

Original Research Article

Response Surface Methodology Driven Systematic Development of a Novel RP-UPLC Method for The Quantification of Aliskiren: A Renin Inhibitor

ABSTRACT

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Aims:The current study envisages experimental design enabled rapid, sensitive, and stability-indicating RP-UPLC method for the quantification of Aliskiren in its pharmaceutical formulations.

Study design:Box-Benken experimental Design using Response surface methodology.

Place and Duration of Study: Department of Analytical Research and Development Brawn laboratories Ltd., Gurugram, India, and Department of Pharmacy KL College Pharmacy, KL Deemed to be University, Vaddesearam, Guntur, Andhra Pradesh, between May 2021 and September 2021.

Methodology:The chromatographic partitioning was achieved on a Waters Acuity H class UPLC system, with BEH 130⁰A, C18 column (100 x 2.1 mm, 1.7 μ m) using isocratic elution with PDA detector. The optimum conditions were delineated, selecting three influential factors (CMPs), i.e., mobile phase composition, flow rate, and injection volume. Systematic optimization was accomplished by 3² Box-Benken design using response surface methodology (RSM).

Results:The selected variables are evaluated for obtained responses (CAAs), i.e., peak area, retention time (Rt), USP Plate count. The optimized method employed, mobile phase composition 0.2 % Glacial acetic acid (pH 3.0) and acetonitrile 50:50 (% v/v) with 0.3 mL min⁻¹ flow rate. The injection volume was maintained as 2 μ L with 2 minutes run time and λ max 280 nm.

Conclusion:Linearity was performed at 5-300 ppm, and R² was found 0.9995. Forced degradation studies were performed, enduring stability profile of drug as per ICH. The short Rt 1.214, minute implies superior robustness, sensitivity, and cost-effective for routine analysis. The results exhibited RSM approach of QbD could be competently used to optimize the RP-UPLC method with fewer experimental trials and error-free investigation.

Keywords: Chromatography; stability; specificity; renin-inhibitor; Design of experiment; Validation.

1. INTRODUCTION

Aliskiren is recognized as a potent drug of choice known as direct orally active nonpeptide renin inhibitors. The drug is applicable predominantly during high blood pressure. Chemically known as N-Methyl-2-[[3-[(1E)-2-(2-pyridinyl) ethenyl]-1H-indazol-6-yl] thio] benzamide (Fig. 1). Aliskiren is oral, potent, and selective inhibitor of vascular endothelial growth factor receptors [1]. For its clinical use, this can exhibit a novel and advantageous pharmacokinetic and pharmacodynamic profile for the long-term treatment of hypertension so-termed as antihypertensive [2]. In association with selected antihypertensive drugs like calcium channel blockers, the medication of Aliskiren might also be applied with thiazides in product form to provide additional hypertension recoveries. Due to most acceptable resolution, rapidity, and sensitivity of analysis, UPLC is considered as budding part of the systematic development of chromatographic science which holds the sensibleness as compared to conventional HPLC techniques. Unification of the three dynamic factors (speed, resolution, and sensitivity) of advanced UPLC systems and with a configuration of UPLC with PDA detection helps in the isolation with the high-speed scan rates and identification of degradation products by reducing the time required to develop stability-indicating methods [3,4]. The modern application of UPLC with the design of experiments (DoE) paradigm is to improve the analysis of the complex mixture of samples, hiccups originated during product development and analytical research. UPLC takes full advantage of chromatographic principles to run the separations using columns packed with smaller particles and higher flow rates for improved speed and sensitivity rather than traditional HPLC development, which is pretty tedious.4 UPLC with QbD served as a proven arena and presiding as an emerging concept based on the robust, rapidity of analysis, regulatory flexibility as well as stability outline of drug products as per ICHQ2R1 guidelines [4],[5] Literature findings revealed for Aliskiren some works with combination dosages have been reported in HPLC, UFLC, etc. But there are limited works that have been reported until now in Response Surface Methodology (RSM) driven Analytical QbD (AQbD) approach, with stability profile analysis. As, QbD based UPLC system has been discovered to produce intense peak capacities with enhanced spectrum quality, separation efficiency, faster elution, and this is found to be quite beneficial in analyzing the complex mixtures rather than depending trial and error basis [4-6].QbD based statistical methodology intensifies analytical design space concept, Risk assessments strategy, MFAT (multiple-factors-at-a-time) approach as a contrast to traditional one-factor-at-a-time (OFAT) operations [2]. Hence, an effort has been made to develop and validate a QbD based precise, cost-effective, sensitive UPLC method [2,3], for the quantification of Aliskiren in its pharmaceutical formulations, which is also stability-signifying as per ICH stability guidelines of ICHQ2R1 and ICHQ8, Q9, Q10 [4,6,7].

