

**Original Research Article**  
**Lethal effects of Red macro algal extracts  
against Tobacco cutworm, *Spodoptera litura*  
Fabricius**

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**ABSTRACT**

**Aims:** To study the toxicity of aqueous and methanol extracts of Red macro algal extracts against a cosmopolitan pest, *Spodoptera litura* Fabricius.

**Place and Duration of Study:** Department of Agricultural Entomology, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu.

**Methodology:** The bioassay was carried by treating aqueous and methanol extracts of three different Red macro algae at 0.7, 1.00, 3.00, 5.00, 7.00 and 10.00 per cent concentration with three replications and ten larvae per treatment and the mortality data was recorded every 24 hours up to adult emergence.

**Results:** The aqueous and methanol extracts of three different Red macroalgae viz., *Acanthophora spicifera*, *Gracilaria corticata* and *Jania rubens* were used to find out their lethal effects against tobacco cutworm, *Spodoptera litura* under laboratory condition. The larval mortalities were observed in both the extracts and the mortalities increase with increase in concentration after 96 hours of exposure period with 60.00, 46.66 and 26.66 per cent in aqueous extracts; and 40.00, 26.66 and 23.33 per cent in methanol extracts at 10 per cent concentration in *A. spicifera*, *G. corticata* and *J. rubens*, respectively. The pre-pupal and pupal mortalities were recorded in all the concentrations showing malformations during the growth. The pupation and the adult emergence were also affected with the highest inhibition by the extracts of *A. spicifera*.

**Conclusion:** It is surmised that Red macroalgae possess insecticidal potential and growth inhibitory properties which can be exploited for eco-friendly management of *Spodoptera litura* and possibly other lepidopteran larvae as well.

**Keywords:** Red algae, *Acanthophora spicifera*, *Gracilaria corticata*, *Jania rubens*, *Spodoptera litura*, Ecofriendly management.

**1. INTRODUCTION**

Tobacco cutworm, *Spodoptera litura* Fabricius is a polyphagous pest of cosmopolitan distribution and defoliate the crops of economic importance viz., groundnut, pulses, castor, cotton, chilli, tobacco etc., [1-2]. The synthetic pesticides are used for their management which invoke insect resistance and resurgence; and cause serious threats to the environment [3-4]. To eradicate the ill effects on usage of synthetic pesticides, replacement with plant derived pesticides proven to be safe and effective is the need of the hour. The plant derived pesticides are environmentally safe

and lower the risk of insect resistance and insect resurgence [5]. There are numerous plants having the insecticide potential with one such alternative is marine macro algae. The marine macro algae, commonly known as seaweeds are one of the dominant creatures that occupy all zones of marine ecosystems. There are three groups of macro algae, Red algae that belongs to the phylum Rhodophyta, Brown algae that belongs to Ochrophyta and Green algae that belongs to Chlorophyta. They are not only the primary producers of marine ecosystem but also the potential sources of bioactive compounds of agricultural importance. Macroalgae have inherently developed defense mechanism with novel bioactive compounds and have amazing capability to survive in the marine environment. Numerous secondary metabolites such as sesquiterpenes, diterpenes, monoterpenoids, steroids are present in seaweeds which showed insecticidal [6-11]; nematocidal [12-13]; and antimicrobial properties [14-15]. With this background, the present study was carried out to explore the lethal effects of aqueous and methanol extracts of Red macro algae viz., *Acanthophora spicifera*, *Gracilaria corticata* and *Jania rubens* against *Spodoptera litura*.

## 2. MATERIAL AND METHODS

### Insect culture

The initial culture as larvae of *Spodoptera litura* were collected from the castor plant in Department of Oil seeds, Tamil Nadu Agricultural University, Coimbatore district (11°1'N, 76°55'E) and reared with castor leaves as natural diet under laboratory condition. They were reared up to pupation and allow to hatch. The emerged healthy adults were released into an oviposition cage and were provided with fresh *Nerium oleander* L leaves for egg laying with petiole immersed in water to prevent the drying of leaves. The adult insects are fed with 10% honey solution fortified with multivitamins to enhance the oviposition rate. The newly emerged F<sub>1</sub> generation larvae were utilized for the experiment.

