

# THE EFFECT OF DIETARY SODIUM ALGINATE ON THE GROWTH AND SURVIVAL OF GIANT FRESHWATER PRAWN (*Macrobrachium rosenbergii*)

## ABSTRACT

The giant freshwater prawn, *Macrobrachium rosenbergii*, is a valuable aquaculture species known for its rapid growth, adaptability to environmental variations, and high nutritional value. However, challenges such as high mortality rates, algal blooms, dissolved oxygen depletion, and pathogen outbreaks constrain its cultivation. The overuse of antibiotics in aquaculture raises significant environmental and health concerns, including antimicrobial resistance. This study explores the potential of sodium alginate, a natural polysaccharide derived from brown algae, as a prebiotic and immunostimulant for improving growth and feed efficiency in *M. rosenbergii*. Juvenile prawns (~0.2 g) were cultured for 60 days and fed experimental diets supplemented with sodium alginate at concentrations of 0 g/kg (control, T0), 1.0 g/kg (T1), 2.0 g/kg (T2), and 3.0 g/kg (T3). Growth performance, feed utilization, and survival rates were assessed bi-weekly. Results showed significant improvements ( $p < 0.05$ ) in growth parameters, including mean weight gain (MWG) and specific growth rate (SGR), in prawns fed a 2.0 g/kg sodium alginate diet (T2). At the end of the experiment, T2 recorded the highest mean weight, MWG, SGR, and lowest feed conversion ratio (FCR,  $2.395 \pm 0.068$ ), demonstrating superior feed utilization efficiency. In contrast, prawns fed a 3.0 g/kg sodium alginate diet (T3) showed reduced growth performance and feed efficiency. This study highlights the efficacy of sodium alginate as a dietary supplement to enhance growth, feed efficiency, and sustainability in freshwater prawn aquaculture, with the optimal dosage identified as 2.0 g/kg. These findings support sodium alginate's potential as an eco-friendly alternative to antibiotics in aquaculture systems.

**Keywords:** *Macrobrachium rosenbergii*, sodium alginate, prebiotic, immunostimulant, aquaculture sustainability

## 1.0 INTRODUCTION:

The giant freshwater prawn, *Macrobrachium rosenbergii*, is a candidate culturable freshwater prawn species with a high growth rate, euryhaline, wide temperature tolerance (15-35 °C), and larger size. In India, freshwater prawns are mostly cultivated in mono- and polyculture systems in either naturally occurring or controlled bodies of water (Mukhopadhyay et al., 2003). The strong demand for this prawn is due to its high nutritional value, including its high levels of protein, polyunsaturated fatty acids, and low-fat levels (D'Abramo & Sheen, 1994). But, apart from these prawns, the major constraint in prawn farming is the high rate of mortality due to higher rate of stocking, algal bloom production, DO depletion, man-made food polluted water, and pathogen diseases (Farook et al., 2019).

The use of antibiotics in aquaculture, especially during larval and growth stages, has raised concerns about environmental and human health impacts, such as bacterial resistance, persistent aquatic diseases, and altered sediment biogeochemistry (Bermúdez-Almada & Espinosa-Plascencia, 2012; Ma et al., 2006). Excessive antibiotic use promotes antimicrobial-resistant bacteria (AMRB) and fosters antibiotic resistance mechanisms (Pruden et al., 2013). To address these challenges, environmentally friendly alternatives like probiotics, immunostimulants, antimicrobial peptides, and quorum sensing interference have been developed (Sivakamavalli et al., 2021). Probiotics and prebiotics have gained attention as more cost-effective solutions compared to antibiotics or vaccines, with prebiotics offering the advantage of being natural feed ingredients that promote beneficial gut bacteria and enhance health (Gatesoupe, 2005; Yousefian & Amiri, 2009). Additionally, medicinal plants, such as marine algae, show promise as natural prebiotics due to their bioactive, antibacterial

properties that are safe for the environment and beneficial in aquaculture (Alghazeer et al., 2013; Dashtiannasab et al., 2016). Algal extracts can be delivered to prawns through injection, immersion, or feed supplementation (Huang et al., 2006; Yeh et al., 2006).