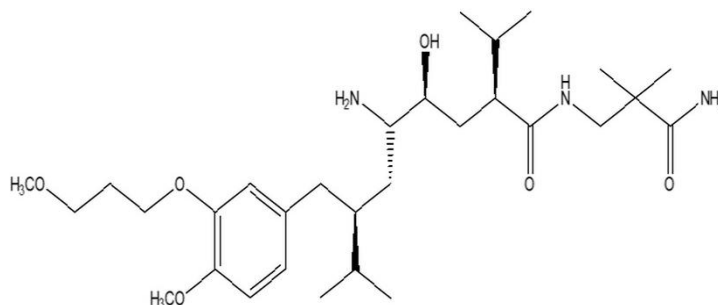


Fig. 1. Chemical structure of Aliskiren

2. METHODOLOGY

2.1.1. Materials and Chemicals

Reference standard or API of Aliskiren (purity 99.4 % w/w) were obtained from Sun Pharmaceutical Laboratories Pvt. Ltd, (Gujarat), India. The commercial pharmaceutical formulations were procured from the local market. The other foremost solvents used for the research include Acetonitrile UPLC Grade (Merck), Mili-Q-Water (Merck) Methanol and Glacial Acetic acid (Spectro chem). The filtration was performed by the Nylon filter (0.22 μm)–Millipore, Mumbai, India. The pH measurements were made using a Metsar Tech. pH meter.

2.1.2. Instrumentation

The chromatographic development was carried out by a Waters Acquity H class UPLC system with BEH (100 x 2.1 mm), with a particle size of 1.7 μm) C18 Column-130 with auto-injector equipped with PDA detector regulated by Empower 2 software. The maximum wavelength was detected with PDA spectra.

2.2. Methods

2.2.1. Statistical Analysis

The advanced statistical software of Design Expert (Ver.12, Stat Ease, Minneapolis, USA) was employed for screening with method optimization for assessing CPPs to obtain CAAs through experimental runs [8]. The calculations for the analysis of regression equation and its ANOVA were premeditated by Microsoft excels 2019 (Microsoft, USA) [1,9].

2.2.2. Preparation of Standard

Aliskiren 75 mg of working standard was systematically weighed and allowed to transfer into a 50 mL volumetric flask, add 30 ml of diluent, ultra-sonicated for 10 minutes, the final volume was made up with diluents to obtain a final concentration of Aliskiren 150 $\mu\text{g mL}^{-1}$. From the stock standard solution, 10 mL was pipetted out in to a 100 ml volumetric flask, and then final volume was made with the diluent. The ensuing chromatogram by injecting blank, standard, and a mixture of excipients (placebo) and formulations are depicted in Fig. 2 (a), (b), (c), and (d) respectively.

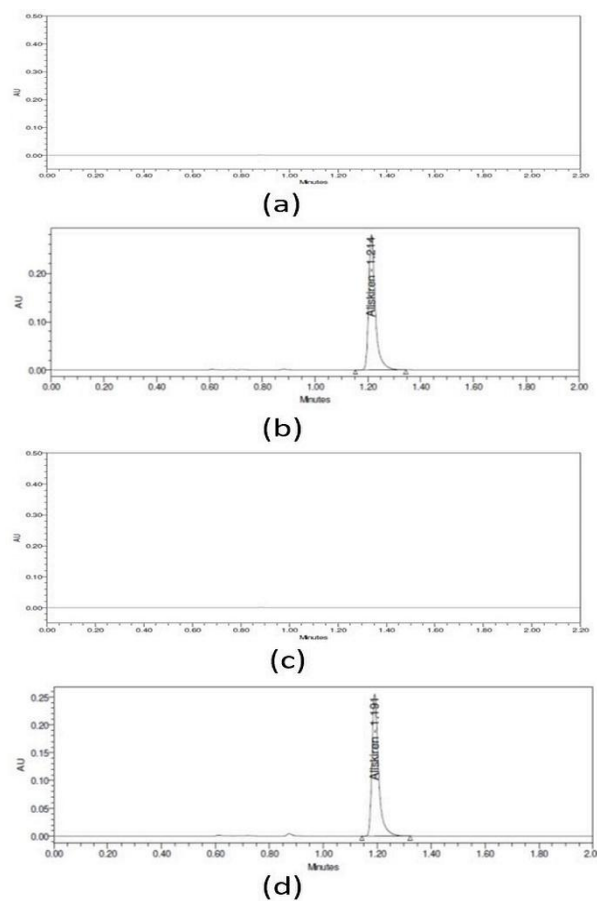


Fig. 2. Optimized Chromatograms of blank (a), standard 150µg/mL (b); mixture of excipients (c) and Formulation (d).

