

Investigation of the Phytochemical Constituents of diverse Epiphytes in the Federal Capital Territory, Nigeria

Abstract

A study on the diversity and classification of epiphytes in the Federal Capital Territory (FCT) was conducted on thirty (30) species of epiphytes. As an extension of the research work, fifteen (15) epiphytic plants were selected on the basis of data collected by administering semi-structured questionnaires to 90 respondents. The interest of this work is to investigate the phytochemical constituents of *Ficus platyphylla*, *Senna mimosifolia*, *Platynerium stemaria*, *Nephrolepis bisserata*, *Calyptrichium emerginatum*, *Frullania dilatata*, *Perperomia pellucida*, *Plagiothecium undulatum*, *Pitrogramma calomelanos*, *Caprinus lagopides*, *Auricularia polytrichia*, *Formitopsis* sp, *Lycopodium clavatum* and *Entodon seductrix* in the FCT, to determine their **ethnobotanical** uses. A qualitative **and quantitative** phytochemical study procedures were **carried out using flavonoids, saponins, alkaloids and tannins**. The result of the qualitative analysis on the diverse epiphytes showed that flavonoids, saponins, alkaloids and tanins were either present or moderately present in each of the epiphytes while that of quantitative indicated that the flavonoid content varied between 4.29 mg/100 and 52.61

mg/100. *Calyptechium emerginatum* has the highest flavonoid content of 52.61 while the lowest was *Entodontopsis nitens* with 4.29.

Keywords: phytochemicals, epiphytes, constituents, medicinal, biological

Introduction:

The classes of compounds found in epiphytes often trigger and encourage the diverse use of epiphytes in the medicinal treatment of diseases such as cancer (Katta *et al.*, 2019). Modern researchers seek to explore the basis of plant use in various areas of human activity. In many habitats, epiphytes have been used as antibiotics, decorations, dyes, natural remedies, stabilizers for fragrance and food for thousands of years (Alamgir, 2017). It is a centuries-old culture and tradition in most African communities to use epiphytes as vegetables, ornamental plants and for the treatment of diseases. FCT, Nigeria is not to be outdone in this regard. The need to develop baseline data on the phytochemical constituents and ethnobotany of **diverse epiphytes in the FCT** cannot be overemphasized; hence the basis of this study.

Epiphytes are sources of food and habitat for other organisms (Migiro 2019). **They are also** good bio-indicators due to their low tolerance to changes in their environment (Akhaltatsi *et al.*, 2014).

In Brazil, the leaves of *Loranthus rotundifolius* Engl. cooked in milk have been used to treat chest diseases (Subhashini *et al.*, 2019). An ointment prepared from the young shoots and leaves of *L. citrocolus* is a well-known remedy for edematous tumours (Shanavaskhan *et al.*, 2012). *Cypripedium parviflora* is widely used as an aphrodisiac and nerve tonic (Singh *et al.*, 2012).

In the mid-eighteenth century, regular crops were severely affected in Europe by frosts and droughts causing famine, and as a result, lichens were used for food due to their easy availability, cheapness, and nutritional value (Luczaj and Pieroni, 2016).

Most lichens are poisonous, although there are some exceptions. *Letharia vulpine* (L.) has been traditionally used to poison predators and treat stomach disorders; more recently, *extracts of L. vulpine* have demonstrated promising antimicrobial properties (Shrestha *et al.*, 2016). According to Gray in GBIF Secretariat (2023), *Cetraria pinastri* (Scop.) Gray is used as a poison for wolves. *Bryoria fremontii* is edible whereas other *Bryoria* species are mildly toxic (Chandler *et al.* 2020). *B. tortuosa* is a well-known toxic lichen rich in vulpinic acid or pinastrinic acid (Spribille *et al.* 2016). They have also been used in traditional foods for millennia and play a critical role in ecosystem functioning and human well-being (Crawford, 2015).

Bryophytes can contribute significantly to total stream metabolism, nutrient cycling, interactions with food webs in streams, and as a direct food source for some invertebrates (Tessler *et al.* 2014). Some species are a great source for herbal medicine (Sabovljevic *et al.*, 2016). However, to date, there has been no attempt to document FCT bryophytes as to their medicinal properties, ethnic and other uses.

Traditional communities depend on wild plants for food, healing, and building materials, firewood, and almost all other material crops (Vira *et al.*, 2015).

Different parts of the world have greatly helped the modern world to benefit from the traditional knowledge systems (Kumar *et al.*, 2021). This has been recognized, mainly in the areas of developing promising life-saving drugs, including plant-based psychotomimetics (Crawford, 2015), by gaining knowledge about the traditional land and plant use model (Bradai *et al.*, 2015), evolving strategies for the conservation of biological diversity and environmental management policies (Santamaria and Mendaz, 2012) and the search for promising new economic plants and land races (Shah, 2014). Tribals living in the area are known to have a great knowledge of the medicinal uses of many of these plants (Mahomoodally, 2013). Because of the difficulty of obtaining the plants from other higher plants, information on these groups is very scarce. No one has so far attempted to record this information. Therefore, the present study is a new endeavor to **establish** the epiphytes of FCT, and record ethnobotanical information from this group of plants.

Materials and Methods



Auricularia polytricha



Caprinus lagopides



Platycerium stemaria



Nephrolepis bisserata



Frullania dilatata

Formitopsis sp



Lycopodium clavatum



Entodon sedatrix



Plagiothecium undulatum



Peperomia pelucida



Nephrolepis undulata and *Ageratum conyzoides*



Ficus platyphylla

Plates: Photos of epiphytes phytochemical analysis was carried out on in the Federal Capital Territory

The Federal Capital Territory (FCT) is situated in the Middle Belt region of Nigeria. It was formed in 1976 from parts of the states of old kwara, Niger, Kaduna and Plateau states (Federal Capital Territory Administration - Facts, 2008).

The FCT is made up of six area councils namely: Abaji, Bwari, Gwagwalada, Kuje, Kwali and Municipal area councils (Figure 1).