Sodium alginate, a natural hydrophilic polysaccharide derived from marine brown algae, is composed of 1,4-d-mannuronic (M) and l-guluronic (G) acids and serves as a structural component in algal cell walls, containing 30–60% alginic acid (Maiti & Kumari, 2016; Santos, 2017). Widely used in aquaculture as a feed additive, it functions as a feed binder to improve pellet stability, a dietary fiber, and a prebiotic promoting gut health in herbivorous and omnivorous fish species. Sodium alginate enhances the intestinal absorption of minerals like calcium and magnesium (Mohapatra et al., 2011) and is recognized as GRAS (generally regarded as safe) by the FDA (Stone et al., 2009). Known for its biocompatibility, nontoxicity, and non-immunogenicity, it also possesses antibacterial, antiviral, antimicrobial, antifungal, antioxidant, immunomodulatory, and anti-inflammatory properties (Sachan et al., 2015; Ngo & Kim, 2013; Guo et al., 2020). Additionally, alginic acid exhibits hypocholesterolemic and antihypertensive effects. Its potential for immunostimulatory and resistance studies, such as in *Macrobrachium rosenbergii* against *Aeromonas hydrophila*, highlights its versatile applications in aquaculture.

## **2.0 MATERIAL & METHODS:**

### **2.1 Sodium Alginate:**

Sodium alginate, which was purchased from Isochem Laboratories in Kochi, Kerala.

### **2.2 Experimental Diet:**

The commercial diet was cooked, sterilized, and powdered. Then, powdered Sodium alginate was added to diets in four doses, including control (T0), 1.00 g/kg (T1), 2.00 g/kg (T2), and 3.00 g/kg (T3). After that, the dough was thoroughly mixed to distribute evenly all the ingredients. Then prepare pellets, and spread them into trays for sun drying. After drying, pellets were packed into airtight plastic containers, labeled, and stored properly.

### **2.3 Experimental Animal & Husbandry Trial**

The experiment was conducted at the Hands-on Training Center, Department of Aquaculture, situated within the College of Fisheries Science, Kamdhenu University, Veraval. *Macrobrachium rosenbergii* juveniles (~0.2 g) were obtained from Dev Fisheries in Valsad, 695 km from the experimental site. Packaged in 15-liter plastic bags with treated freshwater and pure oxygen, sealed bags (1,000 juveniles per bag) were transported in thermocol boxes to minimize stress during transit. Upon arrival, the juveniles underwent prophylactic treatment in a 0.05% potassium permanganate solution following the normal bath method (Cruz-Lacierda et al., 2000) and were transferred to a 500-liter tank for acclimatization. During the 10-day acclimation period, they were fed *Artemia* (live feed) and commercial feed, with continuous aeration to maintain optimal oxygen levels before being moved to experimental tanks.

The experiment was conducted in 16 plastic tanks (2 x 1 x 1 feet, 40 liters), disinfected with 2 ppm  $\text{KMnO}_4$  and rinsed thoroughly. The tanks were divided into four treatment groups with four replicates each. A total of 240 *Macrobrachium rosenbergii* juveniles (~0.2 g initial weight) were acclimatized and randomly distributed (15 prawns per tank) following a completely randomized design (CRD). The prawns were cultured for 60 days, and fed experimental diets twice daily—initially at 10% of body weight for the first month, then reduced to 5% thereafter. Feedings occurred from 9:00–9:30 a.m. and 6:30–7:00 p.m. Weight was monitored biweekly, and feeding rations were adjusted accordingly.

