

Alteration of Some Sperm Proteins, Antioxidants and Prostate-Specific Antigen in the Seminal Plasma of Bacteriospermic Infertile Males

ABSTRACT

Aims: To investigate the alteration of some sperm proteins, antioxidants and prostate-specific antigen in the seminal plasma of bacteriospermic infertile males.

Study design: The study is a case-control design to investigate the semen parameters and sperm proteins in infertile males with bacteriospermia attending urology or fertility clinics in Port Harcourt, Rivers State.

Place and Duration of Study: The study was carried out in the Department of Medical Laboratory Science, Rivers State University, Port Harcourt. However, some of the Laboratory investigations were done in the Chemical Pathology Unit and Medical Microbiology Unit of University of Port Harcourt Teaching Hospital, Port Harcourt. The Study was between the periods of September, 2019 and Feb., 2022.

Methodology: A total of 276 males indicated interest to participate in the study of which 192 male subjects were recruited. Of the 192 subjects recruited, 76 were fertile without bacterial infection, 24 were fertile with bacterial infection, 37 infertile without bacterial infection and 56 infertile with bacterial infection. Infertile males were further classified as asthenozoospermic, oligoasthenozoospermic, teratozoospermic, asthenoteratozoospermic, and oligoasthenoteratozoospermic. HSP70, PKA, and MDA, TAC and GPX (in seminal plasma), PSA and testosterone (in serum) were analysed using ELISA while TAC was done using spectrophotometric methods. Microbial culture was done to isolate and identify bacteria in semen. Results obtained were analysed using GraphPad Prism, Excel and SPSS.

Results: Culture results indicated that 52% of normospermic subjects had no semen infection at the time of investigation. On the other hand, infertile male subjects without semen infections had 18.7%, while infertile males with semen infection were seen to be 29.2%. Likewise, subjects with bacteriospermia as scanty growth had 12.5%, 9.3% moderate, and 7.3% heavy growth. Significantly higher values of HSP70, PKA, PSA and MDA were seen in infertile males with bacteriospermia compared control and infertile males without infection while significantly lower values were seen in TAC, GPX, OPN and testosterone were observed in the infertile males with bacteriospermia compared against control and infertile males without infection..

Conclusion: Conclusively, from our results bacteriospermia significantly increased lipid peroxidation cum oxidative stress leading to altered physiological sperm proteins in the seminal plasma. These alterations could also be responsible for the poor sperm qualities observed in the infertile subjects.

Keywords: Sperm Proteins, Bacterial infection, Male infertility, Port Harcourt, Niger Delta

1. INTRODUCTION

Accurate assessment of male factor infertility is quite difficult due to multi-factorial etiologies or risks as well as due to limited information on male infertility in population. Male factor infertility in Nigeria accounts for up to 50% of all infertility cases [1]. According to the international committee for Monitoring Assisted Reproductive Technology and the World Health Organization (WHO), infertility is a disease of the reproductive system defined by failure to achieve clinical pregnancy after 12 months or more of regular unprotected sexual

intercourse [2]. In the African society, infertility comes with psychological stress and trauma due to strong emphasis on child bearing [2, 3].

Etiologies such as reduced number of spermatozoa (oligozoospermia), reduced sperm motility (asthenozoospermia), reduced sperm vitality (necrozoospermia), abnormal sperm morphology (teratozoospermia) or any combination of these have been reported to induce male infertility [4, 5, 6]. More so, several factors such as genetic factors (deletion, inversion, mutation, aneuploidy, congenital abnormality (cryptorchidism) and translocation), environmental factors (exposure to chemical contaminants which are endocrine disruptors), oxidative stress, life Style (age, obesity, smoking, consumption of alcohol, use of recreational drugs) and bacterial infections have also been implicated in male infertility [1, 2, 7, 8, 9].

Bacterial infection has been observed as one of the major risks of infertility in couples. In males, asymptomatic or symptomatic bacterial infections in the seminal plasma has been reported to affect sperm parameters, viability, and motility resulting in male factor infertility. Several micro-organisms such as Mumps Orchitis, *Chlamydia Trachomatis*, *Streptococcus* species, *Staphylococcus aureus*, *Escherichia coli*, *pseudomonas* sp, and soon have been implicated in male-factor infertility [10, 11]. However, in this study, our interest is on bacterial infection in seminal plasma. Bacterial infection of the seminal plasma has been reported to contribute greatly in reducing sperm count, motility, speed, sperm quality, and playing critical role in inducing morphological derangement such as asthenospermia, and teratospermia thus resulting in the suppression of fertility capacity [1, 11].

In recent times, male factor infertility has been strongly linked with some sperm proteins expression or reduction/absence of such proteins. Therefore in this work, the influence of bacterial infections on some sperm proteins associated with male infertility such as osteopontin (OPN), Heat Shock Protein 70 (HSP70), Protein Kinase A (PKA), and antioxidants such glutathione peroxidase (GPX) and total Antioxidant capacity (TAC) of sperm cells will be our focus. These proteins have been reported to be involved in acrosomic reactions, maintenance of sperm–Oocyte interaction, promote cell survival against environmental stress such as increased scrotal temperature, spermatogenesis, enhanced motility of sperm cells, and in the elimination of reactive oxygen species (ROS) in the seminal plasma.

