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Antioxidant and antibacterial activities of the essential oils of *Synedrella nodiflora*, *Mikania cordata* and *Melanthera scandens* three plants of the Ivorian flora

ABSTRACT

Aims: The objective of this work is to contribute to the valorization of medicinal and aromatic plants of the Ivorian flora. We propose to evaluate the antioxidant and antibacterial activities of the essential oil of three species used in traditional medicine.

Study design: valorization of aromatic and medicinal plants and essential oil.

Methodology: The antioxidant potential of the extracts was evaluated using the Blois method. The antibacterial activity of the different oils at different concentrations was determined for each bacterial strain, by the technique of macro-dilution in solid medium (diffusion in wells). The minimum inhibitory concentration (MIC) and The Minimum Bactericidal Concentration (MBC) were determined. MBC / MIC ratios were calculated. When this ratio is less than 4, the extract is considered to be bactericidal. When it is 4, the extract is considered bacteriostatic.

Results: The antioxidant activity has pointed out the poor antioxidant power of the EO extracted. The EC₅₀ values vary from 15 µg/mL 32 µg/mg. The antibacterial tests have shown that the samples exert an inhibitory effect on Gram (+) bacteria. The diameters of the inhibition zones vary between 14 and 25 mm for the extracts against 35 mm for the gentamycin. The MBC/MIC is 2 for the gentamycin and 4 for all the EO combined versus the resistant *S. aureus meticiline*. Therefore, the essential oil has shown a bacteriostatic effect on this strain. As far as *S. aureus* CIP 483 is concerned, the MBC/MIC has given 1 for the gentamycin, 2 for *M. scandens*. The EO extract of *M. scandens* has a bactericidal action against this bacteria strain.

Conclusion: All the essential oils have less antioxidant activity than that of vitamin C. The antibacterial activity of EO has given satisfactory results on all Gram (+) bacteria. *Melanthera scandens* Essential Oil shows antibacterial potential against *Staphylococcus aureus* CIP 483.

Keywords: *M. cordata*, *M. scandens*, *S. nodiflora*, antioxidant activity, antibacterial activity,

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1. INTRODUCTION

From antiquity to the present day, medicinal and aromatic plants have played a crucial role in the prevention and / or treatment of various human diseases [1]. The use of plants for their medicinal properties is a very ancient practice. It has its origins in the oldest civilizations and has been well preserved over the centuries around the world. Over the past two decades, much attention has been paid to plants as new alternative therapeutic agents due to their natural bioactive compounds [2]. Thus, the search for new molecules, taking into account

20 criteria other than efficiency, has become essential. Biological control through the use of
21 natural antioxidant and antibacterial substances that can be an alternative to chemicals.
22 Among these natural substances are essential oils extracted from aromatic plants [3].
23 Our interest focused on the study of the antibacterial and antioxidant activities of the
24 essential oils of three medicinal and aromatic plants often used in traditional medicine and
25 as a food condiment by the local population in Côte d'Ivoire: *Melanthera scandens*, *Mikania*
26 *cordata* and *Synedrella nodiflora*.
27 To our knowledge, no study of the antioxidant and antibacterial activities has been done on
28 the essential oils of the three Ivorian species. However, studies have been done on the
29 extracts thereof.
30 In the literature, previous studies have been done on the extraction and chemical
31 composition of essential oils from the organs of all three plants [4,5].
32 Antioxidant activity studies were performed on methanolic extracts from the dried leaves of
33 the Nigerian species of *Melanthera scandens*. They show that these have less important
34 antioxidant properties than those of vitamin C. [6] Studies of antibacterial activity have also
35 been carried out in Indonesia and Bangladesh on extracts from the leaves and aerial part of
36 *Mikania cordata* by the diffusion method in a solid medium. In Indonesia, this was carried out
37 against 3 Gram (+) and one Gram (+) bacteria. The results have shown that the ethyl
38 acetate extract have given good activity against *S. aureus* with an inhibition diameter of 14
39 mm [7]. In Bangladesh, it was performed against 8 Gram (+) bacteria and 5 Gram (-)
40 bacteria. It appears that the dichloromethane fraction exerted a strong activity against
41 *Escherichia coli* with a zone of inhibition of 14 mm [8]. Antibacterial and antioxidant activity
42 studies have also been done on extracts of *Synedrella nodiflora*. It shows that the
43 methanolic extract of the leaves has an inhibition zone of 14 mm against *Bacillus cerus* [9].
44 The antioxidant capacities of the methanolic extract and the soluble fractions were tested
45 using DPPH and BHT. The 50% inhibitory concentrations (IC50) in µg / mL of the fractions
46 and extracts vary between 10.52 and 31.25 when that of ascorbic acid and BHT are
47 respectively 5.8 ± 0.21 and 27.5 ± 0.54 . The soluble fraction of dichloromethane has good
48 antioxidant activity. Its IC50 is 10.52 µg / mL [10]. This is why, in this work, in order to
49 contribute to a valuation of the aromatic and medicinal plants of Ivory Coast, we propose to
50 evaluate by spectrophotometry the antioxidant activity of essential oils vis-à-vis the DPPH.
51 and their antibacterial activity.