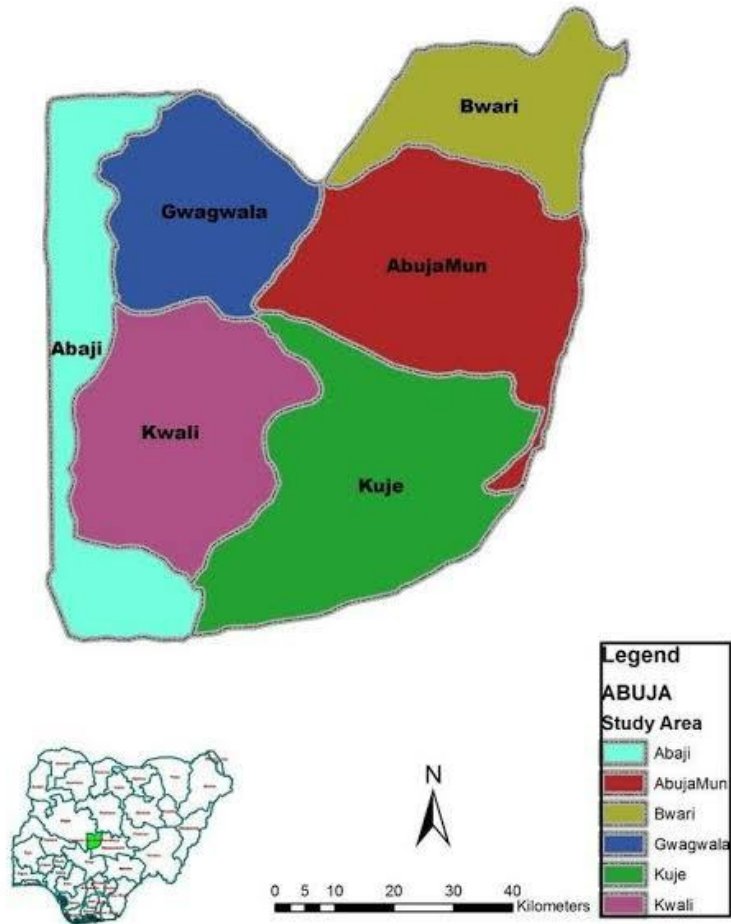


Figure 1: Modified from the Administrative Map of Nigeria: Map of the Federal Capital Territory

Source: Adeiza *et al.*, 2023

According to Ezejiofor *et al.*, (2024), a reconnaissance survey of the study area was carried out from November to December, 2021. The study sites were selected randomly, considering the dry and riparian nature of the region, altitudinal ranges, notable presence of epiphytes diversity and recommendation from respondents.

Data was collected by administering semi-structured questionnaires to 90 respondents for a single purpose face-to-face interview (Ezejiofor *et al.*, 2024).

All the epiphytes were collated and analyzed using ethnobotany and diversity information. This data comprised of the species name, plant family, areas of the ethnobotany (Ezejiofor *et al.*, 2024).

Qualitative and quantitative phytochemical survey procedure of epiphytes were done as described by Yadav and Agarwala (2011).

The Kolmogorov-Smirnov test was used to test if data were normally distributed before any parametric statistics such as the ANOVA, Correlation analysis was deployed.

Test for alkaloids

- a. Mayer's test

To a few ml of plant sample extract, two drops of Mayer's reagent were added along the sides of the tube. The appearance of a creamy white precipitate indicated the presence of alkaloids.

b. Wagner's test

A few drops of Wagner's reagent were added to a few ml of plant extract along the sides of the test tube. A reddish-brown precipitate confirmed that the test was positive.

Test for saponins

The extract (100 mg) was dissolved in 10 ml of distilled water and composed of 20 ml.

The suspension was shaken in a graduated cylinder for 15 minutes. A two-cm layer of foam indicated the presence of saponins. (no picture taken).

Test for phenolic compounds and tannins

a. Ferric Chloride Test

The extract (50 mg) was dissolved in 5 ml of distilled water. To this, a few drops of neutral 5% ferric chloride solution were added. A dark green colour indicated the presence of a phenolic compound. (no picture taken).

b. Lead Acetate Testing

The extract (50 mg) was dissolved in distilled and to which 3 ml of 10% lead acetate solution was added. A large white precipitate indicated the presence of phenolic compounds. (no picture taken).

c. Magnesium and hydrochloric acid reduction

The extract (50 mg) was dissolved in 5 ml of alcohol and a few fragments of magnesium tape and concentrated hydrochloric acid (drip) were added. A pink to crimson color developed, the presence of flavonol glucosides was inferred. (no picture taken).

Quantitative Phytochemical Investigation Procedure

Test for saponins:

The extract (1 g) was macerated with 10 ml of petroleum ether and transferred to a beaker. Another 10 ml of petroleum ether was added to the beaker and the filtrate evaporated until it became dry. The residue was dissolved in 6 ml of ethanol. The solution (2 ml) was put into a test tube and 2 ml of chromagen solution was added. It was left to rest for 30 minutes and the absorbance was read at 550 nm. (no picture taken).

Test for alkaloids

The extract (1g) was macerated with 20ml of ethanol and 20% hydrogen sulphate (H₂SO₄), (1:1 V/V). The filtrate (1 mL) was added to 5 mL of 60% H₂SO₄. After 5 minutes, 5 mL of 0.5% formaldehyde in 60% H₂SO₄ was mixed into the mixture and allowed to sit for 3 hours. The absorbance was read at 565 nm. (no picture taken).

Test for phenolic compounds and tannins

To test for phenolic compounds, the extract (1 g) was macerated with 20 ml of 80% ethanol and then filtered. The filtrate (5 mL) was added to 0.5 mL of folinciocalteus reagent and allowed to sit for 30 minutes, and absorbance was measured at 650 nm. (no picture taken).

To test the tannins, the extract (1g) was macerated with 50ml of methanol and filtered. To the filtrate (5 mL), add 0.3 mL of 0.1N ferric chloride in 0.1N hydrogen chloride (HCl) and 0.3 mL of 0.0008M potassium ferricyanide, and the absorbance was read at 720 nm. (no picture taken)

A qualitative analysis was carried out on the diverse epiphytes and it showed that flavonoids, saponins, alkaloids and tanins were either present or moderately present in each of the epiphytes as indicated in table 1 below.

Table 1: Qualitative Phytochemical Analysis of the Selected Epiphytes (mg/100g)

Epiphytes	Flavonoids	Saponins	Alkaloids	Tannins
<i>Ageratum conyzoides</i>	++	+	++	++
<i>Ficus platyphylla</i>	++	++	++	++
<i>Senna mimosifolia</i>	++	++	++	+
<i>Platycerium stemaria</i>	+	++	++	+
<i>Nephrolepis sp</i>	++	++	++	++
<i>Calypotechium emerginatum</i>	++	++	++	+
<i>Frullaria dilatata</i>	+	++	++	++
<i>Perperomia pelucida</i>	++	++	++	+
<i>Plagiothecium undulatum</i>	+	++	+	++
<i>Pitrogramma sp</i>	++	++	++	++
<i>Caprinus lagopides</i>	+	++	++	+
<i>Auricularia polytrichia</i>	+	++	++	++
<i>Formitopsis sp</i>	+	++	++	++
<i>Lycopodon spadiceus</i>	+	++	++	++
<i>Entodontopsis nitens</i>	+	+	++	+

KEY:

+ = Present

++ = Moderately Present

Table 2: Replicates Quantitative Phytochemical Analysis of Diverse Epiphytes

(mg/100g)