### **2.4 Growth Parameter Analysis**

- **Weight Measurement:** Prawn juveniles' weight was recorded bi-weekly using an electronic balance with a glass beaker of water to reduce stress.
- **Mean Weight (MW):** Average weight per prawn in each tank calculated at bi-weekly intervals.
- **Mean Weight Gain (MWG):** Calculated using the formula:

$$\text{MWG (g)} = \text{Final weight (g)} - \text{Initial weight (g)}$$

- **Specific Growth Rate (SGR):** Calculated using the formula by Manivannan and Saravanan (2012):

$$\text{SGR} = \frac{(\text{Average Final weight} - \text{Average Initial weight}) \times 100}{\text{Number of days}}$$

## 2.5 Feed Utilization Parameter Analysis

- **Feed Conversion Ratio (FCR):** Measures feed efficiency, calculated as:

$$\text{FCR} = \frac{\text{Feed intake}}{\text{Weight gain}}$$

- **Protein Efficiency Ratio (PER):** Assesses protein utilization efficiency, calculated as (on a dry matter basis):

$$\text{PER} = \frac{\text{Wet weight gain}}{\text{Protein intake}}$$

## 2.6 Survival Rate Analysis

- **Survival Monitoring:** Daily inspection of tanks for live and deceased prawns; dead prawns removed and counted.
- **Survival Rate Formula:**

$$\text{Survival rate (\%)} = \frac{\text{No. of prawn stocked}}{\text{No. of prawns survived after rearing}} \times 100$$

## 2.7 Statistical Analysis

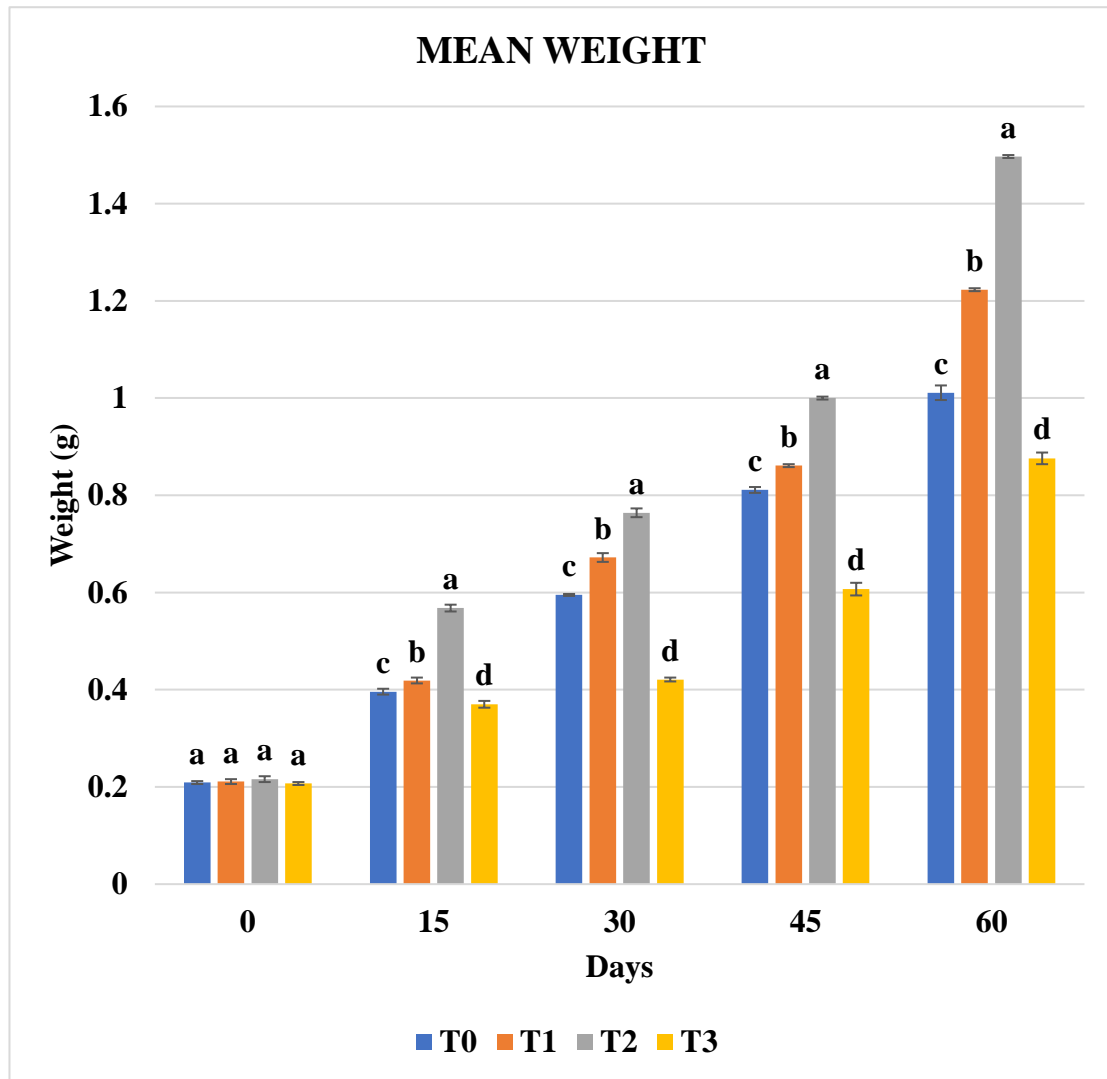
After the experiment, data was evaluated using conventional statistical techniques by Snedecor and Cochran (2014). The analysis was carried out using the SPSS program. To assess "Mean  $\pm$  Standard Error (SE)," one-way analysis of variance (ANOVA) was utilized; to test for significant differences across treatments, the Duncan multiple range test (DMRT) was employed.