52

53 **2. MATERIAL AND METHODS**

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55 **2.1 Equipment**

56 The plant material consists of the EO extracted from the three plants.

57 **Bacterial strains.**

58 Eight (08 strains) of bacteria were used. For the most part, these are reference strains from
59 the laboratory of the Swiss Center for Scientific Research (CSRS) with the names ATCC and
60 SO. Other strains, on the other hand, are clinical strains from the Institut Pasteur with the
61 name CIP. Thus, the antibacterial tests were carried out on the following strains:

62 GRAM (-): *Escherichia coli* ATCC 25922; *Pseudomonas aeruginosa* ATCC 27853;
63 *Salmonella typhimurium* SO66; *Proteus mirabilis* ATCC 14153 and *Proteus vulgaris* CIP 5860
64 GRAM (+): *Staphylococcus aureus* ATCC 25923; *Staphylococcus aureus* CIP 483 and
65 *Staphylococcus aureus* meticilin resistant ATCC 43300.

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67 **2.2. Methods**

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69 **2.2.1. Evaluation of the antioxidant activity of EOs.**

70

71 The antioxidant potential of the extracts was evaluated using the Blois method.
72 The DPPH is dissolved in absolute ethanol to obtain a solution of 0.3 mM molar
73 concentration. The solutions to be tested: are diluted in absolute ethanol in order to have the
74 following concentrations in mg / mL: 0.002; 0.02; 0.125; 0.25; 1 and 2.
75 2.5 ml of test solution are introduced into dry and sterile hemolysis tubes and 1 ml of
76 ethanolic solution of DPPH is added. After shaking, the tubes are placed in the dark for 30
77 min, protected from light.
78 For each solution to be tested, a blank is prepared consisting of 2.5 mL of pure absolute
79 ethanol supplemented with 1 mL of ethanolic solution of DPPH.
80 For the negative control, a solution of DPPH is prepared by diluting 1 mL of the ethanolic
81 solution of DPPH in 2.5 mL of ethanol. For the positive control, a solution of vitamin C
82 (ascorbic acid) is used, the absorbance of which is measured under the same conditions.
83 The measurement of the residual absorbance is carried out at 517 nm. It is translated into
84 percentage inhibition by the following formula [11]:

$$85 \quad \%I = \left(1 - \frac{\text{Abs test}}{\text{Abs DPPH}} \right) \times 100$$

86
87 % I: Percentage inhibition. Abs test: Absorbance of ethanolic solution of EO and DPPH.
88 AbsDPPH: absorbance of blank (ethanolic solution of DPPH).

89 Before each measurement, the absorbance of the blank is measured. For each extract
90 exhibiting an antioxidant potential, the EC50 (effective concentration of the substrate which
91 inhibits the oxidative potential of DPPH by 50%) is determined graphically [12].

92 **2.2.2. Evaluation of the antibacterial activity of Eessential Oils.**

93 ***2.2.2.1 Measurement of the diameters of the zones of inhibition.***

94 The antibacterial activity of the different oils at different concentrations was determined for
95 each bacterial strain, by the technique of macro-dilution in solid medium (diffusion in wells)
96 [13] from a culture of 18 to 20 h (10⁵- 10⁶ CFU / mL). The inoculum of 1 ml is inoculated on
97 the surface of Mueller Hinton (MH) medium previously poured into Petri dishes. After 15 min,
98 6 mm diameter wells were cut using Pasteur pipettes. The bottom of the wells is blocked by
99 a drop of MH agar to limit the diffusion of oils under the agar. Then, 50 µL of the oil at
100 different concentrations and 50 µL of a gentamycin reference are distributed in each well.
101 After diffusion, the cultures are incubated in incubators at 37 ° C. for 24 h. The inhibition
102 halos are measured by a caliper. The activity is considered zero for an inhibition diameter
103 (i.d.) less than or equal to 8 mm; weak for d.i. between 8 and 14 mm, average for i.d.
104 between 14 and 20 mm; strong for i.d.. greater than or equal to 20 mm [13].