2.2.3. Sample Preparation

Twenty tablets of commercial brands of Aliskiren have weighed accurately, and each Tablets' average weight was calculated accurately. After that, the weight equivalent to 300 mg Tablet was transferred into a 100 ml volumetric flask. Diluent of appropriately 50 mL was added and then ultrasonicated for 30 minutes. Further, the volume is made up of diluent and filtered. The filtered solution 1ml was pipette out into a 10 mL volumetric flask and made up to mark with diluent.

2.2.4. Method Development using Box-Benken Design (BBD)

AQbD usually delivers assistance by reducing the time, resources, and efforts to develop robust based methods with pertinency in drug substance analysis, degradation products, and other metabolites.⁹ For development, Box-Behnken experimental design (BBD) was incorporated [10], [11]. to compute independent variables (CMPs) and their capable effects upon, the distinct critical analytical or quality attributes (CQAs) like peak area, retention time (Rt), and USP plate count. The focal, interactions, and quadratic effects upon the influential critical variables like mobile phase ratio, flow rate, and injection volume upon the dependent variables like peak area (Y1), retention time (Y2), USP plate count (Y3), are analyzed with

total 17 experimental runs [12-15]. A method operable design region (MODR) or appropriate design space was earmarked, providing the best method concert via numerical and graphical optimizations over counterplots and by comparing predicted vs. experimental values. The Graphical and statistical chromatographic BBD includes basically the qualitative polynomial equations, 2-D, 3-D counter plot illustrations under the principle of Response surface methodology (RSM) [14],[15].

3. RESULTS AND DISCUSSION

The trail runs aids in the construct of an arithmetical model involving the comprehensive analysis of critical factors¹⁶. Similarly, the 17 experimental runs with three critical factors and its associated responses 3^2 of BBD experimental design have been elucidated. The design matrix of all the encoded critical factors (independent variables i.e., % mobile phase and flow rate, injection volume) and its associated observed responses (CAAs) are summarized in Table 1.

Table 1. Design matrix as per Box-Benkhen design for the optimization of chromatographic method

	Low level (-1)	High level (+1)
Independent factors		
X1: Mobile Phase (% v/v)	30	70
X2: Flow rate (ml/min ⁻¹)	0.2	1
X3: Injection volume (μl)	1	5
Dependent factors (responses)		
Y11: Peak Area		
Y22: Retention Time		
Y3: USP Plate count		

Similarly, the 17 experimental runs with 3 critical factors and its associated responses (3^2) of BBD experimental design has been enlisted in Table 2.

Table 2. Optimization of method by 3² Box–Behnken design using RSM

Experimen tal runs	Organic phase (% v/v)	Flow rate (ml min⁻¹)	Injection volume (µl)	Peak Area (Cm²)	Rt (minute)	USP Plate count
1	30	1	3	1020021	1.911	12762
2	50	0.6	3	1290876	1.208	13985
3	30	0.6	1	509876	1.211	10765
4	50	0.2	5	2423217	1.204	12876
5	70	0.6	5	2208761	0.811	12498
6	30	0.6	5	2406541	2.073	10783
7	70	0.2	3	1001245	1.106	15678
8	30	0.2	3	1301456	2.421	10877
9	70	1	3	1100023	0.916	11097
10	70	0.6	1	309871	1.046	11031
11	50	0.2	1	414527	1.215	14608
12	50	0.6	3	965431	0.912	13985
13	50	1	1	122134	0.829	10876
14	50	0.6	3	804321	0.976	13912
15	50	1	5	576843	0.821	11056
16	50	0.6	3	840654	0.917	13985
17	50	0.6	3	840165	0.914	13980

3.1. Optimized Chromatographic Conditions

The chromatographic column used was, Waters Acuity H class UPLC BEH 130o A, C18, Column (100 x 2.1 mm, 1.7 µm); mobile phase used was 0.2 % GAA: acetonitrile with 50:50 (% v/v), and flow rate was maintained as 0.3 mL min⁻¹ during the study. Similarly, the desired pH was monitored 3.0 and the detector employed was PDA with λ_{max} 255 nm. The ultimate temperature was maintained 30°C during method optimization.

3.1.1 Optimization of chromatographic method using RSM methodology

After selecting optimal chromatographic conditions, the Box-Benkhen Design (BBD) with response surface methodology (RSM), was executed through principles of ANOVA for achieving the enhanced method performance like robustness and leaving scope for continuous enhancement within the specified design space [14],[16],[17]. The multivariate linear regression analysis performed the data optimization analysis to screen out the

tentative responses 2-D counter and 3-D response surface plots. The experimental results or the solutions from the graphical optimization (Experimental run 11) designate that, Organic phase composition (50 % v/v), flow rate (0.2 mL/minute) with injection volume (1 μ L) are the utmost influential variables for the method optimizations affecting the Critical Analytical attributes (CAAs), that elucidate final obtained actual responses, i.e., Peak Area (414527 cm^2), retention time (Rt) 1.215, with USP plate count 14608 (Table 3).