### Collection of macro algae

The three species of Red macro algae, *Acanthophora spicifera* (M.Vahl) Boergesen, *Gracilaria corticata* (J. Agardh) J. Agardh and *Jania rubens* (L.) J.V. Lamouroux were collected from Hare Island located at Lat 9°12'0" N and Long 79°4'48"E which is the part of Gulf of Mannar region, Tuticorin, Tamil Nadu, India during May and October 2019. The algal thalli were collected by hand picking method, washed to remove salts and debris, dried under shade and stored in sealed container at room temperature for further studies. The collected algal specimens were identified and authenticated by Botanical Survey of India, South Regional Centre, Coimbatore and the specimens were deposited in the Department of Agricultural Entomology, Tamil Nadu Agricultural University, Coimbatore.

### Preparation of macroalgal extracts

The dried macro algae were powdered and sieved to uniform particle size of 0.5 mm. The extraction of bioactive compounds was done with ultrapure water (H<sub>2</sub>O) and methanol (MeOH) as solvent at the biomass solvent ratio of 1:10 w/v by using ultra high Frequency sonicator (SONICS®, sonics and materials, Inc.) at 20kHz for 30 minutes followed by centrifugation (Dynamica, VELOCITY 14R) at 10000 rpm (10380xg) for 10 min. The supernatant was collected and the pellets were re-extracted with fresh solvent to enhance the extraction efficiency. The supernatant was pooled and concentrated under reduced pressure at 35°C and 100 rpm using rotary cum vacuum evaporator (Heidolph, Hei-VAP core) and the crude extracts were stored at -20°C until utilization. The extraction yield of crude extracts is calculated by,

$$\text{Extraction yield (\%)} = \text{Yield of crude extracts obtained (g)} / \text{Amount of raw material used (g)} \times 100$$

### Bioassay by leaf dip method

The leaf discs of known size (4 cm diameter) were cut and dipped for 10 seconds in different concentration 0.7, 1.0, 3.0, 5.0, 7.0, 10.00 per cent prepared with aqueous and methanol solution along with 0.1% Triton X-100 as sticking

agent Water and methanol with 0.1% Triton X were used as control. The leaf discs were dried, placed in Petri plates and offered to 4 hours pre-starved second instar larvae of *Spodoptera litura*. Three replications with 10 larvae per replication were designed for toxicity studies and the experiment was Completely Randomized. The mortality due to direct toxicity/growth regulatory activity to the larvae were recorded every 24 hours after treatment up to adult emergence. The dose-response curve was constructed and the lethal concentration (LC<sub>10</sub>, LC<sub>20</sub> and LC<sub>50</sub>) of macro algal extracts was estimated.

### Statistical analysis

The mortality data were *arc sine* transformed before one-way ANOVA, and means were compared using Tukey's Honestly Significance Difference (HSD) ( $p=0.05$ ) test. The concentration mortality data were submitted to Probit analysis at 95% confidence level ( $P < 0.05$ ) to obtain dose-response curve. All the statistical procedures were done with IBM SPSS v. 20 statistical software.

## 3. RESULTS AND DISCUSSION

The Red macroalgae have the potential to act as biopesticide that can be utilized in Integrated Pest Management. The current study involves the mortality bioassays with crude aqueous and methanol extracts of Red macroalgae, *A. spicifera*, *G. corticata* and *J. rubens* that showed significant toxicities against second instar larvae of *S. litura* at different concentrations. Previous reports indicated that members of Rhodophyta have provided an assortment of insecticidal compounds as potential leads for pest management in agriculture [16]. The extraction yield of crude extracts of *A. spicifera*, *G. corticata* and *J. rubens* were 9.6, 6.2, 3.2 and 5.1, 3.2, 2.9 per cent for aqueous and methanol extracts respectively (Table 1).

