

Original Research Article

Induction of Bax and activation of caspases by hydro ethanolic leaf extract of *Citrullus colocynthis* (L)-mediated apoptosis in breast cancer cell line (MCF-7)

Running Title: Leaf extract of *Citrullus colocynthis* on human breast cancer cells

Abstract:

Cancer is the most deadly disease that causes death among the world. It is caused by genome and epigenome abnormalities. Herbal plants are used for treatment of cancer. *Citrullus colocynthis* is valuable medicinal plants used for treatment of cancer. The plant as antidiabetic, anticancer activity, and antioxidant activity. The aim of the study is to find induction of bax and activation of caspases by hydro ethanolic leaf extract of *Citrullus colocynthis* (L)-in Human breast cancer cells (MCF-7). Materials and methods, Human breast cancer cell was purchased from the national centre for cell sciences. Cell line and cell culture were performed and cell viability was done by MTT assay. Total RNA isolation ,cDNA conversion and gene expression analysis by real time PCR. This study clearly indicates that plant extract has a significant role in modulation of intrinsic apoptotic signaling in human breast cancer cells which might be due to the presence of cucurbitacin present in *Citrullus colocynthis*. It is conducted that *Citrullus colocynthis* extract inhibits the growth of the human breast cancer cells (MCF-7) by regulating the expression of pro apoptotic (Bax) and caspases -3 genes. Hence *Citrullus colocynthis* may be served as a potential drug for treatment of human breast cancer cells.

Key words : cancer, genome, *Citrullus colocynthis*, human breast cancer cells, bax, caspases, potential drugs, innovation, novel

Introduction:

Cancer is the most deadly disease that causes death among the world (1), (2). Genome abnormalities cause cancer (3), (4). It is also caused by epigenome (5). There are anticancer drugs for direct effects anthracyclines, cyclophosphamide, cisplatin and for indirect effects taxanes, trastuzumab, sorafenib (6)(7)(8). Cancer cells are resistant against chemotherapeutic drugs. Metformin is a chemotherapeutic drug that kills four different types of breast cancer (9)(10). Breast cancer is the most common type of cancer among women all around the world. New tumours are formed from the breast cancer cells.

Sequential mutation in genes causes cancer (11) (12). Breast cancer is either inherited or acquired through mutations (13)(14). In some countries patients use several herbal plants for cancer treatment (15)(16). *Citrullus colocynthis* plant and its family cucurbitaceae. And its common name is bitter apple.

Citrullus colocynthis is a valuable medicinal plant. The plant contains several bioactive compounds such as cucurbitacin glycosides, flavonoids. Antioxidant activity is also present in these plants (15,17). It is also an antidiabetic property and treated for hemorrhoids (18). The cucurbitacin glycosides have therapeutic value against human breast cancer (19). There are various studies done by scientists by extraction of fruit, root, stem, pulp etc. There is inhibition of cell viability and migration of various cancer cells during pulp extraction. Anti-metastatic potential along with apoptotic activity is present in pulp extraction (20)(21).

The beneficial effect of cucurbitacin glycosides as an effect on chemo prevention of human breast cancer cells (22). In another study fish oil rich in n-3 polyunsaturated fatty acids has beneficial effects in many diseases including cancer also (23). The aim of study is to find induction and activation of caspases by hydroethanolic leaf extract of *Citrullus colocynthis* (L)-in Human breast cancer cells (MCF-7)

Materials and Methods

Dimethyl sulfoxide (DMSO), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were purchased from Sigma Chemical Pvt Ltd, USA. Trypsin-EDTA, fetal bovine serum (FBS), antibiotics-antimycotics, RPMI 1640 medium and phosphate buffered saline (PBS) were purchased from Gibco, Canada. (5,5,6,6-tetrachloro-1,1,3,3-tetraethylbenzimidazolocarboyanine iodide) and Real Time PCR kit was purchased TAKARA (Meadowvale Blvd, Mississauga, ON L5N 5S2, Canada).

