

Assessment of heavy metal pollution in red seaweed collected from intertidal coastal areas in Zanzibar by using EDXRF technique

11 **ABSTRACT**

12

Few studies have been reported the heavy metals concentrations in red seaweed comprises cottonii and spinosum species throughout the world. This study was designed to assess heavy metals (Mn, Zn, Ni, Cu, Fe, As and Pb) contaminations in cottonii and spinosum species commonly farmed in Unguja and Pemba, Zanzibar. The species were collected from intertidal coastal areas and concentration of the assessed metals were carried out using Energy Dispersive X-ray Fluorescence (EDXRF). The samples of cottonii in Unguja had significantly ($p \leq 0.05$) higher concentrations of Fe, Zn, Ni, Cu and Pb than in spinosum, while the samples of spinosum had significantly higher levels of As and Mn. Likewise, the samples of cottonii in Pemba had significantly ($p \leq 0.05$) higher concentrations of Ni, Cu and Pb than in spinosum, whereas the samples of spinosum had significantly higher levels of Fe, As and Zn. The concentrations of heavy metals in cottonii and spinosum in the present study were noticed lower compared to publish results. However, the heavy metals contaminations in cottonii and spinosum along the coastal area in Zanzibar were mainly caused by effluents that directly flow into marine environments.

13
14
15
16

Keywords: Heavy metals; Seaweed; Zanzibar; EDXRF

17 1. INTRODUCTION

18
19
20
21
22
23
24
25
26
27
28
29
30
31

Heavy metals are released into the environment following various industrial activities, domestic wastewater, geological weathering of the earth crust, shipping and harbour activities, and atmospheric deposition [1]. The metals can enter into the aquatic environment and deposited into marine organism via the effects of bioaccumulation and bio-concentration through the food chain process and become toxic when accumulation reaches a substantially high level [2]. The contamination of these metals into aquatic environment posing a major effects of marine plants which is directly related to anthropogenic activities. The adverse effects of heavy metals such as As, Mn, Hg, Zn, Cd, Cu and Pb have raised public concern about the safety due to their toxicity level, non-degradable and persistent in the environment [3-5]. It has been also reported that metals such as As, Cd, Pb and Hg have a high degree of toxicities even at trace levels and ranks among the priority metals that are of public health significant [3]. Heavy metals pollution does not respect any boundaries and may discharge into marine environment by various anthropogenic activities [6, 7]. The contamination of the marine ecosystem by heavy metals pollution negatively affects marine species including seaweeds and may pose considerable environmental risks and concerns [8, 9]. Although seaweeds provide essential nutritional elements, the increasing significance of heavy metals disturbing the environment is especially evident in aquatic systems [10].

32
33
34
35
36

Seaweeds were identifying as bio-indicators of heavy metals contamination in the marine ecosystem for detecting mineralization and anthropogenic impact of coastal marine communities [11-13]. Studies have shown that heavy metals pollution can be found in varying concentrations in marine plants species including the seaweeds [14-16]. Moreover, in the studies of Khalid et al. 2014 and Qari, 2015 reported separately the presence of heavy metals in seaweed by using Atomic Absorption Spectroscopy (AAS) [17, 18]. Mutia et al. 2018 has also found the presence of As, Pb, Cd and Hg in

two seaweed species namely as *Ulva rigida* and *Halimeda opuntia* by using AAS [19]. In Zanzibar, a very limited number of studies have been published about the accumulation of heavy metals of seaweeds. The aim of this study was to assess the heavy metals in *Eucaema denticulatum* (*cottonii*) and *Eucaema spinosum* cultivated in Zanzibar islands by using Energy dispersive X-ray fluorescence (EDXRF). The EDXRF has been shown to be a suitable technique for multi-element analysis, no chemical pre-treatment and small amount of sample is required, minimising sample contamination and the wide range of elements which may simultaneously be detected.

2. MATERIAL AND METHODS

2.1 Description of the study area

The samples analysed in this study were collected from the intertidal coastal areas in Zanzibar Islands with a population of about 1.3 million Peoples [20]. Zanzibar comprises the Zanzibar Archipelago in the Indian Ocean, 25-50 kilometres off the eastern coast of Mainland Tanzania. In the western part of the Indian Ocean, between latitudes 50 and 60 south of the equator and between 390 and 450 east of the Greenwich Meridian. It comprises Unguja and Pemba as main islands and numerous small isles, the first being the biggest of the two. In Zanzibar, peoples lived near to marine environment and seaweed farming is one of the major significant economic activities. The seaweed farming activities mainly take place along intertidal coastal regions and sampling areas includes; Chokocho liko kuu, Chokocho kisiwa panza, Kangaani kuukuu, Wingwi kitaalani, Micheweni shumba mjini, Tumbe, Makunduchi, Bweleo, Paje kikwaju jeuri, Muungoni duta, Jambiani mbuyuni, Kidoti Bondeni, Bwejuu and Uroa (Figure. 1). These sampling areas have been selected based on the richness of the *Spinosum* and *Cottonii* species, proximity to small scale industrial activities and effluents that directly flow into marine environments.