In this study, on treatment with different concentrations *viz.*, 0.7, 1.00, 3.00, 5.00, 7.00 and 10.00 per cent of aqueous and methanol extracts of Red algae, the larval mortality had occurred in all the concentrations used in a dose dependent manner with a highest mortality of 60.00 and 40.00 per cent at 10 per cent concentration and the least larval mortality of 26.66 and 10.00 per cent at 1.00 per cent concentration in aqueous and methanol extracts of *A. spicifera* at the end of 96 hours of exposure to insects. It is followed by *G. corticata* with the maximum mortality of 46.66 and 26.66 per cent and minimum of 10.00 per cent mortality in aqueous and methanol extracts respectively. Among the used red macro algae, *J. rubens* exhibited the least mortality at the highest concentration *i.e.*, 26.66 and 23.33 per cent larval mortality and mere mortality of 3.33 per cent at the lowest used concentration. There is a mere or no mortality had happened at 0.7 per cent concentration in all the used macro algal extracts (Fig 1 and 2). The lethal concentration (LC<sub>10</sub>, LC<sub>20</sub> and LC<sub>50</sub>) of *Acanthophora*, *Gracilaria* and *Jania* were 8.323, 11.542 and 25.100 per cent for aqueous and 16.404, 33.246 and 26.923 per cent for methanol extracts, respectively and the data were provided in table 2. After 96 hours of treatment, the pupation had initiated with the occurrence of pre-pupal mortality of 13.33 per cent in both aqueous and methanol extracts of *A. spicifera* and *J. rubens* whereas the mortality of 20 and 6.66 per cent in aqueous and methanol extracts of *G. corticata* respectively (Fig 3). The least pupation had occurred in *A. spicifera* *i.e.*, 30.00 and 46.66 per cent followed by *G. corticata* *i.e.*, 33.33 and 66.66 per cent and *J. rubens* *i.e.*, 60 and 63.33 per cent in aqueous and methanol extracts respectively. Some of the pupae were failed to emerge as adults with pupal mortality of 13.33 and 16.66 per cent in *A. spicifera* followed by 13.33 and 20.00 per cent in *G. corticata* and 23.33 and 10.00 per cent in *J. rubens* for aqueous and methanol respectively (Fig 4). The Red macro algal extracts have the potential to inhibit the growth of *S. litura*, with the least larval to adult conversion ratio of 1:0.55 and 1:0.42 *A. spicifera* followed by *G. corticata* with the ratio of 1:0.20 and 1:0.46 and *J. rubens* with the ratio of 1: 0.36 and 1:0.53 respectively for aqueous and methanol extracts and it represented in Fig 5.

Previously, it is reported [17-18] that the Red algae, *Plocamium cartilagineum* possess polyhalogenated monoterpenes that have insecticidal and insect feeding deterrence activities against tobacco budworm (*Heliothis virescens*), Tomato moth (*Tuta absoluta*), Southern com rootworm (*Diabrotica undecimpunctata*), European corn borer (*Ostrinia nubilalis*), fall armyworm (*Spodoptera frugiperda*), boll weevil (*Anthonomus grandis*), aster leafhopper (*Macrostelus facifrons*), black bean aphid (*Aphis fabae*), Cereal aphid (*Schizaphis graminum*) and two-spotted spider mite (*Tetranychus urticae*). The members of genus, *Laurencia* have various bioactive metabolites that have an insecticidal potential [19] especially acetylinic sesquiterpene ethers [20]. Researchers compared the effect of crude and active ingredient C<sub>15</sub> acetogenin from the petroleum ether extracts of red alga *Laurencia papillosa* on grub of confused flour beetle, *Tribolium confusum* and found to inhibit their growth [21]. The Red algae, *Liagora ceranoides* demonstrated the insecticidal activity towards *Oryzeaphilus mercator* and *Tribolium castaneum* [22]. Lectins are first identified from the marine algae [23]. The purified lectins from marine red alga, *Gracilaria ornata* showed higher level of mortality to cowpea weevil, *C. maculatus* when incorporated in artificial seeds due to the inhibition of digestion and absorption of food [24]. The repellent action of red flour beetle, *Tribolium castaneum* was observed at 4 per cent concentration of *G. edulis* with acetone, ethyl acetate and chloroform extracts treatments [25].