Cell lines and cell culture

Human breast cancer cell line (MCF-7) was purchased from the National Centre for Cell Sciences (NCCS), Pune, India. Cells were cultured in DMEM medium (Thermo Fisher Scientific, CA, USA) containing 10% fetal bovine serum (Thermo Fisher Scientific, CA, USA), 100 U/ml penicillin and 100 µg/ml streptomycin (Thermo Fisher Scientific, CA, USA) at 37°C with 5% CO₂.

Cell viability by MTT assay

Cell viability was assayed using a modified colorimetric technique that is based on the ability of live cells to convert MTT, a tetrazolium compound into purple formazan crystals by mitochondrial reductases (Mosmann, 1983). Briefly, the cells (1×10^4 /well) were exposed to different concentrations of *Citrullus colocynthis* leaf extract (100-500 μ g/ml) with MCF-7 cells for 48 h. At the end of the treatment, 100 μ l of 0.5 mg/ml MTT solution was added to each well and incubated at 37 °C for an hour. The formed crystals were dissolved in dimethyl sulfoxide (100 μ l) and incubated in dark for an hour. Then the intensity of the color developed was assayed using a Micro ELISA plate reader at 570 nm. The number of viable cells was expressed as the percentage of control cells cultured in serum-free medium. Cell viability in the control medium without any treatment was represented as 100%. The cell viability is calculated using the formula: % cell viability = [A570 nm of treated cells/A570 nm of control cells] \times 100.

Gene expression analysis by Real Time-PCR

Samples from each group were submerged in 2 ml Trizol (Invitrogen, Carlsbad, CA, USA) for RNA extraction and stored at -80°C until further processed. cDNA synthesis was performed on 2 μ g RNA in a 10 μ l sample volume using Superscript II reverse transcriptase (Invitrogen) as recommended by the manufacturer. Real-time PCR array analysis was performed in a total volume of 20 μ l including 1 μ l cDNA, 10 μ l qPCR Master Mix 2x (Takara, USA) and 9 μ l ddH₂O. Reactions were run on an CFX96 Touch Real-Time PCR Detection System (Bio-Rad, USA) using universal thermal cycling parameters (95°C for 5 min, 40 cycles of 15 sec at 95°C, 15 sec at 60°C and 20 sec at 72°C; followed by a melting curve: 5 sec at 95°C, 60 sec at 60°C and continued melting). For quality control purposes, melting curves were acquired for all samples. The specificity of the amplification product was determined by melting curve analysis for each primer pair. The data were analyzed by comparative CT method and the fold change is calculated by $2^{-\Delta\Delta\text{CT}}$ method described by Schmittgen and Livak (2008) using CFX Manager Version 2.1 (Bio Rad, USA).

Statistical analysis

The obtained data were analyzed statistically by one-way analysis of variance (ANOVA) and Duncan's multiple range test with a computer-based software (Graph Pad Prism version 5) to analyze the significance of individual variations among the control and experimental groups. The significance was considered at $p < 0.05$ level in Duncan's test

Results:

This study clearly indicates that plant extract has a significant role in modulation of intrinsic apoptotic signaling in human breast cancer cells which might be due to presence of cucurbitacin present in *Citrullus colocynthis*. The ethanolic extract of *Citrullus colocynthis* inhibits the growth of human breast cancer cells (MCF-7) by regulating the pro apoptotic expression and caspases-3 genes and the result shows that *Citrullus colocynthis* can be used as potential drug for treatment of human breast cancer cells