Specie	Replicates	Flavonoid mg/100g	Saponin mg/100g	Alkaloid mg/100g	Tannin mg/100g
<i>Ageratum conyzoides</i>		44.40	19.75	25.32	17.20
		45.71	20.15	26.57	18.47
		44.89	19.21	25.21	17.65
<i>Ficus platyphylla</i>		33.18	24.85	21.23	26.28
		34.59	25.63	20.16	26.74
		31.74	25.82	21.98	26.00
<i>Senna mimosifolia</i>		16.58	15.48	18.26	19.18
		17.95	15.11	19.47	19.03
		16.20	16.74	18.12	20.15
<i>Platyserium stemaria</i>		9.68	13.29	20.15	20.23
		10.63	14.58	22.37	22.17
		10.87	13.01	20.87	21.10
<i>Nephrolepis bisserata</i>		27.48	26.17	20.41	21.11
		26.58	26.99	23.17	20.98
		28.17	27.65	22.64	22.31
<i>Calytrochilum emerginatum</i>		52.31	15.33	20.38	20.30
		52.04	16.43	21.65	20.87
		53.49	15.10	21.10	21.37
<i>Frullania dilatata</i>		6.87	18.27	16.73	26.36
		6.15	20.18	17.70	28.65
		7.13	19.43	16.34	27.15
<i>Perperomia pellucida</i>		49.84	10.75	21.61	22.95
		51.73	11.32	20.74	23.46
		50.32	10.56	22.15	23.87
<i>Plagiothecium undulatum</i>		10.20	23.28	15.73	24.03
		10.98	24.36	16.34	24.95
		10.04	24.87	17.47	23.87
<i>Pitrgroma cameloanos</i>		45.86	21.91	21.69	18.59
		45.21	22.34	20.98	19.46
		47.27	21.67	21.51	18.43
<i>Caprinus lagopides</i>		4.61	14.36	18.79	27.84
		4.72	14.12	19.74	28.47
		4.24	14.98	18.33	27.54

<i>Auricula</i>	24.27	17.27	20.73	19.83
<i>polytrichia</i>	24.93	18.45	21.42	20.34
	25.16	18.84	20.56	19.31
<i>Formitopsis</i>	16.78	20.18	14.82	26.27
<i>sp</i>	18.47	20.44	14.45	26.23
	17.33	21.31	15.32	27.65
<i>Lycopodon</i>	11.42	16.71	16.73	20.76
<i>clavatum</i>	12.38	17.36	17.84	22.10
	11.87	17.88	16.21	21.27
<i>Entodon</i>	4.02	9.78	19.60	15.88
<i>seductrix</i>	4.64	10.21	20.13	16.48
	4.21	11.32	19.87	15.42

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Table 3: Quantitative Phytochemical Analysis of diverse Epiphytes (mg/100g)

Epiphytes	Flavonoids	Saponins	Alkaloids	Tannins
<i>Ageratum conyzoides</i>	45.00	19.07	25.67	17.77
<i>Ficus platyphylla</i>	33.17	25.43	31.68	26.34
<i>Senna mimifolia</i>	16.91	15.77	18.68	19.45
<i>Platycerium stemaria</i>	10.39	13.62	21.13	21.16
<i>Nephrolepis sp</i>	27.41	26.93	22.07	21.46
<i>Calyptechium emerginatum</i>	52.61	15.62	21.04	20.84
<i>Frullaria dilatata</i>	6.71	19.29	16.92	27.38
<i>Perperomia pelucida</i>	50.63	10.87	21.50	23.42
<i>Plagiothecium</i>	10.40	24.17	16.51	24.28

<i>undulatum</i>				
<i>Pitogramma sp</i>	46.11	52.00	21.39	18.82
<i>Caprinus</i>	4.52	14.48	18.95	27.95
<i>lagopides</i>				
<i>Auricularia</i>	24.78	18.18	20.90	19.82
<i>polytrichia</i>				
<i>Formitopsis sp</i>	17.52	20.64	14.86	26.95
<i>Lycopodon</i>	11.89	17.31	16.92	21.37
<i>spadiceus</i>				
<i>Entodontopsis</i>	4.29	10.43	19.86	15.92
<i>nitens</i>				

Quantitative analysis for **Flavonoids**

Method

Null hypothesis All means are equal

Alternative hypothesis At least one mean is different

Significance level $\alpha = 0.05$

Equal variances were assumed for the analysis.

Factor Information

Factor Levels Values

Factor 15 C1, C2, C3, C4, C5, C6, C7, C8, C9, C10, C11, C12, C13, C14, C15

Analysis of Variance

Source	DF	Adj SS	Adj MS	F-Value	P-Value
Factor	14	1049.17	74.9406	160.12	0.000
Error	30	14.04	0.4680		
Total	44	1063.21			

Model Summary

S	R-sq	R-sq(adj)	R-sq(pred)
0.684131	98.68%	98.06%	97.03%

Means

Factor	N	Mean	StDev	95% CI
C1	3	19.703	0.472	(18.897, 20.510)
C2	3	25.433	0.514	(24.627, 26.240)
C3	3	15.777	0.855	(14.970, 16.583)
C4	3	13.627	0.837	(12.820, 14.433)
C5	3	26.937	0.741	(26.130, 27.743)
C6	3	15.620	0.711	(14.813, 16.427)
C7	3	19.293	0.962	(18.487, 20.100)
C8	3	10.877	0.396	(10.070, 11.683)
C9	3	24.170	0.812	(23.363, 24.977)
C10	3	21.973	0.339	(21.167, 22.780)
C11	3	14.487	0.444	(13.680, 15.293)
C12	3	18.187	0.817	(17.380, 18.993)
C13	3	20.643	0.592	(19.837, 21.450)
C14	3	17.317	0.586	(16.510, 18.123)
C15	3	10.437	0.795	(9.630, 11.243)

Pooled StDev = 0.684131

Quantitative analysis for Saponnins

Method

Null hypothesis All means are equal

Alternative hypothesis At least one mean is different

Significance level $\alpha = 0.05$

Equal variances were assumed for the analysis.

Factor Information

Factor Levels Values

Factor 15 C1, C2, C3, C4, C5, C6, C7, C8, C9, C10, C11, C12, C13, C14, C15

Analysis of Variance

Source	DF	Adj SS	Adj MS	F-Value	P-Value
Factor	14	1049.17	74.9406	160.12	0.000
Error	30	14.04	0.4680		
Total	44	1063.21			

Model Summary

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C5	3	26.937	0.741	(26.130, 27.743)

C6	3	15.620	0.711	(14.813, 16.427)
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C8	3	10.877	0.396	(10.070, 11.683)
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C12	3	18.187	0.817	(17.380, 18.993)
C13	3	20.643	0.592	(19.837, 21.450)
C14	3	17.317	0.586	(16.510, 18.123)
C15	3	10.437	0.795	(9.630, 11.243)

Pooled StDev = 0.684131

Quantitative analysis for alkaloids

Method

Null hypothesis All means are equal

Alternative hypothesis At least one mean is different

Significance level $\alpha = 0.05$

Equal variances were assumed for the analysis.