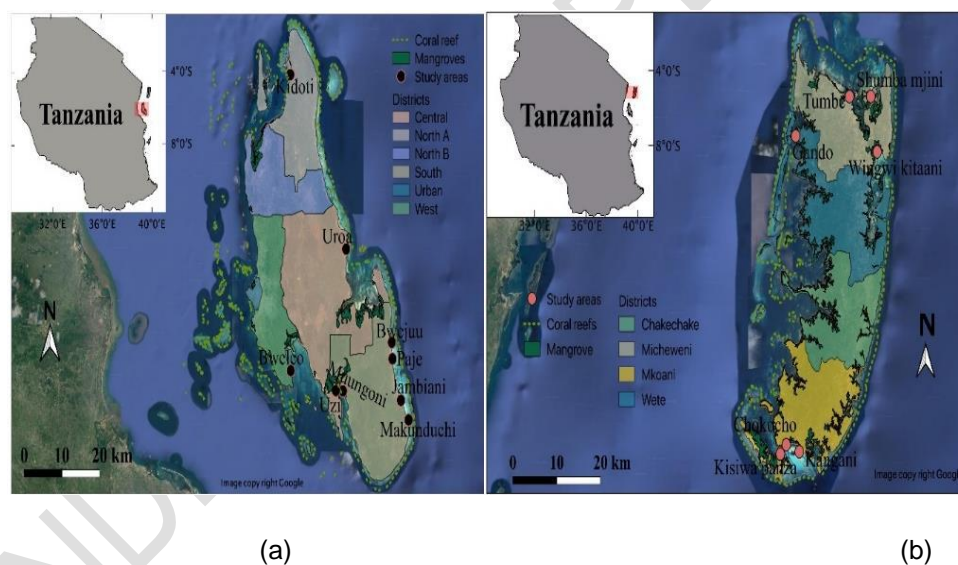


Fig. 1. Map of Zanzibar showing the sampling points (a) Unguja (b) Pemba

1.2 Sampling and Sampling Preparation

Ten samples of *Spinosum* and ten samples of *Cottonii* were collected randomly from 15 different intertidal coastal areas in Zanzibar. The collected samples washed up on the seas water to remove epiphytes and coarse debris and then placed in labelled plastic bags. Upon arrival at the laboratory, the samples were rinsed with distilled water to remove any traces of elements resulting from contamination. Thereafter, all samples were first air dried for 72 hours and then were oven dried for 24 hours at 70 °C to obtain a constant weight. The dried samples were then grinded into fine powder using pestle and mortar. The powdered materials in each sample species were sieved through 2-mm polystyrene sieve.

1.3 XRF preparation

A dry weight of 4 g of each sample with 0.9 g of cellulose binder was put into a bowl together with four spherical balls each with 3 mm radius and fixed to pulveriser which was further grinded and homogenised. The Pulverized machine was set at a speed of 150 revolutions per minutes (rpm) for 10 minutes. By using hydraulic Retsch™ machine, the samples

were compressed into pellets through application of pressure of 15 tones. The pellets were labelled and taken to the EDXRF machine for analysis. The model of the system is Xepos with serial No. 4R0138 and is operated by using X-lab ProTM computer software in which matrix effects was counted for. In this model, X-rays are generated by the X-ray tube built in EDXRF. The instrument use three different secondary targets to increase the excitation sensitivity of elements, the Molybdenum secondary target for (K-Line Cr-Y) and L-Line from (Hf-U). Aluminium oxide (Al₂O₃) polarization target (For L-Lines Zr-Ce) and for High Oriented Pure Graphite (HOPG) Bragg crystal (K-Lines from Na-V). The spectra running time was 30 minutes per sample.

Concentrations of elements in the samples were calculated by the inbuilt software called X-lab ProTM with Turboquant (Tq 9232) algorithm for matrix effect correction [21]. The software corrects for the matrix effects (M_i) and the interference effects (K_i) basing on fundamental parameter methodology. The software corrects also for the background effect on a spectral line intensity (I_i), given as counts per second (cps). After all the corrections, the software converts the intensity into concentration of the element using Equation (1) below [22].

$$C_i = K_i \times I_i \times M_a \quad (1)$$

where C_i is the concentration of a given element i , M_a is the correction factor for matrix effects. K_i is the constant of proportionality, I_i is the intensity of the fluorescent radiation from the element i .