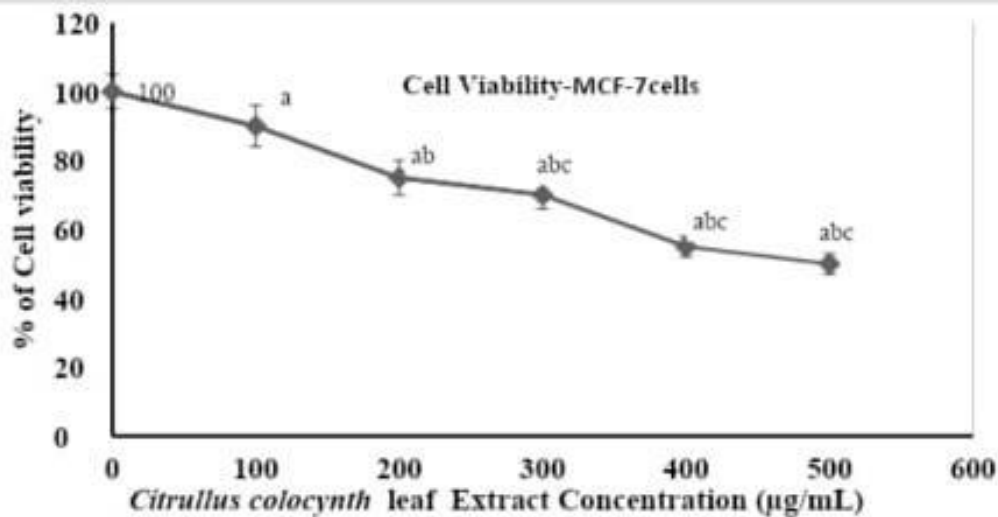


Figure 1: Effect *Citrullus colocynthis* leaf extract on cell viability in MCF-7 cells. Each bar represents a mean \pm SEM of 6 observations. Significance at $p < 0.05$, a-compared with untreated control cells, b-compared with 1nM treated MCF-7 cells.

Gene expression analysis

Bax mRNA expression (Fold change over control)

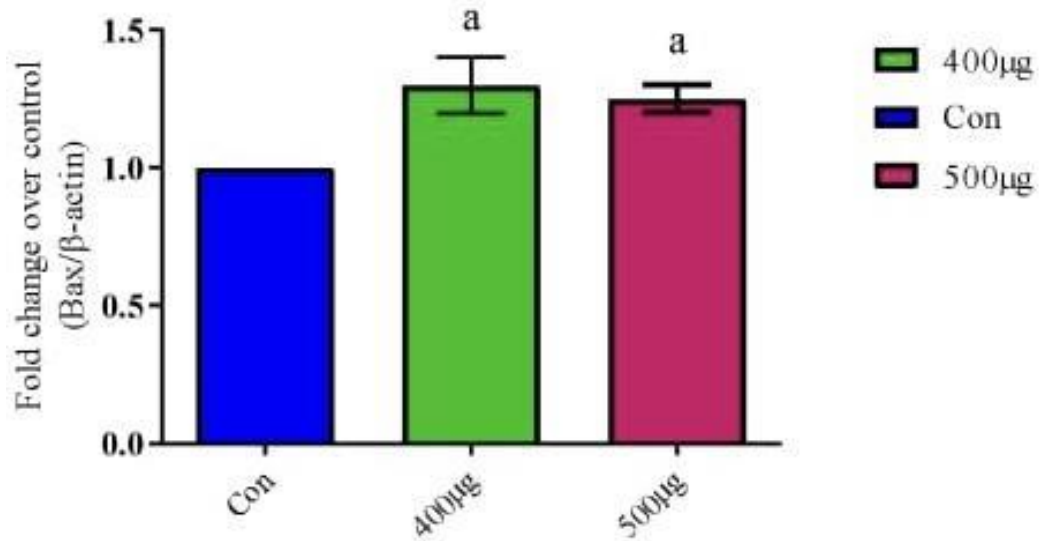


Figure 2: Effect of *Citrullus colocynthis* leaf extract on Bax mRNA expression in MCF-7 cells. Each bar represents a mean \pm SEM of 6 observations: green color represents 400 microgram, blue color represents concentration and red color represents 500 microgram Significance at $p < 0.05$, a-compared with untreated control cells

Caspase-9- mRNA expression (Fold change over control)