Table 3. ANOVA and its significance value with respect to quadratic model post prediction and confirmation data

Source	F value		Peak area (Cm^2)		Retention time (Minute)		USP plate count	
	F value	P- value	F value	P- value	F value	P- value	F-value	P-value
Model	6.12	0.0131*	12.24	*0.0016	13.39	*0.0012		
A- Mobile phase	0.3860	0.5541	56.83	0.0001	9.78	0.0167		
B-Flow rate	5.45	0.0523	8.78	0.0210	25.40	0.0015		
C- injection volume	39.59	0.0004	1.50	0.2597	0.0017	0.9685		
AB	0.2922	0.6056	0.8334	0.3916	31.22	0.0008		
A ²	3.40	0.1078	26.00	0.0014	18.88	0.0034		
B ²	0.8563	0.3856	3.85	0.0906	0.2485	0.6334		
C ²	0.3039	0.5986	2.52	0.1561	27.36	0.0012		
Lack of fit	5.81	0.0612*	3.09	0.1521*	753.9	<0.0001*		
Run 11 Response	Predicted Mean	Predicted Median	Selected values/ Solutions	Observed values	Std Dev.	SE Pred.	95% PI low	95% PI high
Peak Area	3494	3494	12,23561	414527	351676	465223	1096584	1103571
Retention Time	1.12562	1.12562	1.225	1.215	0.175265	0.231853	0.577379	1.67387
USP plate count	13871.4	13871.4	15890	14608	578.595	765.409	12061.5	15681.3

*Significant levels, i.e., less than α value (0.05); *P. I: prediction interval; Std Dev: standard deviation; SE: standard error.

The graphical optimization solutions elucidate that the predicted values are almost close to obtained experimental values. The counter plots (2D and 3D) responses and their significant interactions of three critical factors upon are depicted in Fig. 3.

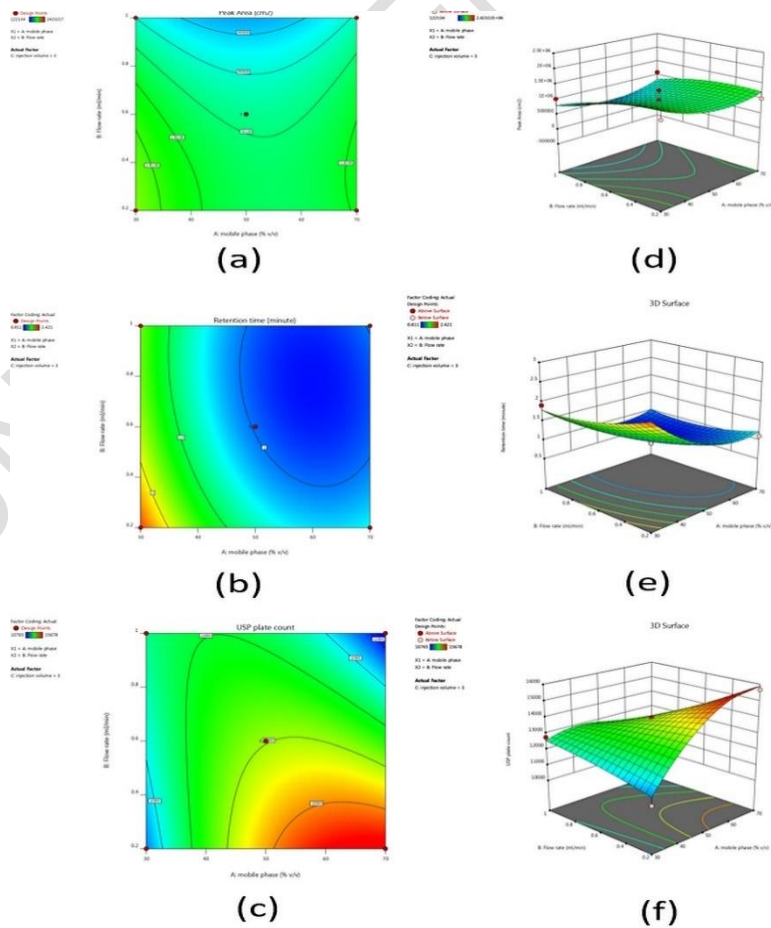
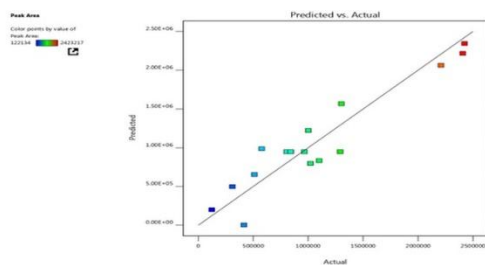
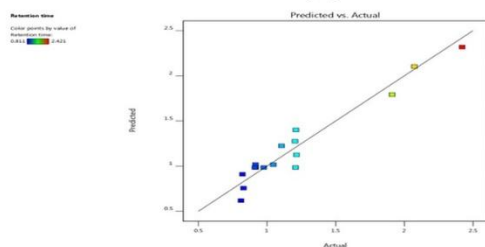