1.4 Quality control

Quality control was carried out using NIST standard reference material Trace and Minor Elements in Tomato Leaves (SRM 1573a) analysed with samples. As Table shows, the analysed elements were within the range of standard reference materials.

Table 1: The comparison between the experimental values (mgkg⁻¹) of standards reference material (SRM 1573a Tomato leaves) and reference values.

Elements	Experimental values	Refence values
Mg	11550	12000
Al	549	598
K	25086	27000
Ca	49351	50500
Cr	1.44	1.99
Mn	275	246
Fe	309	368
Cu	4.17	4.7
Zn	28	30.9
Rb	16.6	14.89
Sr	77	85

1.5 Statistical analysis

One-way ANOVA was used to evaluate the differences among vegetables species. Prior to ANOVA, the homogeneities of the variances were verified using Levene's test. A t-test was used to statistically compare the mean concentrations of elements collected from the two groups (farm site and local market). A probability level of $p < 0.05$ was considered statistically significant. All data were presented as arithmetic mean with standard deviation attached. All statistical analyses were made using the software Excel 2013 and SPSS Version 23, and figures were produced using Origin Version 8.5 software.

3. RESULTS AND DISCUSSION

3.1 Heavy metals concentration in spinosum

The heavy metals in spinosum samples collected from intertidal coastal areas in Unguja and Pemba and their mean concentration values in ($\mu\text{g/g}$) were found. The results at Unguja showed that the mean concentration values of Mn, Zn, Ni, Cu, Fe, As and Pb in spinosum were $44.94 \mu\text{g/g}$, $8.16 \mu\text{g/g}$, $1.76 \mu\text{g/g}$, $2.97 \mu\text{g/g}$, $788.67 \mu\text{g/g}$, $10.59 \mu\text{g/g}$ and $3.83 \mu\text{g/g}$ respectively. The heavy metals detected found in the order of $\text{Fe} > \text{Mn} > \text{As} > \text{Zn} > \text{Pb} > \text{Cu} > \text{Ni}$. The ranges of mean concentration values of these metals were: Mn ($12.8\text{--}44$) $\mu\text{g/g}$, Zn ($1.3\text{--}8.16$) $\mu\text{g/g}$, Ni ($1.24\text{--}1.76$) $\mu\text{g/g}$, Cu ($1.99\text{--}2.97$) $\mu\text{g/g}$, Fe ($180.71\text{--}788.67$) $\mu\text{g/g}$, As ($5.46\text{--}10.59$) $\mu\text{g/g}$ and Pb ($1.45\text{--}3.83$) $\mu\text{g/g}$. Out of seven metals examined, the highest mean concentration of Fe ($180.71\text{--}788.67$) $\mu\text{g/g}$ was observed and the lowest mean value of Ni ($1.24\text{--}1.76$) $\mu\text{g/g}$ was recorded. Likewise, the results at Pemba showed that the mean values of heavy metals in samples of spinosum ranged from $10.98\text{--}53.06 \mu\text{g/g}$ for Mn, $2.33\text{--}11.97 \mu\text{g/g}$ for Zn, $1.04\text{--}1.89 \mu\text{g/g}$ for Ni, $1.42\text{--}2.65 \mu\text{g/g}$ for Cu, $230.20\text{--}1696.86 \mu\text{g/g}$ for Fe, $7.26\text{--}11.25 \mu\text{g/g}$ for As and $1.44\text{--}2.79 \mu\text{g/g}$ for Pb. Moreover, the results showed that the mean concentrations of heavy metals detected in spinosum found in the order of $\text{Fe} > \text{Mn} > \text{As} > \text{Zn} > \text{Pb} > \text{Cu} > \text{Ni}$. The t-test from IBM SPSS software version 23 was used to statistically compare the mean concentrations of heavy metals recorded in samples of spinosum and cottonii from Unguja and Pemba. In this test, the significant value of less than a 0.05 probability was taken.