2. MATERIAL AND METHODS

2.1 Experimental Design and Subject Classification

The study is a case-control design to investigate the semen parameters and sperm proteins in infertile males with bacterial infections attending urology or fertility clinics in Port Harcourt, Rivers State. A total of 276 males indicated interest to participate in the study of which 192 male subjects were recruited. Of the 192 subjects recruited, 76 were fertile without bacterial infection, 24 were fertile with bacterial infection, 37 infertile without bacterial infection and 56 infertile with bacterial infection. Infertile males (case) with progressive motility $\leq 50\%$ and normal morphology $>30\% \leq 50\%$ were classified as asthenozoospermic. Those with progressive motility $\leq 50\%$, normal morphology $>30\% \leq 50\%$ and total count of $\leq 19 \times 10^6/\text{ml}$ were grouped as oligoasthenozoospermic. In addition, those with normal morphology of $\leq 30\%$, progressive motility $\geq 50\%$ were grouped as teratozoospermic while those with normal morphology $\leq 30\%$, progressive motility $\leq 50\%$ were grouped as asthenoteratozoospermic. Finally, infertile males with total count of $< 19 \times 10^6$ cells/ml, normal morphology of $< 30\%$ with progressive motility $< 50\%$ were grouped as oligoasthenoteratozoospermic. Also their semen leukocyte concentration was $< 1 \times 10^6/\text{ml}$ of ejaculate.

2.2 Sample Size Determination

Sample size was determined using the equation as described by Claran & Biswas [12] with precision or absolute error of 5% and 95% confidence interval using the equation $\text{Sample size} = Z^2 P(1-P)/d^2$. A total of 93 was calculated as sample size with the expected proportion of male-factor infertility of 6.5% as reported Sule et al. [13].

2.3 Inclusion and exclusion Criteria

A well-structured questionnaire was issued to all the participants to obtain demographic information, medical history and lifestyle after obtaining consent of participants.

2.3.1 Inclusion Criteria

Subjects used in this study are those attending urology/fertility clinics, without history of hypertension, cardiovascular disorders, osteoporosis, diabetes mellitus or smoking. Omron digital blood pressure kit (Omron healthcare co., Ltd, Japan) was used to check blood pressures while the glucose oxidase method was used to determine their diabetic status. The control subjects were males that have established pregnancies within the period of this study with total sperm count of $\geq 20 \times 10^6$ cells/ml, normal sperm morphology and without bacterial infection. The cases were infertile males that have failed to establish pregnancies after two years, been clinically diagnosed of infertility, and with or without bacterial infections. These subjects also have sperm count between $0 - 19 \times 10^6$ cells/ml.

2.3.2 Exclusion Criteria

Those excluded from the study include subjects that did not give consent, had semen leukocytes (pus cells) $>1 \times 10^6$ /ml, and semen specimen collected without masturbation. More so, subjects that did not abstain for at least 72 hours (3 days) and lifestyle like smoking, alcohol consumption were also excluded. In addition, those with history of hypertension, cardiovascular disorders, osteoporosis, diabetes mellitus and prostate specific antigen (PSA) of $>10\text{ng/ml}$ were also excluded.

2.4 Specimens Sampling and Preparation

2.5.1 Blood and Semen Sampling

Following ethical approval, blood and semen samples were collected from eligible subjects. All semen samples were collected into universal sterile plastic containers by masturbation after an abstinence period of 72 hours. In addition, 5ml of venous blood samples were collected on fasting state from all the participants into plain bottles.

2.5.2 Sample Preparation

Semen samples collected into universal sterile plastic containers were placed on the bench at room temperature of 25°C for 40 minutes to liquifaction before analyses (culture, macroscopic, and microscopic) were performed within 1 hr of collection. Subsequently, the liquefied semen samples were centrifuged at 4500rpm for 15 minutes as described by Conquer et al. [14]. The supernatant seminal plasma was then carefully aliquoted into eppendorf tubes and frozen at -70°C pending biochemical analysis. In terms of the blood samples, the whole blood samples were collected into plain bottles and allowed to stand for 30 minutes to clot. Clotted blood samples were retracted and spun at 3500rpm for 10 minutes to obtain serum for the estimation of testosterone and total prostate-specific antigen.

2.6 Equipment and Reagents

Equipment used includes Olympus binocular microscope, Neubauer haemocytometer, and Auto Elisa P microplate reader (Labtech). Other equipment used includes merment oven, universal bucket centrifuge model 320, Omron digital blood pressure kit (Omron healthcare

co., Ltd, Japan), Haier thermocool deep freezer (China), pipettes and microscopic glass slides and cover slips. Reagents used include Osteopontin (OSP), Heat shock protein 70 (HSP70), and Protein-kinase A (PKA), Malondialdehyde (MDA), Prostate-Specific Antigen (PSA), and testosterone (Testo) enzyme linked immunosorbent assay (ELISA) kits purchased from Bioassay Technology Laboratory (Shanghai, China). Glutathione peroxidase (GPX) ELISA kits and glucose oxidase reagent, as well as total antioxidant capacity (TAC) spectrophotometric kits were purchased from Elabscience (Houston, Texas, USA) and Fortress Diagnostics (Antrim, United Kingdom) respectively.