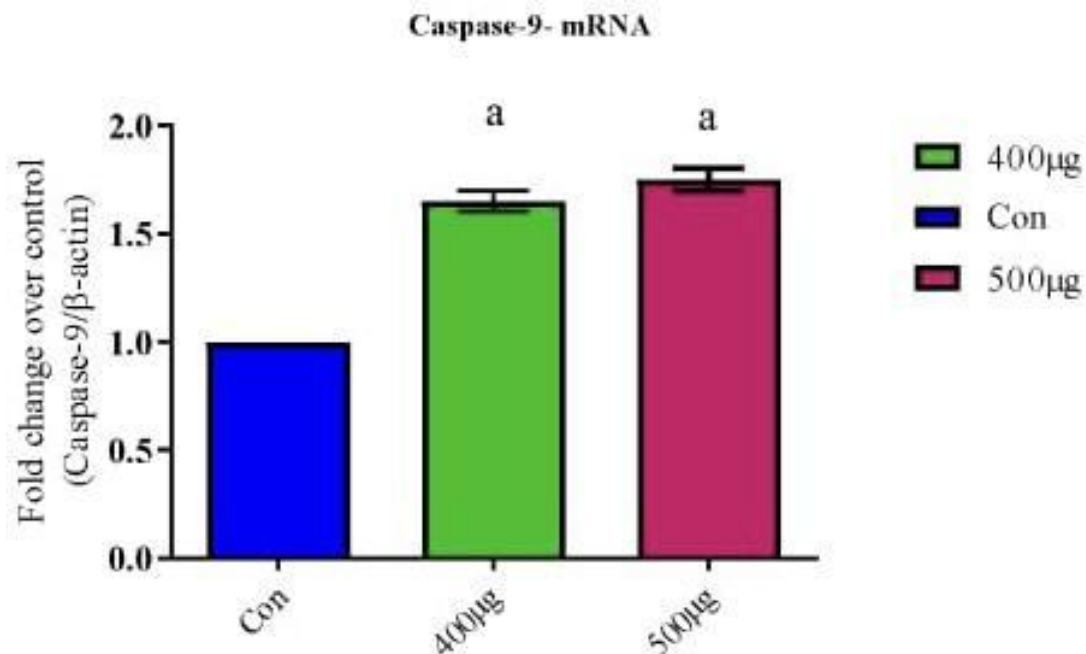


Figure 3: Effect of *Citrullus colocynthis* leaf extract on caspase-9 mRNA expression in MCF-7 cells. Each bar represents a mean \pm SEM of 6 observations: green color represents 400 microgram, blue color represents concentration and red color represents 500 microgram . Significance at $p < 0.05$, a-compared with untreated control cells

Discussion:

This study clearly indicates that plant extract has a significant role in modulation of intrinsic apoptotic signaling in human breast cancer cells which might be due to presence of cucurbitacin present in *Citrullus colocynthis*. This study was aimed to find anti-cancer activity that has been used for diabetic (24)(25). This plant is used as chemotherapeutic drugs for liver and cardiovascular (26). *Citrullus colocynthis* shown as antimicrobial and antifungal effect (27)(28). This plant fruit has cytotoxicity effects (27,29)(30). These are used in chromosomal damage (18)(31). This plant has anti cancer activity against hepatocarcinoma (Hep G2 cells line)(32)(33). It is used in monotherapy (6)(34). *Citrullus colocynthis* fruit is used in treatment of tooth decay (17). Cucurbitacin glycosides extracted from *Citrullus colocynthis* inhibits human breast cancer cell growth(22)(35). *Citrullus colocynthis* has antifungal and antioxidant properties (36). Beta sitosterol present in *Citrullus colocynthis* induces apoptosis in human colon cancer cells and anti-proliferating activity (37)(38) .

