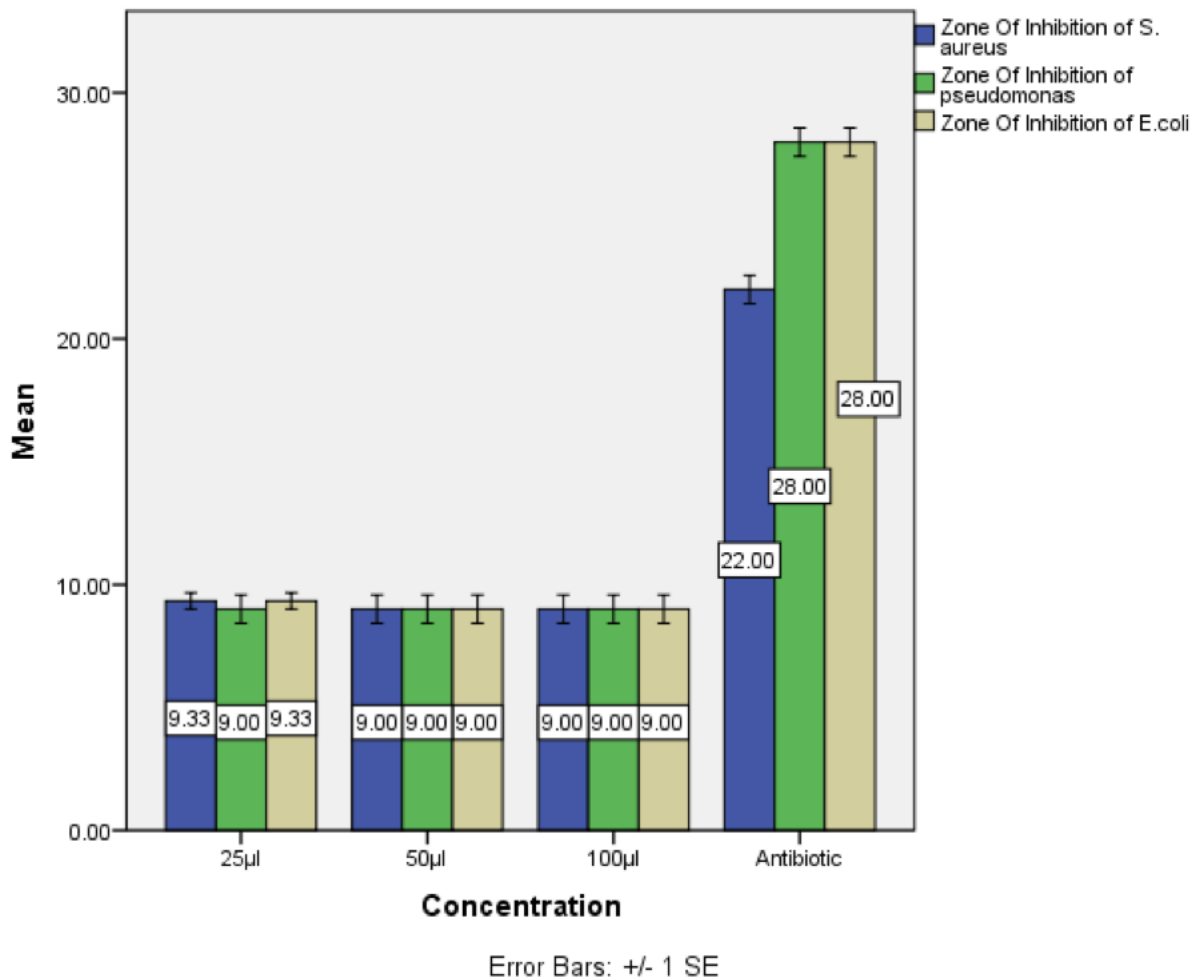


Figure1: The bar graph shows the comparison of the mean Zone of Inhibition of the standard and peppermint and stevia mouth gel at various concentrations. The X-axis represents the concentration in units of μL and antibiotic and the Y-axis represents the mean Zone of Inhibition. Blue represents the Zone of Inhibition of S. aureus, green represents Zone of Inhibition of Pseudomonas and beige represents the mean Zone of Inhibition of E.coli. The graph shows that the efficacy of Stevia and peppermint gel is significantly lesser when compared to the standard at all the concentrations ($p < 0.05$) and there was no significant difference in the zone of Inhibition between the different concentrations ($p > 0.05$). One way Anova followed Tukey's HSD analysis was done for the statistical comparison.

