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3 **Flavonoids, breast cancer prevention, and its treatment: A growing evidence**
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9 **ABSTRACT**

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11 There is mounting evidence linking certain lifestyle factors such as food, weight, and
12 physical activity to an increased risk of breast cancer. Flavonoids are commonly used in
13 traditional medicine. Plenty of studies have investigated the relationship between
14 flavonoid consumption and breast cancer in humans. This review aimed to examine the
15 association between flavonoids, each flavonoid subclass and the risk of breast cancer
16 besides therapeutic use of flavonoids to break the multidrug resistance in breast cancer.
17 Prospective cohort, case-control, and laboratory-based studies published between
18 around 1990's to date and referred to the impact of flavonoids on breast cancer
19 prevention, treatment, or other roles were included. Odd Ratio (OR)/Risk Ratio (RR) and
20 Hazard ratio (HR) along 95% CI were carefully reviewed to reveal the association
21 between different subgroups of flavonoids and breast cancer risk. Other adjustments (e.g.
22 age, menopausal status, food habits, race, BMI, etc.) were also considered. The
23 antioxidant properties of flavonoids, as well as their ability to inhibit apoptosis, suppress
24 estrogen activity, and limit the proliferation of breast cancer cells, all contribute to a
25 significant reduction in the risk of developing breast cancer. There are linings enough that
26 some polyphenolic compounds are found as effective drugs for treating breast cancer.
27

28 **Keywords:** *Polyphenolic compounds; mammary carcinogenesis; antioxidants; multi-drug*
29 *resistance (MDR); phytoestrogen; CYP1A1 inhibitors; ATP-binding cassette.*
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34 INTRODUCTION

35 Breast cancer (or mammary carcinogenesis) is the most frequently detected cancer
36 in women worldwide. It has emerged as a growing global health concern in recent
37 decades (Ganesan et al., 2022). According to GLOBOCAN (2020), the prevalence
38 of new instances of breast cancer among women is expected to be 24.5% (Sung et
39 al., 2021). The number of women who were diagnosed with breast cancer worldwide
40 reached 2.3 million in the year 2020, leading to 685,000 confirmed deaths. With 7.8
41 million confirmed cases in the previous five years, breast cancer exceeded all other
42 cancers in terms of prevalence by the end of 2020 (WHO, 2023). Human mammary
43 carcinoma is the consequence of cumulative exposure of the mammary cells to
44 endogenous estrogens (Colditz, 1998). A wide variety of biological processes,
45 including cell proliferation, cancer progression, apoptosis, and others, are made
46 easier as a result of the interaction between estrogen and the estrogen receptor that
47 is present in breast cells. This phenomenon is the result of this interaction (Thomas
48 and Gustafsson, 2011).

49 Diet is the most pronounced modifiable factor for breast cancer occurrence,
50 recurrence, and mortality (Hamer and Warner, 2017). Several studies in the last few
51 decades have looked at the relationship between breast cancer and certain foods,
52 such as beans, endive, tomatoes, strawberries, grapes, meat, soy products, and a
53 variety of vegetables, fruits, and teas (Peterson and Dwyer, 1998). Jang H. *et al.* in
54 2018 reported an association between anti-inflammatory diets and cancer
55 recurrence as well as overall mortality in breast cancer patients. Experimental
56 evidence suggests that flavonoids can both inhibit and stimulate a wide array of
57 enzyme systems found in mammals. Enzymes like these have an impact on
58 processes including cell proliferation and division, detoxification, platelet
59 aggregation, inflammation, and immunological responses (Franke et al., 1998).
60 Flavonoids have gone under number of investigations for their anti-carcinogenic
61 mechanisms and antiproliferative effects on human lymphocytes as well as breast
62 cancer cells and have also been reported (Peterson et al., 2003). Laboratory and
63 animal studies suggest that dietary flavonoids may be protective against breast
64 cancer risk. On the other hand, there are limited epidemiological studies on this
65 concern. An increased consumption of fruits and vegetables rich in flavonoids has
66 been linked to a decreased risk of cancer, according to data from epidemiological
67 studies. Which bioactive chemicals, if any, are responsible for this correlation
68 remains unknown. This review aims at a clear idea about the 1990-to-date study
69 pattern, interest in flavonoid subclasses, and major findings in both epidemiological
70 and laboratory-based studies.

71

72 **What is breast cancer?**

73 Cancer is a cluster of abnormally grown/growing cells in a tissue. This anomaly is
74 caused by a genetic mutation or other environmental factors that let them
75 divide/grow out of control. These out-of-control cells form ball-shaped lumps of
76 mutated tissue which is termed as tumor(s). When these forms of tumors are from
77 breasts, they're classified as breast cancer. Most breast cancers begin in the milk

78 glands (lobules) or the tubes (ducts) by which the nipple is connected to the milk
 79 glands. According to WHO (2023), the female gender has come up as the strongest
 80 risk factor for the occurrence of breast cancer. But in males, 1% or less of breast
 81 cancer cases are occurred. The main variables that increase the likelihood of breast
 82 cancer, both in terms of incidence and mortality, include being older (40 or older),
 83 having a higher body mass index (BMI), smoking, not being physically active, eating
 84 a diet rich in fat, experiencing menstruation at a young age, having a first full-term
 85 pregnancy late, breastfeeding for shorter periods or not at all, using oral
 86 contraceptives, using menopausal hormone therapy, breast density, and genetic
 87 predisposition (Zhang et al., 2020). A woman's chance of getting breast cancer is
 88 increased by both behavioral and genetic factors. Signs and symptoms of breast
 89 cancer include; thickening or lump in the breast, sometimes without pain, change in
 90 overall appearance (size and shape) of the breast, creek, reddish appearance, pit,
 91 or other changes in the skin, change in the physiognomy of nipples or areola (the
 92 darkish area around the breast), and abnormal fluid (may be bloody) from the nipple
 93 (WHO, 2023).

94
 95 **Molecular subtypes of breast cancer**

96 Physiologically, the human female mammary gland is under the primary control of
 97 different hormones. It is established that among them, estrogen appears to play the
 98 central role. There are mainly four molecular subtypes of breast cancer,
 99 characterized by hormone receptors (HR) in addition to protein involvement (or not
 100 involvement) in each cancer: (I) Luminal A or HR+/HER2- (HR-positive/HER2-
 101 negative), (II) Luminal B or HR+/HER2+ (HR-positive/HER2-positive), (III) HER2
 102 enriched and (IV) Triple-negative (TNBC) or basal-like (HR/HER2-negative)
 103 (Fragomeni et al., 2018). Each of the subtypes of breast cancer is described in Table
 104 1, which includes their characteristics.

| Table 1: Distinctive characteristics of the various subtypes of breast cancer | | | | | |
|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------|------------------|----------------------------|---------------------|---------------------------|
| Attribute | Luminal A | Luminal B | HER2 | TNBC | References |
| Frequency (%) | 50 | 15 | 20 | 15 | Barzaman et al., 2020 |
| ER | Yes | Yes | Some cases | No | Gao and Swain, 2018 |
| PR | Yes | Some cases | Some cases | No | Barzaman et al., 2020 |
| HER | No | No | Yes | No | Engel and Kaklamani, 2007 |
| Mutations | No | BRCA2 | p53 | p53 and BRCA1 | Duffy et al., 2018 |
| Prognosis | Good | Middle | Middle/Bad | Bad | Ahrn et al., 2014 |
| Therapy | Hormonal | Hormonal / Chemo | Hormonal/C hemo/ Herceptin | Chemo/ Experimental | Loibl and Gianni, 2017 |
| ER: estrogen receptor, PR: progesterone receptor, HER: human epidermal receptor, HER2: human epidermal growth factor receptor 2, and TNBC: triple-negative breast cancer | | | | | |

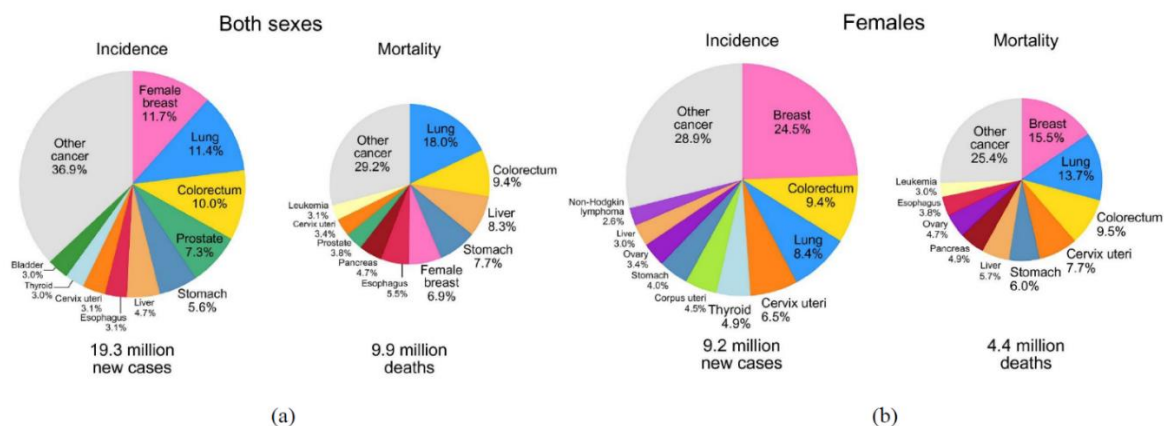
105 **Cancer Growth and Clinical Stages**

106 The female breast consists of 15 to 20 lobes of tissue, each of which contains lobules
 107 that house milk-producing glands and ducts. The nipple receives milk from the
 108 lobules via the ducts. Cancers can begin anywhere. However, the lymph nodes are
 109 often the sites of spreading after beginning in the ducts or lobules. Lymph nodes are
 110 like bus stations for cancer cells; they transport the disease throughout the body.
 111 There are many lymph nodes close to the breasts- around the chest, neck, and
 112 armpits. Cancer can spread regionally to these nodes from the breasts (Regionally
 113 spreading). Metastatic breast cancer spreads beyond those nodes. During diagnosis
 114 with breast cancer, cancer is categorized termed as "staged". Staging lets healthcare
 115 providers to make decision on treatment mechanisms and conditions after treatment.
 116 There are three distinct categories used to classify breast cancer stages: clinical
 117 prognostic, pathologic, and anatomic.