Fig. 3. Schematic diagram indicating 2-D surface contour plot analysis of peak area [Y1] response (a), Retention time [Y2] response (b), USP plate count [Y3] (c); 3-D surface contour plot analysis of peak area [Y1] response (d), Retention time [Y2] response (e), USP plate count [Y3] response.

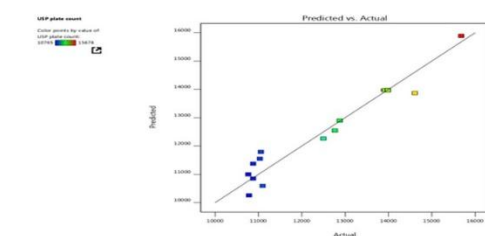
Similarly, the schematic plot indicating predicted values with and actual experimental values are demonstrated in Fig. 4.



(a)



(b)



(c)

Fig. 4. Predicted vs. Actual value for Peak Area [Y1] (a); Predicted vs. Actual value for Retention Time [Y2]; (b) and Predicted vs. Actual value for USP Plate count [Y3] (c).

3.2. Method Validation

Analytical method validation (AMV) proves that an analytical method that affords analytical results is acceptable for the envisioned practice [8],[12],[14]. As per ICH recommended guidelines, and the drug was subjected to various validation parameters like system suitability test (SST), linearity, accuracy, precision (system, intra and interday), robustness, LOD, and LOQ, etc.

3.2.1. Results of method validation parameters

3.2.1.1. Linearity

Linearity of the method was analyzed for the drug concentrations from 5-300 $\mu\text{g mL}^{-1}$, employing an injection volume of 10 μL for each concentration. Regression analysis was performed on the obtained data by correlating concentrations and its responses (Peak Area) using an MS-Excel 2019 spreadsheet (M/s Microsoft Inc., Washington, USA) forcing the line through the origin, and value of pertinent statistical parameters with $Y = 3220X + 32668$, and regression co-efficient (R^2) was obtained 0.9995. The representative linearity plot or calibration curve was depicted in Fig. 5.

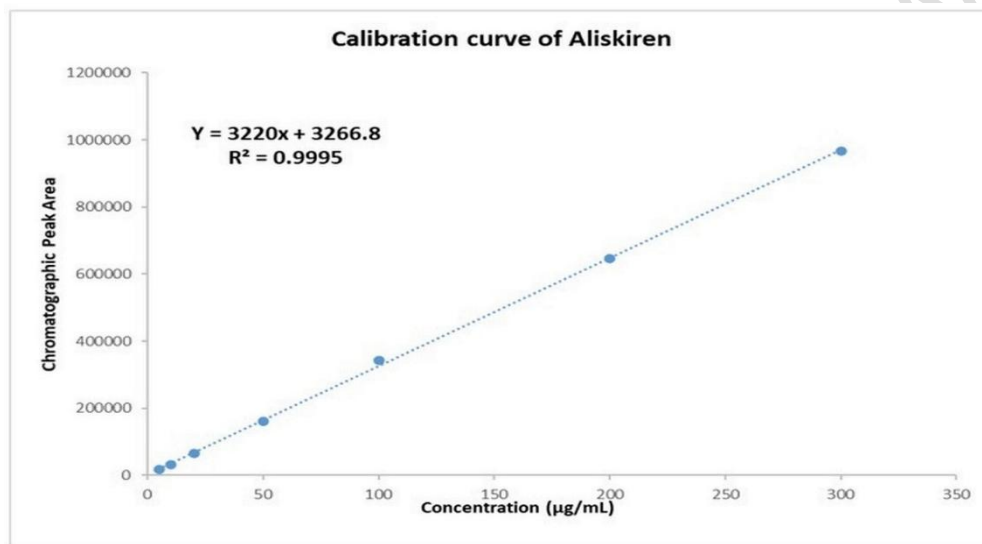


Fig. 5. Schematic diagram of Calibration Plot (a) of Aliskiren

3.2.1.2. Precision

Precision is denoted as the intimacy of preparation (degree of scattering) amongst a series of measurements obtained from multiple samplings of the equal homogeneous sample[8,12]. Precision studies of the drug were carried out by the system, method, and intermediate precision testing. The results of (system, intraday and interday) Precision studies are demonstrated in Table 4.