105 ***2.2.2.2 Determination of minimum inhibitory concentration (MIC) and minimum*** 106 ***bactericidal concentration (MBC)***

107 For the determination of the minimum inhibitory concentration (MIC), a series of 10 sterile
108 hemolysis tubes is used. Using a sterile graduated pipette, 4.6 mL of MH-tween 80 broth are
109 introduced into the first tube and 2.5 mL of the same broth into the other tubes.

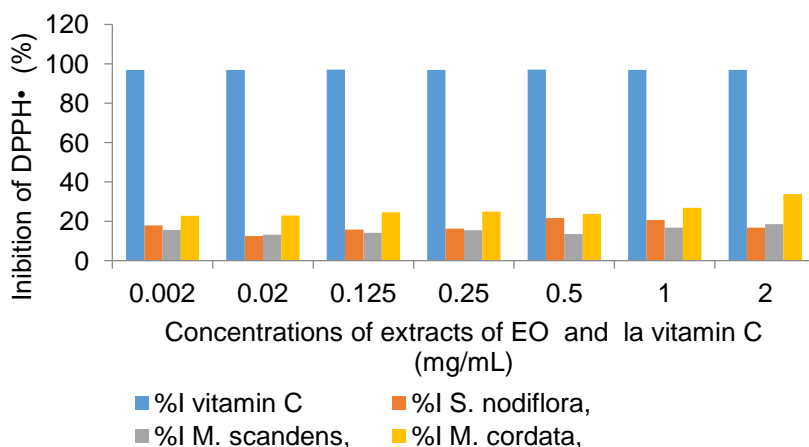
110 Four hundred (400) µL of the E O to be tested are taken and placed in the first sterile tube
111 containing 4.6 mL of BM-H medium, supplemented with Tween 80 (0.01%, v / v). The tube is
112 homogenized by vortexing. Then, a series of dilution in geometric progression is carried out
113 in Mueller -Hinton (BMH) -Tween 80 (0.01%, v / v) broth medium, so as to obtain a range of
114 concentrations of between 80 and 0.3 mg / mL. Finally, 13 µL of a bacterial inoculum, with a
115 density equivalent to Mac Farland standard 0.5 (10⁸ CFU. ML-1), are placed in each of the

116 tubes of the dilution range, which are then placed at 37 ° C under stirring for 24 h. A control
 117 of the bacterial growth for which 13 µL of the standardized inoculum were deposited in BMH-
 118 Tween 80 medium (0.01%, v / v), is also carried out. MIC is the smallest concentration of
 119 extract capable of inhibiting bacterial growth. The smaller is the most effective extract [14].
 120 The Minimum Bactericidal Concentration (MBC) corresponds to the lowest concentration
 121 capable of killing 99.99% of the initial inoculum. The same range of concentrations is used.
 122 Samples are taken in the control tube and in each of the tubes devoid of bacterial pellet and
 123 then deposited in «stria» on Mueller Hinton agar (MHA) . The inoculated dishes are
 124 incubated for 24 hours at 37 ° C [14].
 125 MBC / MIC ratios were calculated. When this ratio is less than 4, the extract is considered to
 126 be bactericidal. When it is 4, the extract is considered bacteriostatic [14].
 127 .