118 **Breast cancer scenario worldwide**

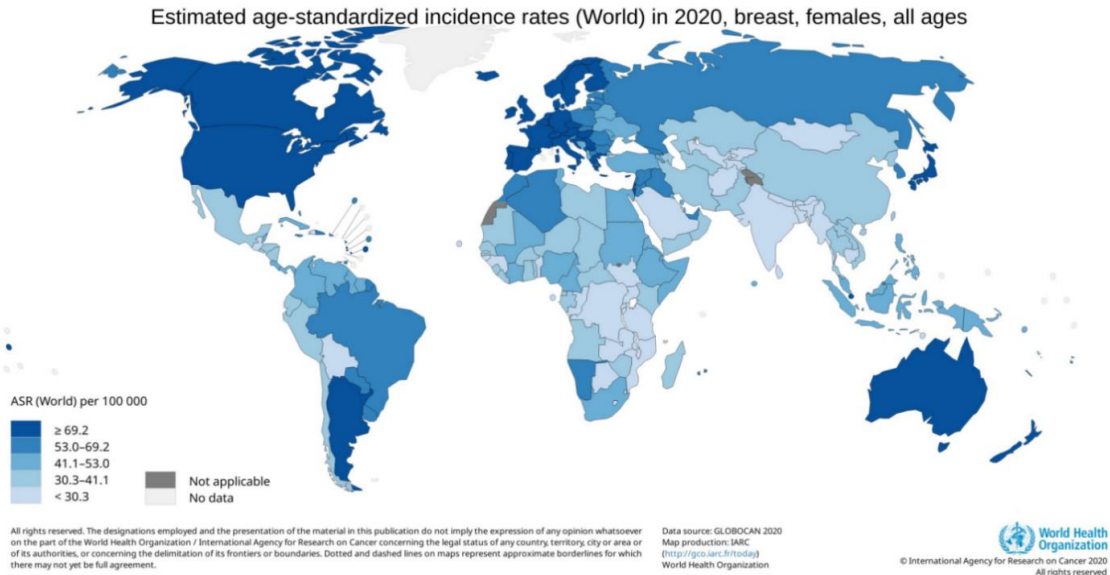
119 Taking aside lung cancer from the title of “most commonly diagnosed cancer
 120 globally”, breast cancer has emerged as the most prevalent cancer type. The year
 121 2020 had an estimated 2.3 million cases and 685,000 fatalities due to breast cancer
 122 and among eight cancer diagnoses one cancer case was diagnosed as breast
 123 cancer case (Sung et al., 2021). Over 2.3 million new instances of breast cancer are
 124 added annually. As more and more instances have been reported, breast cancer has
 125 outpaced all others among adults. More than 95% of countries have determined that
 126 breast cancer is either the first or second major cause of mortality among females
 127 due to cancer (WHO, 2023). The majority of countries throughout the world have
 128 breast cancer as their primary cause of mortality and morbidity in the year 2020.
 129 Breast cancer was responsible for more than a quarter of all cancer diagnoses in
 130 women and approximately fifteen percent of all deaths that were caused by cancer
 131 (Sung et al., 2021). Low- and middle-income nations account for almost 80% of
 132 breast cancer and cervical cancer deaths because of their larger populations (WHO,
 133 2023). By 2070, the anticipated number of breast cancer cases will have risen to 4.4
 134 million (Soerjomataram and Bray, 2021).

135

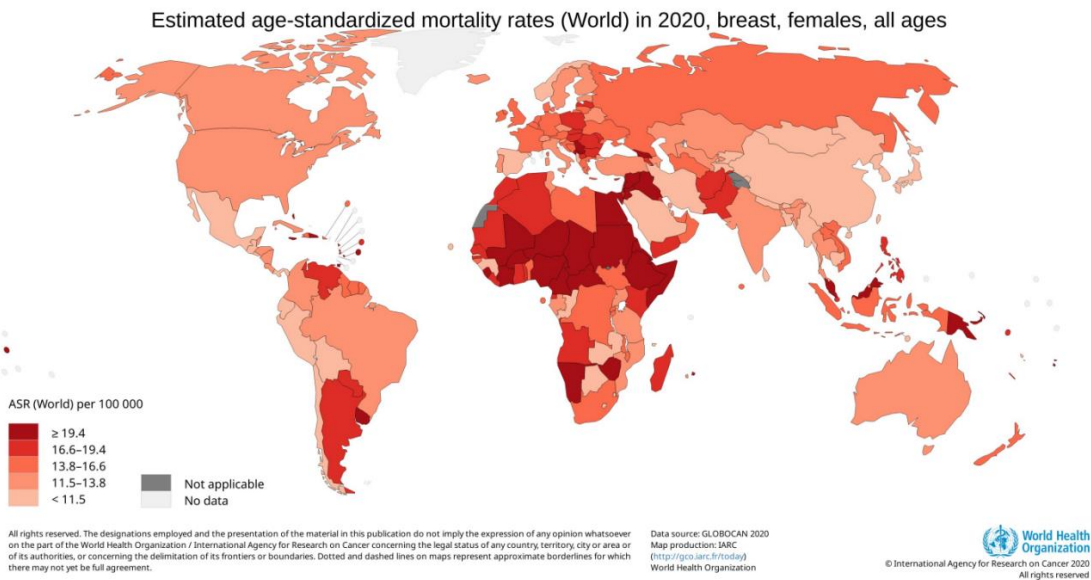


136

137 **Figure 1:** Death and case distributions for the 10 most frequent malignancies in 2020; (a) Both sexes and (b) Females. The percentage of cases or deaths is shown
 138 by the area of the pie chart. The "other" group includes nonmelanoma skin cancers,
 139 which do not include basal cell carcinoma for incidence. Data source: GLOBOCAN
 140 2020. Pie chart: Sung et al., 2021.
 141
 142



143 **Figure 2:** Estimated age-standard female breast cancer incidence rates in 2020.
 144 Data source: GLOBOCAN 2020, Map production: IARC, World Health Organization.
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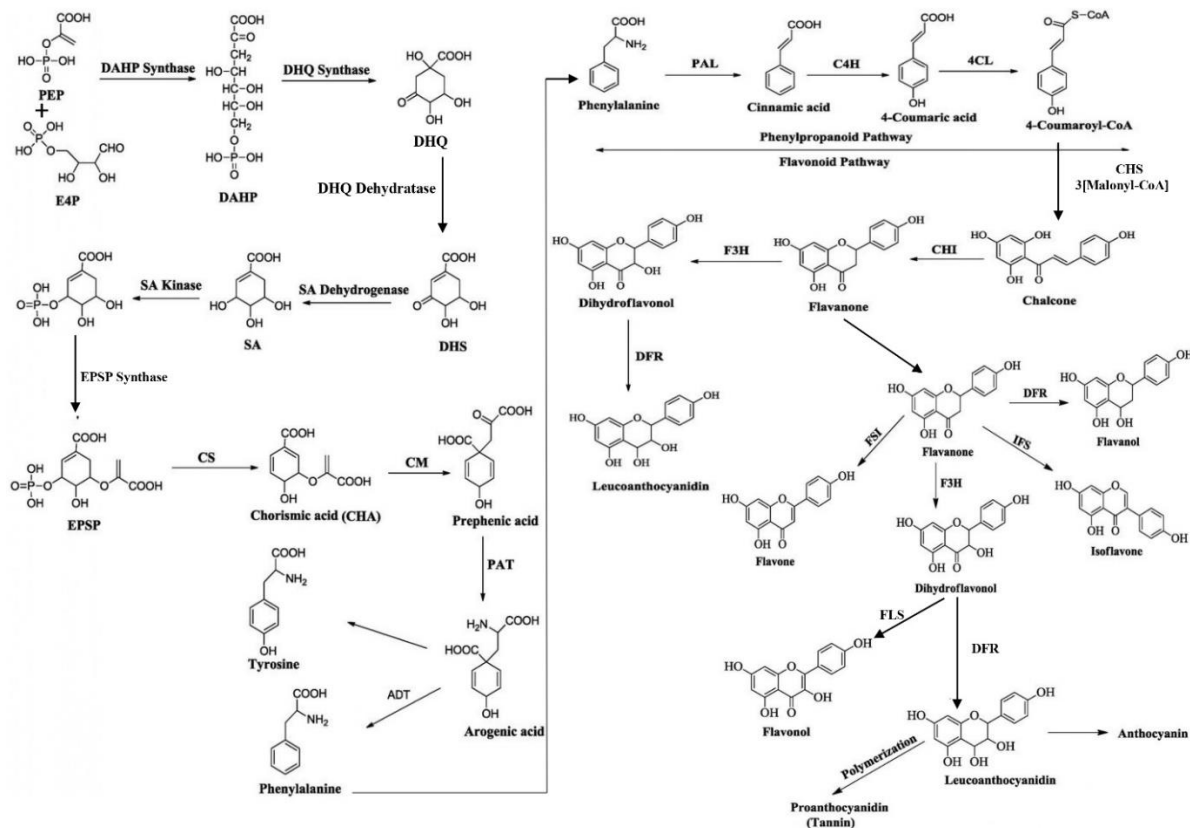
148 **Figure 3:** Estimated age-standard female breast cancer mortality rates in 2020. Data
 149 source: GLOBOCAN 2020, Map production: IARC, World Health Organization.
 150

151 **Flavonoids, Classification, and Distribution**

152 Flavonoids, whose name derives from the Latin word "flavus" signifying yellow, are
153 ubiquitous secondary metabolites found in plants. They are most recognizable as
154 the anthocyanin pigments, which are red, blue, and purple in color, that cover plant
155 tissues (Winkel-Shirley 2000). Flavonoids generally consist of a 15-carbon skeleton
156 comprising two benzene rings joined by a 3-carbon linkage chain (Navabi et al.,
157 2018). They are therefore shown as C6-C3-C6 compounds. The production of these
158 compounds is accomplished by two distinct pathways: the shikimic acid pathway and
159 the acetate pathway (Navabi et al., 2018). Over six thousand distinct flavonoids have
160 been identified. These chemicals are found in many different types of plants and
161 serve multiple purposes, including protecting plants from various biotic and abiotic
162 challenges, acting as signal molecules, detoxifying the body, and fighting microbes
163 (Liu *et al.* 2021). The degree of oxidation, the connection location of ring B, the
164 annularity of ring C, and the position of hydroxy groups in the carbon skeleton all
165 contribute to the classification of flavonoids into several subgroups, which include
166 isoflavones, flavones, flavonols, flavanones, flavanonols, and flavones (Wang et al.,
167 2018). Because of their extraordinary health advantages, foods rich in flavonoids are
168 called superfoods. Beverages, fruits, vegetables, cereals, beans, nuts, and tea make
169 up the vast majority of these plant-based items (Harborne and Williams, 2000).
170 Westerners get most of their flavonoids from wine, while Easterners get most of
171 theirs from tea. Green vegetables, onions, apples, berries, cherries, soybeans, and
172 citrus fruits are also believed to have a high concentration of dietary flavonoids (Butt
173 et al., 2014).