Factor Information

Factor Levels Values

Factor 25.23, 21.23, 18.46, 20.15, 20.41, 20.38, 16.73, 21.61, 15.73, 21.69,
18.79, 20.75, 14.82, 16.72, 19.60

Analysis of Variance

Source	DF	Adj SS	Adj MS	F-Value	P-Value
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Factor	14	218.53	15.6090	21.86	0.000
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Error 15 10.71 0.7141

Total 29 229.24

Model Summary

S R-sq R-sq(adj) R-sq(pred)

0.845035 95.33% 90.97% 81.31%

Means

Factor N Mean StDev 95% CI

25.23 2 25.890 0.962 (24.616, 27.164)

21.23 2 21.070 1.287 (19.796, 22.344)

18.46 2 18.795 0.955 (17.521, 20.069)

20.15 2 21.620 1.061 (20.346, 22.894)

20.41 2 22.905 0.375 (21.631, 24.179)

20.38 2 21.375 0.389 (20.101, 22.649)

16.73 2 17.020 0.962 (15.746, 18.294)

21.61 2 21.445 0.997 (20.171, 22.719)

15.73 2 16.905 0.799 (15.631, 18.179)

21.69 2 21.245 0.375 (19.971, 22.519)

18.79 2 19.035 0.997 (17.761, 20.309)

20.75 2 20.990 0.608 (19.716, 22.264)

14.82 2 14.885 0.615 (13.611, 16.159)

16.72 2 17.025 1.153 (15.751, 18.299)

19.60 2 20.000 0.184 (18.726, 21.274)

Pooled StDev = 0.845035

Quantitative analysis for Tannins

Method

Null hypothesis All means are equal

Alternative hypothesis At least one mean is different

Significance level $\alpha = 0.05$

Equal variances were assumed for the analysis.

Factor Information

Factor Levels Values

Factor 13, 3, 4, 5, 6, 7, 8, 9, 12, 30, 32, unk1, unk2, unk3

Analysis of Variance

Source	DF	Adj SS	Adj MS	F-Value	P-Value
Factor	12	469.17	39.0978	83.52	0.000
Error	26	12.17	0.4681		
Total	38	481.34			

Model Summary

S R-sq R-sq(adj) R-sq(pred)
0.684188 97.47% 96.30% 94.31%

Means

Factor	N	Mean	StDev	95% CI
3	3	19.453	0.608	(18.641, 20.265)
4	3	21.167	0.972	(20.355, 21.979)
5	3	21.467	0.733	(20.655, 22.279)
6	3	20.847	0.535	(20.035, 21.659)
7	3	27.387	1.163	(26.575, 28.199)
8	3	23.427	0.461	(22.615, 24.239)
9	3	24.283	0.583	(23.471, 25.095)
12	3	18.827	0.554	(18.015, 19.639)
30	3	27.950	0.475	(27.138, 28.762)
32	3	19.827	0.515	(19.015, 20.639)
unk1	3	26.950	0.710	(26.138, 27.762)
unk2	3	21.377	0.676	(20.565, 22.189)
unk3	3	15.927	0.532	(15.115, 16.739)

Pooled StDev = 0.684188

A quantitative analysis of the flavonoid components of diverse epiphytes of the FCT was carried out in replicates. The result showed that the flavonoid content of the studied epiphytes ranged between 4.29 mg/100 to 52.61 mg/100. *Calypotechium emerginatum* has the highest flavonoid content of

52.61 while the lowest was *Entodontopsis nitens* with 4.29 as earlier shown in table 3.

Using analysis of variance, the quantitative analysis of the flavonoid contents of the selected epiphytes showed a significant difference among the epiphytes ($P < 0.000$). The F – Value (160.12) and P – Value (0.000) communicating the significance difference.

A quantitative analysis of the saponins components of selected epiphytes of the FCT was carried out in replicates. The result showed that the saponins content of the studied epiphytes ranged between 52.00 to 10.43. *Pitrogramma sp* has the highest saponins content of 52.00 while the lowest was *Entodontopsis nitens* with 10.43 as initially indicated in table 3 above.

Using analysis of variance, the quantitative analysis of the saponins contents of the selected epiphytes showed a significant difference among the epiphytes ($P < 0.000$). The F – Value (160.12) and P – Value (0.000) communicating the significance difference.

A quantitative analysis of the alkaloid components of selected epiphytes of the FCT was carried out in replicates. The result indicated that the alkaloid content of the studied epiphytes ranged between 14.86 to 31.68. *Ficus*

platyphylla has the highest alkaloid content of 31.68 while the lowest was *Formitopsis sp* with 14.86 as shown in table 3 above.

Using analysis of variance, the quantitative comparative analysis of the alkaloid contents of the selected epiphytes showed a significant difference among the epiphytes ($P < 0.000$). The F – Value (21.86) and P – Value (0.000) communicating the significance difference.

A quantitative analysis of the tannins components of diverse epiphytes of the FCT was carried out in replicates. The result indicated that the tannins content of the studied epiphytes ranged between 15.92 to 27.38. *Frullaria dilatata* has the highest tannins content of 27.38 while the lowest was *Entodonpsis nitens* with 15.92 as represented in table 3 above.

Using analysis of variance, the quantitative comparative analysis of the tannins contents of the selected epiphytes showed a significant difference among the epiphytes ($P < 0.000$). The F – Value (83.52) and P – Value (0.000) communicating the significance difference.

Test for alkaloids

a. Mayer's test

To a few ml of plant sample extract, two drops of Mayer's reagent were added along the sides of the tube. The appearance of a creamy white precipitate indicated the presence of alkaloids.

b. Wagner's test

A few drops of Wagner's reagent were added to a few ml of plant extract along the sides of the test tube. A reddish-brown precipitate confirmed that the test was positive.

Test for saponins

The extract (100 mg) was dissolved in 10 ml of distilled water and composed of 20 ml.