**Table 1. Extraction yield of Red macroalgae**

| Macro algae                   | Extraction yield (%)    |                         |
|-------------------------------|-------------------------|-------------------------|
|                               | Aqueous                 | Methanol                |
| <i>Acanthophora spicifera</i> | 9.6 ± 0.09 <sup>d</sup> | 5.1 ± 0.19 <sup>d</sup> |
| <i>Gracilaria corticata</i>   | 6.2 ± 0.13 <sup>e</sup> | 3.2 ± 0.14 <sup>e</sup> |
| <i>Jania rubens</i>           | 3.2 ± 0.05 <sup>f</sup> | 2.9 ± 0.06 <sup>f</sup> |

All values are mean ± SD for triplicate experiments (n=3). Different letters within the same column indicate significant difference between the values (p=.05). Extraction yield (%) calculated as g of dried extract/10 g DW.

**Table 2. Lethal concentration of Red algal extracts after 96 hours of exposure**

| Macro algae        | Extracts         | Lethal concentration n | Estimated value | Confidence interval | Slope ± SE    | χ <sup>2</sup> Value |
|--------------------|------------------|------------------------|-----------------|---------------------|---------------|----------------------|
| <i>Acanthopora</i> | Aqueous          | LC <sub>10</sub>       | 0.976           | 0.000 - 2.058       | 1.243 ± 0.266 | 9.410                |
|                    |                  | LC <sub>20</sub>       | 2.751           | 0.000 - 3.936       |               |                      |
|                    |                  | LC <sub>50</sub>       | 8.323           | 3.708 - 42.30       |               |                      |
|                    | Methanol         | LC <sub>10</sub>       | 1.029           | 0.221 - 1.858       | 1.066 ± 0.278 | 0.221                |
|                    |                  | LC <sub>20</sub>       | 2.662           | 1.287 - 4.190       |               |                      |
|                    |                  | LC <sub>50</sub>       | 16.404          | 8.857 - 84.199      |               |                      |
| <i>Gracilaria</i>  | Aqueous          | LC <sub>10</sub>       | 1.527           | 0.631 - 2.347       | 1.459 ± 0.314 | 2.409                |
|                    |                  | LC <sub>20</sub>       | 3.058           | 1.864 - 4.299       |               |                      |
|                    |                  | LC <sub>50</sub>       | 11.542          | 7.631 - 26.583      |               |                      |
|                    | Methanol         | LC <sub>10</sub>       | 1.804           | 0.385 - 3.114       | 1.013 ± 0.315 | 2.634                |
|                    |                  | LC <sub>20</sub>       | 4.904           | 2.762 - 10.537      |               |                      |
|                    |                  | LC <sub>50</sub>       | 33.246          | 13.672 - 949.680    |               |                      |
| <i>Jania</i>       | Aqueous          | LC <sub>10</sub>       | 3.100           | 1.348 - 4.654       | 1.411 ± 0.400 | 0.980                |
|                    |                  | LC <sub>20</sub>       | 6.356           | 4.194 - 11.972      |               |                      |
|                    |                  | LC <sub>50</sub>       | 25.100          | 12.900 - 207.944    |               |                      |
|                    | LC <sub>10</sub> | 2.832                  | 1.114 - 4.357   |                     |               |                      |

| Methanol | LC <sub>20</sub> | 6.136  | 3.946 - 12.060   | 1.310 ± | 1.469 |
|----------|------------------|--------|------------------|---------|-------|
|          | LC <sub>50</sub> | 26.923 | 13.169 - 284.871 | 0.380   |       |