Table 4. System precision, intraday and interday precision test

System Precision	Intra day	Interday
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Conc. (µg/mL)	Peak Area	USP Tailoring	USP plate count	Conc. (µg/mL)	Peak Area of at different time intervals			Conc. (µg/mL)	Peak Area of at different time intervals		
					(Day 1)				(Day 2)		
					10 A.M	2 P.M	5 P.M		10 A.M	2 P.M	5 P.M
10	48207	1.61	7486	5	241190	240123	241987	5	241554	241487	241442
10	485124	1.62	7453	5	241132	240564	241765	5	241431	241541	241643
10	492661	1.51	7359	5	241087	240221	241889	5	241023	241879	241877
				Average	241136.3	240302.6	241880.3	Average	241336	241635	241654
10	483654	1.53	7422	SD	51.636	231.565	111.253	SD	277.95	212.45	217.70
				% RSD	0.021	0.096	0.045	% RSD	0.115	0.087	0.090
10	482854	1.52	7356	10	482392	482454	482776	10	483018	483968	483765
				10	482129	482146	483543	10	485931	484961	484886
10	486059	1.57	7468	10	482736	486263	483456	10	482625	484066	484134
Average	485404			Average	485228	483621	483258	Average	483858	484331	484261
SD	3842.7			SD	304.3	2293.2	419.9	SD	1805.99	547.21	571.30
% RSD	0.791			% RSD	0.062	0.474	0.086	% RSD	0.062	0.474	0.086

20	970453	970764	971567	20	971732	971837	971728
20	970452	970771	971569	20	971765	971880	971762
20	970437	971732	971498	20	971754	971878	971769
Average	970447.3	970989	971544.7	Average	971750.3	971865	971753
SD	8.9628	383.66	40.426	SD	16.802	24.269	21.931
% RSD	0.00092	0.03951	0.00416	% RSD	0.00172	0.00249	0.00225

**RSD: relative standard deviation; SD: standard deviation*

3.2.1.3. System suitability Testing (SST)

System suitability parameters were studied by injecting the typical standard solution six times, and results were well under the acceptance criteria. Instrumental performance parameters like peak area, retention time, and USP plate count (> 2000) were evaluated and established, which showed that % RSD was not more than 2%. The % RSD for six replicate injections of the standard was to be 0.791 %. The results of system suitability test are demonstrated in Table 5.

3.2.1.4. Robustness

The robustness of an analytical process is about the degree of its capacity to persevere unaffected by a minor, but deliberate disparities in method performance and its parameters, which indicates its reliability during normal usage [12,19]. The study was performed by altering flow rate, wavelength, and % mobile phase composition. The % RSD less than 0.547 indicates a robust method. The results of robustness studies are demonstrated in Table 5.

Table 5. System suitability and robustness data of Aliskiren

SYSTEM SUITABILITY TEST (SST)	ROBUSTNESS STUDY
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Injection No's	Optimized condition (SST)	USP plate count	Tailing Factor	Parameter	Flow Rate [1+ 0.2ml min ⁻¹]	Flow rate [1- 0.2ml min ⁻¹]	Wavelength [254 +2 nm]	Wavelength [254 -2 nm]	Amount of [ACN + 2%v/v]	Amount of [ACN - 2%v/v]
Peak Area (Inj.1)	483356	13090	1.6	Peak Area (Inj.1)	474365	485595	472465	468476	485267	476253
Peak Area (Inj.2)	472178	13143	1.6	Peak Area (Inj.2)	475947	483997	474381	467429	480178	471122
Peak Area (Inj.3)	475965	13069	1.5	Peak Area (Inj.3)	472863	481359	474972	464419	482659	474337
Peak Area (Inj.4)	486824	12962	1.5	Mean	474391.6	483650.33	473939.33	466774.6	482701.3	473904
Peak Area (Inj.5)	487877	13021	1.5	Std	1542.17	2139.17	1310.55	2106.16	2544.76	2592.76
Peak Area (Inj.6)	483257	12964	1.5	% RSD	0.325	0.442	0.276	0.451	0.527	0.547
Mean	481576.16			Difference w.r.t SST (%)	0.90	0.43	1.58	3.07	0.233	1.59
Std	2339.13									
% RSD	0.485									

*RSD: Relative standard deviation, *SD: Standard deviation

3.2.1.5. Accuracy

As per the ICH guidelines about the validation of analytical procedures[19]denotes that accuracy or trueness of experimental observations. Accuracy study was performed at three level (50%, 100 % and 150%) and the results indicate the mean %recovery studies of all, percentage (%) level data are within acceptance level (98-102 %). The results of robustness studies are demonstrated in Table 6.