175 **Flavonoid biosynthesis in plants**

176 The shikimate, phenylpropanoid, and polyketide pathway is the main pathway for higher
177 plant species to synthesize flavonoids. These processes lead to the formation of
178 numerous different types of flavonoids, with chalcones and flavanones serving as
179 intermediates (Rehan, 2021). Plant species and sets of enzymes including hydroxylases,
180 reductases, and isomerases are essential for this principal process to generate different
181 forms of flavonoid skeletons. The flavanone is the progenitor of many flavonoid groups,
182 including anthocyanins, isoflavonoids, and proanthocyanidins, which are condensed
183 tannins (Figure 4). Typically, the phenylpropanoid pathway is where flavonoid synthesis
184 gets started. Following the completion of the shikimate pathway, the phenylpropanoid
185 pathway will eventually start. Erythrose 4-phosphate and phosphoenol pyruvate
186 condense, which is the first step in the shikimate pathway.



(a) Shikimate pathway in the biosynthesis of flavonoids

(b) Phenylpropanoid and Flavonoid pathway

187
188 **Figure 4:** Biosynthesis of various classes of flavonoids through the phenylpropanoid
189 and shikimate pathways (Rehan, 2021).

190
191 **METHODOLOGY**

192
193 **Selection of articles:** Article search was performed in PubMed, Google Scholar and
194 NCBI using combinations of keywords relating to “breast cancer” AND (“dietary
195 flavonoids” OR “flavonoids rich foods” OR “flavonols” OR “flavonones” OR
196 “anthocyanidins” OR “flavones” OR “isoflavones” OR “soy flavonoids” OR “quercetin” OR
197 “flavonoid subclasses” OR “polyphenolic compounds” OR “flavonoids rich foods” OR
198 “flavonoids rich fruits and vegetables”) AND (“preventive role” OR “treatment” OR
199 “flavonoids in drug” OR “flavonoids as chemoprotective agent”) etc. Only original articles
200 both in epidemiological and laboratory-based studies were sorted out for this review.

201 **Inclusion criteria:** The eligible criteria included studies in the English language published
202 between around 1990’s to date. Prospective cohort, case-control, and laboratory-based
203 studies that referred to the impact of flavonoids on breast cancer prevention, treatment,
204 or other roles were included. Titles and abstracts were independently screened. Hence, I
205 included articles that linked flavonoids to (a) breast cancer incidence, (b) breast cancer-
206 specific mortality, (c) the role of flavonoids, (d) therapeutic use of flavonoids, (e) multidrug
207 resistance, etc. The menopausal phase, cancer subtype, and type of anti-cancer
208 medication received by patients were not subject to any restrictions.

209
210 **Data Extraction:** Study characteristics were recorded in Excel spreadsheet as follows:
211 (1) name of the first author and publication year; (2) country or origin; (3) study design;
212 (4) length of follow-up; (5) total cases and controls; (7) exposures to flavonoids; (8) way
213 of flavonoids intakes; (9) Method and cell lines used and (10) RR/HR/OR from the most
214 fully adjusted model for the highest versus the lowest flavonoids exposure and their 95%
215 CI; (10) potential confounders and (11) limitations of studies

216 **Retrieval of information:** Odd Ratio (OR)/Risk Ratio (RR)/Hazard ratio (HR) along 95%
217 CI and p-value were carefully reviewed to reveal the association between different
218 subgroups of flavonoids and breast cancer risk. Other adjustments (e.g. age, menopausal
219 status, food habits, education level, race, BMI, multi-vitamin use, smoking habit, energy
220 intake, age at first live birth, physical activity level, lifetime lactation, etc.) were also
221 considered.

222 223 **How flavonoids fight against breast cancer**

224 Flavonoids, a plant source of polyphenolic compounds have grown its evidence from
225 the 1990's to date that can fight against breast cancer.

226 227 **1. Flavonoids as antioxidants**

228 Flavonoids demonstrate their antioxidant action in a variety of ways, including activating
229 antioxidant enzymes, scavenging reactive oxygen species (ROS), metal-chelating activity
230 etc. Enzymes including catalase, glutathione peroxidase (GPX), and superoxide
231 dismutase (SOD) constitute the backbone of this antioxidant defense system; they
232 deactivate free radicals generated by metabolic processes. (Jeeva et al., 2015). In 1999,
233 Nacia *et al.* demonstrated that flavonoids' intracellular antioxidative effect is dependent
234 on their interaction with GSH-PO, at least in cells expressing the enzyme, and concluded
235 that quercetin and catechin-activate glutathione peroxidase (GSH-PO) clearly.
236 Superoxide dismutase (SOD) levels were increased and glutathione peroxidase (GPx)
237 levels were potently decreased upon intraperitoneal dose of catechins.

238 Flavonoids, being polyphenols, have a high reactivity due to the hydroxyl substituents,
239 which makes them effective at scavenging free radicals through a process called
240 hydrogen atom abstraction (Korkina, and Afanas'Ev, 1997). Flavonoids possess the ability
241 to effectively capture the highest levels of active solar wavelengths (namely, UV-B and
242 UV-A). Not only that, but they are equally adept at halting the creation of ROS and
243 removing them once they've formed (Agati et al., 2012). Troxerutin, a flavonoid, exhibited
244 its cytoprotective efficacy by protecting various cell types (intestinal epithelial cells,
245 fibroblasts, and lymphocytes) against oxidative stress (Panat et al., 2016). The
246 antioxidant properties of silibinin and quercetin stem from their capacity to bind to iron or
247 copper ions and inhibit oxidases, in addition to scavenging free radicals and other
248 oxidizing chemicals (de Groot and Rauen, 2009). Diniz *et al.* in 2015 suggested that
249 flavonoids have the potential for neuroprotection in epilepsy.

250 Rutin and quercetin were found to be more effective in suppressing iron ion-dependent
251 lipid peroxidation systems due to their ability to form stable compounds with iron ions,
252 which are incapable of initiating lipid peroxidation (Afanas'Ev et al., 1989). Apigenin,
253 luteolin, kaempferol, quercetin, myricetin, and rutin, isoflavones (daidzein and genistein),

254 flavanones (taxifolin, naringenin, and naringin; and catechin) were found to have a greater
255 ability to reduce copper ions compared to iron ions (Mira et al., 2012).

256

257 **2. Flavonoids as phytoestrogens:**

258 High consumption of phytoestrogens can result in increased amounts of phytoestrogens
259 in the bloodstream, which have been empirically shown to exhibit estrogen-like actions.
260 Breast cancer risk is lower in women who regularly consume phytoestrogens, especially
261 throughout childhood (Lee et al., 2009). Korde *et al.* in 2009 also concluded a correlation
262 between breast cancer risk reduction with higher soy consumption during childhood,
263 adolescence, and adulthood the most potent protective effect was shown in childhood.
264 According to Rice and Whitehead (2006), phytoestrogens have the ability to change
265 estrogen production by inhibiting aromatase (known as cytochrome P450 19 aromatase
266 (Cyp19)) as well as 17 β -hydroxysteroid dehydrogenases (HSD), estrone sulfatase, and
267 sulfotransferase. In breast tissue, these enzymes play a role in the generation of estradiol,
268 and when they are overexpressed, they are associated with an increased risk of breast
269 cancer. Research has shown that a number of flavonoids can significantly lower estrogen
270 levels in the blood. They may be useful in treating malignancies that are resistant to other
271 treatments, even though their efficacy is lower than that of steroidal aromatase inhibitors
272 often employed in clinical practice.

273

274 **3. Flavonoids as CYP1A1 inhibitors/substrates**

275 Inhibiting CYP1A is a plausible approach to preventing the development of cancer. The
276 selective metabolism of dietary flavonoids by CYP1 enzymes, resulting in the production
277 of conversion products that limit the proliferation of cancer cells, is the key mechanism
278 behind cancer prevention. Cytochrome P450 enzymes (CYP450s) are the primary
279 enzyme group accountable for the metabolism and detoxification of foreign substances,
280 which are commonly referred to as xenobiotics. They are also involved in the biosynthesis
281 as well as metabolism of endogenous compounds like fatty acids and steroids. CYP450s
282 in the liver where they are highly expressed catalyze reactions that are responsible for
283 converting lipophilic compounds into more hydrophilic derivatives. Most drugs follow this
284 biotransformation pathway to be catabolized and eliminated from the body (Gonzalez,
285 1988). But other xenobiotic substances, such as dietary flavonoids, can modulate the
286 CYP450 enzyme function, either increasing or decreasing the catalytic activity. As a
287 result, drug detoxification or functionality could be interrupted (Hodek et al., 2002).

288 It has been established that the CYP1 family is involved in the process of carcinogenesis
289 and the advancement of cancer and demonstrated through several research that
290 flavonoids, such as resveratrol, have the ability to effectively inhibit the activation of CYP1
291 in both in-vivo and ex-vivo environments because of their function as competitive
292 antagonists for the aryl hydrocarbon receptor AhR, which plays a role in the activation of
293 CYP1 expression (Ciolino et al., 1999). For instance, quercetin and myricetin are potent
294 CYP1B1 inhibitors, but they appear to have a lesser impact on the CYP1A1 and CYP1A2
295 variants (Arroo et al., 2009).

296

297 **4. Flavonoids as ABC (ATP-binding cassette) transporter regulators**

298 ATP-binding cassette superfamily comprises P-glycoprotein (P-gp), ABCG2, multidrug
299 resistance proteins 1 and 2 (MRP1 and 2) and breast cancer resistance protein (BCRP).