## 3.0 Results

### 3.1 Growth parameters analysis:

#### 3.1.1 Mean weight analysis:

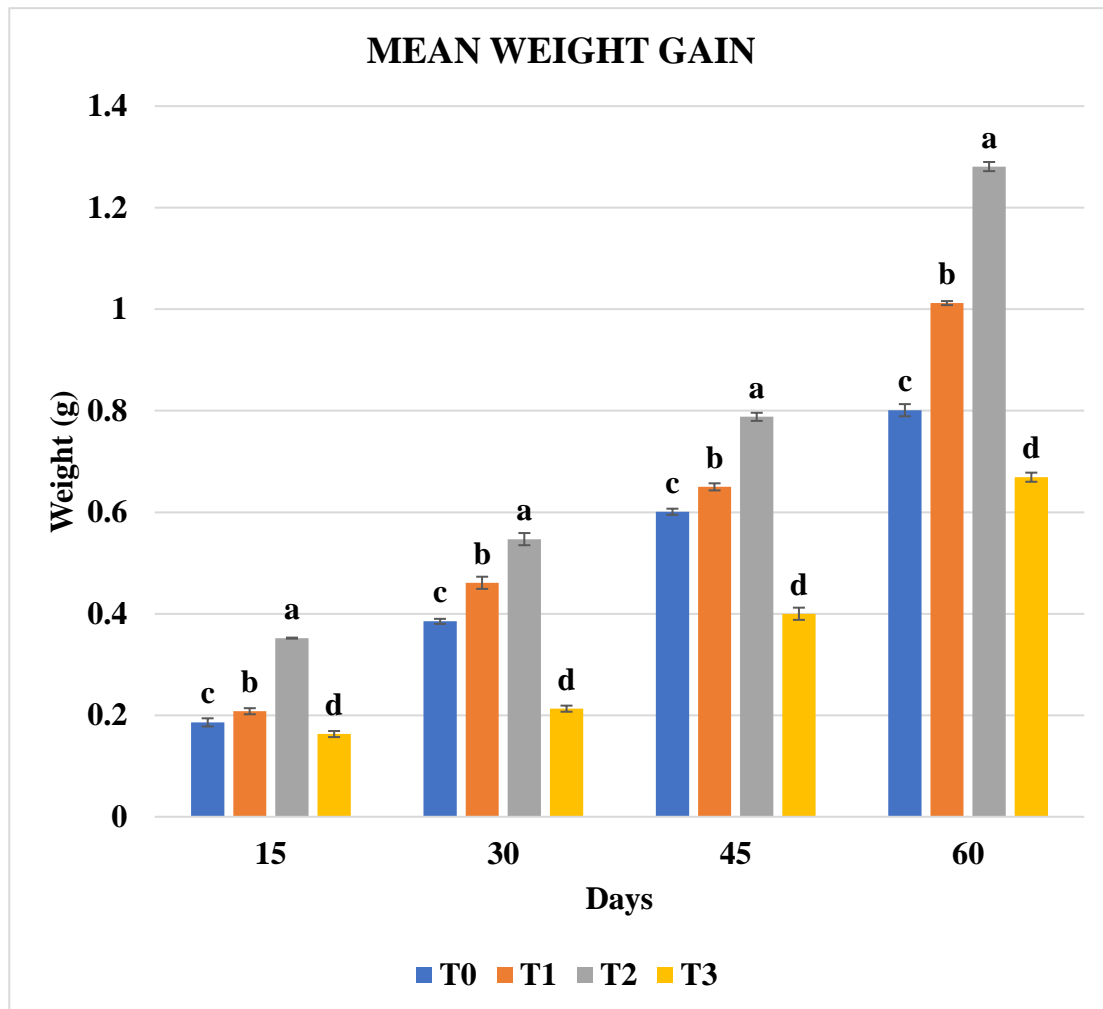
During the 60-day trial, *Macrobrachium rosenbergii* juveniles from each tank were sampled bi-weekly to monitor growth. The initial mean weights of treatments T0, T1, T2, and T3 were  $0.209 \pm 0.003\text{g}$ ,  $0.211 \pm 0.055\text{g}$ ,  $0.216 \pm 0.006\text{g}$ , and  $0.207 \pm 0.003\text{g}$ , respectively, showing no significant differences initially. However, as the experiment progressed, mean weights increased significantly ( $p < 0.05$ ) across treatments, with T2 consistently showing the highest growth, followed by T1, T0, and T3. By day 30, prawns fed a 2.0 g/kg sodium alginate diet (T2) exhibited significantly higher growth. At day 45, T2 maintained superior growth, while T3 showed the lowest growth. At the end of the experiment, the mean weights were  $1.011 \pm 0.015\text{g}$  (T0),  $1.223 \pm 0.003\text{g}$  (T1),  $1.497 \pm 0.003\text{g}$  (T2), and  $0.876 \pm 0.012\text{g}$  (T3), confirming that T2 produced the highest growth ( $p < 0.05$ ), whereas T3 resulted in the lowest.