2.4 Laboratory Analysis of Sperm Parameters and Biochemical Parameters in Seminal Plasma and Serum

2.6.1 Determination of Sperm Volume, pH and Viscosity

The semen ejaculatory volume was determined using graduated glass pasteur pipette while the pH of the semen was determined using combi-9 strip, dipped into the semen and matched against the combi-9 colour chart. For the determination of viscosity was done on the liquefied semen specimens. It was performed by observing the slimy thread formed when a glass rod is placed on semen specimen and withdrawn from the specimen. The length of the slimy threads not less than 0.5cm but not exceeding 2cm was considered normal as described by Vasan [15].

2.6.2 Determination of Sperm Concentration

The sperm concentration (count) was manually determined using Neubauer cell counter. The semen specimens were mixed and diluted 1 in 20 using semen diluting fluid consisting of sodium bicarbonate, formalin and distilled water. The cell counting chamber was charged with the diluted specimen and allowed to settle for 15 minutes. The sperm count was made in the four secondary squares and five tertiary squares of the central secondary squares as described by WHO [16] and Ochei [17]. The total sperm concentration per ml was estimated by multiplying the number of sperm cells counted x dilution factor (20) x multiplication factor (50,000).

2.6.3 Microscopic Evaluation of the Sperm Cells

A wet preparation was made by placing a drop of semen with a pasteur pipette from a well-mixed sample on a clean grease-free slide and covering it with a cover slip. The wet preparation microscopic fields were examined under high powered objectives using x40 objective lens to estimate and quantitate the presence of pus cells, epithelial cells and also to determine the motility, morphology, and viability of the sperm cells as described by WHO [16]. At least a hundred cells were examined at a magnification of x1000, to determine the percentage sperm cell motility as described by WHO [16]. In addition, the morphology of the sperm cells was determined using the methylene blue-eosin staining technique after incubation at 25⁰C with trypsin for 10minutes.

2.6.4 Determination of Bacterial Quality and Identification

The presence and degree of bacterial load in the semen specimens was determined using culture technique. The culture media used were macConkey and blood agar. Loopful of semen specimens were collected under sterile conditions and inoculated into the culture plates and incubated at 37⁰C for 48 hours. After the period of incubation, bacteria were isolated and identified biochemically.

2.7 Assay of Biochemical Parameters

Heat Shock Protein 70 (HSP70), Osteopontin (OPN) and Protein Kinase A (PKA), Glutathione Peroxidase (GPX), testosterone, and Prostate-Specific Antigen (PSA) were determined as described by Engvall & Perlmann [18] ELISA Quantitative Assay Method

while Malondialdehyde (MDA) was determined using ELISA method as described by Moron et al. [19]. More so, Total Anti-oxidant Capacity (TAC) was done using Colorimetric Method as Described by Mencci et al. [20].

2.8 Statistical Analysis

Statistical package used for data analysis were Graphpad Prism 8.0.2 (California, USA), Statistical Package for Social Science (SPSS) version 23.0, and Microsoft Excel version 2018. Descriptive statistics used were mean and standard deviation while inferential statistics used include students't-test, Chi-Square, One-Way ANOVA with Post-Hoc done with Tukey's multiple comparison analysis tests, Pearson's correlation analysis and Odd ratio. Results were presented as Mean±Standard Deviation. Statistical significance was set at $p < 0.05$.

3. RESULTS

3.1 Results on Prevalence and Microscopic Characteristics of Semen/Sperm Cells

Culture results indicated that 52% of normospermic subjects had no semen infection at the time of investigation. However, during interview, 39.6% reported to have never had semen infection 52.1% while 12.5% reported that they have been previously had semen infection and treated accordingly. On the other hand, infertile male subjects without semen infections had 18.7%, while infertile males with semen infection were seen to be 29.2%. The Chi-square results also indicate no significant difference for both the various age intervals (X-square=1, $p=0.607$) and infertile males (X-square=2, $p=0.572$). Likewise, subjects without any microbial infection had frequency of 70.8% those with scanty growth had 12.5%, moderate had 9.3% and heavy growth had 7.3%. The Chi-square results also indicate no significant difference (X-square=2, $p=0.572$). The comparison of their respective % distribution also indicated no significant differences at $p < 0.05$ (table 1).