300 Several xenobiotic-related compounds rely on these membrane proteins for absorption,
301 distribution, and excretion. These transporters are essential for the distribution of
302 pharmaceuticals often used in medical therapy, and they are the principal family
303 responsible for removing drugs from cells (Tan et al., 2013). In recent years, there has
304 been a significant amount of focus placed on the role that ATP-binding cassette (ABC)
305 transporters play role in the biology of cancer cells (Nobili et al., 2019) and for this reason
306 they are the subject of substantial investigation. The development of multidrug resistance,
307 which is predominantly driven by factors related to ABC transporter genes, is the primary
308 obstacle that stands in the way of effective cancer treatment. There are several examples
309 of ATP-binding cassette (ABC) transporters that impart resistance to particular
310 chemotherapy medications. Some of these transporters include breast cancer resistance
311 protein (BCRP), P-glycoprotein (P-gp), and multidrug resistance-associated protein 1
312 (MRP1). Intestinal efflux ABC (ATP binding cassette) transporters, some of which include
313 P-glycoprotein, breast cancer resistance protein, and multidrug resistance-related
314 proteins, perform the function of "pumping doors" to regulate the elimination of flavonoids
315 from intestinal epithelial cells into either the intestinal cavity or the systemic circulation.
316 Breast cancer resistance protein (BCRP) is an ATP-binding cassette transporter that has
317 recently come to light due to the report that it can regulate drug disposition and induce
318 multidrug resistance (MDR) to certain important anticancer treatments. Flavonoids serve
319 as inhibitors of transporters and restrict the ATP hydrolysis activity that leads to the
320 elimination of anticancer drugs from tumor tissues (Zhang et al., 2004). Presumably, this
321 occurs through the interaction with the nucleotide-binding domain (NBD), which is the
322 specific site of action for flavonoids in both BCRP and P-gp (Morris and Zhang, 2006).
323 The complete understanding of the interaction between flavonoids and BCRP, despite
324 their ability to effectively modify medication pharmacokinetics, remains incomplete.

325

326 **5. Flavonoids' impact on cell death, cell cycle reversal, and other signaling** 327 **mechanisms**

328 Flavonoids are well acknowledged for their significance as anti-carcinogenic substances
329 and have shown remarkable efficacy as cytotoxic anti-cancer medicines, triggering
330 apoptosis in most cancer cells. A number of flavonoids have shown promise in inducing
331 cell death in cancers, including breast cancer (Huang et al., 2012) via means of the two
332 primary apoptotic pathways: internal apoptosis involving caspase-9 and mitochondria,
333 and extrinsic apoptosis involving caspase-8 and death receptors (Kamsteeg et al., 2003).
334 Brusselmans *et al.* proposed the ability of flavonoids to inhibit fatty acid synthase (FAS)
335 is closely linked to their potential to cause apoptosis in cancer cells.

336

337 **6. Effect of flavonoids on multi-drug resistance**

338 The primary factor contributing to the failure of the majority of cancer chemotherapy is
339 drug resistance. The fundamental cause of chemoresistance is believed to be the
340 dysregulation of the epigenetic machinery. It was reported that the dysregulation of DNA
341 methylation in resistant cancer cells leads to the abnormal expression of genes involved
342 in cancer proliferation, apoptosis, DNA repair, and drug efflux. The dysregulation of many
343 enzymes that catalyze histone post-translational modifications can also lead to changes
344 in chromatin structure. Moreover, expression of numerous drug-resistance genes
345 is regulated by this imbalance. Alterations in the patterns of microRNA play a role in the

346 development of drug resistance in cancer cells (To Kenneth and Cho, 2021). Curcumin,
347 stilbenes, ellagitannins, phenol carboxylic acids, and flavones are all examples of
348 phytochemicals that possess antioxidant, anti-inflammatory, and anti-cell-growth
349 activities. These features make them useful chemo-preventive agents. In addition to this,
350 they prevent the development of new blood vessels and the spread of cancer cells,
351 regulate immune and inflammatory responses, and deactivate substances that can cause
352 cancer. Furthermore, preclinical and clinical investigations have revealed that these drugs
353 successfully change several pathways, which prevents cancer from developing multidrug
354 resistance (Costea et al., 2020). Researchers are now interested in the effect of
355 flavonoids on drug resistance in chemotherapies. To overcome treatment resistance in
356 breast cancer, flavonoids have been suggested as a potential medication by many
357 studies. According to Rao et al. (2012), luteolin effectively reverses multidrug resistance
358 (MDR) in breast cancer cells that are resistant to mitoxantrone.

359

360 **RESULTS AND DISCUSSION**

361 Flavonoid consumption may have an association with a lower risk of cancer development,
362 according to the findings of the cohort and case-control studies that were reviewed in this
363 article (Table 2). The fact that flavonoids have been shown to have an inverse correlation
364 with cancer risk may be explained by the fact that flavonoids have an effect on a number
365 of critical biological activities. The capacity of flavonoids to scavenge free radicals has
366 been characterized in experimental settings to a satisfactory degree.

367 More recently, flavonoids have been shown to influence the signal transduction pathway
368 (Melnik et al., 2002), accelerate apoptosis (Choi et al., 2001), suppress inflammation and
369 impede proliferation in human cancer cell lines (Manthey et al., 2002). The in vitro and
370 animal model systems were used to publish these findings. Some flavonoids can also
371 enhance the transcription of phase II detoxifying enzymes, which would help eliminate
372 pro-carcinogenic substances. This would reinforce the idea that flavonoids prevent
373 cancer. Flavonoids were shown to dramatically reduce the number of focal areas of
374 dysplasia that were caused by exposure to azoxymethane (AOM), according to the
375 findings of a study that employs mice that were subjected to treatment with azoxymethane
376 (AOM) and fed either a standard diet, a standard diet plus rutin, or a standard diet plus
377 quercetin (Yang et al., 2000). This kind of research is noteworthy because it suggests
378 that flavonoids could be related to a number of early-stage processes in the cascade that
379 result in cancer development.

380 The development of multidrug resistance (MDR) in cancer cells through transporter-
381 mediated active efflux of cytotoxic drugs is one of the processes that has been studied
382 and characterized the most. Previous studies have demonstrated that BCRP (Breast
383 Cancer resistance protein) bestows resistance upon several significant anticancer
384 agents, including doxorubicin, topotecan, mitoxantrone, SN38, methotrexate, and
385 flavopiridol (Doyle et al., 1998). Therefore, to potentially reverse multidrug resistance
386 (MDR), it is necessary to identify inhibitors of BCRP that are both potent and nontoxic.
387 Flavonoids, which have a lengthy history of human ingestion and a stellar safety record,
388 are an essential component of the average diet (Havsteen, 2002). Furthermore, these
389 compounds have been linked to numerous anticancer mechanisms (Havsteen, 2002), as
390 well as synthetic compounds that are analogous to them, including flavone acetic acid
391 and flavopiridol (Senderowicz et al., 1998).

392 In addition to its role in regulating drug distribution, the ATP-binding cassette transporter
393 breast cancer resistance protein (BCRP), which was only recently discovered, has been
394 shown to impart multidrug resistance (MDR) to many important anticancer drugs.
395 Flavonoids, a group of polyphenolic chemicals, are widely present in botanical products
396 and dietary sources. It has been reported that flavonoids interact with P-glycoprotein and
397 multidrug resistance-associated protein 1; nevertheless, the nature of their interaction
398 with BCRP remains unknown. The objective of this study was to evaluate the effects of
399 twenty naturally derived flavonoids on the cytotoxicity and cellular accumulation of
400 mitoxantrone in human cell lines that exhibit BCRP expression but lack BCRP. These
401 investigations utilized human breast cancer cells (MCF-7) that overexpressed BCRP and
402 large-cell lung carcinoma cells (NCI-H460) that lacked BCRP.

403 A considerable number of the flavonoids that were assessed (50 μ M) exhibited an
404 augmentation in mitoxantrone accumulation in cells that overexpressed BCRP, while also
405 effectively reversing the development of mitoxantrone resistance. It is important to note
406 that these flavonoids did not have any effect on the BCRP-negative cells that
407 corresponded to them, which suggests that they act as antioxidants. The impact of these
408 flavonoids on the cellular accumulation and cytotoxicity of mitoxantrone was found to be
409 concentration-dependent, with the majority of the flavonoids exhibiting significant
410 modifications at concentrations below 10 μ M (Hung et al., 2015). Additional research is
411 required to optimize the compounds' bioavailability, biodistribution, and safety in
412 preparation for future human studies. In particular, it is imperative to elucidate how every
413 flavonoid, in accordance with its distinct chemical composition, can traverse the
414 membrane.

415 Based on the available research, it may be concluded that flavonoids have a negative
416 correlation with the chance of developing cancer. Further research utilizing up-to-date
417 dietary databases with HPLC results for flavonoid estimations from food will offer
418 conclusive information about the correlation between plant chemicals and the risk of
419 cancer. Furthermore, conducting feeding experiments that can accurately describe the
420 process of how plant flavonoids are absorbed, metabolized, and eliminated from the body,
421 as well as their interaction with important enzyme systems, would be crucial in
422 comprehending the underlying biological reasons for the negative relationship between
423 flavonoid consumption and the risk of developing cancer. Meanwhile, it is strongly
424 recommended that the public adhere to dietary recommendations for cancer prevention,
425 which involve consuming five or more servings of fruits and vegetables daily. Moreover,
426 the findings of the researcher might offer significant backing for additional exploration into
427 evaluating the effectiveness and safety of flavonoid chemicals whether included in a
428 nutritious diet or used as a supplementary treatment for human malignancies.