Table 1: This table depicts the comparison of zone of Inhibition of Various concentrations of the Stevia and peppermint based gel and control group against S.aureus, E.coli and pseudomonas using One way Anova.

		N	Mean	Std. Deviation	Std. Error	significance
Absorbance of S. aureus	25µl	3	9.33	.577	.333	0.000
	50µl	3	9.00	1.000	.577	
	100µl	3	9.00	1.000	.577	
	Antibiotic	3	22.00	1.000	.577	
	Total	12	12.33	5.883	1.698	
Absorbance of pseudomonas	25µl	3	9.00	1.000	.577	0.000
	50µl	3	9.00	1.000	.577	
	100µl	3	9.00	1.000	.577	
	Antibiotic	3	28.00	1.000	.577	

Absorbance of S. aureus	Tukey HSD	25µl	50µl	.333	.745	.968	-2.05	2.72
			100µl	.333	.745	.968	-2.05	2.72
			Antibiotic	-12.667 [†]	.745	.000	-15.05	-10.28
		50µl	100µl	.000	.745	1.00 0	-2.39	2.39
			Antibiotic	-13.000 [†]	.745	.000	-15.39	-10.61
			Antibiotic	-13.000 [†]	.745	.000	-15.39	-10.61
Absorbance of pseudomonas	Tukey HSD	25µl	50µl	.000	.816	1.00 0	-2.61	2.61
			100µl	.000	.816	1.00 0	-2.61	2.61
			Antibiotic	-19.000 [†]	.816	.000	-21.61	-16.39
		50µl	100µl	.000	.816	1.00 0	-2.61	2.61
			Antibiotic	-19.000 [†]	.816	.000	-21.61	-16.39
			Antibiotic	-19.000 [†]	.816	.000	-21.61	-16.39
Absorbance of E.coli	Tukey HSD	25µl	50µl	.333	.745	.968	-2.05	2.72
			100µl	.333	.745	.968	-2.05	2.72

50µl	Antibiotic	-18.667 [*]	.745	.000	-21.05	-16.28
	100µl	.000	.745	1.00 0	-2.39	2.39
100µl	Antibiotic	-19.000 [*]	.745	.000	-21.39	-16.61
	Antibiotic	-19.000 [*]	.745	.000	-21.39	-16.61

*. The mean difference is significant at the 0.05 level.

DISCUSSION:

Nowadays, due to the excessive usage of antibiotics, the common pathogens have become resistant to them, which is in turn decreasing the efficacy of antibiotics. Due to this reason, it is of utmost importance to find out new antimicrobial agents to combat antibiotic resistance and to aid in better wound healing following an infection(39). The results of this study reveals that the stevia and peppermint based gel is having some antimicrobial properties against staphylococcus aureus, pseudomonas and E. coli but the efficacy was significantly lesser than amoxicillin. The study also revealed that the antimicrobial effect of stevia peppermint gel was not concentration dependent.

The current study evaluated the antimicrobial property of stevia and peppermint based gel on various wound pathogens like Staphylococcus aureus, pseudomonas and E.coli at different concentrations like 25µl, 50µl and 100µl. Staphylococcus aureus is the most common organism associated with surgical wound infections. In a study done by Nwachukwu et al, it was found that 42.3% of infections were found to be caused by Staphylococcus aureus(40). Few other authors found that among the Gram-negative organisms, Escherichia coli were frequently isolated from the wounds(41,42).

In the current study, amoxicillin was used as a standard to compare the antimicrobial activity of Stevia and peppermint gel. Amoxicillin is the most commonly used antibiotic in treating wound infections. Amoxicillin belongs to the beta lactam group of antibiotics(43). Beta-lactams act by binding to penicillin-binding proteins that inhibit a process called transpeptidation (cross-linking process in cell wall synthesis), leading to activation of autolytic enzymes in the bacterial cell wall (44).

In the current study, It was noted that the antimicrobial activity of Stevia and Peppermint mouth gel was lesser than that of the standard (amoxicillin). It was also noted that the antimicrobial activity of stevia and peppermint mouth gel was concentration independent which means that with increase in the concentration, there was no change in the antimicrobial activity of the gel.