Citrullus colocynthis has an anticancer effect, antioxidant and anti metastatic effect(39). The expression of bax gene in *Citrullus colocynthis* and the combination of extract of phycocyanin has higher inhibitory effect on cancer cells (40) but in my research its was found that ethanolic leaf extract showed anticancerous effect on cancerous cell (41)(42) and also has found that *Citrullus lanatus* showed inhibited growth on human leukemia cancer cells (43) (44)but in my research we have used another herbal plants *Citrullus colocynthis* was taken has plant sampling and the study proves that *Citrullus colocynthis* also has anticancer effects. Through this methanolic extract of *Artemisia absinthium* it was found the anticancer effect on colorectal cancer HCT-116 cell line. My research was that ethanolic leaf extract shows not only anticancer effect but also anti inflammatory effect, antioxidant effect and this proves that traditional herbal plants have various medicinal values that can replace chemical drugs (45)(46). It has found that the phytochemical cucurbitacin present in *Citrullus colocynthis* has anti-inflammatory, antihypertensive, anticancer, antihypercholesterolemic and hepatoprotective effects and my research also proves presence of phytochemical cucurbitacin has ability to inhibit cancer cell. *Bryonia dioica* is an herbal plant through which aqueous extract induced apoptosis shows the therapeutic effect against breast cancer cells (47)(48). Limitations of the study are that the time period for the research was too short and Low sample size

Conclusion:

It is concluded that *Citrullus colocynthis* extract inhibits the growth of human breast cancer cells (MCF-7) by regulating the expression of pro apoptotic (Bax) and caspases-3 genes. Hence *Citrullus colocynthis* may be served as a potential drug for treatment of human breast cancer cells.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

REFERENCE:

1. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012 [Internet]. Vol. 65, CA: A Cancer Journal for Clinicians. 2015. p. 87–108. Available from: <http://dx.doi.org/10.3322/caac.21262>
2. Bharath B, Perinbam K, Devanesan S, AlSalhi MS, Saravanan M. Evaluation of the anticancer potential of Hexadecanoic acid from brown algae *Turbinaria ornata* on HT–29 colon cancer cells [Internet]. Vol. 1235, Journal of Molecular Structure. 2021. p. 130229. Available from: <http://dx.doi.org/10.1016/j.molstruc.2021.130229>
3. Balmain A, Gray J, Ponder B. The genetics and genomics of cancer. *Nat Genet.* 2003 Mar;33 Suppl:238–44.
4. Clarizia G, Bernardo P. Diverse Applications of Organic-Inorganic Nanocomposites: Emerging Research and Opportunities: Emerging Research and Opportunities. IGI Global; 2019. 237 p.
5. Feinberg AP, Tycko B. The history of cancer epigenetics. *Nat Rev Cancer.* 2004 Feb;4(2):143–53.
6. Boudjelal A, Henchiri C, Sari M, Sarri D, Hendel N, Benkhaled A, et al. Herbalists and wild medicinal plants in M'Sila (North Algeria): an ethnopharmacology survey. *J*

Ethnopharmacol. 2013 Jul 9;148(2):395–402.