Table 1: Microscopic Characteristics of Semen/Sperm Cells

Subjects/Specimens Characteristics	No	Frequency	% Frequency	Chi-Square	Pvalue
Microbial Quality (Culture)					
a. Fertile males, no growth	76	0.396 } 0.125 } 0.187 } 0.292 }	39.6 } 12.5 } 18.7 } 29.2 }	2.00	0.572
b. Fertile Males, infected before	24				
c. Infertile males, no growth	36				
d. Infertile males, with growth	56				
Bacteria Isolated & Identified					
a. <i>Staphylococcus aureus</i>	35	0.182	18.2	3.333	0.504
b. <i>Escherichia coli</i>	10	0.056	5.6		
c. <i>Klebsiella Pneumoniae</i>	4	0.020	2.0		
d. <i>Pseudomonas aeruginosa</i>	2	0.0104	1.04		
e. Mixed <i>S. aureus</i> + <i>E. coli</i>	5	0.026	2.6		
Degree of Microbial Quality					
a. No Growth	136	0.708	70.8	2.00	0.572
b. Scanty	24	0.125	12.5		
c. Moderate	18	0.093	9.3		
d. Heavy	14	0.073	7.3		

3.2 Results of Sperm Parameters and total sperm count in Infertile Males with Microbial Growth

When infertile male with varying degree (scanty, moderate, and heavy) of bacterial infections were investigated, the results also indicated no significant differences in age, sperm volume, and pH. Although, significantly lower values were seen in % motility and %Active motility when control were compared with infertile males without infection, those with scanty, moderate, and heavy bacterial infections. Also, significantly lower values were seen in %normal sperm morphology of infertile male with moderate and heavy bacterial infections compared with normospermic, infertile males without infection and infertile males with scanty infections. In addition, significantly lower values of testosterone were seen in infertile males without bacterial infection, scanty, moderate and heavy growth. However, infertile male subjects without infections had significantly higher value of testosterone compared against infertile scanty, moderate, and heavy bacterial infections at $p < 0.05$ (Table 2).

3.3 Results of seminal Plasma Proteins, Anti-Oxidants Enzymes, MDA, Testosterone, and PSA in Infertile Males with Microbial Growth

When sperm proteins in seminal plasma were considered in infertile male without or with infections significantly higher values of HP70, PKA, and MDA were seen in the infertile males without infections as well as in the infertile male with infections compared against control. However, no significant differences were between infertile male without infections and infertile males with infection. In PSA, significantly higher values were seen in infertile males with bacterial infections compared control and infertile males without infection. However, no significant differences were seen in PSA values between control and infertile males without infection. Furthermore, significantly lower values were seen in TAC and Testosterone of infertile males without infection and infertile male with infection when compared with control subjects. There was also a significant lower value of TAC in infertile males with infections compared against infertile subjects without infection. However, in OSP and GPX the significantly lower values were only seen in infertile males with bacterial infections when compared with control subjects and infertile males without infections (Table 3).

3.4 Results of seminal Plasma Proteins, Anti-Oxidants Enzymes, MDA, Testosterone, and PSA in Infertile Males with Varying Degree of Microbial Growth

When some sperm proteins in seminal plasma were considered in fertile (control) and infertile subjects with or without bacterial infection of the semen, the results of sperm proteins in seminal plasma indicated significantly lower values in HSP70, PKA, and MDA of normospermic (control, fertile) subjects compared against infertile subjects with scanty, moderate and heavy bacterial infections. However, in PSA, the significantly higher values were observed in the infertile subjects with heavy bacterial growth compared against the normospermic fertile subjects. No significance differences were seen between normospermic subjects and infertile subjects without infection, infertile subject with scanty or moderate bacterial infections. Furthermore, in MDA significantly higher values were seen in infertile subjects with heavy bacterial infection compared against those with scanty and moderate infection. Likewise, those with scanty or moderate infection had significantly higher values of MDA compared against those that are infertile without infection.

In addition, significantly lower values were seen in OSP, GPX, TAC, and testosterone of infertile subjects with scanty, moderate and heavy bacterial infections compared against fertile normospermic subjects without seminal fluid infection at $p < 0.05$ (Table 4). However, in TAC, significantly lowest values were observed in the infertile subjects with scanty bacterial infections when compared infertile subjects without infections at $p < 0.05$ (Table 4).

Table 2: One-Way ANOVA of Sperm parameters in Infertile Males with Varying Degree of Microbial Growth

Parameters	Control (No MG)	Infertile males (No MG)	Infertile Males (SMG)	Infertile Males (MMG)	Infertile Males (HMG)	Fvalue	pvalue	Remark
Age (years)	39.22±7.028	36.67±5.750	41.83±7.39	38.33±6.93	37.00±6.29	0.731	0.5730	NS
Sperm vol. (mL)	2.63±1.51	2.14±0.99	2.68±1.09	2.24±1.37	2.10±1.24	0.376	0.8252	NS
pH	7.95±0.28	7.97±0.12	7.88±0.23	7.94±0.30	7.92±0.34	0.281	0.8890	NS
% Motility	60.51±17.33 ^a	28.00±17.63 ^b	31.25±22.71 ^b	28.89±16.16 ^b	26.71±21.34 ^b	13.13	<0.0001	S
% Active Motility	45.86±21.59 ^a	17.28±11.27 ^b	21.92±18.33 ^b	15.00±13.23 ^b	15.71±15.39 ^b	9.468	<0.0001	S
% Norm. Morph.	82.55±25.21 ^a	64.50±37.78 ^a	75.00±32.86 ^a	44.33±31.04 ^b	28.86±21.06 ^c	10.03	<0.0001	S
% Ab. Morph	17.49±25.71 ^a	35.50±37.78 ^a	25.00±32.86 ^a	55.67±31.04 ^b	71.14±21.06 ^c	9.606	<0.0001	S
TSC (10 ⁶ cells/mL)	42.53±19.75 ^a	16.56±6.519 ^b	9.975±4.621 ^b	8.656±7.385 ^b	4.129±3.324 ^c	24.79	<0.0001	S

Turkey's Post Hoc: Within same row, values with different superscripts differ significantly at p<0.05.