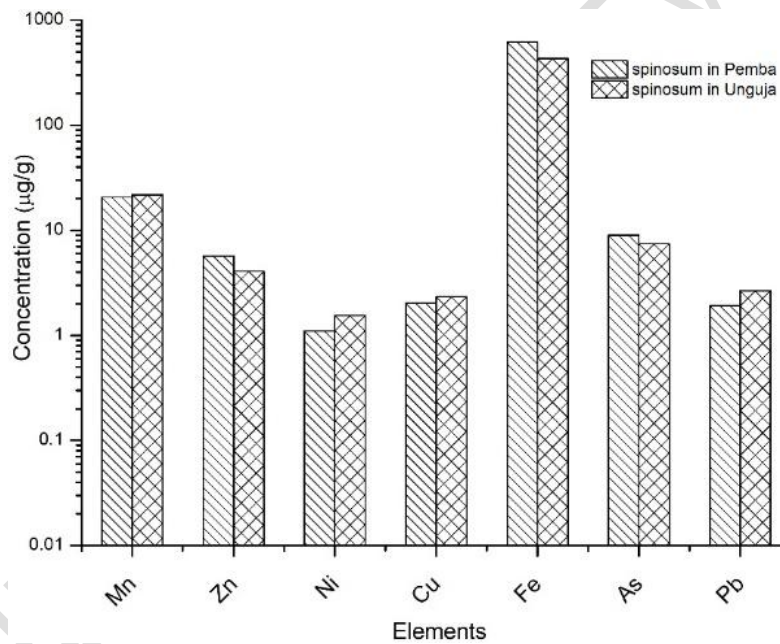


Fig. 2. A histogram showing comparison of heavy metals concentrations in samples from spinosum at Unguja and Pemba.

Samples from Pemba had significant ($p \leq 0.05$) higher mean concentration of Fe, Zn and As, while samples from Unguja had significantly higher concentrations of Mn, Ni, Cu and Pb (Figure 2). Besides Fe, the highest mean concentration values recorded in spinosum was for Mn followed by As, Zn, Pb and Cu and the minimum value was observed for Ni which conforms to the result reported by [23]. Moreover, the mean values of Pb and Cu concentrations were lower than reported in the study of Dadolahi-Sohrab et al. 2011[24]. The lower values in our study might be attributed to the difference in the red seaweed species. Dadolahi-Sohrab et al. 2011 [24] used *acanthophora specifera* species which is also red seaweed while in this study used spinosum to assess heavy metals levels. The total arsenic value ranged from $0.114\text{--}236 \mu\text{g/g}$, for 953 dried seaweed food samples throughout the world was reported by WHO technical report series 959 [25]. In the present study, the highest contents of As found in spinosum and cottonii were $6.77 \mu\text{g/g}$ and $11.25 \mu\text{g/g}$, these values were lower compared to reported value by WHO.

167

168 3.2 Heavy metals concentration in cottonii

169 The concentrations of Mn, Zn, Ni, Cu, Fe, As and Pb in the samples of cottonii collected from Unguja and Pemba are
 170 given in Tables 2 and 3. The levels of heavy metals in Unguja were found to vary from 18.27–19.48 µg/g for Mn,
 171 4.7–10.62 µg/g for Zn, 1.78–2.16 µg/g for Ni, 2.34 –3.36 µg/g for Cu, 425.54–666.77 µg/g for Fe, 3.5–6.16 µg/g for As
 172 and 1.83–4.62 µg/g for Pb; while in Pemba, the levels were ranged from 11.41–32.09 µg/g for Mn, 2.61–6.32 µg/g for Zn,
 173 1.37–1.92 µg/g for Ni, 2.56 –3.24 µg/g for Cu, 401.89–828.44 µg/g for Fe, 4.97–6.77 µg/g for As and 3.75–4.18 µg/g for
 174 Pb.

175 The heavy metals distribution pattern decreased in the order: Fe > Mn > As > Zn > Pb > Cu > Ni. Samples from Unguja
 176 (S1, S2 and S3) had significant ($p \leq 0.05$) higher mean concentrations of Mn, Zn, Ni, Cu, Pb and Fe, while samples from
 177 Pemba (S4, S5 and S6) had significantly higher mean concentrations of As (Figure 3). However, the concentrations of Pb,
 178 Cu and Ni from samples in Pemba and Unguja were lower compared with the study of Dadolahi-Sohrab et al. 2011 [24].
 179 Higher concentrations of Cu and Zn particular in samples S1, S2 and S3 have been attributed to the discharge of
 180 municipal wastewater to the marine environment.