Table 6. Accuracy data of Aliskiren

% Level	Amount Spiked (µg/mL)	Amount Recovered (µg/mL)	% Recovery	Mean % Recovery
50%	75	75.02	100.03	100.26 %
	75	75.31	100.42	
	75	75.62	100.83	
100%	150	150.3	100.20	
	150	149.13	99.42	
	150	150.49	100.33	
	225	223.74	99.54	
150%	225	223.80	99.47	
	225	226.86	100.83	

3.2.1.6. LOD and LOQ

The limit of detection (LOD) and limit of quantification (LOQ)of the current studies were evaluated from the baseline noise of Aliskirenthrough findings of calculated signals of samples with known concentrations of analyte with that of the blank by (signal-to-noise) S/N

ratio 3:1 (LOD) & 10:1 (LOQ) as per ICHQ2B guidelines [14],[19]. The calculated values of LOD and LOQ were obtained as $0.48 \mu\text{g mL}^{-1}$ and $1.45 \mu\text{g mL}^{-1}$ respectively.

3.2.1.7. Specificity

Specificity is the capability to evaluate the analyte explicitly in the occurrence of components i.e., degradants, matrix, which may be anticipated to be present. The specificity of the method can be studied by performing stress testing or forced degradation studies. The mixture of excipients was injected to check the interference with the main peak. The results established that, there is no interference was detected from the mixture of excipients which are depicted in Fig. 6 (a-d).

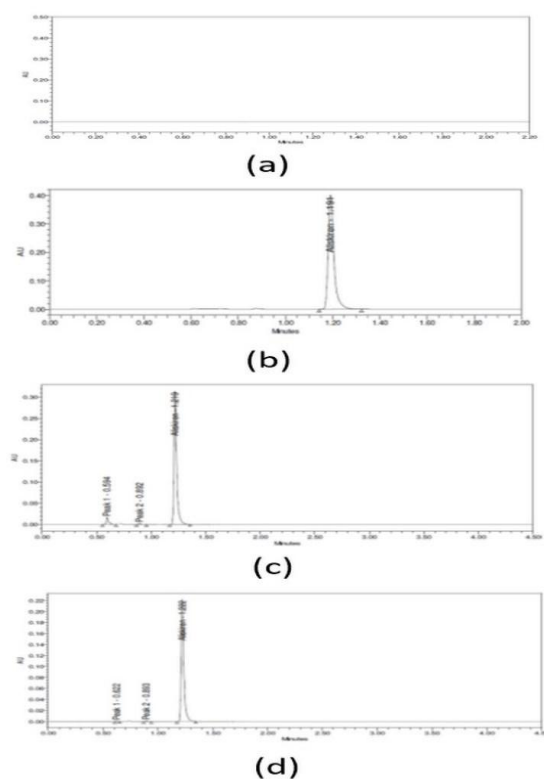


Fig. 6. (a-d). Schematic diagram indicating (a) mixture of excipients, (b) sample acidic degradation, (c) alkali degradation and (d) Peroxide degradation.

3.3. Forced Degradation Studies

Forced degradation studies was carried out to patterned the stability of the drug substance and drug product as per recommendations of ICHQ2R1[12],[19]. The drug was subjected to different stress conditions as per ICH. Acidic degradation was performed by taking 1 mL of stock s solution of Aliskiren, and to this 1mL of 2N Hydrochloric acid was

added and refluxed for 10 minutes at 60°C. Alkali Degradation studies were carried out by taking 1 mL of stock solution Aliskiren, and to this 1 mL of 2N sodium hydroxide (NaOH) was added and allowed to reflux for 30 mins at 60°C. Peroxide degradation was carried out by taking 1 mL of stock solution of Aliskiren, and to it 1 mL 3% H₂O₂, hydrogen peroxide (H₂O₂) was added separately. Finally, the ensuing solutions of acidic, alkali and peroxide degradations were diluted to obtain 150 µg mL⁻¹ and 2.0 µL were injected into the UPLC system. The chromatograms with results of degradations studies were recorded Fig. 6 (a-d) and Table 7.

