

# Cancer Diagnostic and Therapeutic Considerations in Dogs: A Review

---

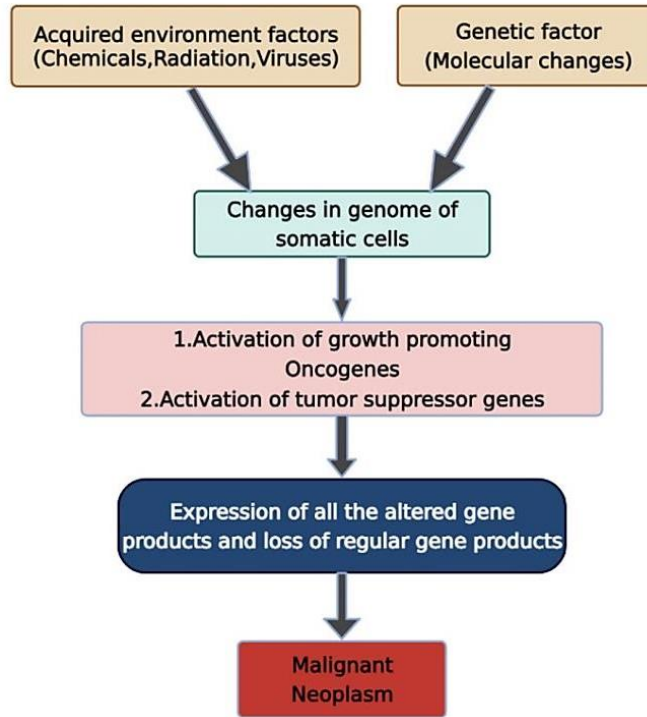
## ABSTRACT

Cancer presents a significant health concern in dogs, akin to its impact on humans, and stands as a primary contributor to canine mortality. The elevated incidence of cancer in dogs, relative to other domesticated animals, may be attributed to their intimate companionship with humans, advanced medical diagnostics, and elevated healthcare standards that have extended their lifespans beyond typical expectations, albeit accompanied by a heightened susceptibility to cancer. Effective diagnosis and treatment of canine cancer hinge upon a comprehensive understanding of tumor varieties, their underlying pathophysiology, and molecular alterations affecting genes and proteins. A range of diagnostic tools is employed for detecting cancer in dogs, encompassing blood profiles, cytological evaluations, histopathology, immunohistochemistry, and various imaging methods such as computed tomography, magnetic resonance imaging, positron emission tomography, and single-photon emission computed tomography. Additionally, molecular techniques play a crucial role. Therapeutic options for addressing canine cancer encompass chemotherapy, surgical procedures, and radiation therapy. Furthermore, promising developments including cancer vaccines and novel anti-cancer medications are currently undergoing clinical trials, aiming to enhance the management of cancer in dogs. This article provides an overview of both current and emerging diagnostic and treatment approaches, whether currently available or in the experimental stage, for effectively managing cancer in dogs.

*Keywords: Dog cancers, Cancer biomarkers, Cancer vaccines, Chemotherapy, Nanoparticles, Surgery*

## 1. INTRODUCTION

The term "tumor" denotes the uncontrolled growth and proliferation of cells, often resulting from the loss of contact inhibition phenomena (Lisanti *et al.*, 2010). The alteration in DNA due to several factors leads to the development of cancers (Fig. 1). In the canine world, cancer is a pressing health issue, affecting approximately four million dogs annually (Gardner *et al.*, 2016). Tumors are classified into two primary categories: benign and malignant (Sarver *et al.*, 2022). Benign tumors remain localized at their site of origin and refrain from invading surrounding tissues, blood vessels, or lymph nodes. In contrast, malignant tumors display invasive behavior, spreading to nearby tissues, blood vessels, or lymph nodes. Malignant tumors are commonly referred to as "cancer" (Chow, 2010). The incidence of cancer in dogs is influenced by several predisposing factors, including breed, gender, age, and geographical location. Older dogs, particularly those aged ten or older, exhibit a higher susceptibility to cancer (Gardner *et al.*, 2016). Gender also plays a role, with female dogs experiencing a higher incidence due to the prevalence of mammary cancer (Merlo *et al.*, 2008).



**Fig. 1. Cancer development. Overview of factors and mechanisms inducing development of neoplasm**

Specific dog breeds are at a higher risk of developing particular types of cancers (Table 1) (Raghavan *et al.*, 2005; Dobson, 2013; Erich *et al.*, 2013; Seim-Wikse *et al.*, 2013; Edmunds *et al.*, 2021; Labadie *et al.*, 2022). Common tumor types include meningiomas, gliomas, hemangiosarcoma, pituitary tumors, lymphoma, metastatic carcinoma, and histiocytic sarcoma (Fig. 2) (McGrath, 1962; Song *et al.*, 2013). The accurate and early diagnosis of cancers is very important to choose appropriate therapeutic measures. At present, a wide range of diagnostic and therapeutic considerations are available for the management of cancers in dogs. This review article summarized different diagnostic and therapeutic considerations in dog cancers including both traditional as well as more advanced ones.

**Table 1. Dog breeds with a higher incidence of certain cancers**

Dog breed	Susceptibility for cancer
Boxer	Mast-cell cancer and Gliomas
Rottweilers and Greyhounds	Osteosarcoma
Golden Retrievers	Lymphoma and Osteosarcoma
Scottish Terriers	Transitional-cell carcinoma of the bladder
Flat-Coat Retrievers, Bernese	Histiocytic sarcomas

---

Mountain Dogs

Chow Chows

Gastric carcinoma and Melanoma.

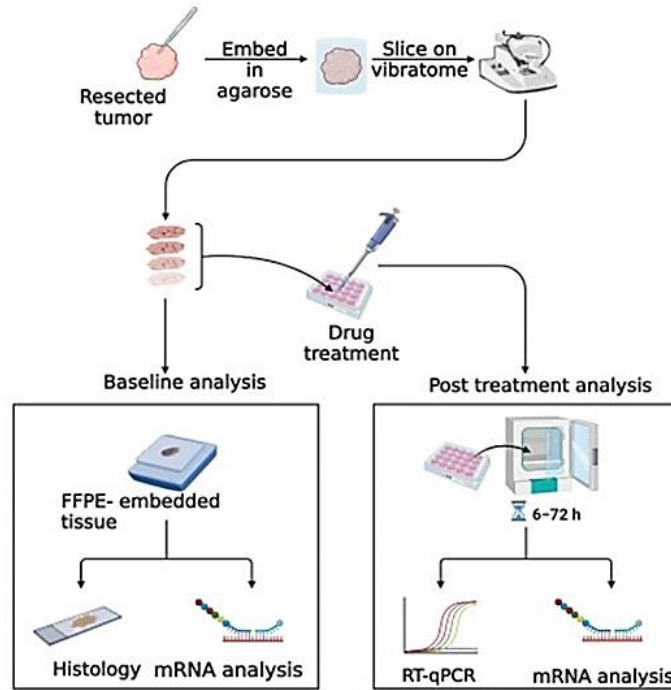
---



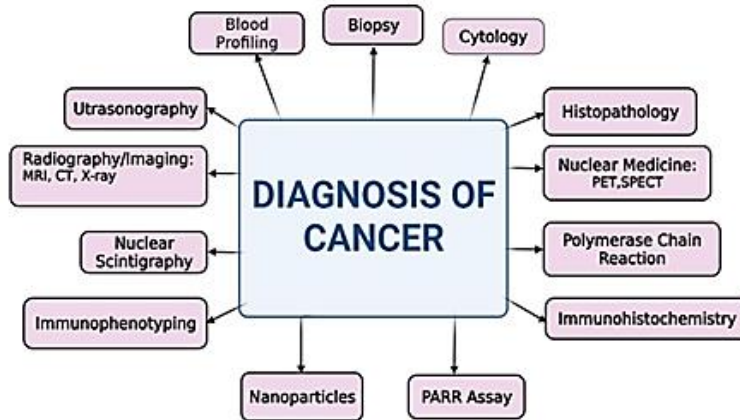
**Fig. 2. Common cancers in dogs. The highly prevalent common cancers in dogs irrespective of their breeds.**