181

182

183

184

185

186

187

188

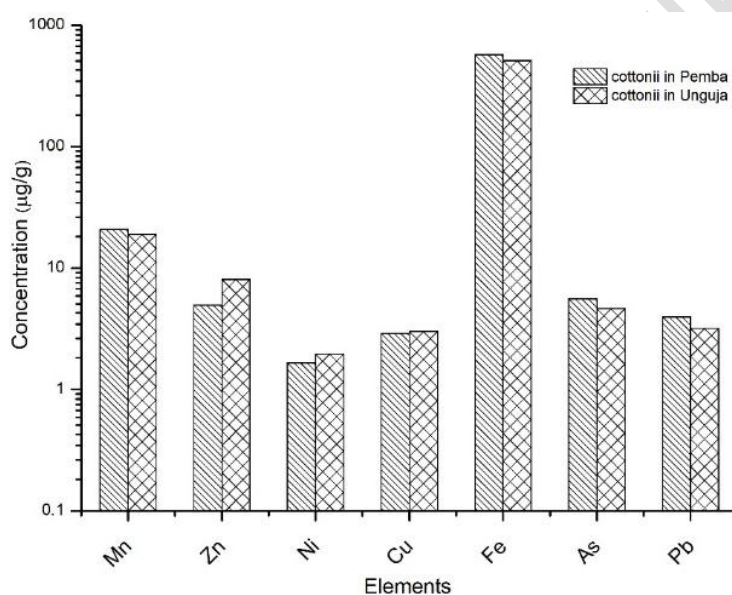
189

190

191

192

193



194 **Fig. 3. A histogram showing a comparison of heavy metals concentrations in samples from cottonii at Unguja**
 195 **and Pemba.**

196

197 The concentrations of As in samples of cottonii in the present study lie between 3.50– 6.16 µg/g in Unguja and 4.97–6.77
 198 µg/g in Pemba. As indicated in Table 2, the highest levels of 6.16 µg/g was detected at Uzi in Unguja, while 6.77 µg/g was
 199 detected at chokocho kisiwa panza in Pemba as shown in Table 3. The values of As concentrations in samples of cottonii
 200 were strongly supported by the findings ranged from 1.72–9.46 µg/g which was reported by Ryu et al., 2009 [26].

201

202 **Table 2: Mean ± SD concentrations (µg/g) of heavy metals in cottonii from Bweleo (S4), Muongoni duta (S5) and**
 203 **Uzi (S6), Unguja.**

Site	Elements						
	Mn	Zn	Ni	Cu	Fe	As	Pb
S4	11.41±1.09	2.61±0.73	1.37±0.97	BDL	401.89±12.56	6.77±0.54	BDL
S5	32.09±7.75	6.32±0.90	1.68±0.26	2.56±0.11	828.44±11.81	5.01±0.40	4.18±0.13
S6	19.03±7.20	5.92±0.84	1.94±0.01	3.40±1.09	487.18±9.88	4.97±1.13	3.75±1.36

204 **Note:** BDL=below detection limit

205
206
207
208
209

Table 3: Mean \pm SD concentrations ($\mu\text{g/g}$) of heavy metals in cottonii from Chokocho kisiwa panza (S1), Kangaani kuukuu (S2) and Chokocho liko kuu (S3), Pemba.

Site	Elements						
	Mn	Zn	Ni	Cu	Fe	As	Pb
SI	19.22 \pm 6.84	8.91 \pm 0.58	1.78 \pm 0.28	2.34 \pm 0.41	446.81 \pm 10.82	4.27 \pm 1.38	3.13 \pm 1.01
S2	18.27 \pm 4.04	4.70 \pm 0.52	2.16 \pm 0.44	3.36 \pm 0.50	425.54 \pm 9.58	3.50 \pm 1.03	4.62 \pm 0.29
S3	19.48 \pm 5.16	10.62 \pm 0.69	1.94 \pm 0.37	3.33 \pm 0.89	666.77 \pm 13.07	6.16 \pm 1.13	1.83 \pm 1.29

210
211
212
213
214
215
216
217
218
219

Figures 4 (a) and (b) compare the mean concentrations of metals in cottonii and spinosum collected from Pemba and Unguja. The t-test shows that samples of cottonii in Unguja had significantly ($p \leq 0.05$) higher mean concentrations of Fe, Zn, Ni, Cu and Pb than in spinosum. The samples of spinosum had significantly higher mean concentrations of As and Mn as shown in Figure 4 (a). Likewise, samples of cottonii in Pemba had significantly ($p \leq 0.05$) higher mean concentrations of Ni, Cu and Pb than in spinosum. Samples of spinosum had significantly higher mean concentrations of Fe, As and Zn as shown in Figure 4 (b). In both samples from Unguja and Pemba, mean concentrations of Fe were much higher compared to Zn; for instant, the mean concentrations of Fe in samples of cottonii and spinosum from Unguja were 63.5 and 105.7 times higher than that of Zn.