128 3. RESULTS AND DISCUSSION

129 3.1 Results of antioxidant activity

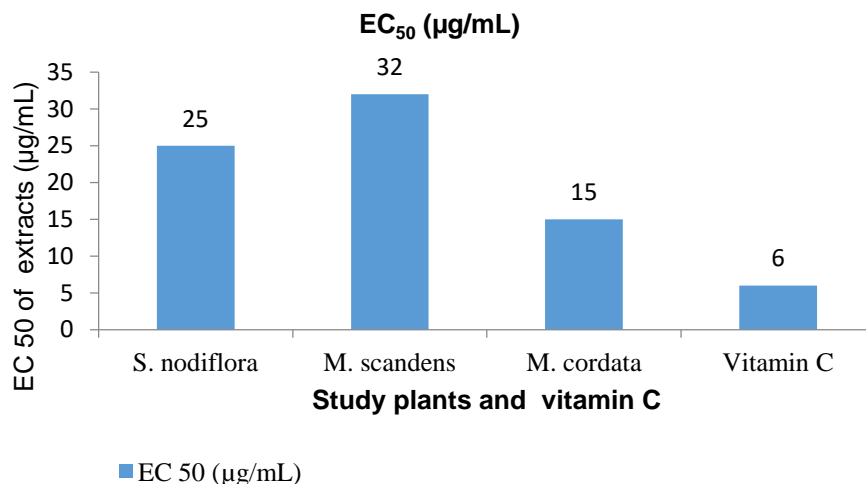
131 These results (Figure 1) show that the extracted EOs exhibit relatively low antioxidant
 132 activity compared to vitamin C; This is proven by the determination of the EC50 (figure 2).
 133 Indeed, the EC50 is defined as the concentration of the substrate which causes the loss of
 134 50% of the activity of DPPH• [12]. The lower this concentration, the more effective the
 135 extract.



136
 137 **Figure 1: Inhibition of DPPH• as a function of the concentration of EOs and vitamin C**

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139 The Essential Oils have a capacity of reduction of the free radical. The concentrations
 140 required for the neutralization of DPPH• vary between 32 µg / mL (Figure 2).



141

142 EC₅₀: effective substrate concentration

143

144 **Figure 2: EC₅₀ of EO from study plants and vitamin C**

145

146 The EC₅₀ of *M. cordata* extract is twice that of *M. scandens*. Therefore, the antioxidant
 147 activity of EO from *M. cordata* is twice as effective as that of EO from *M. scandens*. The
 148 EC₅₀ values show that the EO of *M. cordata* is more anti-free radical than that of *S. nodiflora*
 149 which is also more antioxidant than the EO extract of *M. scandens*. Furthermore, the EC₅₀
 150 of the extracted EOs was compared with that of vitamin C (6 µg / mL) determined by
 151 N'gaman [15] because the EC₅₀ of vitamin C could not be determined for this study. It is two
 152 times smaller than that of *Mikania cordata*, four times smaller than that of *Synedrella*
 153 *nodiflora* and five times smaller than that of *M. scandens*. It follows that ascorbic acid
 154 (vitamin C) is twice as effective as extract of *M. cordata*, four times more anti-free radical
 155 than EO of *S. nodiflora* and six times more antioxidant than extract of *M. scandens*.

156 The relative antioxidant activity of EOs could justify the use of the plants from which they are
 157 derived in traditional therapy. This antioxidant activity is linked to the presence of terpenes
 158 because they are endowed with antioxidant power.

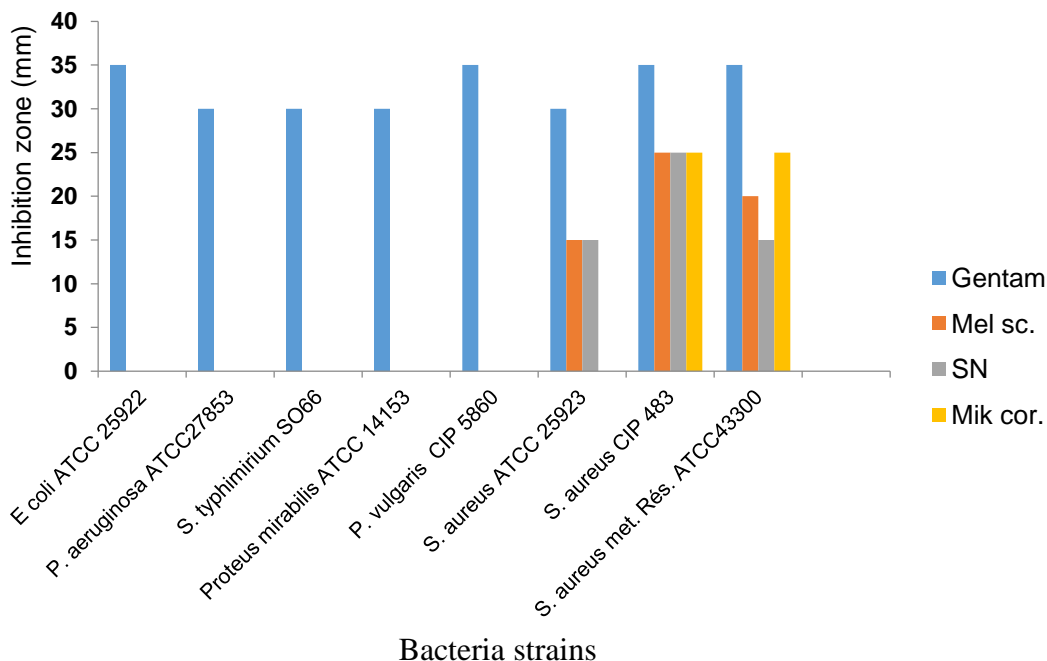
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160 **3-2-Results of antibacterial activity.**