The suspension was shaken in a graduated cylinder for 15 minutes. A two-cm layer of foam indicated the presence of saponins. (no picture taken)

Test for phenolic compounds and tannins

has. Ferric Chloride Test

The extract (50 mg) was dissolved in 5 ml of distilled water. To this, a few drops of neutral 5% ferric chloride solution were added. A dark green colour indicated the presence of a phenolic compound. (no picture taken)

b. Lead Acetate Testing

The extract (50 mg) was dissolved in distilled and to which 3 ml of 10% lead acetate solution was added. A large white precipitate indicated the presence of phenolic compounds. (no picture taken)

c. Magnesium and hydrochloric acid reduction

The extract (50 mg) was dissolved in 5 ml of alcohol and a few fragments of magnesium tape and concentrated hydrochloric acid (drip) were added. A pink to crimson color developed, the presence of flavonol glucosides was inferred. (no picture taken)

Quantitative Phytochemical Investigation Procedure

Test for saponins:

The extract (1 g) was macerated with 10 ml of petroleum ether and transferred to a beaker. Another 10 ml of petroleum ether was added to the beaker and the filtrate evaporated until it became dry. The residue was dissolved in 6 ml of ethanol. The solution (2 ml) was put into a test tube and 2 ml of chromagen solution was added. It was left to rest for 30 minutes and the absorbance was read at 550 nm. (no picture taken)

Test for alkaloids

The extract (1g) was macerated with 20ml of ethanol and 20% hydrogen sulphate (H₂SO₄), (1:1 V/V). The filtrate (1 mL) was added to 5 mL of 60% H₂SO₄. After 5 minutes, 5 mL of 0.5% formaldehyde in 60% H₂SO₄ was mixed into the mixture and allowed to sit for 3 hours. The absorbance was read at 565 nm. (no picture taken)

Test for phenolic compounds and tannins

To test for phenolic compounds, the extract (1 g) was macerated with 20 ml of 80% ethanol and then filtered. The filtrate (5 mL) was added to 0.5 mL of folinciocalteus reagent and allowed to sit for 30 minutes, and absorbance was measured at 650 nm. (no picture taken)

To test the tannins, the extract (1g) was macerated with 50ml of methanol and filtered. To the filtrate (5 mL), add 0.3 mL of 0.1N ferric chloride in 0.1N hydrogen chloride (HCl) and 0.3 mL of 0.0008M potassium ferricyanide, and the absorbance was read at 720 nm. (no picture taken)

Result

The qualitative analysis on the diverse epiphytes showed that flavonoids, saponins, alkaloids and tanins were either present or moderately present in each of the epiphytes as represented in table 1 above.

Using analysis of variance, quantitative analysis of the flavonoid content of the selected epiphytes showed a significant difference between epiphytes ($P < 0.000$). The F-value (160.12) and the P-value (0.000) communicate the difference in significance.

Certains des épiphytes à savoir ; *Ageratum conyzoides*, *Ficus platyphylla*, *Senna mimosifolia*, *Platycerium stemaria*, *Nephrolepis sp*, *Calypotechium emerginatum*, *Frullaria dilatata*, *Perperomia pelucilia*, *Plagiothecium undulatum*, *Pitrogramma sp*, *Caprinus lagopides*, *Auricularia polytrichia*, *Formitopsis sp*, *Lycopodon spadiceus* and *Entodon nitens* were subjected to qualitative and quantitative phytochemical analysis to determine their phytochemical constituents. These epiphytes were selected based on data collected by administering semi-structured questionnaires to 90 respondents for a single-purpose face-to-face interview.

A quantitative analysis of the flavonoid components of some FCT epiphytes was performed in replicates. The result showed that the flavonoid content of the epiphytes studied varied between 4.29 mg/100 and 52.61 mg/100. *Calypotechium emerginatum* has the highest flavonoid content (52.61), while the lowest was *Entodontopsis nitens* (4.29), as shown in Table 2 below.

Using analysis of variance, quantitative analysis of the flavonoid content of the selected epiphytes showed a significant difference between epiphytes ($P < 0.05$).

The F-value (160.13) and the P < value 0.05 communicate the difference in significance.

Discussion

For many years, epiphytes have been a source of medicinal agents and a whole bunch of modern medicines have been isolated from epiphytic plants. This is because plants have the ability to produce a wide variety of secondary metabolites such as saponins, tannins, phenols, alkaloids, triterpens, and phytosterols. In the current qualitative analysis of diverse epiphytes in the Federal Capital Territory, the results indicated the presence of flavonoids, saponins, alkaloids and tannins in all plants (Table: 1). It has been reported that tannins have various physiological effects such as anti-irritant, antisecretolytic, Antiphlogistics, antimicrobials and antiparasitics (Hassanpour *et al.*, 2011). In addition, there are reports showing that tannin contains plants with phytotherapeutic effect. Phytochemical analysis of some plant extracts revealed the presence of constituents known to have medicinal values as well as physiological activity. Phytochemicals such as flavonoids, saponins, alkaloids and tannins have been associated with medicinal properties (Rabizadeh *et al.*, 2022). Phytochemicals are known to be the basic source for the

establishment of several pharmaceutical industries. Constituents present in plants play a crucial role in the identification of crude drug references. In addition, phytochemical screening is a very useful tool for identifying the new source of therapeutically and industrially important compounds. Epiphytes play an important role in the treatment of certain diseases, beautification and nutrition. This has undoubtedly led to other uses of *Ageratum conyzoides* L. as an analgesic, against fungal, inflammatory, anticoagulant activity, wound healing, dysentery, pesticides, and herbicides (Sivakrishnan and Kavitha, 2017). *Ficus platyphylla* Delile is used as a drug for the treatment of diarrhea, chest pain, cough, seizures, and pain (Ugwah-Oguejiofor *et al.*, 2021). *Senna mimosifolia* Mill is used to treat gastrointestinal diseases, it causes rectal cancer when used excessively (Oladeji *et al.* 2021). It is also used for weight loss by drinking tea made from its fresh leaves. Prolonged use of *Senna mimosifolia* tea can lead to liver degradation and cause bowel dysfunction (Oluwole *et al.*, 2021). According to the International Plant Names Index and the World Checklist of Vascular Plants (2024), *Platyserium stemaria* (P. Beauv.) Desv. Used in the treatment of diseases associated with blood circulation, liver disease, genital stimulation or depression, asthma and infectious diseases. The curled young leaves of *Nephrolepis bisserata* (Sw.) Schott are used as food for humans, medicines, biofertilizers, and ornaments (Shah *et al.* 2014).

Calyptrichilum emerginatum (Afzel.ex Sw.) Schltr. is used in the treatment of cough (Mathias *et al.*, 2006), tuberculosis and malaria (Okhale *et al.* 2014).