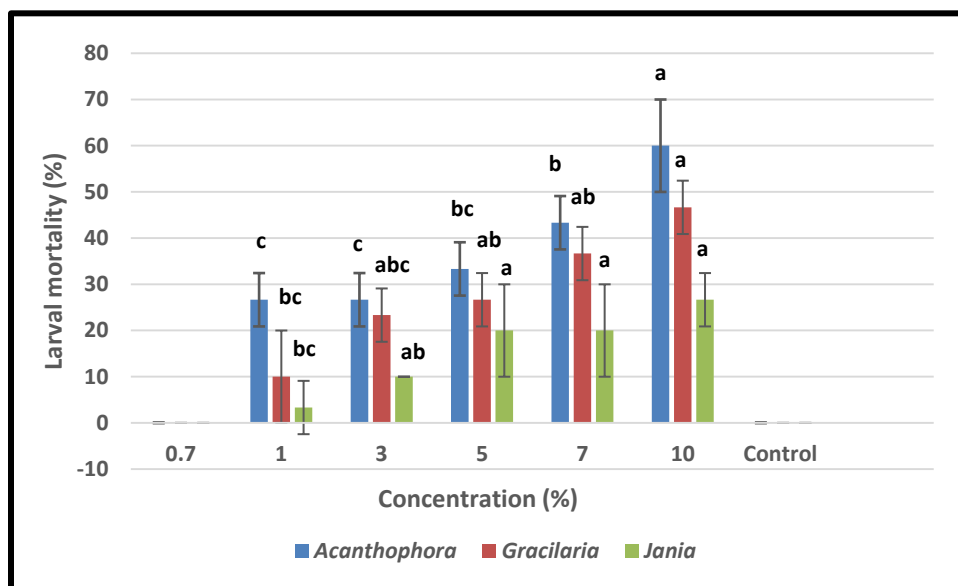


Fig. 1. Larval mortality of *Spodoptera litura* on treatments with aqueous extracts of different Red algae. Treatment means ( $\pm$  SEM) with different letters show significant differences by Tukey's Honestly Significant Difference Test at  $p=.05$  level.

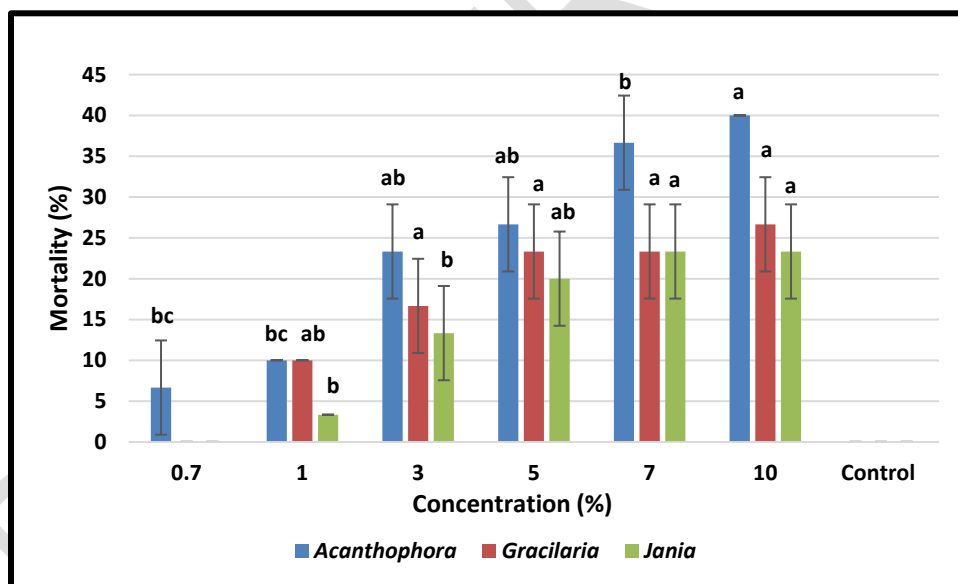


Fig. 2. Larval mortality of *Spodoptera litura* on treatments with methanol extracts of different Red algae. Treatment means ( $\pm$  SEM) with different letters show significant differences by Tukey's Honestly Significant Difference test at  $p=.05$  level.