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162 Antibacterial screening was performed. The diameters of the zones of inhibition are
 163 highlighted (Figure 3).

164 These results show that not all Gram (-) bacteria are sensitive to the EO extracts tested
 165 because there is no zone of inhibition of microorganisms. However, gentamycin has a strong
 166 inhibitory action on the growth of all the strains tested (d> 20 mm). All EO extracts are active
 167 against *S. aureus* CIP 483 and *S. aureus* methicilin resistant ATCC 43300, although it is
 168 resistant to methicilin.



169

170 Gentam: gentamicyn; Mel sc: Eo of *Melanthera scandens*; SN: EO of *synedrella nodiflora*;
 171 Mik cor: EO of *Mikania cordata*.

172

173 **Figure 3: Zones of inhibition of bacteria by EOs and gentamycin at 1 mg / mL**

174

175 The diameters of the zones of inhibition vary between 14 and 25 mm. It follows that EO
 176 extracts have remarkable activities on these bacteria. It should be noted that for the
 177 concentration of 1 mg / mL, all the EOs have a strong activity on *S. aureus* CIP 483 (d = 25
 178 mm). Furthermore, *S. Aureus* ATCC 25923 is only sensitive to extracts of *Melanthera*
 179 *scandens* and *Synedrella nodiflora*. The diameter of the zones of inhibition is 15 mm
 180 compared to 30 mm for the reference antibiotic. So these EOs have an average activity
 181 against *S. aureus* ATCC 25923 (14 <d <20 mm). The EO of *Mikania cordata* is inactive on *S.*
 182 *aureus* ATCC 25923. The comparison of the chemical composition of the different extracts of
 183 EO reveals the absence of oxygenated monoterpenes in the EO of *M. cordata* when they are
 184 low. Amounts in extracts of *M. Scandens* (0.58%) and *S. nodiflora* (0.5%). The absence of
 185 EO activity of *Mikania cordata* on *S. aureus* strain ATCC 25923 is believed to be due to the
 186 absence of oxygenated monoterpenes. Moreover, concerning the majority compounds, in
 187 the extract of EO of *Mikania cordata*, the levels of α -caryophyllene (6.95%) and β -
 188 caryophyllene (8.45%) are low compared to the levels in the extracts. EO of *M. scandens*
 189 and *S. nodiflora*. Antibacterial activity could also depend on the percentage of major milk
 190 constituents.

191 Not all Gram (-) bacterial strains are sensitive to essential oil extracts. This would be due to
 192 the structure of their outer membrane. Indeed, the outer membrane of Gram (-) bacteria is
 193 rich in lipopolysaccharide. It makes the bacteria more hydrophilic, which prevents
 194 hydrophobic terpenes from adhering to it [16].

195 The results, of the MICs and MBCs determined and of the MBC / MIC ratios calculated of
 196 the extracted EOs and of gentamycin on the strains *S. aureus* CIP 483 and *S. aureus*
 197 methicilin resistant ATCC 43300, are given in Table1

198

199

200

201 **Table 1: CMI, CMB and CMB / CMI ratios of extracts from EO and gentamycin**

202

Souche	S. aureus methicilin resistant			S. aureus CIP 483		
	CMI mg/mL	CMB mg/mL	CMB/CMI	CMI mg/mL	CMB mg/mL	CMB/MC
Mel sc	2.5	10	4	10	20	2
Mik cor	5	20	4	5	40	8
SN	5	20	4	10	80	8
Gentamicine	0,005	0,01	2	0,0025	0,0025	1