## **2. DIAGNOSTIC JOURNEY: FROM BLOOD PROFILING TO MOLECULAR TOOLS**

Accurate and timely cancer diagnosis plays a pivotal role in enabling clinicians to select appropriate treatment modalities (such as surgery, chemotherapy, radiation therapy, etc.), thereby influencing the cancer prognosis. The process of diagnosing a suspected cancer case commences with a comprehensive patient history review and a thorough physical examination of the canine, encompassing meticulous scrutiny for any external growths on the dog's body surface, alongside hematological and biochemical profiling—a straightforward and economically viable testing procedure. Nonetheless, these diagnostic assessments are rudimentary and cannot provide definitive and early confirmatory diagnosis. Certain cancers manifest subtly, necessitating more advanced diagnostic techniques such as cytology, histopathology, detection of cancer biomarkers, immunohistochemistry, molecular tools (such as PARR assay for diagnosis of lymphoma), flowcytometry (for detection of the type of cancer), etc. (Fig. 3 & Fig. 4). These diagnostic modalities are discussed below in succeeding sections.



**Fig. 3. Overview of tumor analysis. Common methodologies applied across different labs for analysis of tumors**



**Fig. 4. Diagnosis of cancer. Different sets of techniques available for diagnosis of cancers in different settings**

### 2.1 Blood Profiling

Blood profiling includes both hematological (blood cells and bone marrow) and biochemical examination. Blood profiling is crucial to diagnose blood malignancies, particularly leukemia. Before approaching another diagnosis, the initial complete blood count of a dog can provide valuable details about the overall health of the patient and potential recommendations for

further investigation (Wilson-Robles *et al.*, 2020). In this test, blood cancers are diagnosed by assessing the number of blood cells or abnormal cells whether increasing or decreasing in blood. Later, a bone marrow biopsy can help in confirming the diagnosis of blood cancer.

The VDI TKcanine+ test measures thymidine kinase (TK1) and canine C-reactive protein (cCRP) levels, shedding light on both cancer and inflammation. The Nu.Q Vet Cancer test detects nucleosomes, a marker associated with various cancers, including lymphoma and hemangiosarcoma (McAnena *et al.*, 2017).

## **2.2 Cytological Examination**

Cytology involves the microscopic examination of cells obtained through fine needle aspiration (FNA) or smear impression. It offers fundamental information about tumor type and assists in classifying tumors as mesenchymal or epithelial in origin (Ayele *et al.*, 2017). Cytological examinations have numerous advantages, including minimal sample requirements, no need for sophisticated equipment, and the ability to perform tests without sedation or anesthesia. Veterinarians often use cytology for on-the-spot assessments to determine the necessary steps (Chibuk *et al.*, 2021). The section covers cytological examination applications, sensitivity, and specificity for various cancers.

## **2.3 Biopsy**

Biopsies involve the microscopic examination of tissue samples obtained through various techniques for cancer diagnosis (Howlett *et al.*, 2016). A biopsy sample should ideally contain both normal and tumor tissue to evaluate microscopic invasiveness accurately. The fast and cost-effective fine needle aspiration biopsy technique, which does not require anesthesia, is highlighted. The importance of histological evaluation in mammary tumor diagnosis is discussed, emphasizing its gold standard status. Liquid biopsy, a non-invasive technique, detects circulating tumor DNA (ctDNA) in the blood. Its applications in diagnosing urinary tract cancers and c-kit gene mutations in mast cell tumors are examined (Schillaci *et al.*, 2019). The concept of ctDNA, its significance, and its potential use for early cancer detection are explored. Liquid biopsy's role in challenging cases where tissue biopsies are impractical, such as lung and brain cancers, is emphasized (Flory *et al.*, 2022).

## **2.4 Histopathology**

Histopathology involves the examination of excised tissue samples under a microscope after various processing steps. This method provides detailed insights into tissue architecture and is particularly valuable for distinguishing between benign and malignant tumors (Withrow *et al.*, 2013). Histological examination is considered the gold standard for cancer diagnosis and provides a definitive diagnosis compared to cytological methods. Pathologists identify tumor cell histogenesis based on microscopic features, such as nuclear shape, cytoplasmic characteristics, and the presence or absence of stromal components (Prasad *et al.*, 2021). Histochemical stains, including methyl green, toluidine blue, and periodic acid-Schiff, are commonly employed to determine the cell of origin in cancer development. While histopathology offers unparalleled accuracy, it requires substantial time for tissue fixation and preparation, particularly in the case of bone tumors necessitating decalcification. In contrast, cytopathology through fine needle puncture allows for quicker diagnoses.

## **2.5 Immunohistochemistry (IHC)**

IHC is a valuable adjunct to histopathology, aiding in the confirmation of histological diagnoses and determining the origin of poorly differentiated tumors. IHC involves the use of labeled monoclonal or polyclonal antibodies to identify specific antigens on cell surfaces. These antibodies bind to target antigens, forming antibody-antigen complexes that can be visualized, often using enzyme-based reactions or fluorophores (Berg and Fishman, 2020). In cancer diagnosis, IHC helps pathologists verify histological diagnoses made from standard stained sections. It also plays a crucial role in identifying the cell of origin for poorly differentiated tumors. Specific biomarkers, as listed in Table 2 (Altmannsberger *et al.*, 1985; Ramos-Vara *et al.*, 2007; Rao, 2010; Nieves *et al.*, 2014; Paździór-Czapula *et al.*, 2015; Usman *et al.*, 2021), assist in characterizing various cancers.

**Table 2. Tumour markers that are commonly identified using IHC**

Marker (antigen)	Significance and usage
Vimentin	Identifies all mesenchymal cells. Often used to help confirm diagnosis of a tumour of mesenchymal origin.
Cytokeratin	Identifies most epithelial cells. Often used to help confirm diagnosis of a tumour of epithelial origin
CD3	A marker of T-cell origin. Together with CD79a, used to help distinguish between T- and B-cell lymphomas.
CD79a	A marker of B-cell origin. Used in a similar manner to CD3. Most plasma cells do not express CD79a, making it of less use in diagnosing plasmacytomas
CD18	A marker of leucocytes. Very useful in helping to diagnose tumours of monocyte/macrophage origin.
Desmin	A marker of muscle origin. Is expressed in most tumours of smooth muscle or striated muscle origin.