In a study done by Das et al. the antimicrobial activity of aqueous, methanolic, and ethanolic extracts of *S. rebaudiana* leaves was evaluated. All individual extracts showed potential antimicrobial activity compared to standard ampicillin, but the activities were lower than standard which was similar to the results found in our study(45).

In a study done by Gosh et al., evaluated the antimicrobial activity of *S. rebaudiana* leaf extracts (i.e., petroleum ether, cyclohexane, chloroform, water, acetone, and ethanol) against 10 pathogenic as well as food-spoilage fungal (*Alternaria solani*, *Helminthosporium solani*, *Aspergillus niger*, and *Penicillium chrysogenum*) and pathogenic bacterial (*Escherichia coli*, *Bacillus subtilis*, *Enterococcus faecalis*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus*) isolates, using streptomycin and cotrimazole as controls. These authors found that petroleum ether extracts at 250 mg/mL (MIC) inhibit the growth of *E. coli* and *S. aureus* (by the plate dilution method) among bacteria and *P. chrysogenum* among fungi. Among all extracts, petroleum ether exhibited the best antimicrobial potential followed by water, chloroform, cyclohexane, acetone, and ethanol. This shows that extracts of *S. rebaudiana* act on a wide spectrum of microorganisms(46). The medicinal properties are attributed to the primary and secondary metabolites synthesized by the plant(47).

The principal constituents of peppermint include monoterpene alcohols mainly menthol (38-48%), ketones mainly menthones (20-30%), some monoterpenes and oxides. The metabolites of stevia includes, steviol and its glycosides, stevioside, rebaudioside (A to F), steviolbioside, dihydro isosteviol, rubusoside, and dulcoside A. these metabolites contribute to its antioxidant, antibacterial, anti-spasmodic and antiviral properties(48–52).

Even Though the present results reveal that the stevia and peppermint gel is not as good as the amoxicillin even at 100 microlitre concentration, further studies using higher concentrations or different extract preparation methods may add more value to this study. Moreover the other

medicinal properties of stevia such as its antioxidant and anti inflammatory effects will add additional benefits while using this gel(53) (54) (55) (56) (57) (58) (59) (60) (61) (62) (63) ((63,64) (65) (66). Due to the increase in the antibiotic sensitivity, herbal formulation will be a viable alternative to the conventional antibiotics. These herbal formulations can also be used as an adjuvant with the antimicrobials, increasing their efficacy Further research on isolating different components may be undertaken and may be incorporated into existing anti-inflammatory herbal compositions to improve their efficacy. Promising results regarding the antimicrobial effect of Stevia peppermint gel can be further validated with future in vivo studies to find the safe and effective concentration for clinical usage.

Conclusion:

From the present study, it may be concluded that stevia and peppermint based mouth gel have antimicrobial properties but are comparatively lesser when compared with the antibiotic amoxicillin .

REFERENCES:

1. Bowler PG, Duerden BI, Armstrong DG. Wound microbiology and associated approaches to wound management. *Clin Microbiol Rev.* 2001 Apr;14(2):244–69.
2. Politis C, Schoenaers J, Jacobs R, Agbaje JO. Wound Healing Problems in the Mouth. *Front Physiol.* 2016 Nov 2;7:507.
3. Schultz GS, Chin GA, Moldawer L, Diegelmann RF. 23 Principles of Wound Healing. *Mechanisms of vascular disease: a reference book for vascular specialists.* 2011;423.
4. Broughton G, Janis JE, Attinger CE. Wound Healing: An Overview [Internet]. Vol. 117, *Plastic and Reconstructive Surgery.* 2006. p. 1e – S. Available from: <http://dx.doi.org/10.1097/01.prs.0000222562.60260.f9>
5. Cohen IK, Die-gelmann RF, Lindblad WJ, Hugo NE. Wound Healing: Biochemical and Clinical Aspects. *Plast Reconstr Surg.* 1992 Nov;90(5):926.
6. Cooper DM. Wound healing: new understandings. *Nurse Pract Forum.* 1999 Jun;10(2):74–86.
7. Sundin GW, Bender CL. Dissemination of the strA-strB streptomycin-resistance genes among commensal and pathogenic bacteria from humans, animals, and plants. *Mol Ecol.* 1996 Feb;5(1):133–43.
8. Encarnación Dimayuga R, Keer Garcia S. Antimicrobial screening of medicinal plants from

Baja California Sur, Mexico. *J Ethnopharmacol.* 1991 Feb;31(2):181–92.