7. Gambardella J, Trimarco B, Iaccarino G, Sorriento D. Cardiac Nonmyocyte Cell Functions and Crosstalks in Response to Cardiotoxic Drugs. *Oxid Med Cell Longev*. 2017 Oct 22;2017:1089359.
8. Egbuna C, Mishra AP, Goyal MR. Preparation of Phytopharmaceuticals for the Management of Disorders: The Development of Nutraceuticals and Traditional Medicine. Academic Press; 2020. 574 p.
9. Hirsch HA, Iliopoulos D, Tsiichlis PN, Struhl K. Metformin selectively targets cancer stem cells, and acts together with chemotherapy to block tumor growth and prolong remission. *Cancer Res*. 2009 Oct 1;69(19):7507–11.
10. Ezhilarasan D. Critical role of estrogen in the progression of chronic liver diseases. *Hepatobiliary Pancreat Dis Int*. 2020 Oct;19(5):429–34.
11. Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci U S A*. 2003 Apr 1;100(7):3983–8.
12. Barabadi H, Mojab F, Vahidi H, Marashi B, Talank N, Hosseini O, et al. Green synthesis, characterization, antibacterial and biofilm inhibitory activity of silver nanoparticles compared to commercial silver nanoparticles [Internet]. Vol. 129, *Inorganic Chemistry Communications*. 2021. p. 108647. Available from: <http://dx.doi.org/10.1016/j.inoche.2021.108647>
13. Dontu G, Al-Hajj M, Abdallah WM, Clarke MF, Wicha MS. Stem cells in normal breast development and breast cancer. *Cell Prolif*. 2003 Oct;36 Suppl 1:59–72.
14. Gowhari Shabgah A, Ezzatifar F, Aravindhyan S, Olegovna Zekiy A, Ahmadi M, Gheibihayat SM, et al. Shedding more light on the role of Midkine in hepatocellular carcinoma: New perspectives on diagnosis and therapy. *IUBMB Life*. 2021 Apr;73(4):659–69.
15. Alves-Silva JM, Romane A, Efferth T, Salgueiro L. North African Medicinal Plants Traditionally Used in Cancer Therapy. *Front Pharmacol*. 2017 Jun 26;8:383.
16. J PC, Pradeep CJ, Marimuthu T, Krithika C, Devadoss P, Kumar SM. Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study [Internet]. Vol. 20, *Clinical Implant Dentistry and Related Research*. 2018. p. 531–4. Available from: <http://dx.doi.org/10.1111/cid.12609>
17. Kafshgari HS, Yazdanian M, Ranjbar R, Tahmasebi E, Mirsaeed SRG, Tebyanian H, et al. The effect of *Citrullus colocynthis* extracts on *Streptococcus mutans*, *Candida albicans*, normal gingival fibroblast and breast cancer cells [Internet]. Vol. 92, *Journal of Biological Research - Bollettino della Società Italiana di Biologia Sperimentale*. 2019. Available from: <http://dx.doi.org/10.4081/jbr.2019.8201>

18. Shokrzadeh M, Chabra A, Naghshvar F, Ahmadi A. The mitigating effect of *Citrullus colocynthis* (L.) fruit extract against genotoxicity induced by cyclophosphamide in mice bone marrow cells. *ScientificWorldJournal*. 2013 Nov 7;2013:980480.
19. Al-Snafi AE. Immunological Effects of Medicinal Plants: A Review (Part 2) [Internet]. Vol. 16, *Immunology, Endocrine & Metabolic Agents in Medicinal Chemistry*. 2016. p. 100–21. Available from: <http://dx.doi.org/10.2174/1871522216666161014155814>
20. Chowdhury K, Sharma A, Kumar S, Gunjan GK, Nag A, Mandal CC. Colocynth Extracts Prevent Epithelial to Mesenchymal Transition and Stemness of Breast Cancer Cells. *Front Pharmacol*. 2017 Sep 5;8:593.
21. Kamath SM, Manjunath Kamath S, Jaison D, Rao SK, Sridhar K, Kasthuri N, et al. In vitro augmentation of chondrogenesis by Epigallocatechin gallate in primary Human chondrocytes - Sustained release model for cartilage regeneration [Internet]. Vol. 60, *Journal of Drug Delivery Science and Technology*. 2020. p. 101992. Available from: <http://dx.doi.org/10.1016/j.jddst.2020.101992>
22. Tannin-Spitz T, Grossman S, Dovrat S, Gottlieb HE, Bergman M. Growth inhibitory activity of cucurbitacin glucosides isolated from *Citrullus colocynthis* on human breast cancer cells. *Biochem Pharmacol*. 2007 Jan 1;73(1):56–67.
23. Shah BK, Mandal R. Survival trends in metastatic bladder cancer in the United States: a population based study. *J Cancer Res Ther*. 2015 Jan;11(1):124–8.
24. Zaini NAM, Anwar F, Hamid AA, Saari N. Kundur [*Benincasa hispida* (Thunb.) Cogn.]: A potential source for valuable nutrients and functional foods [Internet]. Vol. 44, *Food Research International*. 2011. p. 2368–76. Available from: <http://dx.doi.org/10.1016/j.foodres.2010.10.024>
25. Nambi G, Kamal W, Es S, Joshi S, Trivedi P. Spinal manipulation plus laser therapy versus laser therapy alone in the treatment of chronic non-specific low back pain: a randomized controlled study. *Eur J Phys Rehabil Med*. 2018 Dec;54(6):880–9.
26. King PD, Perry MC. Hepatotoxicity of Chemotherapy [Internet]. Vol. 6, *The Oncologist*. 2001. p. 162–76. Available from: <http://dx.doi.org/10.1634/theoncologist.6-2-162>
27. Marzouk B, Marzouk Z, Décor R, Edziri H, Haloui E, Fenina N, et al. Antibacterial and anticandidal screening of Tunisian *Citrullus colocynthis* Schrad. from Medenine [Internet]. Vol. 125, *Journal of Ethnopharmacology*. 2009. p. 344–9. Available from: <http://dx.doi.org/10.1016/j.jep.2009.04.025>
28. Mudigonda SK, Murugan S, Velavan K, Thulasiraman S, Krishna Kumar Raja VB. Non-suturing microvascular anastomosis in maxillofacial reconstruction- a comparative study. *Journal of Cranio-Maxillofacial Surgery*. 2020 Jun 1;48(6):599–606.
29. Hedberg I, Staugård F. *Traditional Medicinal Plants*. 1989. 324 p.