Table 2: Summary of epidemiological studies investigating the association of flavonoid intake with breast incidence and mortality.

| Author(s) | Method | Subject(s) | Cancer type | Exposer | Conclusion |
|------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------|---------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Zheng et al., 1999 | Case-control study Structured questionnaire Urinary excretion rates In-person interviews | Sixty (60) breast cancer cases and individually matched controls examined for rates of excretion in urine Face-to-face interviews conducted for 746 urbane Chinese women in Shanghai | Breast cancer | Total phenols and five major isoflavonoids (glycitein, daidzein, genistein, equol and O-desmethylangol ensin) | <ul style="list-style-type: none"> Breast cancer cases had lower urinary total phenols and all isoflavonoids. The disparity between the case and control groups became more apparent when comparing the median levels of these compounds Individuals with the greatest levels of daidzein, glycitein, and total isoflavonoids had around 50% lower risk of cancer compared to those with the lowest levels. |
| Horn-Ross et al., 2001 | Case-control study Food Frequency Questionnaire (FFQ) Ontario phytoestrogen database Multivariate logistic regression | 1,326 cases and 1,657 controls non-Asian US women (35–79 years) residing in the San Francisco Bay Area | Breast cancer | Isoflavones, total isoflavones, lignans, total lignans, total phytoestrogens | <ul style="list-style-type: none"> Phytoestrogen was found not to be protective against breast cancer risk (odds ratio = 1.0 and 95% CI = 0.80, 1.3 for the highest versus lowest quartile) |
| Peterson et al., 2003 | Case-control study in Greece Semi-quantitative food frequency questionnaire US Department of Agriculture-Iowa State University Database | 820 women diagnosed with breast cancer against 1548 control women | Breast cancer | Flavones, flavonols, flavanones, flavan-3-ols, isoflavones and anthocyanidins | <ul style="list-style-type: none"> A significant negative correlation between flavone consumption and breast cancer was observed. The odd ratio for an increment in daily flavone consumption equal to one standard deviation (0.5 mg/day) was 0.87 (95% CI = 0.77, 0.97). |
| Silva et al., 2004 | Case-control study Face-to-face interview with food frequency questionnaire (FFQ) Conditional logistic regression models | 240 South Asians diagnosed with breast cancer residing in England against 477 population-based controls matched for age | Breast Cancer | Total isoflavones, (genistein, daidzein) as well as total lignans (secoisolariciresinol, matairesinol) | <ul style="list-style-type: none"> After controlling for known and established risk factors for breast cancer, there is moderate evidence of a dose-response relationship between isoflavone consumption and the likelihood of developing breast cancer (P value for trend 0.08). |

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|-------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------|-------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Bosetti et al., 2005 | Case-control study Standard structured questionnaire Food Frequency Questionnaire (FFQ) | 2,569 women with histologically confirmed breast cancer and 2,588 hospital controls (age range 23-74 years, median age 55) from six Italian areas | Breast Cancer | Flavanones, flavones, anthocyanidins, flavan-3-ols, flavonols, and isoflavones | <ul style="list-style-type: none"> ● Increased intake of flavones reduced the risk of breast cancer (OR = 0.81 for the highest versus the lowest quintile with a P-trend of 0.02). ● Significant association of risk reduction of breast cancer was not found for other flavonoids |
| Fink et al., 2006 | Case-control study Interviewed about known as well as suspected risk factors Food Frequency Questionnaire (FFQ) | 1,434 female cases (English-speaking and newly diagnosed with breast cancer) and 1,440 control. Both are from Long Island, New York, USA | Breast Cancer | Flavonols, flavan-3-ols, flavones, flavanones, isoflavones, lignans anthocyanidins, | <ul style="list-style-type: none"> ● Flavonoid intake was associated with breast cancer risk reduction. ● The reduction was most pronounced among postmenopausal women for flavonols (odds ratio (OR) ¼ 0.54 with 95% CI = 0.40, 0.73). |
| Cotterchio et al., 2007 | Case-control study Block food frequency questionnaire (FFQ) expanded to include foods containing phytoestrogen | 3,063 breast cancer cases (age range 25-74 years and diagnosed in 2002 and 2003) and 3430 controls were identified by using the Ontario Cancer Registry | Breast cancer | Lignans and isoflavones | <ul style="list-style-type: none"> ● Lignan intake was found to be effective in reducing breast cancer risk. ● A significant reduction in breast cancer risk was observed in overweight premenopausal women who had a high consumption of phytoestrogens |
| Cutler et al., 2008 | Prospective cohort Food frequency questionnaire Food composition databases Baseline questionnaire (1985) and 5 follow-up questionnaires (1987, 1989, 1992, 1997, 2004) Followed for cancer incidence from 1986 through 2004 | 34,708 postmenopausal women (55-69 years) in the Iowa Women's Health Study | Pancreatic, lung, breast, colorectal and upper aerodigestive cancer | Seven flavonoid subclasses along with total flavonoids | <ul style="list-style-type: none"> ● Isoflavone intake and overall cancer incidence were inversely associated (HR = 0.93; 95% CI = 0.86–1.00). |
| Torres-Sanchez | Case-control study | A total of 198 women with breast cancer (age range 21–79 | Breast cancer | Onion, lettuce, tea, apple, and spinach | <ul style="list-style-type: none"> ● The adjusted odds ratio was 0.27 (95% CI = 0.16, 0.47) with a statistically significant |

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|-----------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------|---------------|----------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| et al., 2009 | Semiquantitative food frequency questionnaire (FFQ) Logistic regression models | years) and age-matched control (± 3 yr) for each case | | | trend ($P < 0.001$) for the consumption of more than one slice of onion per day. <ul style="list-style-type: none"> Consumption of lettuce and spinach had a substantial preventive impact. |
| Ha et al., 2009 | Cross-sectional study Dietary questionnaire | 128 women aged 40–79 year and newly diagnosed with invasive breast cancer. | Breast cancer | Isoflavonoids and lignans | <ul style="list-style-type: none"> The chance of getting a cancer diagnosis at any stage other than stage 1 was 32% lower in women who consumed a higher level of phytoestrogens. |
| Pantavos et al., 2014 | Prospective cohort study Semiquantitative FFQ Ferric reducing antioxidant potential (FRAP) Follow-up of 17 years Crude and multivariate Cox proportional hazard models | Women aged 55 years and older ($n = 3,209$) and 199 breast cancer cases, identified from the Rotterdam study | Breast cancer | Vitamin A, C and E, selenium, flavonoids and carotenoids | <ul style="list-style-type: none"> No correlation between total antioxidant intake and the risk of breast cancer was found. A reduced consumption of flavonoids was linked to an increased risk of breast cancer in women aged 70 and above (HR = 1.80; 95% CI = 1.09, 2.99). |
| Wang et al., 2014 | Prospective cohort study Self-administered questionnaires Modified Willett FFQ Cox proportional hazards regression | 56,630 postmenopausal women from the CPS-II Nutrition Cohort were included in the analytical cohort | Breast cancer | Total flavonoids anthocyanidins flavan-3-ols flavanones flavones etc | <ul style="list-style-type: none"> Total flavonoid intake was not associated with breast cancer risk. Despite this, the total risk of breast cancer was somewhat inversely related to flavone consumption. |
| Feng et al., 2020 | Case-control study Food frequency questionnaire (Face-to-face) Multivariable logistic regression models. | 1522 breast cancer cases and 1547 frequency-matched controls (both cases and controls are selected Chinese population) | Breast cancer | Total flavonoids anthocyanidins flavanols flavan-3-ol etc. | <ul style="list-style-type: none"> After considering possible influencing factors, a clear inverse relationship was shown between the risk of developing breast cancer and the intake of various types of flavonoids, |

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| Author | Cell line | Model | Flavonoids exposers | Conclusion |
|----------------|-------------------|---------------------------------------------------|------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Miksicek, 1993 | MCF7, HeLa, COS-7 | Transfection studies, competition binding studies | Chalcones, flavanones, | <ul style="list-style-type: none"> Selected hydroxylated flavonoids interacted directly with the estrogen receptor based on their ability to |

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|-----------------------|-----------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| | | | flavones, and isoflavones | <p>compete for binding 17β-[3H]estradiol to the receptor in the free cell extracts.</p> <ul style="list-style-type: none"> ● These substances exhibited lower activity, when measured on a molar basis, compared to 17β-[3H]estradiol or the synthetic dihydroxystilbene estrogens. |
| So et al., 1996 | Human breast carcinoma cell line, MDA-MB-435. | Proliferation assay, growth curve at iC_{50} , MTT viability assay, tumorigenicity experiments | Two citrus flavonoids along with four noncitrus flavonoids | <ul style="list-style-type: none"> ● Among the six individual flavonoids tested, baicalein had the highest potency to effectively suppress the proliferation of the cells. ● Hesperetin and naringenin (found in citrus fruits) exhibited moderate inhibitory effects. |
| Zand et al., 2000 | BT-474 human breast cancer cells | ELISA-type immunofluorometric assays, | 72 flavonoids and structurally related compounds. | <ul style="list-style-type: none"> ● Flavonoids exhibited significant steroid hormone activity. Based on the findings they concluded that flavonoids may have an effect on cancer risk, prevention and cancer therapeutics. |
| Imai et al., (2004) | K562 (K562/BCRP), LLC/BCRP, K562/MDR, and KB/MRP Cell Lines. | Cell growth inhibition assay, cellular [3 H]genistein accumulation in K562/BCRP Cells, transcellular transport assay of [3 H]genistein | Estrone, kaempferol, naringenin, naringenin-7-glucoside | <ul style="list-style-type: none"> ● Phytoestrogens/flavonoids, including genistein, naringenin, acacetin, and kaempferol, boosted the toxicity of SN-38 and mitoxantrone in BCRP-transduced K562 (K562/BCRP) cells. |
| Zhang et al., 2004 | MCF-7, NCI-H460 | Western blot analysis of BCRP, P-gp, and MRP1, mitoxantrone accumulation studies, mitoxantrone cytotoxicity studies., | Apigenin, biochanin a, chrysin, daidzein, fisetin, morin, myricetin, genistein etc. | <ul style="list-style-type: none"> ● Many of the tested flavonoids (50 M) are BCRP inhibitors. ● Chrysin and biochanin A exhibited the highest level of potency as inhibitors of BCRP. |
| Katayama et al., 2007 | Human leukemia K562 cells and human epidermoid carcinoma KB-3-1 cells | Growth inhibition assay, topotecan uptake | Flavone, flavonol, isoflavone, chalcone | <ul style="list-style-type: none"> ● 3,4,7-trimethoxyXavone demonstrated the most potent anti-BCRP activity for SN-38 and mitoxantrone respectively. ● 3,4,7-trimethoxyXavone and acacetin, exhibited minimal anti-P-gp activity, while the others did not demonstrate any inhibitory effects on P-gp. |
| Chen et al., 2007 | MDA-MB-231 breast cancer cell cultures and xenografts | Proteasomal chymotrypsin-like and caspase-3/caspase-7 activity assays, MTT assay, western blot analysis and human breast tumor xenograft experiments | Apigenin | <ul style="list-style-type: none"> ● Apigenin suppresses the proteasomal chymotrypsin-like activity and induces apoptosis in both cultured MDA-MB-231 cells and MDA-MB-231 xenografts. |