9. Wele M, Kirkman L, Diarra N, Goita Y, Doumbia M, Traore K, et al. Antiplasmodial Potential and Phytochemical Screening of Ten Plants Used as Antimalarial in Mali [Internet]. Vol. 19, *European Journal of Medicinal Plants.* 2017. p. 1–9. Available from: <http://dx.doi.org/10.9734/ejmp/2017/34523>
10. Obinna O, John A, Peter OO. The effect of extraction protocol on the phytochemical and antimicrobial activities of *Lantana camara* leaf extract found within a local environment. *Journal of Basic and Applied Chemistry.* 2013;3(1):5–10.
11. Venkanna L, Estari M. In vitro antimicrobial activity of some medicinal plants used by tribes in Warangal district (Andhra Pradesh), India. *Biol Med .* 2012 Apr;4(2):85–8.
12. Anitha R, Aneesa N, Varghese S. Antidiabetic activity of ajwain oil in different in vitro models [Internet]. Vol. 11, *Journal of Pharmacy And Bioallied Sciences.* 2019. p. 142. Available from: http://dx.doi.org/10.4103/jpbs.jpbs_128_18
13. Kritika Jangid , Jayakumar ND, Sheeja S Varghese. Achievable therapeutic effect of *Myristica Fragrans* (NUTMEG) on Periodontitis A short Review. *Int J Pharm Pharm Sci.* 2014 May 15;6(5):591–4.
14. Fahimi S, Mortazavi SA, Abdollahi M, Hajimehdipour H. Formulation of a Traditionally Used Polyherbal Product for Burn Healing and HPTLC Fingerprinting of Its Phenolic Contents. *Iran J Pharm Res.* 2016 Winter;15(1):95–105.
15. Pinheiro CE, de Oliveira SS, da Silva SM, Poletto MI, Pinheiro CF. Effect of guaraná and *Stévia Rebaudiana* Bertoni (leaves) extracts, and stevioside, on the fermentation and synthesis of extracellular insoluble polysaccharides of dental plaque. *Rev Odontol Univ Sao Paulo.* 1987 Oct;1(4):9–13.
16. Takahashi K, Matsuda M, Ohashi K, Taniguchi K, Nakagomi O, Abe Y, et al. Analysis of anti-rotavirus activity of extract from *Stevia rebaudiana*. *Antiviral Res.* 2001 Jan;49(1):15–24.
17. Taware AS, Mukadam DS, Chavan AM, Taware SD, Others. Comparative studies of in vitro and in vivo grown plants and callus of *Stevia rebaudiana* (Bertoni). *Int J Integr Biol.* 2010;9(1):10–5.
18. CRAMMER, B. Sweet glycosides from the *Stevia* plant. *Chem Br.* 1986;22:915–8.
19. Ramesh A, Varghese S, Jayakumar ND, Malaiappan S. Comparative estimation of sulfiredoxin levels between chronic periodontitis and healthy patients - A case-control study. *J Periodontol.* 2018 Oct;89(10):1241–8.
20. Paramasivam A, Priyadharsini JV, Raghunandhakumar S, Elumalai P. A novel COVID-19 and its effects on cardiovascular disease. *Hypertens Res.* 2020 Jul;43(7):729–30.