203 *MIC: Minimum inhibitory concentration; MBC: Minimum bactericidal concentration; Mel sc:*
204 *EO extract from M. scandens ; SN: EO extract from S. nodiflora; Mikcor: EO extract from M.*
205 *cordata*

206

207 All EOs extracts have a bacteriostatic effect on the methicilin resistant *Staphylococcus*
208 *aureus* strain (CMB / CMI = 4) while the *Melanthera scandens* extract has a bactericidal
209 effect (CMB / CMI = 2) on the *S. aureus* strain CIP 483. This activity is similar to that of
210 gentamycin on the resistant strain. These results would explain the use of *M. scandens*
211 leaves in traditional medicine to treat certain pathologies such as malaria [17], diarrhea,
212 dysentery, other gastrointestinal diseases [18] and diseases caused by fungi [7]. Although
213 there are no studies in the literature on the antimicrobial activity of the essential oil of
214 *Melanthera scandens*, the profile of the volatile compounds it contains, according to various
215 studies, confirms the activity that it can exercise.

216 Indeed, according to Ultée et al., Mustafa et al, the antibacterial activity of EOs is mainly
217 linked to the nature of their major compounds [19, 20]. But according to Delaquis et al. [21],
218 the antimicrobial activity of certain EOs could be due to the presence of minority
219 components. These compounds could exhibit an antibacterial activity by phenomena of
220 synergy between the various constituents, much more pronounced than that foreseeable of
221 the majority constituents. Thus, the antibacterial effects of EOs could be explained mainly by
222 the presence of terpenes (monoterpenes and sesquiterpenes) and phenolic compounds.
223 This is because the hydroxyls of the phenolic compounds are able to bind to the active sites
224 of the target enzymes by hydrogen bonds. The presence of sesquiterpene alcohols in the
225 EO of *M. scandens* could also explain this activity. Terpene alcohols are known for their
226 antimicrobial power due to their solubility in water; which gives them an ability to penetrate
227 bacterial cells [22]. The potential antibacterial activity of the essential oil of *Melanthera*
228 *scandens*, obviously, seems to be linked to the presence of secondary metabolites [23-25].

229 Staphylococci are known for their involvement in food contamination and as agents causing
230 many pathologies. Their antibiotic resistance is also known. It therefore appears imperative
231 to find alternative antibiotics which could not only be used to eradicate certain pathologies
232 but also as preservatives for certain foods such as juices and yoghurts [26]. EO from *M.*
233 *scandens'* organs could also be used as an alternative to reduce these risks of food
234 contamination.

235

236 **4. CONCLUSION**

237

238 The study of antioxidant activity by the DPPH•test has shown that the EOs analyzed, all
239 have less antioxidant activity than that of vitamin C, taken as a reference antioxidant.

240 The evaluation of the antibacterial activity of EO has given satisfactory results on all Gram
241 (+) bacteria. The diameters of the zones of inhibition vary between 15 and 25 mm. In

242 contrast, not all extracts have activity against all Gram (-) bacteria. *Melanthera scandens* EO
243 shows antibacterial potential against *Staphylococcus aureus* CIP 483.

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251

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256

257 **COMPETING INTERESTS**

258

259 Authors have declared that no competing interests exist. The products used for this research
260 are commonly and predominantly use products in our area of research and country. There is
261 absolutely no conflict of interest between the authors and producers of the products because
262 we do not intend to use these products as an avenue for any litigation but for the
263 advancement of knowledge. Also, the research was not funded by the producing company
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265

266 **AUTHORS' CONTRIBUTIONS**

267

268 This work was carried out in collaboration among all authors. Author KNS managed the
269 bibliographical searches, wrote the protocol and the first edition of the manuscript. Author
270 KKV help to evaluated the antioxidant activity of essential oils.. Author KBA provided the
271 material and technical assistance for the study. Authors MBJA is the scientific supervisor.
272 BY-A is the scientific director. All the authors read and approved the final manuscript.

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276 **NOTE:**

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278 **THE STUDY HIGHLIGHTS THE EFFICACY OF " TRADITIONAL MEDICINE "**
279 **WHICH IS AN ANCIENT TRADITION, USED IN SOME PARTS OF INDIA. THIS**
280 **ANCIENT CONCEPT SHOULD BE CAREFULLY EVALUATED IN THE LIGHT OF**
281 **MODERN MEDICAL SCIENCE AND CAN BE UTILIZED PARTIALLY IF FOUND**
282 **SUITABLE.**

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