220
221

222
223
224
225
226
227
228
229
230
231
232
233
234
235
236
237
238
239

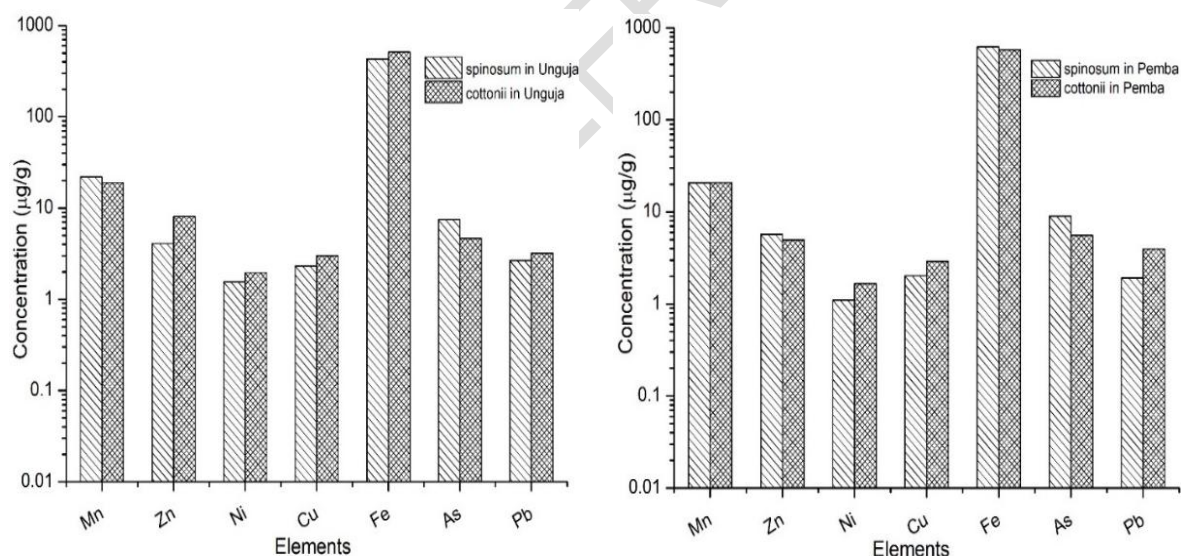


Fig. 4. The histograms showing the comparison of heavy metals concentrations in samples from cottonii and spinosum at Unguja and Pemba.

240
241
242
243
244
245
246
247
248
249
250

In this study, the heavy metals of these species of red seaweed commonly farmed in Zanzibar were detected. The levels of Pb in samples of cottonii collected from Unguja and Pemba were much higher compared to spinosum, this was because of the lubricating oil from diesel and engine from boat [27]. Generally, the levels of heavy metals contaminations in both samples along coastal area in Zanzibar might be caused by waste effluents that directly flow into the marine environment. Thus, the waste effluents from various anthropogenic activities were shown to contain heavy metals including Fe, Zn, Ni, Cu, As and Pb [28-30]. Ni and Mn, were not reported in any of the literature reviewed, however, they were found in low concentrations in both samples.

251 **CONCLUSION**

252
253 The present study illustrates the application of the EDXRF technique in assessment of heavy metals contamination in red
254 seaweed comprises cottonii and spinosum species collected from intertidal coastal areas in Zanzibar. The t-test from IBM
255 SPSS computer software version 23 was used to statistically compare the mean concentrations of heavy metals recorded
256 in samples of spinosum and cottonii from Unguja and Pemba. In this test, the significant value of less than a 0.05
257 probability was taken. The levels of heavy metals contaminations in cottonii and spinosum in this study were noticed
258 generally lower compared to published results. However, samples of cottonii in Unguju had significantly ($p \leq 0.05$) higher
259 mean concentrations of Fe, Zn, Ni, Cu and Pb than in spinosum while the samples of spinosum had significantly higher
260 concentrations of As and Mn. Likewise, the samples of cottonii in Pemba had significantly ($p \leq 0.05$) higher mean
261 concentrations of Ni, Cu and Pb than in spinosum, whereas the samples of spinosum had significantly higher mean
262 concentrations of Fe, As and Zn. The concentrations of Pb in samples of cottonii collected from Unguja and Pemba were
263 much higher compared to spinosum, this might be caused by lubricating oil from diesel and engine from the boat (Aballah
264 et al., 2005). Generally, the heavy metals contaminations detected in cottonii and spinosum along the coastal area in
265 Zanzibar were mainly caused by waste effluents that directly flow into marine environments. Thus serious measures
266 should be taken to reduce the flow of pollutants into the marine ecosystem.

267
268 **ACKNOWLEDGEMENTS**

269
270 This work was supported by the Tanzania Atomic Energy Commission (TAEC). The authors are highly acknowledged and
271 express their gratitude to high support from TAEC management and technicians that provided necessary laboratory
272 facilities.
273

COMPETING INTERESTS

Authors have declared that no competing of interests exist.