**Figure 1: Impact of sodium alginate diet on *M. rosenbergii* mean weight (MW) after the experiment**

### 3.1.2 Mean Weight Gain (MWG) Analysis:

The fortnightly growth performance of *Macrobrachium rosenbergii* juveniles was assessed through mean weight gain, presented in Figure 2. On day 15, the mean weight gains for T0, T1, T2, and T3 were  $0.186 \pm 0.008\text{g}$ ,  $0.208 \pm 0.006\text{g}$ ,  $0.352 \pm 0.001\text{g}$ , and  $0.163 \pm 0.006\text{g}$ , respectively, with T2 showing the highest gain ( $p < 0.05$ ). By day 30, T2 continued to outperform with  $0.547 \pm 0.012\text{g}$ , followed by T1 ( $0.461 \pm 0.012\text{g}$ ), T0 ( $0.385 \pm 0.005\text{g}$ ), and T3 ( $0.213 \pm 0.006\text{g}$ ). On day 45, T2 maintained the highest gain ( $0.788 \pm 0.008\text{g}$ ), while T0, T1, and T3 showed lower gains. By day 60, mean weight gains were  $0.801 \pm 0.012\text{g}$  (T0),  $1.012 \pm 0.004\text{g}$  (T1),  $1.281 \pm 0.009\text{g}$  (T2), and  $0.669 \pm 0.009\text{g}$  (T3). The results confirm that T2 had significantly higher weight gain throughout the culture period.