No MG= No Microbial Growth, SMG=Scanty Microbial Growth, MMG=Moderate Microbial Growth, HMG=Heavy Microbial Growth on culture media, TSC=Total Sperm Count.

S= Significant, NS=Not Significant at p<0.05.

UNDER PEER REVIEW

Table 3: Results of One-Way ANOVA of Seminal Plasma Proteins, Anti-Oxidants Enzymes, MDA, Testosterone, and PSA in Infertile Males with Microbial Growth

Parameters	Control (No MG)	Infertile males (No MG)	Infertile males (MG)	Fvalue	pvalue	Remark
HSP70 (ng/ml)	12.17±4.59 ^a	15.77±3.51 ^b	16.47±7.48 ^b	6.554	0.0022	S
OPN (ng/ml)	4.39±2.14 ^a	3.15±1.27 ^b	3.05±1.27 ^b	6.428	0.0024	S
PKA (ng/ml)	7.54±3.04 ^a	9.73±2.14 ^b	9.67±4.19 ^b	5.093	0.0080	S
GPX (ng/ml)	8.32±1.07 ^a	7.707±1.77 ^a	6.16±1.79 ^b	19.83	<0.0001	S
MDA (ng/ml)	52.15±17.29 ^a	70.14±37.05 ^a	102.6±36.74 ^b	28.14	<0.0001	S
TAC (mmol/L)	7.49±2.0 ^a	4.49±3.07 ^b	2.16±0.78 ^c	65.11	<0.0001	S
TESTO (ng/ml)	4.91±1.41 ^a	3.06±1.58 ^b	2.55±1.10 ^b	30.55	<0.0001	S
PSA (ng/ml)	4.17±3.07 ^a	4.82±3.0 ^{ac}	5.96±2.59 ^{bc}	3.390	0.0380	S

Turkey's Post Hoc: Within same row, values with different superscripts differ significantly at p<0.05. No MG= No Microbial Growth.

S= Significant at p<0.05.

3.5 One-Way ANOVA of seminal Plasma Proteins, Anti-Oxidants Enzymes, MDA, Testosterone, and PSA in Infertile Males with Microbial Growth and Sperm Cells Deformities

More so, when some sperm proteins in seminal plasma were considered in infertile subjects with bacterial infection of the semen and subjects sperm cell deformities were statistically analyzed, the ANOVA results of sperm proteins in seminal plasma indicated significantly higher values were seen in HSP70, PKA, PSA, and MDA of infertile subjects with scanty, moderate and heavy bacterial infections of the semen and those with sperm cells deformities compared against normospermic (control, fertile) subjects. However, the MDA values with significantly highest values were seen in oligoasthenoteratozoospermic (OAT) subjects compared against infertile subjects with varying degree of seminal plasma infections as well as those with sperm cells deformities such as asthenozoospermic, oligoasthenozoospermic, teratozoospermic and asthenoteratozoospermic at p<0.05

Also, significantly lower values were seen in OSP, GPX, TAC, and testosterone of infertile subjects with scanty, moderate and heavy bacterial infections of the semen as well as subjects with sperm cells deformities such as asthenozoospermic, oligoasthenozoospermic, teratozoospermic, asthenoteratozoospermic and oligoasthenoteratozoospermic when compared against fertile normospermic subjects without seminal fluid infection at p<0.05 (Table 5).

Table 4: One-Way ANOVA of seminal Plasma Proteins, Anti-Oxidants Enzymes, MDA, Testosterone, and PSA in Infertile Males with Varying Degree of Microbial Growth