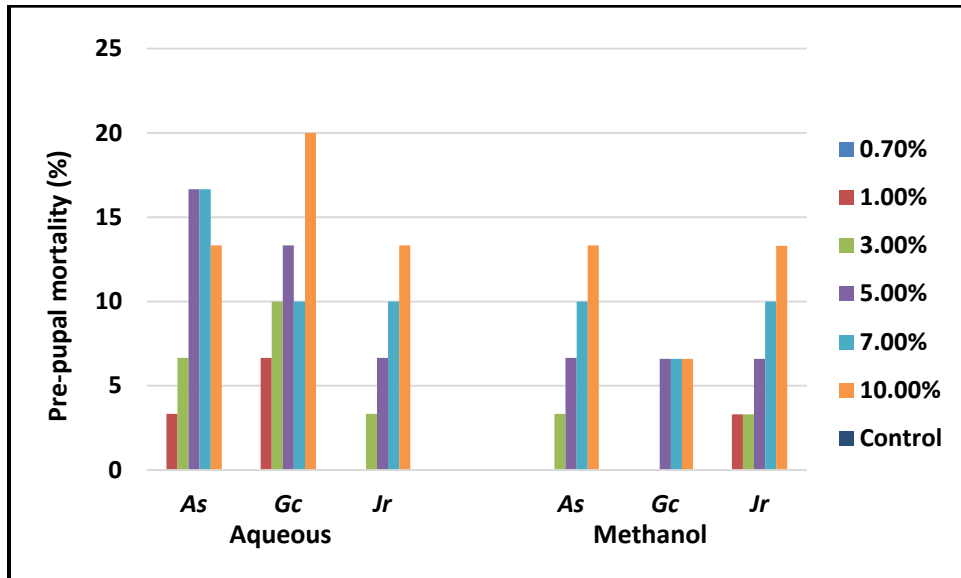


Fig. 3. Pre-pupal mortality of *Spodoptera litura* on treatments with aqueous and methanol extracts of different Red algae.

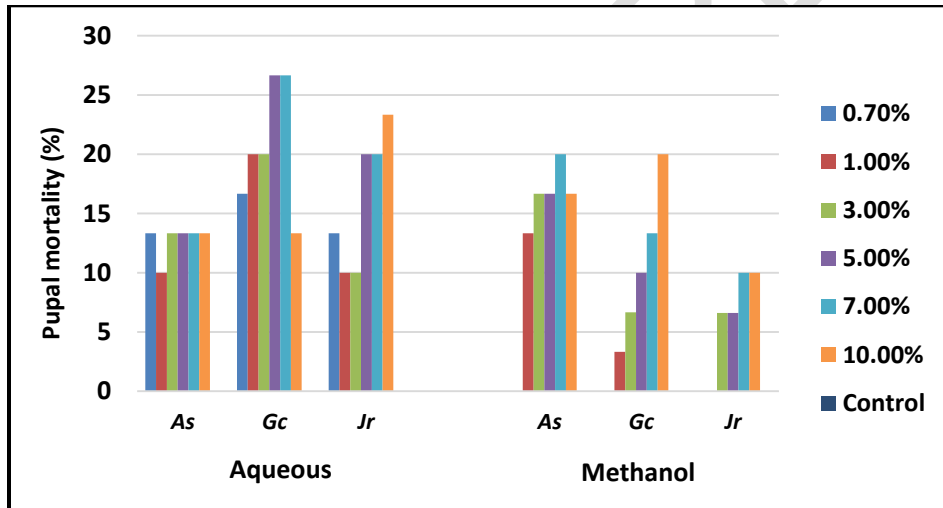


Fig. 4. Pupal mortality of *Spodoptera litura* on treatments with aqueous and methanol extracts of different Red algae.