According to Plant Basel (2023), *Frullania dilatata* (L.) Dumort. causes intense allergic contact dermatitis *Perperomia pellucida* (L.) Kunth serves as a remedy for insect bites, sexually transmitted diseases, fever, cough, smallpox, measles, and kidney infections (Keat Lam *et al.*, 2022). *Plagiothecium undulatum* (Hedw.) Schimp. is used in the treatment of malaria in north-central Nigeria (Egbuomwan *et al.*, 2023). The leaves of *Pitrogramma calomelanos* (L.) Link ex Britton and Millsp. are used externally to heal wounds and stop bleeding (DeFilipps *et al.*, 2024). An infusion of the whole plant is used to strengthen the backs of men; i.e., increasing male sexual stamina and treating female bleeding (DeFilipps *et al.*, 2024). It is also used to treat asthma, coughs, colds, pneumonia, tuberculosis, and pertussis (DeFilipps *et al.*, 2024). *Caprinus lagopides* P.Karst. is not toxic (N'Douba Amako *et al.*, 2022). Its edibility is unknown, but it is considered too small to be worthwhile (Davis and Sommer 2014). *Auricularia polytrichia* (Mont.) Sacc. is used as an antioxidant, antitumor, immunomodulatory, hyperlipidemic, antidiabetic, anticoagulant, and hepatoprotectant (Miao *et al.*, 2020). *Formitopsis* sp (Sw.) P.Karst. has therapeutic effects, including anti-inflammatory, cytotoxic, and antimalarial effects (Muszynsk *et al.*, 2020). According to Plant Resources of Tropical Africa (2022), *Lycopodium clavatum* (L.) stimulates peristaltic

movements of the intestine and contraction of the uterus. The whole plant is chewed to induce vomiting after poisoning or sharp stomach pains and it is applied externally on skin diseases, wounds, ulcers, and irritations. *Entodon seductrix* (Hedw.) Müll.Hal. causes skin reactions, contact dermatitis, can be used to treat liver disorders, cardiovascular disease, fever, and wounds (Bandyopadhyay and Dey, 2022).

Research indicates that *Ageratum conyzoides* L. is used as an analgesic, against fungal, inflammatory, anticoagulant activity, wound healing, dysentery, pesticides and herbicides (Sivakrishnan and Kavitha, 2017). *Ficus platyphylla* Delile is used as a drug for the treatment of diarrhea, chest pain, cough, seizures, and pain (Ugwah-Oguejiofor *et al.*, 2021). *Senna mimosifolia* Mill is used to treat gastrointestinal diseases, it causes rectal cancer when used excessively (Oladeji *et al.*, 2021). It is also used for weight loss by drinking tea made from its fresh leaves. Prolonged use of *Senna mimosifolia* tea can lead to liver degradation and cause bowel dysfunction (Oluwole *et al.*, 2021). According to the International Plant Names Index and the World Checklist of Vascular Plants (2024), *Platyserium stemaria* (P. Beauv.) Desv. Used in the treatment of diseases associated with blood circulation, liver disease, genital stimulation or depression, asthma and infectious diseases. The curled young leaves of *Nephrolepis bisserata* (Sw.) Schott are used as food for humans, medicines, biofertilizers, and ornaments (Shah *et al.* 2014).

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contraction of the uterus. The whole plant is chewed to induce vomiting after poisoning or sharp stomach pains and it is applied externally on skin diseases, wounds, ulcers, and irritations. *Entodon seductrix* (Hedw.) Müll.Hal. causes skin reactions, contact dermatitis, can be used to treat liver disorders, cardiovascular disease, fever, and wounds (Bandyopadhyay and Dey, 2022). (This is the first investigation on epiphytes done in the Federal Capital Territory).

Conclusion

The present research concludes that some epiphytes are important medicinal plants and contain various active phytoconstituents. Phytochemicals such as flavonoids, saponins, alkaloids and tannins have been associated with medicinal properties. Qualitatively, they are either present or moderately present in each of the epiphytes understudied. Phytochemicals are also known to be the basic source for the establishment of several pharmaceutical industries. In addition, phytochemical screening is a very useful tool for identifying the new source of therapeutically and industrially important compounds. Constituents present in plants play a crucial role in the identification of crude drug references. These constituents have made some epiphytic plants poisonous, while others serve as food, medicine, or ornaments. It is therefore necessary to develop further research on these epiphytes to determine their medicinal value.

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REFERENCES

Adeiza, A., Sani, N., Daniel, N., Godwin, E. and Okoli, Elizabeth (2023). Assessment of Cryptosporidium Burden in Cattle in Federal Capital Territory, Nigeria. VL 4. DOI: 10.31559VMPH.4.1.1. *Veterinary Medicine and Public Health Journal*.

Alamgir, A. N. M. (2017). Therapeutic use of medicinal plants and their extracts:

Tome 1. <https://doi.org/10.1007/978-3-319-63862-1>

Akhalkatsi, M., Arabuli, G., & Loren, R. (2014). Orchids as indicator species of forest disturbances on a limestone quarry in Georgia (South Caucasus) VL – 46. *Journal Europaischer Orchideen*.

Atherton, I., Bosanquet, S., & Lawley, M. (2010). Mosses and liverworts of Britain and Ireland - a field guide. British Organic Society. <https://www.britishbryologicalsociety.org.uk>

Bandyopadhyay, A. and Dey, A. (2022). Ethno-medicinal and pharmacological Bryophyte Attributes: A Review. *Phytomedicine Plus* Volume 2, Issue 2.

Bohan, D.A. and Vanbrngen, A.J. (2021). The future of agricultural landscapes, Part 11 Volume 64, pages 2-368.

Bradai, L., Neffar, S., Amrani, k., Bissati, S. et Chenchouni, H. (2015). Ethnomycological investigation of the traditional use and indigenous knowledge of desert truffles among the indigenous peoples of the Sahara Desert in Algeria. *J. Ethnopharmacol.* 13 March; Rev. 162:31-8.

Chandler, A., Meredith, C.R., McMullin, T. et Allen, J. (2020). "Wila - *Bryoria fremontii*" IUCN Red List of Threatened Species. e.T1757095A175710662.

Crawford, S.D. (2015). Lichens used in traditional medicine. DOI: 10.1007/978 – 3 – 319 – 13374 – 4 _ 2.

Davis, R., & Sommer, M.J.A. (2014). Western Mushroom Field Guide North America. Berkley: University of California Press. P. 207.

DeFilipps, R.A., Maina, S.L., & Crepin, J. (2024). Ken Tropical Plant Database Fern. Tropical. Ferns .info. 08 – 15 (viewtropical.php ?id=*Pitrogramma calomelanos*)

Derzhavina, N.M. (2020). Ecological morphology of the proto-epiphytic fern *Lemmaphyllum microphyllum* C. presl and its relationship to adaptogenesis. Contemporary Problems of Ecology Number 3.

DOI : <https://doi.org/10.1134/S19954255200>

Dinerstein, E., Olson, D., Joshi, A., Noss, R., Hassen, M., Locke, H., Ellis, E.C., Jones, B., Barber, C.V., Hayes, R., Kormos, C. Martin, V., Crist, E., Sechrest, W., & Saleem, M. (2017). An ecoregion-based approach to protect half of the terrestrial domain. *Bioscience*, 67(6), 534-545. <https://doi.org/1093/biosci/bix014>

Ebika, S.T.N., Morgan, D., Sanz, C. et Harris, D.J. (2015). *Ficus* h mi- piphyte (Moraceae) in a Congolese forest. *Plant Ecology and Evolution* 148(3):377-386, <http://dx.doi.org/10.5091/plecevo.1024>

Ellis, C.J. (2016). The climates of oceanic and temperate tropical forests and their

indicators in Great Britain. *Ecological indicators* 70: 125 – 133.