## 2.6 Radiographs/Imaging

Radiographic and imaging techniques have evolved significantly, offering diverse modalities for cancer detection. Conventional methods, such as X-ray, ultrasonography (USG), CT-scan, and magnetic resonance imaging (MRI), provide information on cancer location, anatomical changes, and tissue appearance alterations (Edmunds *et al.*, 2021). Functional imaging techniques, including dynamic CT, functional MRI, PET, SPECT, contrast-enhanced ultrasound, and more, detect cancers at earlier stages by assessing pathophysiological changes within tumors. These changes encompass altered blood flow, increased glucose metabolism, and changes in cellularity (Marcus *et al.*, 2009; Bai *et al.*, 2023). Functional imaging also supports monitoring the therapeutic response in cancer patients.

## 2.7 Immunophenotyping

Immunophenotyping relies on flow cytometry (FC) to identify and categorize cell populations based on specific protein expression. FC rapidly and accurately assesses the presence or absence of cell-surface markers (Song *et al.*, 2012). While immunophenotyping is valuable in diagnosing various diseases, including blood malignancies like lymphoma and leukemia, it is often used in conjunction with other diagnostic tools (Ghisleni *et al.*, 2006). FC enhances the accuracy of diagnosis by evaluating cell lineage, antigen levels, clonality, and minimal residual disease. It is also instrumental in monitoring treatment responses and aiding in the development of new therapeutic strategies (Ghisleni *et al.*, 2006; Kruse *et al.*, 2014).

## 2.8 Molecular diagnosis

In response to limitations associated with conventional diagnostic techniques like imaging, histology, and biochemical evaluation, molecular and immunophenotyping methods have gained prominence for early-stage cancer diagnosis. These methods are facilitated by user-friendly molecular analysis tools, enabling applications in cancer diagnosis, staging, immunophenotyping, and the detection of minimal residual disease (MRD) genes (Regan, 2017). Notable molecular tools in cancer diagnosis include PCR-based detection of c-kit gene mutations, PARR assay for lymphoma detection (Burnett *et al.*, 2003), real-time PCR for identifying low BRCA2 gene expression in canine mammary tumors (Yoshikawa *et al.*, 2015), and FISH for detecting BCR-ABL chromosomal abnormalities in chronic myeloid leukemia (Culver *et al.*, 2013). These tools have significantly contributed to accurate early cancer diagnosis and represent the evolution of cancer diagnostic techniques (Mochizuki *et al.*, 2015; Settawongsin *et al.*, 2016).

## 3. THERAPEUTICS FOR DOG CANCERS

Effective therapeutic strategies for treating dog cancers encompass chemotherapy, surgery, radiation therapy, immunotherapy, electro-chemotherapy, gene therapy, and nanoparticles (Fig. 5) (Rassnick *et al.*, 2000). Each of these therapeutic approaches is discussed below.



**Fig. 5. Methods of cancer therapy. Various modalities available to the clinicians/researchers for therapeutic interventions against different cancers**

### 3.1 Chemotherapy

Recent decades have witnessed significant progress in canine cancer chemotherapy, targeting rapidly dividing cells. While these drugs effectively kill cancer cells, they also harm rapidly dividing normal cells, leading to side effects such as nausea, vomiting, and anemia (Kamble *et al.*, 2021). Noteworthy drugs include Toceranib phosphate (Palladia®, Zoetis) for

mast cell tumors, Vinorelbine for various malignancies, and histamine-receptor blockers like diphenhydramine. Combination therapy, such as cyclophosphamide, vincristine, and prednisolone, is used for multicentric lymphoma (Addissie and Klingemann, 2021). Chemotherapy is administered through three methods. First, conventional chemotherapy follows maximum-tolerated doses, leading to serious complications. Second, metronomic chemotherapy, administered continuously at low doses, offers unique benefits including anti-angiogenic properties and fewer side effects. Third, targeted chemotherapy, such as tyrosine kinase inhibitors like Toceranib phosphate and Masitinib mesylate, focuses on specific cellular pathways (Simeonov and Stoikov, 2006). Tyrosine kinase inhibitors target abnormal cell proliferation and have shown promise in treating canine cancers, with drugs like Ibrutinib (Imbruvica®) approved for various hematopoietic malignancies (Robak *et al.*, 2022). Verdinexor (KPT-335) inhibits nuclear export protein Exportin 1 in canine osteosarcoma (Breitbach *et al.*, 2021), while STA-1474 has exhibited biological activity in dogs with various tumors (Weissleder *et al.*, 1989).

### **3.2 Surgery**

Surgical interventions aim to remove tumors with surrounding healthy tissue. Factors like tumor type, location, and invasiveness dictate surgical approaches. These include complete tumor removal, debulking (if total removal poses risks), and alleviating pain due to tumor pressure. Surgical interventions may complement chemotherapy for better prognosis (Nordlinger *et al.*, 2009). Achieving curative-wide surgical margins (>5cm) is crucial to prevent tumor recurrence (Hao *et al.*, 2018).

### **3.3 Radiation Therapy**

Radiation therapy is an increasingly accessible treatment option, using ionizing radiations like X-rays, gamma rays, electron beams, or protons. These radiations harm cancer cells or alter their genetic makeup, leading to cell death. It is used as a standalone treatment for certain tumors like skin cancers, prostate carcinomas, and lymphomas, but often works best when combined with surgery or chemotherapy. Radiation therapy has shown success in treating mast cell tumors, lymphomas, and thymoma (Meleo, 1997; Ladue *et al.*, 1998; Mayer, 2006). In some cases, radiation therapy can reduce tumor size and improve neurological symptoms, as seen in pituitary tumors (Dow *et al.*, 1990; Bley *et al.*, 2005; Mayer, 2006). However, the prognosis depends on the tumor's severity, histopathologic grade, clinical stage, location, and the patient's overall health. Side effects may manifest after treatment completion (Bateman *et al.*, 1994).

### **3.4 Immunotherapy for Canine Cancer**

Immunotherapies designed to harness the immune system's potential to combat tumors and employ tumor-associated antigens as vaccines have emerged as a promising approach in canine oncology. In particular, CD4+ and CD8+ T cells play pivotal roles in orchestrating anticancer immune responses (Hammerstorm *et al.*, 2011).

One notable example of an anticancer vaccine is sipuleucel-T, FDA-approved for prostate cancer treatment in humans. It consists of autologous T-lymphocytes incubated with a fusion protein linking a common prostate cancer antigen (prostatic acid phosphatase) to an adjuvant (granulocyte-macrophage colony-stimulating factor) (Hammerstorm *et al.*, 2011). In dogs, the USDA-approved Oncept vaccine, a DNA vaccine, is used specifically to treat stage II or III oral melanoma (Wouda *et al.*, 2015). Further enhancing T-lymphocyte efficacy, researchers are exploring chimeric antigen receptors (CARs) that specifically recognize

tumor-associated surface antigens. Although the FDA has approved six CAR T-cell treatments for human blood malignancies, such as lymphomas and leukemias, the application of CAR-T cells in canine cancer treatment is still in its early stages (Lucroy *et al.*, 2020).