21. S G, T G, K V, Faleh A A, Sukumaran A, P N S. Development of 3D scaffolds using nanochitosan/silk-fibroin/hyaluronic acid biomaterials for tissue engineering applications. *Int J Biol Macromol*. 2018 Dec;120(Pt A):876–85.
22. Del Fabbro M, Karanxha L, Panda S, Bucchi C, Nadathur Doraiswamy J, Sankari M, et al. Autologous platelet concentrates for treating periodontal infrabony defects. *Cochrane Database Syst Rev*. 2018 Nov 26;11:CD011423.
23. Paramasivam A, Vijayashree Priyadharsini J. MitomiRs: new emerging microRNAs in mitochondrial dysfunction and cardiovascular disease. *Hypertens Res*. 2020 Aug;43(8):851–3.
24. Jayaseelan VP, Arumugam P. Dissecting the theranostic potential of exosomes in autoimmune disorders. *Cell Mol Immunol*. 2019 Dec;16(12):935–6.
25. Vellappally S, Al Kheraif AA, Divakar DD, Basavarajappa S, Anil S, Fouad H. Tooth implant prosthesis using ultra low power and low cost crystalline carbon bio-tooth sensor with hybridized data acquisition algorithm. *Comput Commun*. 2019 Dec 15;148:176–84.
26. Vellappally S, Al Kheraif AA, Anil S, Assery MK, Kumar KA, Divakar DD. Analyzing Relationship between Patient and Doctor in Public Dental Health using Particle Memetic Multivariable Logistic Regression Analysis Approach (MLRA2). *J Med Syst*. 2018 Aug 29;42(10):183.
27. Varghese SS, Ramesh A, Veeraiyan DN. Blended Module-Based Teaching in Biostatistics and Research Methodology: A Retrospective Study with Postgraduate Dental Students. *J Dent Educ*. 2019 Apr;83(4):445–50.
28. Venkatesan J, Singh SK, Anil S, Kim S-K, Shim MS. Preparation, Characterization and Biological Applications of Biosynthesized Silver Nanoparticles with Chitosan-Fucoidan Coating. *Molecules* [Internet]. 2018 Jun 12;23(6). Available from: <http://dx.doi.org/10.3390/molecules23061429>
29. Alsubait SA, Al Ajlan R, Mitwalli H, Aburaisi N, Mahmood A, Muthurangan M, et al. Cytotoxicity of Different Concentrations of Three Root Canal Sealers on Human Mesenchymal Stem Cells. *Biomolecules* [Internet]. 2018 Aug 1;8(3). Available from: <http://dx.doi.org/10.3390/biom8030068>
30. Venkatesan J, Rekha PD, Anil S, Bhatnagar I, Sudha PN, Dechsakulwatana C, et al. Hydroxyapatite from Cuttlefish Bone: Isolation, Characterizations, and Applications. *Biotechnol Bioprocess Eng*. 2018 Aug 1;23(4):383–93.
31. Vellappally S, Al Kheraif AA, Anil S, Wahba AA. IoT medical tooth mounted sensor for monitoring teeth and food level using bacterial optimization along with adaptive deep learning neural network. *Measurement*. 2019 Mar 1;135:672–7.
32. PradeepKumar AR, Shemesh H, Nivedhitha MS, Hashir MMJ, Arockiam S, Uma Maheswari TN, et al. Diagnosis of Vertical Root Fractures by Cone-beam Computed

Tomography in Root-filled Teeth with Confirmation by Direct Visualization: A Systematic Review and Meta-Analysis. *J Endod.* 2021 Aug;47(8):1198–214.