30. Prakash AKS, Devaraj E. Cytotoxic potentials of *S. cumini* methanolic seed kernel extract in human hepatoma HepG2 cells [Internet]. Vol. 34, Environmental Toxicology. 2019. p. 1313–9. Available from: <http://dx.doi.org/10.1002/tox.22832>
31. Rajakumari R, Volova T, Oluwafemi OS, Rajesh Kumar S, Thomas S, Kalarikkal N. Grape seed extract-soluplus dispersion and its antioxidant activity. Drug Dev Ind Pharm. 2020 Aug;46(8):1219–29.
32. I DOO, Ochwang I DO, Kimwele CN, Oduma JA. Phytochemical Screening of Medicinal Plants of the Kakamega Country, Kenya Commonly Used against Cancer [Internet]. Vol. 05, Medicinal & Aromatic Plants. 2016. Available from: <http://dx.doi.org/10.4172/2167-0412.1000277>
33. R H, Hannah R, Ramani P, Ramanathan A, Jancy MR, Gheena S, et al. CYP2 C9 polymorphism among patients with oral squamous cell carcinoma and its role in altering the metabolism of benzo[a]pyrene [Internet]. Vol. 130, Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology. 2020. p. 306–12. Available from: <http://dx.doi.org/10.1016/j.oooo.2020.06.021>
34. Santhakumar P, Roy A, Mohanraj KG, Jayaraman S, Durairaj R. Ethanolic Extract of Capparis decidua Fruit Ameliorates Methotrexate-Induced Hepatotoxicity by Activating Nrf2/HO-1 and PPAR γ Mediated Pathways. Ind J Pharm Educ. 2021 Mar 19;55(1s):s265–74.
35. Wadhwa R, Paudel KR, Chin LH, Hon CM, Madheswaran T, Gupta G, et al. Anti-inflammatory and anticancer activities of Naringenin-loaded liquid crystalline nanoparticles in vitro. J Food Biochem. 2021 Jan;45(1):e13572.
36. Hadizadeh I, Peivastegan B, Kolahi M. Antifungal activity of nettle (*Urtica dioica* L.), colocynth (*Citrullus colocynthis* L. Schrad), oleander (*Nerium oleander* L.) and konar (*Ziziphus spina-christi* L.) extracts on plants pathogenic fungi. Pak J Biol Sci. 2009 Jan 1;12(1):58–63.
37. Choi YH, Kong KR, Kim Y-A, Jung K-O, Kil J-H, Rhee S-H, et al. Induction of Bax and activation of caspases during beta-sitosterol-mediated apoptosis in human colon cancer cells. Int J Oncol. 2003 Dec;23(6):1657–62.
38. Wahab PUA, Abdul Wahab PU, Madhulaxmi M, Senthilnathan P, Muthusekhar MR, Vohra Y, et al. Scalpel Versus Diathermy in Wound Healing After Mucosal Incisions: A Split-Mouth Study [Internet]. Vol. 76, Journal of Oral and Maxillofacial Surgery. 2018. p. 1160–4. Available from: <http://dx.doi.org/10.1016/j.joms.2017.12.020>
39. Abdulridha MK, Al-Marzoqi A-H, Ghasemian A. The Anticancer Efficiency of *Citrullus colocynthis* Toward the Colorectal Cancer Therapy. J Gastrointest Cancer. 2020 Jun;51(2):439–44.
40. Hamdan NT, Jwad BAAA, Jasim SA. Synergistic anticancer effects of phycocyanin and *Citrullus colocynthis* extract against WiDr, HCT-15 and HCT-116 colon cancer cell lines