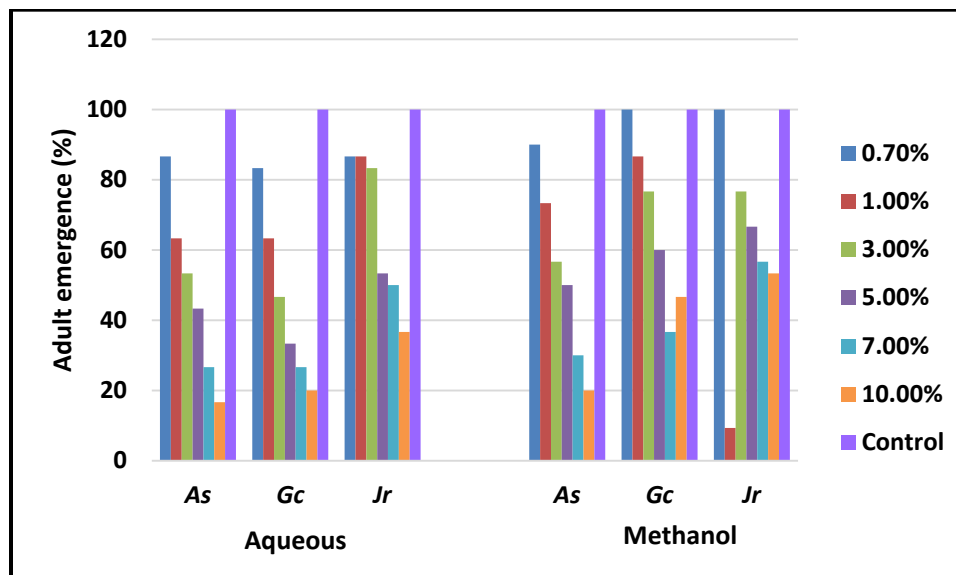


Fig. 5. Adult emergence of *Spodoptera litura* on treatments with aqueous and methanol extracts of different Red algae.

#### 4. CONCLUSION

The Red macroalgal extracts of *A. spicifera*, *G. corticata* and *J. rubens* using aqueous and methanol extracts showed potentials of insecticidal activity through contact/ feeding toxicity supported with growth regulatory activity against *S. litura* in this study. The  $LC_{50}$  values of both aqueous and methanol extracts of the Red algae suggests that it can be improved by using the insecticidal compounds/ principles through nano-formulation/ controlled release formulation thereby full potential of the Red algae can be realized. Hence, Red macroalgae would be a befitting alternative to synthetic pesticides on the grounds of green pesticides for organic farming.

#### COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

#### REFERENCES

1. Yooboon T, Pengsook A, Ratwatthananon, A. A plant-based extract mixture for controlling *Spodoptera litura* (Lepidoptera: Noctuidae). Chemical and Biological Technologies in Agriculture. 2019; 6: 5.
2. Datta R, Kaur A, Saraf I, Singh IP, Kaur S. Effect of crude extracts and purified compounds of *Alpinia galanga* on nutritional physiology of a polyphagous lepidopteran pest, *Spodoptera litura* (Fabricius). Ecotoxicology and Environmental Safety. 2019; 168: 324-329.
3. Tripathy MK, Singh, HN. Inheritance of resistance through generations in *Helicoverpa armigera* Hubner at Varanasi, Uttar Pradesh. Indian Journal of Agricultural Research. 2000; 34(3):164-167.