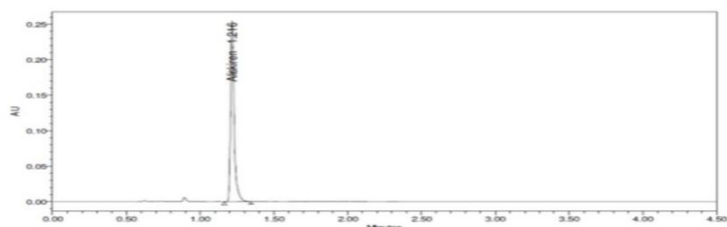
Table 7. Forced degradations and solution stability data of Aliskiren

Stress conditions	Chromatographic Peak Area	*Drug Recovered (%)	*Drug decomposed (%)
Aliskiren standard (Control)	418252	99.8	---
Acidic degradation 1 ml of 2N Hydrochloric acid 60°C, 10 minutes	373215	94.53	5.27
Alkali degradation 1 ml of 2N sodium hydroxide NaOH, 60°C, 30 mins	412367	95.65	4.15
Peroxide degradation 1 ml 3% H ₂ O ₂ , room temperature, 10 minutes	486381	96.66	3.14
Thermal degradation 60°C 6 hours	492666	98.22	1.58
Photolytic degradation 365 nm, 3 hours in UV Chamber	495459	99.41	0.39

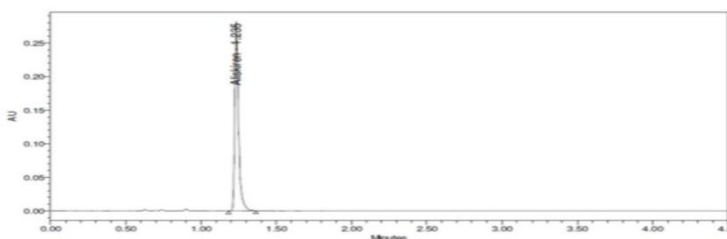
Solution stability data of Aliskiren

Time in Hrs	Standard Peak Area	% Difference
Initial	485404	---
4	484521	0.18
6	484235	0.24
8	482565	0.58
12	481256	0.85
24	478569	1.41
28	477856	1.55
32	476589	1.82
36	476025	1.93

Likewise, thermal and photolytic degradations of the drug were also premeditated by exposing the 150 µg mL⁻¹ solution to UV light by keeping the beaker in UV Chamber for one day with 200-Watt hours/m² in photostability chamber (Fig. 7a & 7b).



(a)



(b)

Figure 7 (a-b). Schematic diagram indicating thermal degradation (a), and UV degradations (b)

Finally, the subsequent solution was diluted to obtain 150 µg mL⁻¹ solutions, and 0.2µL were injected into the UPLC system fitted out with PDA detector. The resultant chromatograms were recorded to assess the stability of the sample [12],[18],[19]and the results are enlisted in Table 7.

3.3.1. Stability of Analytical Solutions

Stability of the analytical solution was calculated by monitoring the standard and sample solution at 25 ± 2 °C for the diverse time intervals. Eventually, to assess the stability

of the sample, the standard drug and samples were monitored carefully, which signifies those solutions will be stable for up to 36 hours demonstrated in Table 7 [18-20].

3.4. Assay of Pharmaceutical Formulations

The measured values of % assay of two different marketed formulations of Aliskiren are embodied in Table 8. The results demonstrate that all the values of marketed formulations are within the acceptance limit i.e., 98-102 % (Table8).

Table 8. Assay of Formulations

Sample No	Brands	Label claims (mg)	% Drug obtained	% Recovery
1	Rasilez, Novartis	300	298.69	99.56
2	Aliskiren Tablets, PAR Formulations Pvt. Ltd.	300	300.08	100.02

4. CONCLUSION

The present article productively reveals the efficiency of Response surface methodology (RSM) through AQbD approach, and is to enhance the UPLC chromatographic method for the analysis with an improved understanding of the critical factor-response relationship for expanding the method performance. As, AQbD is widely being accepted as a scientifically-sound and legitimate paradigm, that possess significant strategies for its execution, mostly when there is not precisely a regulatory need. The overall stress degradation studies practicing the RSM based, estimation of the Aliskiren in tablet dosage forms ensured and stability indicating systematic holistic method which is sensitive, precise, and highly robust. These indicate that, quality assurance will be guaranteed in the developed method with regulatory flexibility. Ultimately, the method also can find practical application to the quality control labs for routine analysis.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

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LIST OF ABBREVIATIONS

ICH:International Conference on Harmonization

λ_{max} : Maximum Wavelength

RSD: Relative standard deviation

UPLC: Ultra Performance Liquid Chromatography

ANOVA: Analysis of Variance

% RSD: % Relative Standard Deviation

2D: Two dimensional

3D: Three dimensional

BBD: Box-Benkhen Design

DoE: Design of Experiment

FDA: Food and Drug Administration

CMPs: Critical Method Parameters

MODR: Method Operable design region

ATP: Analytical Target Profile

AQbD: Analytical Quality By design

UNDER PEER REVIEW