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|------------------------------|-----------------------------------------------|---------------------------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Yang et al., 2012 | MCF-7, MDA-MB-231, MCF-10A cells | MTT colorimetric assay, Luminescence ATP detection assay, Western blot analysis | Fisetin (3,3',4',7-tetrahydroxyflavone) | <ul style="list-style-type: none"> ● Fisetin exhibited greater cytotoxicity in MCF-7 human breast cancer cells compared to MDA-MB-231 cells. ● Fisetin can trigger a novel atypical apoptosis in caspase-3-deficient MCF-7 cells. |
| Shan et al., 2013 | MCF-7 and MDA-MB-231 | MTT, clonogenicity assay, apoptosis assay, western blot analysis | Triticuside A from wheat bran | <ul style="list-style-type: none"> ● Triticuside A dose-dependently suppressed the growth of human breast cancer cells (MCF-7 and MDA-MB-231). |
| Pradhan et al., 2015 | MCF-7 cell lines and MCF-7 xenografts | Proteasomal chymotrypsin-like and caspase-3/ caspase-7 activity assays, cell viability assay, xenograft experiments | Quercetin | <ul style="list-style-type: none"> ● Quercetin suppresses the proteasomal chymotrypsin-like activity and triggers apoptosis in both cultivated MCF-7 cell lines and MCF-7 xenografts. |
| Vrhovac Madunic et al., 2017 | ER-positive MCF-7, triple-negative MDA MB-231 | MTT assay, cell viability, comet and lipid peroxidation assays, | Apigenin | <ul style="list-style-type: none"> ● Apigenin caused cell death in both cell lines resulting in significant toxicity and apoptosis. ● No significant cytogenotoxic effects were detected in normal cells. |
| Santes-Palacios et al., 2019 | <i>Escherichia coli</i> DH5 α cells | Ethoxyresorufin O-deethylase activity (EROD) assay, bacterial mutagenicity assay | Quercetin, myricetin, luteolin, fisetin, kaempferol, 5-hydroxyflavone, flavone etc. | <ul style="list-style-type: none"> ● 5-hydroxyflavone, 3 hydroxy-flavone and flavone exerted the most potent inhibitory activity with IC50 values of 0.07, 0.10 and 0.08 μM respectively. |

LIMITATIONS

This article only reviewed some of open access articles available in online. This narrowed down the number of articles reviewed. Some articles reviewed in this study may not adequately account for all confounding factors that could influence cancer risk. Few studies showed negative finding in the context of breast cancer prevention and treatment and need more investigation.

CONCLUSION

There is evidence that dietary flavonoids can lower the risk of cancer, especially breast cancer. This review briefly discussed about that the different sources of flavonoid and their subclasses which are the polyphenolic compounds mainly present in plant sources like fruits, vegetables, and cereals. They mainly reduce the risk of breast cancer by antioxidant activity, anti-carcinogenic effect, inhibiting apoptosis, reducing estrogen activity, and inhibiting the growth of breast cancer cells. Researchers are enthusiastic about revealing the therapeutic properties of flavonoids to treat breast cancer cases. There are linings enough that some polyphenolic compounds are found as effective drugs for treating breast cancer.

COMPLETING INTERESTS

The authors of this manuscript certify that they have NO affiliations with or involvement in any organization or entity with any financial interest (such as honoraria; educational grants; participation in speakers' bureaus; membership, employment, consultancies, stock ownership, or other equity interest; and expert testimony or patent-licensing arrangements), or non-financial interest (such as personal or professional relationships, affiliations, knowledge or beliefs) in the subject matter or materials discussed in this manuscript.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

REFERENCES

1. Ganesan, K., Du, B., & Chen, J. (2022). Effects and mechanisms of dietary bioactive compounds on breast cancer prevention. *Pharmacological Research* 178: 105974. [https://DOI: 10.1016/j.phrs.2021.105974](https://doi.org/10.1016/j.phrs.2021.105974)
2. Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., & Bray, F. (2021). Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA A Cancer J Clin* 71: 209-249. [https://doi: 10.3322/caac.21660](https://doi.org/10.3322/caac.21660)

3. WHO (2023). Breast cancer. Link: <https://www.who.int/news-room/fact-sheets/detail/breastcancer#:~:text=Overview,producing%20lobules%20of%20the%20breast>.
4. Colditz, G. A. (1998). Relationship between estrogen levels, use of hormone replacement therapy, and breast cancer. *Nat Rev Cancer* 90: 814-32. <https://doi.org/10.1093/jnci/90.11.814>
5. Thomas, C., & Gustafsson, J. A. (2011). The different roles of ER subtype in cancer biology and therapy. *Journal of Nature Reviews Cancer* 11: 597-608. <https://doi.org/10.1038/nrc3093>.
6. Hamer, J. & Warner, E. (2017). Lifestyle modifications for patients with breast cancer to improve prognosis and optimize overall health. *CMAJ* 189: E268–e274. <https://doi.org/10.1503/cmaj.160464>.
7. Peterson, J. & Dwyer, J. (1998). Flavonoids: dietary occurrence and biochemical activity. *Nutr Res.* 18: 1995-2018.
8. Jang, H., Chung, M. S., Kang, S. S., & Park, Y. (2018). Association between the dietary inflammatory index and risk for cancer recurrence and mortality among patients with breast cancer. *Nutrients* 10: 1095. <https://doi.org/10.3390/nu10081095>.
9. Franke, A. A., Cooney, R. V., Custer, L. J., Mordan, L. J., & Tanaka, Y. (1998). Inhibition of neoplastic transformation and bioavailability of dietary flavonoid agents. *Adv Exp Med Biol.* 439: 237-48. https://doi.org/10.1007/978-1-4615-5335-9_17
10. Peterson, J., Lagiou, P., Samoli, E., Lagiou, A., Katsouyanni, K., La Vecchia, C. *et al.* (2003). Flavonoid intake and breast cancer risk: a case-control study in Greece. *British Journal of Cancer* 89: 1255–1259. <https://doi.org/10.1038/sj.bjc.6601271>
11. Who (2023). WHO launches new roadmap on breast cancer. Link: <https://www.who.int/news/item/03-02-2023-who-launches-new-roadmap-on-breast-cancer>
12. Zhang, Y., Zhou, Y., Mao, F., Yao, R., & Sun, Q. (2020). Ki-67 index, progesterone receptor expression, histologic grade and tumor size in predicting breast cancer recurrence risk: A consecutive cohort study. *Cancer Commun (Lond)* 40(4): 181-93.
13. Fragomeni, S. M., Sciallis, A., & Jeruss, J. S. (2018). Molecular subtypes and local-regional control of breast cancer. *Surg Oncol Clin N Am.* 27(1): 95-120. <https://doi.org/10.1016/j.soc.2017.08.005>
14. Barzaman, K., Karami, J., Zarei, Z., Hosseinzadeh, A., Kazemi, M. H., Moradi-Kalbolandi, S., ... & Farahmand, L. (2020). Breast cancer: Biology, biomarkers, and treatments. *International immunopharmacology*, 84, 106535. PMID: 32361569. <https://doi.org/10.1016/j.intimp.2020.106535>
15. Gao, J. J., & Swain, S. M. (2018). Luminal a breast cancer and molecular assays: a review. *Oncologist.* 23(5): 556–565. <https://doi.org/10.1634/theoncologist.2017-0535>
16. Engel, R. H. & Kaklamani, V. G. (2007). HER2-positive breast cancer: current and future treatment strategies. *Drugs.* 67(9): 1329-1341. <https://doi.org/10.2165/00003495-200767090-00006>
17. Duffy, M. J., Synnott, N. C., & Crown, J. (2018). Mutant p53 in breast cancer: potential as a therapeutic target and biomarker. *Breast Cancer Res Treat.* 170(2): 213-219. <https://doi.org/10.1007/s10549-018-4753-7>