Natural Killer (NK) cells, which are gaining attention as cellular therapeutics for canine cancer, hold potential as well (Comiskey *et al.*, 2018). While only one clinical trial using NK cells in dogs has been reported to date, it yielded promising results. In this trial, NK cells were isolated from blood lymphocytes, and dogs with locally advanced, non-metastatic osteosarcoma received radiation followed by intra-tumor NK cell injections. Some dogs also received human IL-2. Tumor shrinkage was observed, with no pulmonary metastasis in 5 out of 10 patients at the 6-month endpoint (Stoica *et al.*, 2009). The combination of pre-treatment radiation with NK cell therapy enhances its antitumor effects, inducing adhesion molecules on tumor cells that bind to NK cells. Additionally, intra-tumor injection of NK cells has demonstrated efficacy in inducing tumor regression and lasting responses (Suzuki *et al.*, 2018).

Researchers are exploring various avenues, including the development of a *Listeria* vector vaccine for osteosarcoma in both dogs and humans. This vaccine uses an attenuated strain of *Listeria monocytogenes* carrying tumor-associated antigens. Attenuated *Listeria* strains are preferred for their ability to trigger potent CTL (cytotoxic T lymphocyte) responses (Oladejo *et al.*, 2021). This vaccine is currently undergoing clinical trials and has shown promise in treating malignancies.

### 3.5 Electro-Chemotherapy (ECT)

Electrochemotherapy (ECT) is an innovative treatment approach that aims to enhance the uptake of chemotherapy drugs within tumor cells by applying electric pulses. These electric pulses significantly increase drug toxicity and efficacy, potentially by up to 1000 times (Probst *et al.*, 2018). ECT has demonstrated remarkable effectiveness against various tumors, particularly those located in or just under the skin, including melanoma, squamous cell carcinoma, soft tissue sarcomas, and localized cutaneous lymphoma in dogs (Yeom *et al.*, 2021).

ECT was first introduced in veterinary medicine in Europe in 1997 (Tellado *et al.*, 2022). A study comparing ECT to surgery for mast cell tumors in dogs found that ECT achieved complete responses in 70% of patients, while surgery achieved complete responses in only 50% of cases, highlighting ECT's efficacy as a treatment comparable to surgery (Leibman *et al.*, 2001).

ECT is particularly effective in cases of drug-resistant tumors, as it enhances the permeability of tumor cell membranes to chemotherapy drugs (Tellado *et al.*, 2022). Commonly used drugs in electrochemotherapy include bleomycin and cisplatin (De Virgilio *et al.*, 2018). Unlike physical methods like surgery, high-frequency focused ultrasound, hyperthermia, or radiotherapy, which cannot differentiate between malignant and healthy cells, ECT offers a precise and selective approach to killing tumor cells when combined with electro-permeabilization and appropriate drugs (Vižintin *et al.*, 2021).

ECT has been successfully applied in the treatment of various primary cancers and metastases in both dogs and cats (Tozon *et al.*, 2016). It can be used as a curative treatment or as an adjunct to surgery for solitary, multiple, or subcutaneous tumor nodules. Recent developments in electrode designs and technological breakthroughs show great

promise in treating deep-seated cancers through electro-chemotherapy (Tozon *et al.*, 2016). While ECT achieves up to 80% local tumor control, its impact on distant metastases is limited (Kotnik *et al.*, 2019). Thus, combining ECT with other therapies may be necessary to achieve a systemic response.

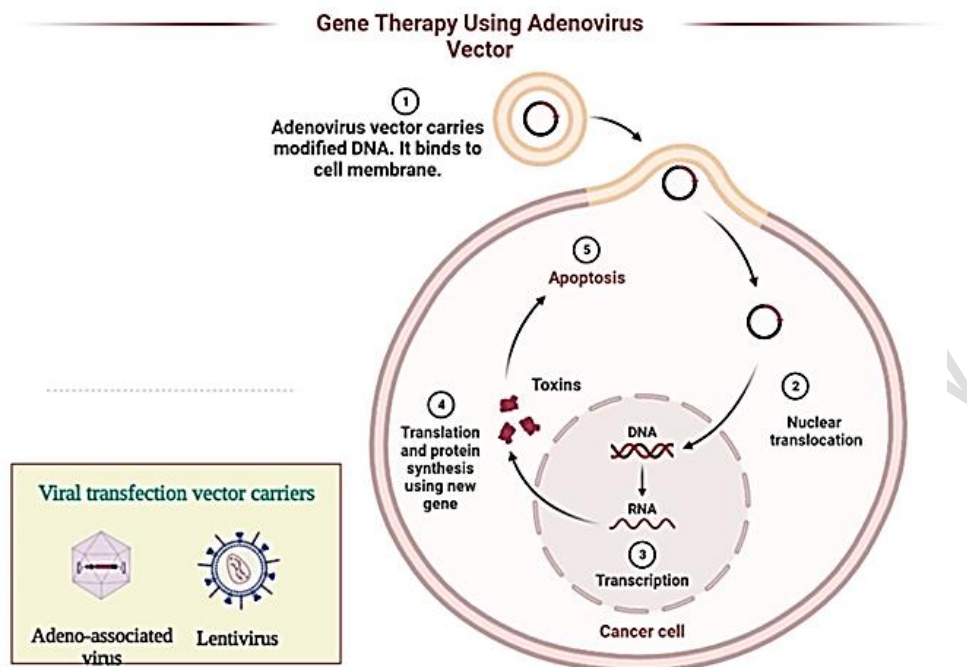
### 3.6 Gene Therapy in Canine Cancer

Gene therapy has emerged as a promising approach in treating cancer, targeting both inherited and somatic mutations in proto-oncogenes and tumor suppressor genes (Matlashewski *et al.*, 1984; Wiman, 1993; Vogelstein *et al.*, 2000; Shanker *et al.*, 2011). The core idea behind gene therapy is to replace defective genes with functional copies, thereby restoring normal cellular function. Compared to other treatment modalities, gene therapy offers a more specific approach (Das *et al.*, 2015).

The foundation for gene therapy was laid in the 1960s when it was observed that viruses could integrate their genetic material into the genomes of infected cells, potentially leading to malignant transformation. Edward Tatum outlined the therapeutic potential of using viruses for genetic modification of somatic cells in 1966 (Das *et al.*, 2015). Various approaches are under investigation for gene therapy in cancer treatment, including: a) Expressing genes to induce apoptosis or increase tumor sensitivity to conventional therapies. b) Introducing wild-type tumor suppressor genes to compensate for their loss or deregulation. c) Using antisense (RNA/DNA) techniques to prevent oncogene expression. d) Enhancing tumor immunogenicity to promote immune cell activity (Das *et al.*, 2015).

Viruses, particularly morbilliviruses and adenoviruses, have been widely studied as vectors for gene therapy due to their restricted host range and the availability of species-specific subtypes (Arendt *et al.*, 2009). The mechanism of action of gene therapy against cancer cells using adenovirus is illustrated in Fig. 6. However, it's important to consider the toxicity associated with viral vectors in gene therapy (Arendt *et al.*, 2009).