33. R H, Ramani P, Tilakaratne WM, Sukumaran G, Ramasubramanian A, Krishnan RP. Critical appraisal of different triggering pathways for the pathobiology of pemphigus vulgaris-A review. *Oral Dis* [Internet]. 2021 Jun 21; Available from: <http://dx.doi.org/10.1111/odi.13937>
34. Ezhilarasan D, Lakshmi T, Subha M, Deepak Nallasamy V, Raghunandhakumar S. The ambiguous role of sirtuins in head and neck squamous cell carcinoma. *Oral Dis* [Internet]. 2021 Feb 11; Available from: <http://dx.doi.org/10.1111/odi.13798>
35. Sarode SC, Gondivkar S, Sarode GS, Gadbail A, Yuwanati M. Hybrid oral potentially malignant disorder: A neglected fact in oral submucous fibrosis. *Oral Oncol.* 2021 Jun 16;105390.
36. Kavarthapu A, Gurumoorthy K. Linking chronic periodontitis and oral cancer: A review. *Oral Oncol.* 2021 Jun 14;105375.
37. Vellappally S, Abdullah Al-Kheraif A, Anil S, Basavarajappa S, Hassanein AS. Maintaining patient oral health by using a xeno-genetic spiking neural network. *J Ambient Intell Humaniz Comput* [Internet]. 2018 Dec 14; Available from: <https://doi.org/10.1007/s12652-018-1166-8>
38. Aldhuwayhi S, Mallineni SK, Sakhamuri S, Thakare AA, Mallineni S, Sajja R, et al. Covid-19 Knowledge and Perceptions Among Dental Specialists: A Cross-Sectional Online Questionnaire Survey. *Risk Manag Healthc Policy.* 2021 Jul 7;14:2851–61.
39. Ionescu MI. Are Herbal Products an Alternative to Antibiotics? In: Sahra, editor. *Bacterial Pathogenesis and Antibacterial Control.* Rijeka: IntechOpen; 2018.
40. Nwachukwu NC, Orji FA, Okike UM. Antibiotic susceptibility patterns of bacterial isolates from surgical wounds in Abia State University Teaching Hospital (ABSUTH), Aba--Nigeria. *Research Journal of Medicine and Medical Sciences.* 2009;4(2):575–9.
41. Afroz H, Fakruddin M, Masud MR. Incidence of and risk factors for hospital acquired infection in a tertiary care hospital of Dhaka, Bangladesh. *Bangladesh Journal of* [Internet]. 2017; Available from: <https://www.banglajol.info/index.php/BJMS/article/view/32847>
42. Aman S. Bacteriological analysis of wound infection in Mayo hospital, Lahore. *J Pak Med Assoc* [Internet]. 1982; Available from: <https://www.jpma.org.pk/PdfDownload/6510>
43. Handsfield HH, Clark H, Wallace JF, Holmes KK, Turck M. Amoxicillin, a new penicillin antibiotic. *Antimicrob Agents Chemother.* 1973 Feb;3(2):262–5.
44. Kapoor G, Saigal S, Elongavan A. Action and resistance mechanisms of antibiotics: A guide for clinicians. *J Anaesthesiol Clin Pharmacol.* 2017 Jul;33(3):300–5.

45. Das K, Dang R. Influence of Biofertilizers on Protein, Moisture and Ash Content in Relation to Swelling Property, Water Absorption Capacity, Mineral Elements, Total Phenolic Level of Stevia (*Stevia rebaudiana* Bert.) Plant Grown under Acidic Soil Zone of South India [Internet]. Vol. 6, International Letters of Natural Sciences. 2013. p. 78–97. Available from: <http://dx.doi.org/10.18052/www.scipress.com/ilns.6.78>
46. Ghosh S, Subudhi E, Nayak S. Antimicrobial assay of Stevia rebaudiana Bertoni leaf extracts against 10 pathogens. *Int J Integr Biol*. 2008;2(1):27–31.
47. Gupta E, Purwar S, Sundaram S, Rai GK. Nutritional and therapeutic values of Stevia rebaudiana: A review. *Journal of Medicinal Plants Research*. 2013;7(46):3343–53.
48. Thosar N, Basak S, Bahadure RN, Rajurkar M. Antimicrobial efficacy of five essential oils against oral pathogens: An in vitro study. *Eur J Dent*. 2013 Sep;7(Suppl 1):S071–7.
49. Ratan R. Handbook of aromatherapy: a complete guide to essential & carrier oils, their application & therapeutic use for holistic health & wellbeing. Institute of Holistic Health Science; 2006.
50. Dawes C, Macpherson LM. Effects of nine different chewing-gums and lozenges on salivary flow rate and pH. *Caries Res*. 1992;26(3):176–82.
51. Savita SM, Sheela K, Sunanda S, Shankar AG, Ramakrishna P. Stevia rebaudiana – A Functional Component for Food Industry. *J Hum Ecol*. 2004 Apr 1;15(4):261–4.
52. Shivanna N, Naika M, Khanum F, Kaul VK. Antioxidant, anti-diabetic and renal protective properties of Stevia rebaudiana. *J Diabetes Complications*. 2013 Mar;27(2):103–13.
53. Danda AK. Comparison of a single noncompression miniplate versus 2 noncompression miniplates in the treatment of mandibular angle fractures: a prospective, randomized clinical trial. *J Oral Maxillofac Surg*. 2010 Jul;68(7):1565–7.
54. Robert R, Justin Raj C, Krishnan S, Jerome Das S. Growth, theoretical and optical studies on potassium dihydrogen phosphate (KDP) single crystals by modified Sankaranarayanan–Ramasamy (mSR) method [Internet]. Vol. 405, *Physica B: Condensed Matter*. 2010. p. 20–4. Available from: <http://dx.doi.org/10.1016/j.physb.2009.08.015>
55. Krishnan V, Lakshmi T. Bioglass: A novel biocompatible innovation. *J Adv Pharm Technol Res*. 2013 Apr;4(2):78–83.
56. Soh CL, Narayanan V. Quality of life assessment in patients with dentofacial deformity undergoing orthognathic surgery—A systematic review [Internet]. Vol. 42, *International Journal of Oral and Maxillofacial Surgery*. 2013. p. 974–80. Available from: <http://dx.doi.org/10.1016/j.ijom.2013.03.023>
57. Lekha L, Kanmani Raja K, Rajagopal G, Easwaramoorthy D. Schiff base complexes of rare earth metal ions: Synthesis, characterization and catalytic activity for the oxidation of aniline and substituted anilines [Internet]. Vol. 753, *Journal of Organometallic Chemistry*.