AUTHORS' CONTRIBUTIONS

This work was carried out in collaboration between both authors. Author SAS, SKA and MKA have designed the study, performed the statistical analysis and literature searches and wrote the first draft of the manuscript. Authors AAM and FOM supervised the analyses of the samples, reviewed the first draft and wrote the final manuscript. All authors read and approved the final manuscript.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

REFERENCES

1. Yilmaz F. The comparison of heavy metal concentrations (cd, cu, mn, pb, and zn) in tissues of three economically important fish (*Anguilla anguilla*, *Mugil cephalus* and *Oreochromis niloticus*) Inhabiting Koycegiz Lake-Mugla (Turkey). *Turkish Journal of Science and Technology*. 2009; 4(1): 7-15.
2. Huang WB. Heavy metal concentrations in the common benthic fishes caught from the coastal waters of eastern Taiwan. *Journal of Food and Drug Analysis*. 2003; 11(4): 324-330.
3. Tchounwou PB, Yedjou CG, Patlolla AK, Sutton DJ. Heavy metal toxicity and the environment. *Molecular, clinical and environmental toxicology*. Springer, basel. 2012; 101: 133-164. DOI: 10.1007/978-3-7643-8340-4_6.
4. Xiong T, Dumat C, Pierart A, Shahid M, Kang Y, Li N, Bertoni G, Laplanche C. Measurement of metal bioaccessibility in vegetables to improve human exposure assessments: field study of soil–plant–atmosphere transfers in urban areas, south china. *Environmental geochemistry and health*. 2016; 38(6): 1283-1301. DOI: 10.1007/S10653-016-9796-2.
5. Khalid S, Shahid M, Niazi NK, Murtaza B, Bibi I, Dumat C. A comparison of technologies for remediation of heavy metal contaminated soils. *Journal of geochemical exploration*. 2017; 182: 247-268. <https://doi.org/10.1016/j.gexplo.2016.11.021>.
6. Al naggar Y, Khalil MS, Ghorab MA. Environmental pollution by heavy metals in the aquatic ecosystems of Egypt. *Open acc j of toxicol*. 2018; 3(1): 555603. DOI: 10.19080/OAJT.2018.03.555603.
7. Wu B, Wang G, Wu J, Fu Q, Liu C. Sources of Heavy Metals in Surface Sediments and an Ecological Risk Assessment from Two Adjacent Plateau Reservoirs. *Plos One*. 2014; 9(7): e102101. <https://doi.org/10.1371/journal.pone.0102101>.
8. Singh A, Singh DR, Yadav HK. Impact and assessment of heavy metal toxicity on water quality, edible fishes and sediments in lakes: a review. *Trends. Biosci*. 2017; 10(8); 1551-1560.
9. Tornero V, Hanke G. Chemical contaminants entering the marine environment from sea-based sources: a review with a focus on European seas. *Marine Pollution Bulletin*. 2016; 112 (1-2): 17-38. <https://doi.org/10.1016/j.marpolbul.2016.06.091>
10. Sudharsan S, Seedeve P, Ramasamy P, Subhapradha N, Vairamani S, Shanmugam A. Heavy metal accumulation in seaweeds and sea grasses along southeast coast of india. *Journal of chemical and pharmaceutical research*. 2012; 4(9): 4240-4244.
11. [11] AKCALI, I.; KUCUKSEZGIN, F. A biomonitoring study: heavy metals in macroalgae from eastern Aegean coastal areas. *Marine pollution bulletin*. v. 62, p. 637-645, 2011.
12. Søndergaard J, Bach I, Gustavson K. Measuring bioavailable metals using diffusive gradients in thin films (DGT) and transplanted seaweed (*Fucus vesiculosus*), blue mussels (*Mytilus edulis*) and sea snails (*Littorina saxatilis*) suspended from monitoring buoys near a former lead–zinc mine in West Greenland. *Marine pollution bulletin*. 2014; 78 (1-2): 102-109. <https://doi.org/10.1016/j.marpolbul.2013.10.054>.
13. Chakraborty S, Owens G. Metal distributions in seawater, sediment and marine benthic macroalgae from the South Australian coastline. *International Journal of Environmental Science and Technology*. 2014; 11(5): 1259-1270. <https://doi.org/10.1007/s13762-013-0310-4>.