Parameters	Control (No MG)	Infertile males (No MG)	Infertile Males (SMG)	Infertile Males (MMG)	Infertile Males (HMG)	Fvalue	pvalue	Remark
HSP70 (ng/ml)	12.17±4.59 ^a	15.77±3.51 ^b	18.73±10.48 ^b	18.22±4.62 ^b	20.70±2.90 ^b	7.545	<0.0001	S
OPN (ng/ml)	4.39±2.14 ^a	3.15±1.27 ^b	2.62±0.74 ^b	2.40±0.45 ^b	2.643±0.72 ^b	5.705	0.0004	S
PKA (ng/ml)	7.54±3.04 ^a	9.73±2.14 ^b	11.13±2.66 ^b	11.18±4.02 ^b	12.07±3.67 ^b	7.471	0.0001	S
GPX (ng/ml)	8.32±1.07 ^a	7.707±1.77 ^a	7.02±1.52 ^b	5.74±1.79 ^b	4.21±1.0 ^b	19.08	<0.0001	S
MDA (ng/ml)	52.15±17.29 ^a	70.14±37.05 ^a	103.4±38.97 ^b	80.19±17.17 ^b	134.8±28.78 ^c	21.59	<0.0001	S
TAC (mmol/L)	7.49±2.0 ^a	4.49±3.07 ^{bc}	1.79±0.81 ^{bd}	2.60±0.73 ^b	2.36±0.57 ^b	31.93	<0.0001	S
TESTO (ng/ml)	4.91±1.41 ^a	3.06±1.58 ^b	2.89±1.15 ^b	2.81±0.91 ^b	2.04±0.82 ^b	15.08	<0.0001	S
PSA (ng/ml)	4.17±3.07 ^a	4.82±3.0 ^a	5.84±2.45 ^a	5.91±2.56 ^a	7.84±1.83 ^b	3.238	0.0157	S

Turkey's Post Hoc:

Within same row, values with different suoverscripts differ significantly at p<0.05.

No MG= No Microbial Growth, SMG=Scanty Microbial Growth, MMG=Moderate Microbial Growth, HMG=Heavy Microbial Growth on culture media, TSC=Total Sperm Count.
S= Significant, NS=Not Significant at p<0.05.

Table 5: One-Way ANOVA of seminal Plasma Proteins, Anti-Oxidants Enzymes, MDA, Testosterone, and PSA in Infertile Males with Microbial Growth and Sperm Cells Deformities

Parameters	Control (No MG)	Infertile males (No MG)	Infertile males (MG)	TZS	ATS	OAT	Fvalue	pvalue	Remark
HSP70 (ng/ml)	12.17±4.59 ^a	15.77±3.51 ^b	16.47±7.48 ^b	18.11±3.71 ^b	15.78±4.11 ^b	18.20±3.98 ^b	9.335	<0.0001	S
OPN (ng/ml)	4.39±2.14 ^a	3.15±1.27 ^b	3.05±1.27 ^b	2.52±0.76 ^b	2.59±0.62 ^b	2.27±0.87 ^b	11.28	<0.0001	S
PKA (ng/ml)	7.54±3.04 ^a	9.73±2.14 ^b	9.67±4.19 ^b	10.34±3.11 ^b	10.60±2.77 ^b	13.57±3.21 ^b	9.071	<0.0001	S
GPX (ng/ml)	8.32±1.07 ^a	7.707±1.77 ^b	6.16±1.79 ^b	4.79±1.25 ^b	4.56±0.97 ^b	4.35±0.86 ^b	37.50	<0.0001	S
MDA (ng/ml)	52.15±17.29 ^a	70.14±37.05 ^b	102.6±36.74 ^b	109.2±33.65 ^b	83.44±31.63 ^b	129.1±29.74 ^c	20.17	<0.0001	S
TAC (mmol/L)	7.49±2.0 ^a	4.49±3.07 ^b	2.16±0.78 ^b	2.21±1.14 ^b	2.43±1.23 ^b	1.331±0.27 ^c	62.25	<0.0001	S
TESTO (ng/ml)	4.91±1.41 ^a	3.06±1.58 ^b	2.55±1.10 ^b	2.30±1.14 ^b	2.08±0.96 ^b	1.96±0.69 ^b	31.36	<0.0001	S
PSA (ng/ml)	4.17±3.07 ^a	4.82±3.0 ^a	5.96±2.59 ^b	5.82±2.75 ^b	6.49±2.71 ^b	7.67±2.13 ^b	3.845	0.0028	S

Turkey's Post Hoc:

Within same row, values with different superscripts differ significantly at p<0.05.

No MG= No Microbial Growth, SMG=Scanty Microbial Growth, MMG=Moderate Microbial Growth, HMG=Heavy Microbial Growth on culture media, TSC=Total Sperm Count.
S= Significant, NS=Not Significant at p<0.05.

4. DISCUSSION

Bacterial infections in seminal plasma after culture indicated prevalence of 29.2 % in infertile males of which 12.5%, 9.3% and 7.3% indicate scanty, moderate, and heavy bacterial growth. The organisms isolated and identified were *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Escherichia coli* and *Pseudomonas aeruginosa*. Of the organisms, *Staphylococcus aureus* had the highest prevalence with 18.2%, followed by *Escherichia coli* with 5.2%, *Klebsiella pneumoniae* with prevalence of 2.0%, and *Pseudomonas aeruginosa* with 1.04% as well as mixed growth of *Staphylococcus aureus* and *Escherichia coli* with 2.6%. Our findings are similar to the observations made in Port Harcourt by Oyeyipo et al. [21]. They also reported microbial infections (and prevalence) such as *Staphylococcus aureus* (30%), *Klebsiella pneumoniae* (10%), *Escherichiacoli* (2%), *Proteus mirabilis* (10%), and *Pseudomonas aeruginosa* (7%) isolated from seminal plasma.