Clinical trials have shown promise, with nine lung cancer patients directly injected with a retroviral vector containing the p53 gene exhibiting tumor regression in some cases (Roth *et al.*, 1996). Similarly, intratumoral injection of an adenovirus vector encoding wild-type p53 complementary DNA (Ad-p53) led to tumor regression in preclinical investigations in animal models (Swisher *et al.*, 1999). The use of interleukin-12 (IL-12) gene therapy has demonstrated potent anticancer effects in various tumors in animal models (Cross and Burmester, 2006; Pavlin *et al.*, 2012). While the results of ongoing clinical trials for gene therapy in cancer treatment are promising, further validation is required before it can be widely used in the treatment of cancer in both dogs and humans.



**Fig. 6. Gene therapy for treatment of cancer using adenovirus vector. Different viruses are being tested for viral-vector based gene therapy modalities. These vectors may carry therapeutic transgene to target and eliminate cancer cells**

### 3.7 Nanoparticles in Cancer Therapy

Nanoparticles have shown significant potential in cancer treatment, primarily as vehicles for the targeted delivery of chemotherapy drugs to tumor sites. Key properties of nanoparticles, including size, charge, and surface chemistry, impact their uptake, intracellular fate, and cytotoxicity (Abbasi *et al.*, 2023).

Nanotechnology's role in cancer treatment is still in its early stages of research and development. Promising clinical trials have been conducted with hyaluronan cisplatin nanoparticles, nanocrystals of cisplatin, and paclitaxel for the effective treatment of various canine cancers, such as oral melanoma, oral sarcoma, and anal gland adenocarcinoma (Zabielska-Koczywas and Lechowski, 2017).

One noteworthy example is liposome-encapsulated muramyl tripeptide-phosphatidylethanolamine (L-MTP-PE), a synthetic analog of muramyl tripeptide found in mycobacterium cell walls (Nardin *et al.*, 2006). L-MTP-PE enhances the anticancer activity of monocytes and macrophages by inducing the release of inflammatory cytokines such as TNF- $\alpha$  and IL-6 (Asano and Kleinerman, 1993). Clinical trials in dogs with hemangiosarcoma have shown significantly longer survival periods for those receiving L-MTP-PE compared to dogs receiving chemotherapy and placebo liposomes (Barnes *et al.*, 2022). Mifamurtide, a liposomal muramyl tripeptide phosphatidyl ethanolamine, is licensed in Europe for the treatment of cancers in humans (Barnes *et al.*, 2022).

Other liposome-based nanoparticles have been employed for delivering various chemotherapy drugs, including curcumin, doxorubicin, cisplatin, and untargeted tumor RNA (Ambrosio *et al.*, 2022). Additionally, doxorubicin conjugated to glutathione-stabilized gold nanoparticles demonstrated higher cytotoxicity against canine osteosarcoma cell lines compared to the free drug (Malek *et al.*, 2021).

Although several studies have explored nanoparticle-based cancer therapies in dogs, none of these nano-drugs are currently FDA-approved for canine cancer treatment. In contrast, a few nano-drugs have gained approval for cancer treatment in humans in the USA and Europe (Dang and Guan, 2003).

#### **4. CONCLUSION**

Dogs face a heightened susceptibility to cancer compared to other domesticated animals, mirroring the cancer incidence seen in humans. This presents a significant challenge for veterinary oncologists striving to effectively manage these malignancies in canines. Traditional diagnostic tools, such as X-rays, ultrasonography, cytology, and histopathology, prove effective in identifying larger tumors or those with evident structural and morphological changes. However, they fall short in detecting early-stage, cell-originating, or inconspicuous cancers. To overcome these limitations, advanced diagnostic techniques, including functional imaging methods, molecular tools, and immunophenotyping, play a crucial role.

In the realm of canine cancer treatment, three primary avenues—chemotherapy, surgery, and radiation therapy—are at the disposal of veterinary professionals. The most remarkable outcomes are achieved when these modalities are combined rather than used in isolation. Nonetheless, these therapies lack the ability to distinguish between healthy and cancerous cells. In contrast, cutting-edge therapeutic approaches like electrochemotherapy (ECT), gene therapy, cancer vaccines, and nanoparticle-based treatments show promise in precisely targeting and eliminating cancer cells while sparing healthy ones. However, the majority of these advanced therapeutic techniques are still in various stages of clinical trials, holding the potential to revolutionize canine cancer management in the future.

## REFERENCES

1. Abbasi, R., Shineh, G., Mobaraki, M., Doughty, S., & Tayebi, L. (2023). Structural parameters of nanoparticles affecting their toxicity for biomedical applications: a review. *Journal of Nanoparticle Research*, 25(3), 43.
2. Addissie, S., & Klingemann, H. (2018). Cellular immunotherapy of canine cancer. *Veterinary sciences*, 5(4), 100.
3. Altmannsberger, M., Weber, K., Droste, R., & Osborn, M. (1985). Desmin is a specific marker for rhabdomyosarcomas of human and rat origin. *The American journal of pathology*, 118(1), 85.
4. Ambrosio, N., Voci, S., Gagliardi, A., Palma, E., Fresta, M., & Cosco, D. (2022). Application of biocompatible drug delivery nanosystems for the treatment of naturally occurring cancer in dogs. *Journal of Functional Biomaterials*, 13(3), 116.
5. Arendt, M., Nasir, L., & Morgan, I. M. (2009). Oncolytic gene therapy for canine cancers: teaching old dog viruses new tricks. *Veterinary and comparative oncology*, 7(3), 153-161.
6. Asano, T., & Kleinerman, E. S. (1993). Liposome-encapsulated MTP-PE: a novel biologic agent for cancer therapy. *Journal of immunotherapy*, 14(4), 286-292.
7. Ayele, L., Mohammed, C., & Yimer, L. (2017). Review on diagnostic cytology: Techniques and applications in veterinary medicine. *Journal of Veterinary Science and Technology*, 8(408), 2.
8. Bacci, G., Picci, P., Ferrari, S., Orlandi, M., Ruggieri, P., Casadei, R., et al. (1993). Prognostic significance of serum alkaline phosphatase measurements in patients with osteosarcoma treated with adjuvant or neoadjuvant chemotherapy. *Cancer*, 71(4), 1224-1230.
9. Bai, J. W., Qiu, S. Q., & Zhang, G. J. (2023). Molecular and functional imaging in cancer-targeted therapy: current applications and future directions. *Signal Transduction and Targeted Therapy*, 8(1), 89.
10. Barnes, D.J., Dutton, P., Bruland, Ø., Gelderblom, H., Faletti, A., Bühnemann, C., van Maldegem, A., Johnson, H., Poulton, L., Love, S. and Tiemeier, G., 2022. Outcomes from a mechanistic biomarker multi-arm and randomised study of liposomal MTP-PE (Mifamurtide) in metastatic and/or recurrent osteosarcoma (EuroSarc-Memos trial). *BMC cancer*, 22(1), 629.
11. Bateman, K. E., Catton, P. A., Pennock, P. W., & Kruth, S. A. (1994). 0–7–21 radiation therapy for the palliation of advanced cancer in dogs. *Journal of Veterinary Internal Medicine*, 8(6), 394-399.
12. Berg, E. A., & Fishman, J. B. (2020). Labeling antibodies using colloidal gold. *Cold Spring Harbor Protocols*, 2020(4), pdb-prot099333.
13. Bley, C. R., Sumova, A., Roos, M., & Kaser-Hotz, B. (2005). Irradiation of brain tumors in dogs with neurologic disease. *Journal of Veterinary Internal Medicine*, 19(6), 849-854.
14. Breitbach, J. T., Louke, D. S., Tobin, S. J., Watts, M. R., Davies, A. E., & Fenger, J. M. (2021). The selective inhibitor of nuclear export (SINE) verdinexor exhibits biologic activity against canine osteosarcoma cell lines. *Veterinary and Comparative Oncology*, 19(2), 362-373.