Ellis, C.J., Eaton, S., Theodoropoulos, M. et Elliot, K. (2015). *Épiphyte Indicator communities and species: an ecological guide for Scotland's forests*. Royal Botanic Garden, Edinburgh.

Evbuomwan, I.O., Stephen, A.O. and Oluba, O.M. (2023). *Médecine Autochtone Plants used in traditional medicine for the treatment of malaria in Kwara State, Nigeria: an ethnobotanical study*. *BMC Supplement Med Ther.* 23, 324. DOI: <https://doi.org/10.1186/s/2906> - 023-04131-4

Ezejiolor, F.U.C., Ndana, R.W., Ogunlade-Anibasa, G.O. and Madara, A.A. (2024). *Diversity and Taxonomic Classification of Epiphytes in the Federal Capital Territory, Nigeria*. *Annual Research and Review in Biology*. Volume 39, Issue 10, Page 52-65.

Federal Capital Territory Administration - Facts (2008).

Gao, Y., Skutsch, M., Masera, O., & Pacheco, P. (2011). *A global analysis of deforestation due to the development of biofuels*. Working Paper 68. CIFOR, Bogor, Indonesia. www.cifor.org

Getaneh, Z. A. and Gamo, F.W. (2016). "Vascular epiphytes in Doshke and

Kurpaye: A Comparative Study, Gamo Gofa, Ethiopia". *International Journal of Biodiversity*. <https://doi.org/115/9482057>

Govaerts, R., Luther, H.E., & Grant, J. (2013). World list of Bromeliads.

Kew: Royal Botanic Gardens. Available at: <http://apps.kew.org/wcsp/>

Gray in GBIF Secretariat (2023), GBIF Backbone Taxonomy. Checklist dataset <https://doi.org/10.15468/39omei> accessible via [GBIF.org](https://www.gbif.org)

Hassanpour, S., Maheri-Sis, N., Eshratkhah, B. and Mehmandar, F.B. (2011). Plants and secondary metabolites (Tannins): A Review, *International Journal of Forest, Soil and Erosion*, 1 (1): 47 – 53.

Hengameh, P. and Rajkumar, H.G. (2017). Physiological and chemical analysis for Identification of some additional lichen. *Journal of Pharmacognosy and Phytochemistry*. 6(5): 2611–2621. Available online at www.phytojournal.com

Igbere and Ogbole (2018). Ethnobotanical survey of plants used in the treatment of

Typhoid and its complications in Esan North East Local Government Area, Uromi, Edo State. *Nig. J. Pharm.* 14 (2), 175 to 188.

Katta J, Rampilla V, Khasim SM. A Study on Phytochemical and Anticancer Activities of Epiphytic Orchid *Aerides odorata* Lour. *Euro. J. Med. Plants*.

[Internet]. 2019 Jul. 23 [cited 2024 Dec. 31];28(3):1-21. Available from:
<https://journalejmp.com/index.php/EJMP/article/view/780>

Keat, L.H., Phaik, H.Y., Chee, W.W., Umah, R.K., Chek T.N., Festo, M. et Zhi, X. N. (2022). A journal on phytochemistry, pharmacology and toxicology. *Journal of Integrative Medicine. Volume 20, Number 4, Pages 292 - 304*

Kumar, A., Kumar, S., Komal, Ramchiary, N. et Singh, P. (2021). Rôle de Traditional ethnobotanical knowledge and indigenous communities in achieving the Sustainable Development Goals. *A Durability 13, 3062.*
<https://doi.org/10.3390/su13063062>

Luczaj, T. and Pieroni, A. (2016). Nutritional ethnobotany in Europe: from from emergency foods to healthy folk kitchens and contemporary foraging trends. DOI: 10.1007/978-1-4939-3329-7-3

Mahomoodally, M.F. (2013). "Traditional Medicines in Africa: An Evaluation of Ten powerful, complementary, evidence-based African medicinal plants and Alternative Medicine, vol. 2013, 14 pages.
<https://doi.org/10.1155/2013/617459>

Mathias, S.N., Ilyas, I., & Musa, K.Y. (2006). Ethnobotanical survey and

preliminary phytochemical studies of plants used in the practice of traditional medicine within the Takkad-speaking community of southern Kaduna-Nigeria. Chem. Class J. 4: 70 to 75.

Miao, J., Regenstein, J.M., Qui, J., Zhang, J., Zhang, X., Li, H., Zang, H. et Wang, Z. (2020). Isolation, structural characterization and bioactivities of polysaccharides and their ear derivatives – A review: *International Journal of Biological Molecules*. Volume 150, pages 102 to 113.

Migiro, G. (2019). What is Epiphyte? World Atlas. The original Ressource OnlineGeography.

Moravcova, L., Pysek, P., Jarosik, V., & Pergl, J. (2015). Choosing the right one Characteristics: Reproductive and dispersal characteristics predict the invasiveness of herbaceous plant species.

PLoS ONE 10(4):e0123634.doi:10.1371/journal.

Morquecho-Contreras, A., Carmen, Z. and Sanchez-Sanchez, H. (2018). Plant Defense against herbivores various environments. Pure and applied biogeography. DOI: 10.577/intechopen-70418.

N'Douba Amako, P., Kouassi Kouadio, C.K., Koffi N'Dono Boni, C., Douira,

A. and Ayolie, K. (2022). "Coprophilic fungi of the city of Daloa: New species for the fungal flora of Côte d'Ivoire" GSC Biological and Pharmaceutical Sciences. 20 (3): 251 – 260. doi:10.30574/gscbps.220.3.0362.

Muszynska, B., Fijalkowska, A., Sulkowska-Ziaja, K. Wlodarczyk, A., Kaczmarczyk, P., Nogaj, E., & Pietka, J. (2020). *Formitopsis officinalis*: a species of autochthonous fungus with promising biological and medicinal properties. Chem Biodivers 17 (6): e2000213.doi:101002/cbdv.00213.

Okhale, S.A., Ugbabe, G. A., Bamidele, O., Ajoku, G.A. et Egrevba, Yes.O. (2014). Phytochemical and antimicrobial studies on extracts of *Calypochilum emerginatum* (SW) Schltr (Orchidaceae) growing in Nigeria. *J. Med. Plants Res.* Vol. 8 (4), pp. 223-228. DOI: 10.5897/JMPR2013.5212.