The high prevalence of *Staphylococcus aureus* could be attributed to normal flora contamination of the glans penis or orifice of the urethral opening. However, their heavy presence (*Staphylococcus aureus*) in seminal plasma alongside scanty or moderate growth of *Escherichia coli* observed as mixed growth could suggest actual pathogenic bacterial infection of the seminal plasma with significance. Seminal plasma infection has contributed to male infertility affecting sperm cells viability and motility.

The non-significant differences observed in sperm volume and pH of the control subjects compared against the infertile subjects with bacterial infection (table 2) is in line with the findings of Palani & Alahmar [22]. They also reported no significant difference in the ejaculatory volume of semen of bacterial infected infertile male and control. However, the significantly lower values in sperm motility, active (progressive) motility, normal morphology and total sperm counts were observed in the infertile subjects with bacterial infection (table 2) are similar with previous works done in Port Harcourt by Oyeyipo et. al. [21] and Green & Nwachuku [6], they also gave similar findings as seen in our results in their respective studies. This further reveals that sperm volume and pH are never good indicators of microbial infections inspite of the severity of the infections, significantly lowered total sperm counts, sperm motility, active (progressive) motility, and morphological abnormalities.

When the degree of infection were considered, significant differences were not seen in the motility, and active motility amongst those infertile male with scanty, moderate and heavy infections (table 2). However, significant lower values were observed in the normal morphology of sperm cells, sperm viability, and total sperm count in infertile males with heavy bacterial infection compared against infertile males with scanty and moderate infection (table 2). It was also observed that those (infertile males) with scanty infection had higher values of morphologically normal sperm cells compared to those with moderate and heavy bacterial infection. Our findings were not really in line with the reports of Oyeyipo et al. [21] and Green & Nwachuku [6] because, the degrees of infection were not considered in their respective study. However, our results are similar to the reports of Liu et al. [23], who reported that the degree and type of bacterial infection in seminal plasma correlates negatively with the level of sperm cells viability, motility and progressive motility. The significantly lower values of motility, active motility, normal morphology, and total sperm count could be as a result of the activities of the infectious agents directly or indirectly on the sperm cells. Toxins released by these organisms stimulate the defense mechanism of the host system releasing interleukins, tumor necrotic factors- alpha, and interferons. The activities of these inflammatory substances can result in the production of reactive oxygen species and subsequently oxidative stress. Furthermore, due to the susceptibility of sperm cells to oxidative stress as a result of their limited cytoplasm to tackle reactive species and high levels of polyunsaturated fatty acids (PUFAs), the bacterial induced oxidative stress can lead to poor motility, sperm cells deformation and eventually dead. Infections affecting sperm cell

motility has been reported as one of the major causes of infertility. Our results further suggest that the higher the degree of infection, the higher the levels of sperm cell deformities, the lower the levels of active motility, the lower the sperm concentration in every ejaculate and vice versa. The cumulative effect of these results in male infertility.

When the influence of bacterial infections on sperm proteins, GPX, and TAC in the seminal plasma as well as testosterone and PSA in the serum were considered (table 3 and table 4), the significant higher values of HSP70 observed in the infertile males with bacterial infection are in line with findings of Kosecka-Strojek et al., [11], Yu et al., [24], and Njemini et al. [25]. They all reported in their respective study that bacterial infection of the seminal plasma such as *Staphylococcus aureus*, *Escherichia coli*, *klebsiella pneumoniae*, etc induced significantly higher values of HSPs family including HSP70. Kosecka-Strojek et al. [11], further reported that these changes were also observed in other mammals like chickens while Yu et al. [24], also explained viral infections have also been seen to induce increased levels of HSP70. In addition, the findings of significantly lower levels of OPN in the infertile males and those infertile with bacterial infection still support the results of Waheed et al. [26] who documented that higher levels of OPN are associated with fertility. More so, the significantly higher levels of PKA seen in infertile subjects are in line with the reports of Blommaert et al. [27]. They also reported significantly higher values of Anchor Kinase Activated Protein (AKAPs) and PKA in subjects with infertility. Again, the higher values of MDA and lower values of GPX and TAC in our findings are similar to the findings of Liu et al. [23], when values of antioxidants were compared in subjects with bacterial infections compared against controls. The levels of testosterone were also found to be lower in subjects with infection while PSA was observed to be higher in infertile subjects with bacterial infections. The higher value observed in HSP70 in our study could also be related to the protective role of these proteins in mitigating infections. An infection that triggers the defense system of the body and one of such is the HSP70 alongside interferons, interleukins, and lymphocytes. HSP70 could have been induced through changes in the ROS concentration and degradative enzymes such as coagulase, hyaluronidase, etc produced by the infectious organisms. HSPs are known to conserve and protect spermatozoa and other cells against deformities and as such may be involved in the activation of cytotoxic T-lymphocytes without the assistance of T-helper lymphocytes. In 2010, Guo and colleagues [29] also reported that HSP70 in the cellular response to infections and are actively produced when monocytes and macrophages are exposed to bacterial toxins. The significant reduction in GPX and TAC as well as the increase in MDA observed clearly suggests oxidative stress induced by the infectious pathogens. Oxidative stress has been reported to cause death of sperm cells, deformities, and reduced sperm cell motility. These factors could be one of the major causes of infertility among males.