4. Chavan SM, Sushilkumar, Arve SS. Efficacy and economics of various pest management modules against tomato fruit borer, *Helicoverpa armigera* (Hubner). *Agricultural Science Digest*. 2012; 32(4): 296-300.
5. Souto AL, Sylvestre M, Tolke ED, Tavares JF, Barbosa-Filho JM, Cebrian-Torrejon G. Plant-Derived Pesticides as an Alternative to Pest Management and Sustainable Agricultural Production: Prospects, Applications and Challenges. *Molecules* (Basel, Switzerland). 2021; 26(16): 4835. <https://doi.org/10.3390/molecules26164835>.
6. Biju B, Jacob M, Padmakumar K, Muraleedhran D. Effect of extract of the seaweed *Bryopsis plumosa* (Huds.) (Ag) on the feeding rate and protein profile of haemolymph and fat body of *Hyblaea puera* (Cramer) (Lepidoptera: Hyblacidae). *Entomon*. 2004; 29: 271-276.
7. Cetin H, Gokoglu M, Oz E. Larvicidal activity of the extract of seaweed, *Caulerpa scalpelliformis*, against *Culex pipiens*. *Journal of the American Mosquito Control Association*. 2010; 26(4): 433-436.
8. Sahayaraj K, Kalidas S. Evaluation of nymphicidal and ovicidal effect of a seaweed, *Padina pavonica* (Linn.) (Phaeophyceae) on cotton pest, *Dysdercus cingulatus* (Fab.). *Indian Journal of Geo Marine Sciences*. 2011; 40(1): 125-129.
9. Sahayaraj K, Maryjeeva Y. Nymphicidal and ovipositional efficacy of seaweed *Sargassum tenerrimum* (J. agardh) against *Dysdercus cingulatus* (Fab.) (Pyrrhocoridae). *Chilean Journal of Agricultural Research*. 2012; 72(1): 152 - 156.
10. Gowthish, K. and Kannan, R. (2018). Bioefficacy of brown algal seaweed *Sargassum cristaefolium* C. against cosmopolitan pest, *Spodoptera litura* Fabricius (Lepidoptera: Noctuidae). *Multilogic in Science*, 8(D): 56 - 57.
11. Dharanipriya N, Kannan R, Ayyasamy R. Potential Toxicity and Insect Growth Regulator Activity of Red Algal Seaweeds against a Polyphagous Pest, *Spodoptera litura* (Fab.). *Annals of Biology*. 2020; 36(1): 88-93.
12. El-Ansary MSM, Hamouda RA. Biocontrol of Root Knot Nematode Infected Banana Plants by Some Marine Algae. *Russian Journal of Marine Biology*. 2014; 40(2): 140-146.
13. Khan AM, Naz S, Abid M. Evaluation of marine red alga *Melanothamnus unilateralis* against *Meloidogyne incognita*, fungus and as fertilizing potential on okra. *Pakistan Journal of Nematology*. 2016; 34(1): 91-100.
14. Fialho MB, deMoraes MHD, Tremocoldi AR, Pascholati SF. Potential of antimicrobial volatile organic compounds to control *Sclerotinia sclerotiorum* in bean seeds. *Perquisa Agropecuaria Brasileira*. 2011; 46(2): 137-142.
15. Ibraheem BMI, Hamed SM, Abdelrhman AA, Farag FM, Abdel-Raouf N. Antimicrobial activities of some brown macro algae against some soil borne plant pathogens and in vivo management of *Solanum melongena* root diseases. *Australian Journal of Basic Applied Sciences*. 2017; 11: 157-168.
16. El Sayed KA, Dunbar DC, Perry TL, Wilkins SP, Hamann MT, Greenplate JT, Wideman MA. Marine natural products as prototype insecticidal agents. *Journal of Agricultural and Food Chemistry*. 1997; 45: 2735-2739.
17. Martin AS, Negret R, Roviroso J. Insecticidal and acaricidal activity of polyhalogenated monoterpenes from Chilean *Plocamium cartilagineum*. *Phytochemistry*. 1991; 30(7): 2165-2169.
18. Argandona V, Pozol TD, Martin AS, Roviroso J. Insecticidal activity of *Plocamium cartilagineum* monoterpenes. *Journal of the Chilean Chemical Society*. 2000; 45(3): 1-6.
19. Watanabe K, Miyakado M, Ohno N, Okada A, Yanagi K, Moriguchi K. A polyhalogenated insecticidal monoterpene from the Red alga, *Plocamium telfairiae*. *Phytochemistry*. 1989; 28: 77-78.
20. Fukuzawa A, Masamune T. Laurepinnacin and isolaurepinnacin: new acetylenic cyclic ethers from the marine alga *Laurencia pinnata* Yamada. *Tetrahedron Letters*. 1981; 22: 4081 - 4084.

21. Abou-Elnaga ZS, Alarif WM, Al-lihaibi SS. New Larvicidal Acetogenin from the Red Alga, *Laurencia papillosa*. CLEAN - Soil, Air, Water. 2011; 39(8): 787-794.
22. Pasdaran A, Hamed A, Mamedov N. Antibacterial and insecticidal activity of volatile compounds of three algae species of Oman Sea. International Journal of Secondary Metabolite. 2016; 3(2): 66-73.
23. Boyd WC, Almodovar LR, Boyd LG. Agglutinin in marine algae for human erythrocytes. Transfusion. 1996; 6: 82-83.
24. Leite YFM, Silva LMC, Amorim RCN, Freire EA, Jorge DMM, Grangeiro TB, Benevides NMB. Purification of a lectin from the marine red alga *Gracilaria ornata* and its effect on the development of the cowpea weevil *Callosobruchus maculatus* (Coleoptera: Bruchidae). Biochimica et Biophysica Acta (BBA) - General Subjects. 2005; 724(1-2): 137-145.
25. Aswathi EM, Jamila P. Antifungal phytochemical Screening and Repellent Effect in Seaweed *G. edulis*. International Journal of Recent Scientific Research. 2014; 5(11): 2037-2042.

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