Oladeji, O.S., Adelowo, F.E., Oluyori, A.P. et Bankole, D.T. (2020). Ethnobotanical description and biological activities of *Senna alata*. Evid based on the Alternat Med. Doi supplement: 10.1155/2580259

Oluwole, O., Funmilayo, E.A. and Abimbola, O. (2021). The *Senna* genre (Fabaceae): A journal on its traditional uses, botany, phytochemistry, pharmacology and toxicology. DOI: 10.1016/jsajb.2020.11.017.

Basel plant (2023). Published online. 12(9) : 1877 Doi : 10 – 3390/plants
12091877

Petruzzelo, M. (2020). Epiphyte. Encyclopedia Britannica,

<https://www.britannica.com/plant/epiphyte>

Rabizadeh, F., Mirian, M.S., Dusti, R., Kiani-Anbuhi, R. et Iftekhari, A. (2022).

Phytochemical classification of medicinal plants used in the treatment of kidney diseases based on traditional Persian medicine. Evid Based Complement Alternat Med. doi : 10.1155/8022599.

Sabovljevic, M., Sabovljevic, A, Ikram, N, Peramuna, A., Bae, H. et Simonsen, H. (2016). Bryophytes – an emerging source of herbal remedies and chemical production. Plant genetic resources, 14 (4), 314-327.

Santamaria, L. and Mendez, P.F. (2012). Biodiversity Policy Developments – Current gaps and future needs. *Scalable Applications* 5(2): 202 – 218. doi:10.1111/j.1752-4571.2011.00229.

Shah, North Carolina (2014). Lichens of commercial importance in India. *Scitech J.* 01:32-6

Shah, M.D., Gnanaraj, C., Haque, A.E., & Iqbal, M. (2014). Antioxidant and

chemopreventive effects of *Nephrolepis bisserata* against carbon tetrachloride (CCL4) – induced oxidative stress and Biology, 53(1), 31–39. <https://doi.org/10.3109/13880209.909502>.

Shanavskhan, A.E., Sivadasan, M., Alfarhan, A.H. E.T. Thomas, J. (2012). Ethnomedicinal aspects of angiospermic epiphytes and parasites from Kerala, India. *Indian Journal of Traditional Knowledge. Vol. II (20)*, pp. 250-258.

Subhashini, K., Kumar, P.K.R. E. T. Gaddeiah, G. (2019). Une reviews about *Dendrophthoe falcate* (L.F.) Ettingsh. (Loranthaceae). *Tropical Plant Research* 6 (3): 514–520.

Shrestha, G., Thompson, A., Robison, R., St Clair L.L. (2016). *Letharia vulpine*, un Lichen containing vulpinic acid targets cell limb and cell division processes in methicillin-resistant *Staphylococcus aureus*. 54(3) :413-8.doi : 10.3109/13880209.2015.1038754.Epub.

Singh, A.G., Kumar, A., & Tawari, D. (2012). An ethnobotanical survey on medicinal plants used in the Terai Forest in western Nepal. *Journal of Ethnobiology and Ethnomedicine* 8(1):19

Sivakrishnan, S. and Kavitha, J. (2017). Traditional Uses of *Ageratum conyzoides*

and its Bioactivities – A Brief Review. *Journal of Emerging Technologies and Innovative Research (JETIR)*. www.jetir.org p. 229. Volume 4, Number 7.

Spribille, T., Tuovinen, V., Resl, P., Vanderpool, D., Wolinsk, H., Aime, M.C., Schneider, K., Stabentheiner, E., Toome-Heller, M., Thor, G., Mayrhofer, H.J., & McCutcheon, J.P. (2016). Basidiomycete yeast in the cortex of ascomycete macrolichens. *Science*: 29; 353 (6298): 488 – 492. Doi : 10.1126/science.aaf8287

Stanton, D.E., Chavez, J.H., Villegas, L., Villansante, F., Armesto, J., Hedin, L.O. and Horn, H. (2014). "Epiphytes improve water use by modifying the microenvironment." *Functional ecology*. 28(5): 1274 to 1283.doi: 10.1111/1365 to 2435.12249

Stanton, D.E., Chavez, J.H., Villegas, L., Hedin, L.O. et Horn, H. (2014). Epiphytes improve the host plant's use of water by altering the microenvironment. <https://doi.org/10.1111/1365-2435.12249>

Tessler, M., Kam, M.T., Bliss-Moreau, M., & Wehr, J.O. (2014). Diversity and Distribution of stream bryophytes: is pH important? *Freshwater Sciences*, Volume 33, Number 3. <https://www.journals.uchicago>

The International Index of Plant Names and the World List of Vascular Plants (2024).

Published on the Internet at

<http://www.ipni.org> et <https://powo.science.kew.org/>

Ugwah-Oguejiofor, C.J., Ibrahim, S.G., Mshelia, U., Mohammed, U. et Adebisi, I.M. (2021). Department of Pharmacology and Toxicology, Faculty of Pharmaceutical Sciences, Usmanu Danfodiyo University, Sokoto. Department of Pharmacology and Ethnopharmacy, Faculty of Pharmaceutical Sciences, Usmanu Danfodiyo University, Sokoto. *Nigerian Journal of Basic and Applied Sciences* 29 (2): 67-75.

Vira, B., Wildburger, C., & Mansourian, S. (2015). Forests and food: Fighting hunger and nutrition in sustainable landscapes. <http://books.openedition.org/obp/2729>

Wagner, K., Mendieta-Leiva, G., & Zotz, G. (2015). Host specificity in the vascular Epiphytes: A review of methodology, empirical evidence, and potential mechanisms. *AoB PANTS*, Volume 7, plu092, <https://doi.org/10.1093/aobpla/plu092>

Westwood, J.H. (2020). "Parasitic plant". *Encyclopedia Britannica*, <https://britannia.com/plant/parasitic-plant>

Wester, S. and Zotz, G. (2010). Growth and survival of *Tillandsia flexuosa* on

electrical cables in Panama. *Journal of Tropical Ecology* 26(01):123-126.

DOI: 10.1017/S0266477400000459

William, C. (2022). Useful plant resources of tropical Africa.

<https://www.prota4u.org/database>

Woods, C., Cardelus, C., & Dewalt, S. (2014). Microhabitat associations of vascular epiphytes in a tropical rainforest canopy. *Journal of Ecology*. 103 (2)

DOI: 10.1111/1365-2745.12357.

Yadav, R.N.S. and Agarwala (2011). Phytochemical analysis of some

plants. *Journal of Plant Science* 3 (12): 10 to 14. <http://journal-phytology.com/>

Yang, Y.Y. and Kim, J.G. (2016). The optimal balance between sexual and asexual reproduction in a variable environment: a systematic review. *J Ecology and the environment* 40, 12. <https://doi.org/10.1186/s41610-016-0013-0>