- 331 14. Ali AHA, Hamed MA, Abd El-Azim H. Heavy metals distribution in the coral reef ecosystems of the Northern Red
332 Sea. *Helgoland marine research*. 2011; 65 (1): 67-80. DOI 10.1007/s10152-010-0202-7.
- 333 15. Bai I, Liu XI, Hu J, Li J, Wang ZI, Han G, Li S, Liu CQ. Heavy metal accumulation in common aquatic plants in rivers
334 and lakes in the Taihu Basin. *International journal of environmental research and public health*. 2018; 15(12): 2857.
335 DOI: 10.3390/ijerph15122857.
- 336 16. Ali H, Khan E, Ilahi I. Environmental chemistry and ecotoxicology of hazardous heavy metals: environmental
337 persistence, toxicity, and bioaccumulation. *Journal of chemistry*. 2019; 1-14. <https://doi.org/10.1155/2019/6730305>.
- 338 17. Khaled A, Hessein A, Abdel-halim AM, Morsy FM. Distribution of heavy metals in seaweeds collected along Marsa-
339 Matrouh beaches, Egyptian Mediterranean Sea. *The Egyptian Journal of Aquatic Research*. 2014; 40(4): 363-371.
340 <https://doi.org/10.1016/j.ejar.2014.11.007>
- 341 18. Qari R. Heavy metals concentrations in brown seaweed *Padina Pavonia* (L.) and *P. tetrastromatica* at different
342 beaches of Karachi Coast. *Indian Journal of Geo-Marine Sciences*. 2015; 44(8): 1200-1206.
- 343 19. Mutia GM, Matern MS. Analysis of bio-accumulation of heavy metals in seaweeds *ulva rigida* and *halimeda opuntia* in
344 validation of their safety for use in aquaculture feeds in Kenya. *IOSR Journal of Environmental Science, Toxicology*
345 *and Food Technology*. 2018; 12(8): 56-63. DOI: 10.9790/2402-1208015663.
- 346 20. Tanzania population and housing census. 2012.
- 347 21. Schramm R, Heckel J. Fast analysis of traces and major elements with EDXRF using polarized X-rays: turboquant. *J.*
348 *Phys*. 1998; 8(PR5): 335–342. <https://doi.org/10.1051/jp4:1998542>.
- 349 22. Rousseau RM, Bouchard M. Fundamental algorithm between concentration and intensity in XRF analysis 2: practical
350 application. *X-ray Spectrom*. 2005; 13(3):121-125.
- 351 23. Qari R, Siddiqui SA. A comparative study of heavy metal concentrations in red seaweeds from different coastal areas
352 of Karachi, Arabian Sea. *Indian Journal of Marine Sciences*. 2010; 39(1): 27-42.
- 353 24. Dadolahi-Sohrab A, Nikvarz A, Nabavi SMB, Safahyeh A, Ketal-Mohseni M. Environmental monitoring of heavy
354 metals in seaweed and associated sediment from the Strait of Hormuz, IR Iran. *World Journal of Fish and Marine*
355 *Sciences*. 2011; 3(6): 576-589.
- 356 25. World Health Organization (WHO), 2011. WHO Technical Report Series, No. 959. Evaluation of certain contaminants
357 in food. 72 nd report of the Joint FAO/WHO Expert Committee on Food Additives. Available at: <https://tinyurl.com/y954aryj>.
- 358 26. Ryu KY, Shim SI, Hwang IM, Jung MS, Jun SN, Seo HY, Park JS, Kim HY, Om AS, Park KS, Kim KS. Arsenic
359 speciation and risk assesment of Hijiki (*Hizikia fusiforme*) by HPLC-ICP-MS. *Korean Journal of Food Science and*
360 *Technology*. 2009; 41(1): 1-6.
- 361 27. Abdallah AMA, Abdallah MA, Beltagy AI. Contents of heavy metals in marine seaweeds from the Egyptian coast of
362 the Red Sea. *Chemistry and Ecology*. 2005; 21(5): 399-411. <https://doi.org/10.1080/02757540500290222>.
- 363 28. Alalwan HA, Kadhom MA, Alminshid AH. Removal of heavy metals from wastewater using agricultural
364 byproducts. *Journal of Water Supply: Research and Technology-AQUA*. 2020; 69(2): 99-112.
365 <https://doi.org/10.2166/aqua.2020.133>.
- 366 29. Burakov AE, Galunin EV, Burakova IV, Kucherova AE, Agarwal S, Tkachev AG, Gupta VK. Adsorption of heavy
367 metals on conventional and nanostructured materials for wastewater treatment purposes: A review. *Ecotoxicology and*
368 *environmental safety*. 2018; 148: 702-712. <https://doi.org/10.1016/j.ecoenv.2017.11.034>.
- 369 30. Baysal A, Ozbek N, Akman S. Determination of trace metals in waste water and their removal processes. *Intech Open*
370 *Science, Wastewater treatment technologies and recent analytical developments. Waste water*. 2013; 20: 145-171.
371 DOI: 10.5772/52025.
- 372
- 373