15. Burnett, R. C., Vernau, W., Modiano, J. F., Olver, C. S., Moore, P. F., & Avery, A. C. (2003). Diagnosis of canine lymphoid neoplasia using clonal rearrangements of antigen receptor genes. *Veterinary Pathology*, 40(1), 32-41.
16. Chibuk, J., Flory, A., Kruglyak, K. M., Leibman, N., Nahama, A., Dharajiya, N., et al. (2021). Horizons in veterinary precision oncology: fundamentals of cancer genomics and applications of liquid biopsy for the detection, characterization, and management of cancer in dogs. *Frontiers in Veterinary Science*, 8, 664718.
17. Chow, A. Y. (2010). Cell cycle control by oncogenes and tumor suppressors: driving the transformation of normal cells into cancerous cells. *Nature Education*, 3(9), 7.
18. Cohen, M., Bohling, M. W., Wright, J. C., Welles, E. A., & Spano, J. S. (2003). Evaluation of sensitivity and specificity of cytologic examination: 269 cases (1999–2000). *Journal of the American Veterinary Medical Association*, 222(7), 964-967.
19. Comiskey, M. C., Dallos, M. C., & Drake, C. G. (2018). Immunotherapy in prostate cancer: teaching an old dog new tricks. *Current oncology reports*, 20, 1-10.
20. Cross, D., & Burmester, J. K. (2006). Gene therapy for cancer treatment: past, present and future. *Clinical Medicine & Research*, 4(3), 218-227.
21. Culver, S., Ito, D., Borst, L., Bell, J. S., Modiano, J. F., & Breen, M. (2013). Molecular characterization of canine BCR-ABL–positive chronic myelomonocytic leukemia before and after chemotherapy. *Veterinary Clinical Pathology*, 42(3), 314-322.
22. Dang, Y., & Guan, J. (2020). Nanoparticle-based drug delivery systems for cancer therapy. *Smart Materials in Medicine*, 1, 10-19.
23. Das, S. K., Menezes, M. E., Bhatia, S., Wang, X. Y., Emdad, L., Sarkar, D., & Fisher, P. B. (2015). Gene therapies for cancer: strategies, challenges and successes. *Journal of Cellular Physiology*, 230(2), 259-271.
24. Davis, B. W., & Ostrander, E. A. (2014). Domestic dogs and cancer research: a breed-based genomics approach. *ILAR journal*, 55(1), 59-68.
25. De Virgilio, A., Ralli, M., Longo, L., Mancini, P., Attanasio, G., Atturo, F., et al. (2018). Electrochemotherapy in head and neck cancer: A review of an emerging cancer treatment. *Oncology Letters*, 16(3), 3415-3423.
26. Dobson, J. M. (2013). Breed-predispositions to cancer in pedigree dogs. *International Scholarly Research Notices*, 2013(1), 941275.
27. Dow, S. W., LeCouteur, R. A., Rosychuk, R. A. W., Powers, B. E., Kemppainen, R. J., & Gillette, E. L. (1990). Response of dogs with functional pituitary macroadenomas and macrocarcinomas to radiation. *Journal of Small Animal Practice*, 31(6), 287-294.
28. Edmunds, G. L., Smalley, M. J., Beck, S., Errington, R. J., Gould, S., Winter, H., et al. (2021). Dog breeds and body conformations with predisposition to osteosarcoma in the UK: a case-control study. *Canine medicine and genetics*, 8, 1-22.
29. Erich, S. A., Rutteman, G. R., & Teske, E. (2013). Causes of death and the impact of histiocytic sarcoma on the life expectancy of the Dutch population of Bernese mountain dogs and Flat-coated retrievers. *The Veterinary Journal*, 198(3), 678-683.
30. Flory, A., Kruglyak, K. M., Tynan, J. A., McLennan, L. M., Rafalko, J. M., Fiaux, P. C., et al. (2022). Clinical validation of a next-generation sequencing-based multi-cancer early detection “liquid biopsy” blood test in over 1,000 dogs using an independent testing set: The CANcer Detection in Dogs (CANDiD) study. *PLoS One*, 17(4), e0266623.

31. Gardner, H. L., Fenger, J. M., & London, C. A. (2016). Dogs as a model for cancer. *Annual Review of Animal Biosciences*, 4(1), 199-222.
32. Ghisleni, G., Roccabianca, P., Ceruti, R., Stefanello, D., Bertazzolo, W., Bonfanti, U., & Caniatti, M. (2006). Correlation between fine-needle aspiration cytology and histopathology in the evaluation of cutaneous and subcutaneous masses from dogs and cats. *Veterinary Clinical Pathology*, 35(1), 24-30.
33. Grüntzig, K., Graf, R., Hässig, M., Welle, M., Meier, D., Lott, G., et al. (2015). The Swiss Canine Cancer Registry: a retrospective study on the occurrence of tumours in dogs in Switzerland from 1955 to 2008. *Journal of comparative pathology*, 152(2-3), 161-171.
34. Hao, Y., Yang, C., & He, J. (2018). The accurate surgical margin before surgery for malignant musculoskeletal tumors: a retrospective study. *American Journal of Translational Research*, 10(8), 2324.
35. Howlett, D. C., & Triantafyllou, A. (2016). Evaluation: Fine Needle Aspiration Cytology, Ultrasound-Guided Core Biopsy and Open Biopsy Techniques. *Advances in Oto-rhinolaryngology*, 78, 39-45.
36. Kamble, M., Devangan, R., Sharda, R., Tiwari, S. K., Kalim, M. O., Gumasta, P., ... & Yadav, D. (2021). Successful surgical management of mammary tumor in a dog: Case report.
37. Kotnik, T., Rems, L., Tarek, M., & Miklavčič, D. (2019). Membrane electroporation and electroporabilization: mechanisms and models. *Annual review of biophysics*, 48(1), 63-91.
38. Kruse, M. A., Holmes, E. S., Balko, J. A., Fernandez, S., Brown, D. C., & Goldschmidt, M. H. (2013). Evaluation of clinical and histopathologic prognostic factors for survival in canine osteosarcoma of the extracranial flat and irregular bones. *Veterinary pathology*, 50(4), 704-708.
39. Labadie, J., Swafford, B., DePena, M., Tietje, K., Page, R., & Patterson-Kane, J. (2022). Cohort profile: The golden retriever lifetime study (GRLS). *Plos one*, 17(6), e0269425.
40. Ladue, T., Price, G. S., Dodge, R., Page, R. L., & Thrall, D. E. (1998). Radiation therapy for incompletely resected canine mast cell tumors. *Veterinary Radiology & Ultrasound*, 39(1), 57-62.
41. Leibman, N. F., Kuntz, C. A., Steyn, P. F., Fettman, M. J., Powers, B. E., Withrow, S. J., & Dernell, W. S. (2001). Accuracy of radiography, nuclear scintigraphy, and histopathology for determining the proximal extent of distal radius osteosarcoma in dogs. *Veterinary Surgery*, 30(3), 240-245.
42. Lisanti, M. P., Martinez-Outschoorn, U. E., Chiavarina, B., Pavlides, S., Whitaker-Menezes, D., Tsigos, A., ... & Sotgia, F. (2010). Understanding the "lethal" drivers of tumor-stroma co-evolution: emerging role (s) for hypoxia, oxidative stress and autophagy/mitophagy in the tumor microenvironment. *Cancer biology & therapy*, 10(6), 537-542.
43. Lucroy, M. D., Clauson, R. M., Suckow, M. A., El-Tayyeb, F., & Kalinauskas, A. (2020). Evaluation of an autologous cancer vaccine for the treatment of metastatic canine hemangiosarcoma: a preliminary study. *BMC Veterinary Research*, 16, 1-12.
44. Marcus, C. D., Ladam-Marcus, V., Cucu, C., Bouché, O., Lucas, L., & Hoeffel, C. (2009). Imaging techniques to evaluate the response to treatment in oncology: current standards and perspectives. *Critical reviews in oncology/hematology*, 72(3), 217-238.