The significant fall in testosterone concentration in the infertile males could also be as a result of the influence of generated ROS as a result of the infections on the Sertoli and Leydig cells of the testis. Oxidative stress has been reported to influence negatively the steroidogenesis and spermatogenesis in males. The increase in PSA among infertile males (table 3) especially among those with heavy bacterial infection (table 4) could also suggest inflammation of the prostate resulting in infectious prostatitis. Healthy prostate has been reported to contribute majorly in the seminal plasma. Therefore, higher values of PSA suggest inflammation of the prostate which in turn may affect sperm quality and fertility. Our results further suggest that though scanty and moderate bacterial infection could be complaisant in inflammation of the prostate but heavy bacterial infection elicit more inflammation of the prostate as reflected by the PSA results (table 4).

More so, our findings further suggested that the degree of infection did not cause any significant change in the values of HSP70, PKA, OPN, GPX, and TAC in seminal plasma as

well as testosterone in the serum among infertile males with bacterial infection (table 4). This also implies that scanty bacterial infection can elicit similar effect as moderate or heavy bacterial infection on sperm proteins such as HSP70, PKA, OPN, and GPX as well as TAC in the seminal plasma including testosterone in the serum. However, the values of MDA were significantly increased as the based on the degree of infection. This could be due to the fact that MDA are metabolic product of lipid peroxidation and the higher the degree of infection, the higher the tendency of lipid peroxidation viz-a-viz MDA concentration in the seminal plasma. Our findings contradict the findings of Cedenho et al. [29] and Coa et al. [30]. Though, we are in agreement that HSPs family including HSP70 were altered significantly in infertility but they (HSPs) are not better predictors or markers of sperm motility dysfunction rather MDA is a better predictor because of the changes in their concentration in the seminal plasma is proportional to the changes observed in the level of ROS/OS as indicated by the value of TAC.

When Sperm abnormalities were considered, significantly higher values of HSP70, PKA, MDA, and PSA while significantly lower values were seen in OPN, GPX, TAC and testosterone in our study when infertile males without bacterial infection, infertile subjects with bacterial infections, and subjects with sperm abnormalities (teratozoospermic, asthenoteratozoospermic and oligoasthenoteratozoospermic subjects) were compared against fertile subjects (control) (table 5). However, significant differences were not observed in the comparison amongst the infertile without bacterial infection, infertile subjects with bacterial infections, and subjects with sperm abnormalities (teratozoospermic, asthenoteratozoospermic and oligoasthenoteratozoospermic subjects) except PSA and MDA (table 4). Our findings suggest that both infection and sperm deformities may exert similar pathophysiological changes in the seminal plasma levels of sperm proteins HSP70, PKA, OPN, GPX, and TAC as well as in the values of testosterone but not necessarily the same for PSA and MDA. PSA values were observed to significantly lower in infertile males without bacterial infection unlike the sperm proteins and anti-oxidant proteins in the seminal plasma. This result further suggest that infection of the prostate or inflammatory response of the prostate as indicated the values of PSA in the serum play significant role in exacerbating the degree or problem of infertility amongst males. Further in the case of MDA it was also seen that MDA values were significantly highest in oligoasthenoteratozoospermic (OAT) subjects compared to teratozoospermic and asthenoteratozoospermic (ATS) as well as infertile subjects with bacterial infection. oligoasthenoteratozoospermic (OAT) has been previous reported by Agarwal et al. [32] and Cologar et al. [32] as the one of the main causes of male infertility due to its three-fold negative dimension of sperm quality affecting the sperm concentration, progressive motility and normal morphology. This MDA results could suggest that OAT subjects are more prone to oxidative damage than other sperm cells deformities and infection. These results further suggest MDA as a better predictor of sperm cells damages and infertility because of the changes in their concentration in the seminal plasma is proportional to the changes observed in the level of ROS/OS as indicated by the value of TAC.

5. CONCLUSION

The results of bacterial infection of seminal plasma indicated significantly increased lipid peroxidation cum oxidative stress leading to altered physiology of the sperm proteins in the seminal plasma. In addition, the infections (especially those with heavy growth), induced significant reduction and increase in plasma testosterone and PSA levels respectively. These alterations could also be responsible for the poor sperm qualities and their infertile state as observed in the case-subjects.

ETHICAL APPROVAL

Before the start of the study, ethical approval was sort and obtained from the Ethical Review Boards of the Rivers State Government, through the Rivers State Ministry of Health covering the Rivers State University teaching Hospital and Primary Healthcare Centres in the State with approval no of MH/PRS/391/VOL.2/726 and MH/PRS/391/VOL.2/727 respectively. In addition, written informed consents were gotten from all the participants before being recruited for the study. We hereby declare that all experiments have been examined and approved by the appropriate ethics committee and have therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

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