[Internet]. Vol. 22, Gene Reports. 2021. p. 100972. Available from: <http://dx.doi.org/10.1016/j.genrep.2020.100972>

41. Itoh T, Ono A, Kawaguchi K, Teraoka S, Harada M, Sumi K, et al. Phytol isolated from watermelon (*Citrullus lanatus*) sprouts induces cell death in human T-lymphoid cell line Jurkat cells via S-phase cell cycle arrest. *Food Chem Toxicol*. 2018 May;115:425–35.
42. Saraswathi I, Saikarthik J, Senthil Kumar K, Madhan Srinivasan K, Ardhanaari M, Gunapriya R. Impact of COVID-19 outbreak on the mental health status of undergraduate medical students in a COVID-19 treating medical college: a prospective longitudinal study. *PeerJ*. 2020 Oct 16;8:e10164.
43. Nazeri M, Mirzaie-Asl A, Saidijam M, Moradi M. Methanolic extract of *Artemisia absinthium* prompts apoptosis, enhancing expression of Bax/Bcl-2 ratio, cell cycle arrest, caspase-3 activation and mitochondrial membrane potential destruction in human colorectal cancer HCT-116 cells. *Mol Biol Rep*. 2020 Nov;47(11):8831–40.
44. Vivekanandhan K, Shanmugam P, Barabadi H, Arumugam V, Raj DDRD, Sivasubramanian M, et al. Emerging Therapeutic Approaches to Combat COVID-19: Present Status and Future Perspectives [Internet]. Vol. 8, *Frontiers in Molecular Biosciences*. 2021. Available from: <http://dx.doi.org/10.3389/fmolb.2021.604447>
45. Gacem MA, El Hadj-Khelil AO, Boudjemaa B, Gacem H. Phytochemistry, Toxicity and Pharmacology of *Pistacia lentiscus*, *Artemisia herba-alba* and *Citrullus colocynthis* [Internet]. *Sustainable Agriculture Reviews* 39. 2020. p. 57–93. Available from: http://dx.doi.org/10.1007/978-3-030-38881-2_3
46. Sridharan G, Ramani P, Patankar S, Vijayaraghavan R. Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma. *J Oral Pathol Med*. 2019 Apr;48(4):299–306.
47. Benarba B, Elmallah A, Pandiella A. *Bryonia dioica* aqueous extract induces apoptosis and G2/M cell cycle arrest in MDA- MB 231 breast cancer cells. *Mol Med Rep*. 2019 Jul;20(1):73–80.
48. Tahmasebi S, Qasim MT, Krivenkova MV, Zekiy AO, Thangavelu L, Aravindhan S, et al. The effects of oxygen-ozone therapy on regulatory T-cell responses in multiple sclerosis patients. *Cell Biol Int*. 2021 Jul;45(7):1498–509.