45. Matlashewski, G., Lamb, P., Pim, D., Peacock, J., Crawford, L., & Benchimol, S. (1984). Isolation and characterization of a human p53 cDNA clone: expression of the human p53 gene. *The EMBO journal*, 3(13), 3257-3262.
46. Mayer, M. N. (2006). Radiation therapy for canine mast cell tumors. *The Canadian Veterinary Journal*, 47(3), 263.
47. McAnena, P., Brown, J. A., & Kerin, M. J. (2017). Circulating nucleosomes and nucleosome modifications as biomarkers in cancer. *Cancers*, 9(1), 5.
48. McGrath, J. T. (1962). Intracranial pathology of the dog. In *Symposium Über Vergleichende Neuropathologie: Abgehalten von der Arbeitsgemeinschaft für Vergleichende Neuropathologie Während des IV. Internationalen Kongresses für Neuropathologie Vom 4.–8. September 1961 in München* (pp. 3-4). Springer Berlin Heidelberg.
49. Meleo, K. A. (1997). The role of radiotherapy in the treatment of lymphoma and thymoma. *Veterinary Clinics: Small Animal Practice*, 27(1), 115-129.
50. Merlo, D. F., Rossi, L., Pellegrino, C., Ceppi, M., Cardellino, U., Capurro, C., et al. (2008). Cancer incidence in pet dogs: findings of the Animal Tumor Registry of Genoa, Italy. *Journal of Veterinary Internal Medicine*, 22(4), 976-984.
51. Mochizuki, H., Shapiro, S. G., & Breen, M. (2015). Detection of BRAF mutation in urine DNA as a molecular diagnostic for canine urothelial and prostatic carcinoma. *PloS one*, 10(12), e0144170.
52. Moore, A. S., London, C. A., Wood, C. A., Williams, L. E., Cotter, S. M., L'Heureux, D. A., & Frimberger, A. E. (1999). Lomustine (CCNU) for the treatment of resistant lymphoma in dogs. *Journal of Veterinary Internal Medicine*, 13(5), 395-398.
53. Nardin, A., Lefebvre, M. L., Labroquere, K., Faure, O., & Abastado, J. P. (2006). Liposomal muramyl tripeptide phosphatidylethanolamine: Targeting and activating macrophages for adjuvant treatment of osteosarcoma. *Current cancer drug targets*, 6(2), 123-133.
54. Nieves, S., Apellaniz, D., Tapia, G., Maglia, A., Mosqueda-Taylor, A., & Bologna-Molina, R. (2014). Cytokeratins 14 and 19 in odontogenic cysts and tumors: a review. *Odontoestomatología*, 16(24), 45-55.
55. Nordlinger, B., Van Cutsem, E., Gruenberger, T., Glimelius, B., Poston, G., Rougier, P., et al. (2009). Combination of surgery and chemotherapy and the role of targeted agents in the treatment of patients with colorectal liver metastases: recommendations from an expert panel. *Annals of Oncology*, 20(6), 985-992.
56. Oladejo, M., Paterson, Y., & Wood, L. M. (2021). Clinical experience and recent advances in the development of listeria-based tumor immunotherapies. *Frontiers in Immunology*, 12, 642316.
57. Pavlin, D., Cemazar, M., Sersa, G., & Tozon, N. (2012). IL-12 based gene therapy in veterinary medicine. *Journal of Translational Medicine*, 10, 1-11.
58. Paździor-Czapula, K., Rotkiewicz, T., Otrocka-Domagala, I., Gesek, M., & Śmiech, A. (2015). Morphology and immunophenotype of canine cutaneous histiocytic tumours with particular emphasis on diagnostic application. *Veterinary Research Communications*, 39, 7-17.
59. Prasad, M., Ghosh, M., Patki, H. S., Kumar, S., Brar, B., Sindhu, N., et al. (2021). *Imaging Techniques in Veterinary Disease Diagnosis*. In *Advances in Animal Disease Diagnosis* (pp. 103-145). CRC Press.