2014. p. 72–80. Available from: <http://dx.doi.org/10.1016/j.jorganchem.2013.12.014>
58. Dhinesh B, Isaac JoshuaRamesh Lalvani J, Parthasarathy M, Annamalai K. An assessment on performance, emission and combustion characteristics of single cylinder diesel engine powered by *Cymbopogon flexuosus* biofuel [Internet]. Vol. 117, *Energy Conversion and Management*. 2016. p. 466–74. Available from: <http://dx.doi.org/10.1016/j.enconman.2016.03.049>
 59. PradeepKumar AR, Shemesh H, Jothilatha S, Vijayabharathi R, Jayalakshmi S, Kishen A. Diagnosis of Vertical Root Fractures in Restored Endodontically Treated Teeth: A Time-dependent Retrospective Cohort Study. *J Endod*. 2016 Aug;42(8):1175–80.
 60. Vijayakumar GNS, Nixon Samuel Vijayakumar G, Devashankar S, Rathnakumari M, Sureshkumar P. Synthesis of electrospun ZnO/CuO nanocomposite fibers and their dielectric and non-linear optic studies [Internet]. Vol. 507, *Journal of Alloys and Compounds*. 2010. p. 225–9. Available from: <http://dx.doi.org/10.1016/j.jallcom.2010.07.161>
 61. Kavitha M, Subramanian R, Narayanan R, Udhayabanu V. Solution combustion synthesis and characterization of strontium substituted hydroxyapatite nanocrystals [Internet]. Vol. 253, *Powder Technology*. 2014. p. 129–37. Available from: <http://dx.doi.org/10.1016/j.powtec.2013.10.045>
 62. Sahu D, Kannan GM, Vijayaraghavan R. Size-Dependent Effect of Zinc Oxide on Toxicity and Inflammatory Potential of Human Monocytes [Internet]. Vol. 77, *Journal of Toxicology and Environmental Health, Part A*. 2014. p. 177–91. Available from: <http://dx.doi.org/10.1080/15287394.2013.853224>
 63. Neelakantan P, Cheng CQ, Mohanraj R, Sriraman P, Subbarao C, Sharma S. Antibiofilm activity of three irrigation protocols activated by ultrasonic, diode laser or Er:YAG laser in vitro [Internet]. Vol. 48, *International Endodontic Journal*. 2015. p. 602–10. Available from: <http://dx.doi.org/10.1111/iej.12354>
 64. Lekha L, Kanmani Raja K, Rajagopal G, Easwaramoorthy D. Synthesis, spectroscopic characterization and antibacterial studies of lanthanide(III) Schiff base complexes containing N, O donor atoms [Internet]. Vols. 1056-1057, *Journal of Molecular Structure*. 2014. p. 307–13. Available from: <http://dx.doi.org/10.1016/j.molstruc.2013.10.014>
 65. Gopalakannan S, Senthivelan T, Ranganathan S. Modeling and Optimization of EDM Process Parameters on Machining of Al 7075-B4C MMC Using RSM [Internet]. Vol. 38, *Procedia Engineering*. 2012. p. 685–90. Available from: <http://dx.doi.org/10.1016/j.proeng.2012.06.086>
 66. Parthasarathy M, Isaac JoshuaRamesh Lalvani J, Dhinesh B, Annamalai K. Effect of hydrogen on ethanol-biodiesel blend on performance and emission characteristics of a direct injection diesel engine. *Ecotoxicol Environ Saf*. 2016 Dec;134(Pt 2):433–9.