18. Ahern, T. P., Lash, T. L., Damkier, P., Christiansen, P. M., & Cronin-Fenton, D. P. (2014). Statins and breast cancer prognosis: evidence and opportunities. *The lancet oncology*, 15(10), e461-e468. PMID: PMC4167822. [https://doi.org/10.1016/S1470-2045\(14\)70119-6](https://doi.org/10.1016/S1470-2045(14)70119-6)
19. Loibl, S. & Gianni, L. (2017). HER2-positive breast cancer. *Lancet*. 389(10087): 2415-2429. [https://doi.org/10.1016/S0140-6736\(16\)32417-5](https://doi.org/10.1016/S0140-6736(16)32417-5)
20. Soerjomataram, I. & Bray, F. (2021). Planning for tomorrow: global cancer incidence and the role of prevention 2020-2070. *Nat Rev Clin Oncol*. 18(Suppl. 1). <https://doi.org/10.1038/s41571-021-00514-z>
21. Winkel-Shirley, B. (2001). Flavonoid biosynthesis. A colorful model for genetics, biochemistry, cell biology, and biotechnology. *Plant physiology*, 126(2), 485-493. <https://doi.org/10.1104/pp.126.2.485>
22. Nabavi, S. M., Šamec, D., Tomczyk, M., Milella, L., Russo, D., Habtemariam, S. *et al.* (2018). Flavonoid biosynthetic pathways in plants: Versatile targets for metabolic engineering. *Biotechnol. Ad*. 38: 107316. [https://DOI: 10.1016/j.biotechadv.2018.11.005](https://DOI:10.1016/j.biotechadv.2018.11.005)
23. Liu, W., Feng, Y., Yu, S., Fan, Z., Li, X., Li, J., & Yin, H. (2021). The Flavonoid Biosynthesis Network in Plants. *Int J Mol Sci*. 22(23):12824. [https://DOI: 10.3390/ijms222312824](https://DOI:10.3390/ijms222312824)
24. Wang, T. Y., Li, Q., & Bi, K. S. (2018). Bioactive flavonoids in medicinal plants: Structure, activity and biological fate. *Asian J. Pharm. Sci*. 13:12-23. doi: 10.1016/j.ajps.2017.08.004.
25. Harborne, J.B. & Williams, C. A. (2000). Advances in flavonoid research since 1992. *Phytochemistry* 55: 481-504. [https://DOI: 10.1016/s0031-9422\(00\)00235-1](https://DOI:10.1016/s0031-9422(00)00235-1)
26. Butt, M. S., Imran, A., Sharif, M. K., Ahmad, R. S., Xiao, H., Imran, M., & Rsool, H. A. (2014). Black Tea Polyphenols: A Mechanistic Treatise. *Critical Reviews in Food Science and Nutrition*, 54(8), 1002–1011. <https://doi.org/10.1080/10408398.2011.623198>
27. Rehan, M. (2021). Biosynthesis of Diverse Class Flavonoids via Shikimate and Phenylpropanoid Pathway. *Intech Open*. [https://doi: 10.5772/intechopen.96512](https://doi:10.5772/intechopen.96512)
28. Jeeva, J. S., Sunitha, J., Ananthalakshmi, R., Rajkumari, S., Ramesh, M., & Krishnan, R. (2015). Enzymatic antioxidants and its role in oral diseases. *J Pharm Bioallied Sci*. 7(Suppl 2): S331-3. [https://doi: 10.4103/0975-7406.163438](https://doi:10.4103/0975-7406.163438)
29. Korkina, L. G, Afanas'ev, I. B (1997). Antioxidant and chelating properties of flavonoids. *Adv Pharmacol*. 38:151-63. doi: 10.1016/s1054-3589(08)60983-7.
30. Nacaia, H., Takekoshi, S., Takagi, T., Honma, T., & Watanabe, K. (1999). Antioxidative Action of Flavonoids, Quercetin and Catechin, Mediated by the Activation of Glutathione Peroxidase. *Tokai J Exp Clin Med*. 24(1): 1-11. PMID: 10530620
31. Agati, G., Azzarello, E., Pollastri, S., & Tattini, M. (2012). Flavonoids as antioxidants in plants: location and functional significance. *Plant science*, 196, 67-76. PMID: 23017900. [https://doi: 10.1016/j.plantsci.2012.07.014](https://doi:10.1016/j.plantsci.2012.07.014)
32. Panat, N. A., Maurya, D. K., Ghaskadbi, S. S., & Sandur, S. K. (2016). Troxerutin, a plant flavonoid, protects cells against oxidative stress-induced cell death through radical scavenging mechanism. *Food Chemistry* 94: 32-45. [https://DOI: 10.1016/j.foodchem.2015.07.078](https://DOI:10.1016/j.foodchem.2015.07.078)
33. de Groot, H. D. & Rauen, U. (1998). Tissue injury by reactive oxygen species and the protective effects of flavonoids. *Fundamental and clinical Pharmacology* 12(3): 249-255. [https://DOI: 10.1111/j.1472-8206.1998.tb00951.x](https://DOI:10.1111/j.1472-8206.1998.tb00951.x)

34. Diniz, T. C., Silva, J. C., Lima-Saraiva, S. R. G. D., Ribeiro, F. P. R. D. A., Pacheco, A. G. M. *et al.* (2015). The role of flavonoids on oxidative stress in epilepsy. *Cell Longev* 2015:171756. <https://doi:10.1155/2015/171756>
35. Afanas' ev, I. B., Dcrozshko, A. I., Brodskii, A. V., Kostyuk, V. A., & Potapovitch, A. I. (1989). Chelating and free radical scavenging mechanisms of inhibitory action of rutin and quercetin in lipid peroxidation. *Biochemical pharmacology*, 38(11), 1763-1769. PMID: 2735934. [https://doi:10.1016/0006-2952\(89\)90410-3](https://doi:10.1016/0006-2952(89)90410-3)
36. Mira, L., Tereza Fernandez, M., Santos, M., Rocha, R., Helena Florêncio, M., & Jennings, K. R. (2002). Interactions of Flavonoids with Iron and Copper Ions: A Mechanism for their Antioxidant Activity. *Free Radical Research* 36(11): 1199-1208. <https://doi:10.1080/1071576021000016463>
37. Lee, S. A., Shu, X. O., Li, H., Yang, G., Cai, H., Wen, W. *et al.* (2009). Adolescent and adult soy food intake and breast cancer risk: results from the Shanghai Women's Health Study. *The American Journal of Clinical Nutrition* 89 (6): 1920-6. <https://doi:10.3945/ajcn.2008.27361>
38. Korde, L. A., Wu, A. H., Fears, T., Nomura, A. M., West, D. W., Kolonel, L. N. *et al.* (2009). Childhood soy intake and breast cancer risk in Asian American women. *Cancer Epidemiology, Biomarkers & Prevention* 18(4): 1050-9. <https://doi:10.1158/1055-9965.EPI-08-0405>.
39. Rice, S. & Whitehead, S. (2006). Phytoestrogens and breast cancer--promoters or protectors? *Endocr. Relat. Cancer* 13: 995-1015. <https://doi:10.1677/erc.1.01159>.
40. Gonzalez, F. J. (1988). The molecular biology of cytochrome P450s. *Pharmacol. Rev.* 40: 243-288. PMID: 3072575
41. Hodek, P., Trefil, P., & Stiborová, M. (2002). Flavonoids-potent and versatile biologically active compounds interacting with cytochromes P450. *Chem. Biol. Interact.* 139: 1-21. [https://DOI:10.1016/s0009-2797\(01\)00285-x](https://DOI:10.1016/s0009-2797(01)00285-x)
42. Ciolino, H. P., Daschner, P. J., & Yeh, G. C. (1999). Dietary flavonols quercetin and kaempferol are ligands of the aryl hydrocarbon receptor that affect CYP1A1 transcription differentially. *Biochem. J.* 340: 715-722. PMID: 10359656. PMCID: PMC1220303
43. Arroo, R. R., Androutsopoulos, V., Beresford, K., Ruparelia, K., Surichan, S. *et al.* (2009). Phytoestrogens as natural prodrugs in cancer prevention: Dietary flavonoids. *Phytochem. Rev.* 8: 375-386.
44. Tan, K. W., Li, Y., Paxton, J. W., Birch, N. P., & Scheepens, A. (2013). Identification of novel dietary phytochemicals inhibiting the efflux transporter breast cancer resistance protein (BCRP/ABCG2). *Food Chem.* 138: 2267-74. <https://doi:10.1016/j.foodchem.2012.12.021>
45. Nobili, S., Lapucci, A., Landini, I., Coronello, M., Roviello, G., & Mini, E. (2020). Role of ATP-binding cassette transporters in cancer initiation and progression. *Semin Cancer Biol.* 60: 72-95. doi: <https://doi.org/10.1016/j.semcancer.2019.08.006>
46. Zhang, S., Yang, X., & Morris, M. E (2004). Flavonoids are inhibitors of breast cancer resistance protein (ABCG2)-mediated transport. *Molecular pharmacology* 65(5): 1208-1216. <https://doi.org/10.1124/mol.65.5.1208>
47. Morris, M. E., & Zhang, S. (2006). Flavonoid-drug interactions: effects of flavonoids on ABC transporters. *Life Sci.* 78: 2116-30. <https://DOI:10.1016/j.lfs.2005.12.003>

48. Huang, C., Chen, X., Guo, B., Huang, W., Shen, T., Sun, X., ... & Zhou, Q. (2012). Induction of Apoptosis by Icariside II through Extrinsic and Intrinsic Signaling Pathways in Human Breast Cancer MCF7 Cells. *Biosci. Biotechnol. Biochem.* 76: 1322–1328. <https://DOI: 10.1271/bbb.120077>
49. Kamsteeg, M., Rutherford, T., Sapi, E., Hanczaruk, B., Shahabi, S., Flick, M. *et al.* (2003). Phenoxodiol - An isoflavone analog - Induces apoptosis in chemoresistant ovarian cancer cells. *Oncogene* 22; 2611–2620. <https://DOI: 10.1038/sj.onc.1206422>
50. Brusselmans, K., Vrolix, R., Verhoeven, G., & Swinnen, J. V. (2005). Induction of cancer cell apoptosis by flavonoids is associated with their ability to inhibit fatty acid synthase activity. *Journal of Biological Chemistry*, 280(7), 5636-5645. <https://doi 10.1074/jbc.M408177200>.
51. To, K. K., & Cho, W. (2021). Flavonoids Overcome Drug Resistance to Cancer Chemotherapy by Epigenetically Modulating Multiple Mechanisms. *Current Cancer Drug Targets* 21(4): 289-305(17). <https://doi: 10.2174/1568009621666210203111220>.
52. Costea, T., Vlad, O. C., Miclea, L. C., Ganea, C., Szöllősi, J., & Mocanu, M. M. (2020). Alleviation of Multidrug Resistance by Flavonoid and Non-Flavonoid Compounds in Breast, Lung, Colorectal and Prostate Cancer. *International Journal of Molecular Sciences* 21(2): 401. <https://doi.org/10.3390/ijms21020401>
53. Rao, P. S., Satelli, A., Moridani, M., Jenkins, M., & Rao, U. S. (2012). Luteolin induces apoptosis in multidrug resistant cancer cells without affecting the drug transporter function: Involvement of cell line-specific apoptotic mechanisms. *Int. J. Cancer* 130: 2703-2714. <https://doi: 10.1002/ijc.26308>
54. Melnik, L. I., Collins-Burow, B. M., Pace, D. K., McLachlan, J. A., Burow, M. E., Frigo, D. E. *et al.* (2002). Flavonoid phytochemicals regulate activator protein-1 signal transduction pathways in endometrial and kidney stable cell lines. *J Nutr.* 132: 1848-185. <https://DOI: 10.1093/jn/132.7.1848>
55. Choi, J. A., Kim, J.Y., Lee, J.Y., Kang, C.M., Kwon, H.J., Yoo, Y.D., Kim, T.W., Lee, Y.S., Lee, S. J. (2001). Induction of cell cycle arrest and apoptosis in human breast cancer cells by quercetin. *Int J Oncol.*19(4):837-44. <https://doi :10.3892/ijo.19.4.837>. PMID: 11562764.
56. Manthey, J. A., & Guthrie, N. (2002). Antiproliferative activities of citrus flavonoids against six human cancer cell lines. *J Agril Food Chem.* 50: 5837-5843. <https://DOI: 10.1021/jf020121d>
57. Yang, K., Lamprecht, S. A., Liu, Y., Shinozaki, H., Fan, K., Leung, D. *et al.* (2000). Chemoprevention studies of the flavonoids: quercetin and rutin in normal and azoxymethane-treated mouse colon. *Carcinogenesis* 21: 1655-1660. <https://doi: 10.1093/carcin/21.9.1655>.
58. Doyle, L. A., Yang, W., Abruzzo, L. V., Krogmann, T., Gao, Y., Rishi, A. K., & Ross, D. D. (1998). A multidrug resistance transporter from human MCF-7 breast cancer cells. *Proc Natl Acad Sci USA* 95:15665-15670. <https://DOI: 10.1073/pnas.95.26.15665>
59. Havsteen, B. H. (2002). The biochemistry and medical significance of the flavonoids. *Pharmacol Ther.* 96: 67202. [https://DOI: 10.1016/s0163-7258\(02\)00298-x](https://DOI: 10.1016/s0163-7258(02)00298-x)
60. Senderowicz, A. M., Headlee, D., Stinson, S. F., Lush, R. M., Kalil, N., Villalba, L. *et al.* (1998). Phase I trial of continuous infusion flavopiridol, a novel cyclin-dependent kinase