60. Probst, U., Fuhrmann, I., Beyer, L., & Wiggermann, P. (2018). Electrochemotherapy as a new modality in interventional oncology: a review. *Technology in cancer research & treatment*, 17, 1533033818785329.
61. Raghavan, M., Knapp, D. W., Bonney, P. L., Dawson, M. H., & Glickman, L. T. (2005). Evaluation of the effect of dietary vegetable consumption on reducing risk of transitional cell carcinoma of the urinary bladder in Scottish Terriers. *Journal of the American Veterinary Medical Association*, 227(1), 94-100.
62. Ramos-Vara, J. A., Miller, M. A., & Valli, V. E. O. (2007). Immunohistochemical detection of multiple myeloma 1/interferon regulatory factor 4 (MUM1/IRF-4) in canine plasmacytoma: comparison with CD79a and CD20. *Veterinary Pathology*, 44(6), 875-884.
63. Rao, I. S. (2010). Role of immunohistochemistry in lymphoma. *Indian Journal of Medical and Paediatric Oncology*, 31(04), 145-147.
64. Rassnick, K. M., Mauldin, G. E., Al-Sarraf, R., Mauldin, G. N., Moore, A. S., & Mooney, S. C. (2002). MOPP chemotherapy for treatment of resistant lymphoma in dogs: a retrospective study of 117 cases (1989–2000). *Journal of veterinary internal medicine*, 16(5), 576-580.
65. Regan, D. (2017). Pharmacological Characterization of Losartan as a Ccr2 Antagonist and Pre-Clinical and Pharmacodynamic Assessment as a Potential Anti-Metastatic Therapy (Doctoral dissertation, Colorado State University).
66. Robak, T., Witkowska, M., & Smolewski, P. (2022). The role of Bruton's kinase inhibitors in chronic lymphocytic leukemia: current status and future directions. *Cancers*, 14(3), 771.
67. Roth, J. A., Nguyen, D., Lawrence, D. D., Kemp, B. L., Carrasco, C. H., Ferson, D. Z., et al. (1996). Retrovirus-mediated wild-type P53 gene transfer to tumors of patients with lung cancer. *Nature Medicine*, 2(9), 985-991.
68. Sarver, A. L., Makielski, K. M., DePauw, T. A., Schulte, A. J., & Modiano, J. F. (2022). Increased risk of cancer in dogs and humans: A consequence of recent extension of lifespan beyond evolutionarily determined limitations?. *Aging and Cancer*, 3(1), 3-19.
69. Schillaci, O., Scimeca, M., Toschi, N., Bonfiglio, R., Urbano, N., & Bonanno, E. (2019). Combining diagnostic imaging and pathology for improving diagnosis and prognosis of cancer. *Contrast Media & Molecular Imaging*, 2019(1), 9429761.
70. Seim-Wikse, T., Jörundsson, E., Nødtvedt, A., Grotmol, T., Bjornvad, C. R., Kristensen, A. T., & Skancke, E. (2013). Breed predisposition to canine gastric carcinoma—a study based on the Norwegian canine cancer register. *Acta Veterinaria Scandinavica*, 55, 1-6.
71. Shanker, M., Jin, J., Branch, C. D., Miyamoto, S., Grimm, E. A., Roth, J. A., & Ramesh, R. (2011). Tumor suppressor gene-based nanotherapy: from test tube to the clinic. *Journal of drug delivery*, 2011(1), 465845.
72. Simeonov, R., & Stoikov, D. (2006). Study on the correlation between the cytological and histological tests in the diagnostics of canine spontaneous mammary neoplasms. *Bulg J Vet Med*, 9(3), 211-219.
73. Singh, P., Pandit, S., Mokkaapati, V. R. S. S., Garg, A., Ravikumar, V., & Mijakovic, I. (2018). Gold nanoparticles in diagnostics and therapeutics for human cancer. *International journal of molecular sciences*, 19(7), 1979.
74. Song, J. Y., Filie, A. C., Venzon, D., Stetler-Stevenson, M., & Yuan, C. M. (2012). Flow cytometry increases the sensitivity of detection of leukemia and lymphoma cells in

- bronchoalveolar lavage specimens. *Cytometry Part B: Clinical Cytometry*, 82(5), 305-312.
75. Song, R. B., Vite, C. H., Bradley, C. W., & Cross, J. R. (2013). Postmortem evaluation of 435 cases of intracranial neoplasia in dogs and relationship of neoplasm with breed, age, and body weight. *Journal of veterinary internal medicine*, 27(5), 1143-1152.
  76. Stoica, G., Lungu, G., Martini-Stoica, H., Waghela, S., Levine, J., & Smith III, R. (2009). Identification of cancer stem cells in dog glioblastoma. *Veterinary pathology*, 46(3), 391-406.
  77. Suzuki, D. O., Berkenbrock, J. A., Frederico, M. J., Silva, F. R., & Rangel, M. M. (2018). Oral mucosa model for electrochemotherapy treatment of dog mouth cancer: ex vivo, in silico, and in vivo experiments. *Artificial Organs*, 42(3), 297-304.
  78. Swisher, S. G., Roth, J. A., Nemunaitis, J., Lawrence, D. D., Kemp, B. L., Carrasco, C. H., et al. (1999). Adenovirus-mediated p53 gene transfer in advanced non-small-cell lung cancer. *Journal of the National Cancer Institute*, 91(9), 763-771.
  79. Tellado, M., Mir, L. M., & Maglietti, F. (2022). Veterinary guidelines for electrochemotherapy of superficial tumors. *Frontiers in Veterinary Science*, 9, 868989.
  80. Tozon, N., Tratar, U. L., Znidar, K., Sersa, G., Teissie, J., & Cemazar, M. (2016). Operating procedures of the electrochemotherapy for treatment of tumor in dogs and cats. *Journal of visualized experiments: JoVE*, (116).
  81. Usman, S., Waseem, N. H., Nguyen, T. K. N., Mohsin, S., Jamal, A., Teh, M. T., & Waseem, A. (2021). Vimentin is at the heart of epithelial mesenchymal transition (EMT) mediated metastasis. *Cancers*, 13(19), 4985.
  82. Vižintin, A., Marković, S., Ščančar, J., & Miklavčič, D. (2021). Electroporation with nanosecond pulses and bleomycin or cisplatin results in efficient cell kill and low metal release from electrodes. *Bioelectrochemistry*, 140, 107798.
  83. Vogelstein, B., Lane, D., & Levine, A. J. (2000). Surfing the p53 network. *Nature*, 408(6810), 307-310.
  84. Weissleder, R., Elizondo, G., Stark, D. D., Hahn, P. F., Marfil, J., Gonzalez, J. F., et al. (1989). The diagnosis of splenic lymphoma by MR imaging: value of superparamagnetic iron oxide. *American Journal of Roentgenology*, 152(1), 175-180.
  85. Wilson-Robles, H., Miller, T., Jarvis, J., Terrell, J., Dewsbury, N., Kelly, T., et al. (2020). Evaluation of nucleosome concentrations in healthy dogs and dogs with cancer. *PLoS One*, 15(8), e0236228.
  86. Wiman, K. G. (1993). The retinoblastoma gene: role in cell cycle control and cell differentiation. *The FASEB journal*, 7(10), 841-845.
  87. Vail, D. M., Thamm, D. H., & Liptak, J. M. (2019). *Withrow and MacEwen's small animal clinical Oncology-E-Book*. Elsevier Health Sciences.
  88. Wouda, R. M., Miller, M. E., Chon, E., & Stein, T. J. (2015). Clinical effects of vinorelbine administration in the management of various malignant tumor types in dogs: 58 cases (1997–2012). *Journal of the American Veterinary Medical Association*, 246(11), 1230-1237.
  89. Yeom, S. C., Song, K. H., & Seo, K. W. (2021). The application of electrochemotherapy in three dogs with inoperable cancers. *Korean journal of veterinary research*, 61(1), 9-1.

90. Yoshikawa, Y., Morimatsu, M., Ochiai, K., Ishiguro-Oonuma, T., Wada, S., Orino, K., & Watanabe, K. (2015). Reduced canine BRCA2 expression levels in mammary gland tumors. *BMC Veterinary Research*, 11, 1-8.
91. Zabielska-Koczywaś, K., & Lechowski, R. (2017). The use of liposomes and nanoparticles as drug delivery systems to improve cancer treatment in dogs and cats. *Molecules*, 22(12), 2167.

UNDER PEER REVIEW