- inhibitor, in patients with refractory neoplasms. *J Clin Oncol.* 16: 2986-2999. [https://doi: 10.1200/JCO.1998.16.9.2986](https://doi.org/10.1200/JCO.1998.16.9.2986)
61. Hung, C. H., Chan, S. H., Chu, P. M., & Tsai, K. L. (2015). Quercetin is a potent anti-atherosclerotic compound by activation of SIRT1 signaling under oxLDL stimulation, *Mol. Nutr. Food Res.* 59: 1905-1917. [https://DOI: 10.1002/mnfr.201500144](https://doi.org/10.1002/mnfr.201500144)
 62. Zheng, W., Dai, Q., Custer, L. J., Shu, X. O., Wen, W. Q., Jin, F., & Franke, A. A. (1999). Urinary excretion of isoflavonoids and the risk of breast cancer. *Cancer Epidemiol Biomarkers Prev.* 8(1) :35-40.
 63. Horn-Ross, P. L., John, E. M., Lee, M., Stewart, S. L., Koo, J., Sakoda, L. C. *et al.* (2001). Phytoestrogen consumption and breast cancer risk in a multiethnic population: the Bay Area Breast Cancer Study. *Am J Epidemiol.* 154(5): 434-441. [https://doi:10.1093/aje/154.5.434](https://doi.org/10.1093/aje/154.5.434)
 64. Silva, I. D. S., Mangtani, P., McCormack, V., Bhakta, D., McMichael, A. J., & Sevak, L. (2004). Phyto-oestrogen intake and breast cancer risk in South Asian women in England: findings from a population-based case-control study. *Cancer causes & control: CCC* 15(8): 805–818. [https://doi: 10.1023/B:CACO.0000043431.85706.d8](https://doi.org/10.1023/B:CACO.0000043431.85706.d8).
 65. Bosetti, C., Spertini, L., Parpinel, M., Gnagnarella, P., Lagiou, P., Negri, E., ... & La Vecchia, C. (2005). Flavonoids and breast cancer risk in Italy. *Cancer Epidemiology Biomarkers & Prevention*, 14(4), 805-808. [https://doi:10.1158/1055-9965.EPI-04-0838](https://doi.org/10.1158/1055-9965.EPI-04-0838).
 66. Fink, B. N., Steck, S. E., Wolff, M. S., Britton, J. A., Kabat, G. C., Schroeder, J. C. *et al.* (2007). Dietary flavonoid intake and breast cancer risk among women on Long Island. *American journal of epidemiology* 165(5): 514-523. <https://doi.org/10.1093/aje/kwk033>
 67. Cotterchio, M., Boucher, B. A., Kreiger, N., Mills, C. A., & Thompson, L. U. (2008). Dietary phytoestrogen intake--lignans and isoflavones--and breast cancer risk (Canada). *Cancer causes & control: CCC* 19(3): 259-272. <https://doi.org/10.1007/s10552-007-9089-2>
 68. Cutler, G. J., Nettleton, J. A., Ross, J. A., Harnack, L. J., Jacobs Jr, D. R., Scrafford, C. G. *et al.* (2008). Dietary flavonoid intake and risk of cancer in postmenopausal women: the Iowa Women's Health Study. *International journal of cancer* 123(3): 664-671. <https://doi.org/10.1002/ijc.23564>
 69. Torres-Sanchez, L., Lopez--Carrillo, L., Lo--Cervantes, M., Rueda-Neria, C., & Wolff, M. S. (2000). Food sources of phytoestrogens and breast cancer risk in Mexican women. *Nutrition and cancer* 37(2): 134-139. <https://doi.org/10.1207/S15327914NC372-3>
 70. Ha, T. C., Lyons-Wall, P. M., Moore, D. E., Tattam, B. N., Boyages, J. *et al.* (2006). Phytoestrogens and indicators of breast cancer prognosis. *Nutrition and cancer* 56(1): 3-10. [https://DOI: 10.1207/s15327914nc5601_2](https://doi.org/10.1207/s15327914nc5601_2)
 71. Pantavos, A., Ruiter, R., Feskens, E. F., de Keyser, C. E., Hofman, A., Stricker, B. H. *et al.* (2015). Total dietary antioxidant capacity, individual antioxidant intake and breast cancer risk: the Rotterdam Study. *International journal of cancer* 136(9): 2178-2186. [https://DOI: 10.1002/ijc.29249](https://doi.org/10.1002/ijc.29249)
 72. Wang, Y., Gapstur, S. M., Gaudet, M. M., Peterson, J. J., Dwyer, J. T., & McCullough, M. L. (2014). Evidence for an Association of Dietary Flavonoid Intake with Breast Cancer Risk by Estrogen Receptor Status Is Limited. *J. Nutr.* 144: 1603-1611. [https://doi: 10.3945/jn.114.196964](https://doi.org/10.3945/jn.114.196964)

73. Feng, X. L., Ho, S. C., Mo, X. F., Lin, F. Y., Zhang, N. Q., Luo, H. *et al.* (2020). Association between flavonoids, flavonoid subclasses intake and breast cancer risk: a case-control study in China. *Eur J Cancer Prev.* 29(6): 493-500. <https://DOI:10.1097/CEJ.0000000000000561>
74. Miksicek, R. J. (1993). Commonly occurring plant flavonoids have estrogenic activity. *The American Society of Pharmacology. Molecular Pharmacology* 44(1): 37-43. PMID: 8341277
75. So, F. V., Guthrie, N., Chambers, A. F., Moussa, M., & Carroll, K. K. (1996). Inhibition of human breast cancer cell proliferation and delay of mammary tumorigenesis by flavonoids and citrus juices. *Nutrition and cancer* 26(2): 167-181. <https://doi:10.1080/01635589609514473>.
76. Zand, R. S. R., Jenkins, D. J., & Diamandis, E. P. (2000). Steroid hormone activity of flavonoids and related compounds. *Breast cancer research and treatment* 62(1): 35-49. <https://doi.org/10.1023/a:1006422302173>
77. Imai, Y., Tsukahara, S., Asada, S., & Sugimoto, Y. (2004). Phytoestrogens/flavonoids reverse breast cancer resistance protein/ABCG2-mediated multidrug resistance. *Cancer research* 64(12): 4346-4352. <https://doi.org/10.1158/0008-5472.CAN-04-0078>
78. Katayama, K., Masuyama, K., Yoshioka, S., Hasegawa, H., Mitsuhashi, J., & Sugimoto, Y. (2007). Flavonoids inhibit breast cancer resistance protein-mediated drug resistance: transporter specificity and structure-activity relationship. *Cancer chemotherapy and pharmacology* 60(6): 789-797. <https://doi.org/10.1007/s00280-007-0426-7>
79. Chen D, Landis-Piowar KR, Chen MS, Dou QP (2007). Inhibition of proteasome activity by the dietary flavonoid apigenin is associated with growth inhibition in cultured breast cancer cells and xenografts. *Breast cancer research: BCR* 9(6): R80. <https://doi.org/10.1186/bcr1797>
80. Yang, P. M., Tseng, H. H., Peng, C. W., Chen, W. S., & Chiu, S. J. (2012). Dietary flavonoid fisetin targets caspase-3-deficient human breast cancer MCF-7 cells by induction of caspase-7-associated apoptosis and inhibition of autophagy. *International journal of oncology* 40(2): 469-478. <https://doi.org/10.3892/ijo.2011.1203>
81. Shan, Y., Cheng, Y., Zhang, Y., Guan, F. Q., Sun, H. *et al.* (2013). Triticuside A, a dietary flavonoid, inhibits proliferation of human breast cancer cells via inducing apoptosis. *Nutrition and cancer* 65(6): 891-899. <https://doi.org/10.1080/01635581.2013.802001>
82. Pradhan, D., Pradhan, R. K., Tripathy, G., & Pradhan, S. (2015). Inhibition of proteasome activity by the dietary flavonoid Quercetin associated with growth inhibition in cultured breast cancer cells and xenografts. *Journal of Young Pharmacists* 7(3). <https://doi:10.5530/jyp.2015.3.13>
83. Vrhovac Madunić, I., Madunić, J., Antunović, M., Paradžik, M., Garaj-Vrhovac, V., Brejčak, D. *et al.* (2018) Apigenin, a dietary flavonoid, induces apoptosis, DNA damage, and oxidative stress in human breast cancer MCF-7 and MDA MB-231 cells. *Naunyn-Schmiedeberg's archives of pharmacology* 391(5): 537-550. <https://doi.org/10.1007/s00210-018-1486-4>
84. Santes-Palacios, R., Marroquín-Pérez, A. L., Hernández-Ojeda, S. L., Camacho-Carranza, R., Govezensky, T., & Espinosa-Aguirre, J. (2020). Human CYP1A1 inhibition

by flavonoids. Toxicology in vitro: an international journal published in association with
BIBRA. Toxicology in Vitro 62: 104681. <https://doi.org/10.1016/j.tiv.2019.104681>