**Figure 2: Impact of sodium alginate diet on *M. rosenbergii* mean weight gain (MWG) after the experiment**

### 3.1.3 Specific Growth Rate Analysis:

The specific growth rate (SGR) of *M. rosenbergii* juveniles at the end of the study is shown in Figure 3. The lowest SGR ( $2.399 \pm 0.009$ ) was observed in T3, where prawns were fed a 3.0 g/kg sodium alginate diet. The highest SGR ( $3.224 \pm 0.049$ ) was recorded in T2, fed a 2.0 g/kg sodium alginate diet, followed by T1 (1.0 g/kg sodium alginate) and T0 (fish meal-based diet). All treatments showed significant differences ( $p < 0.05$ ), with T2 demonstrating the most favorable growth performance.

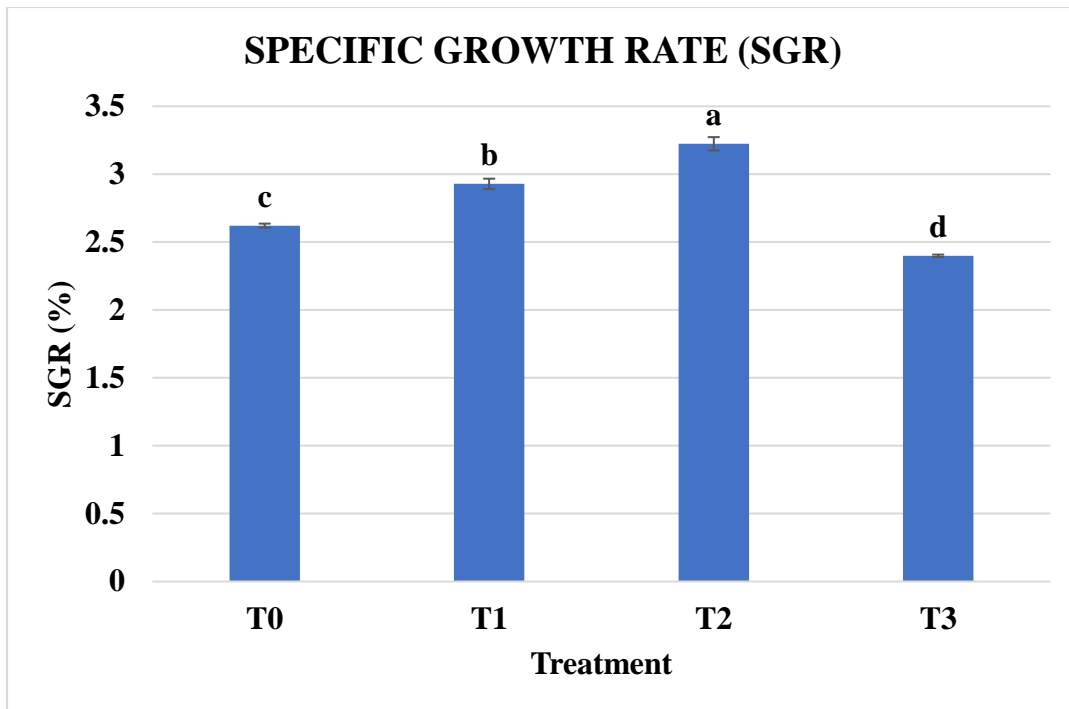


Figure 3: Impact of sodium alginate diet on *M. rosenbergii* specific growth rate (SGR) after the experiment

### 3.2 Feed Utilization Parameter Analysis:

#### 3.2.1 Feed Conversion Ratio (FCR) Analysis:

At the end of the 60-day study, the feed conversion ratio (FCR) of *M. rosenbergii* juveniles was calculated and presented in Figure 4. The FCR values for treatments T0, T1, T2, and T3 were  $2.878 \pm 0.081$ ,  $2.612 \pm 0.204$ ,  $2.395 \pm 0.068$ , and  $3.106 \pm 0.089$ , respectively. The lowest FCR was observed in T2 (2.0 g/kg sodium alginate diet), indicating the most efficient feed utilization, while the highest FCR was in T3 (3.0 g/kg sodium alginate diet). Significant differences ( $p < 0.05$ ) were observed among all treatments.

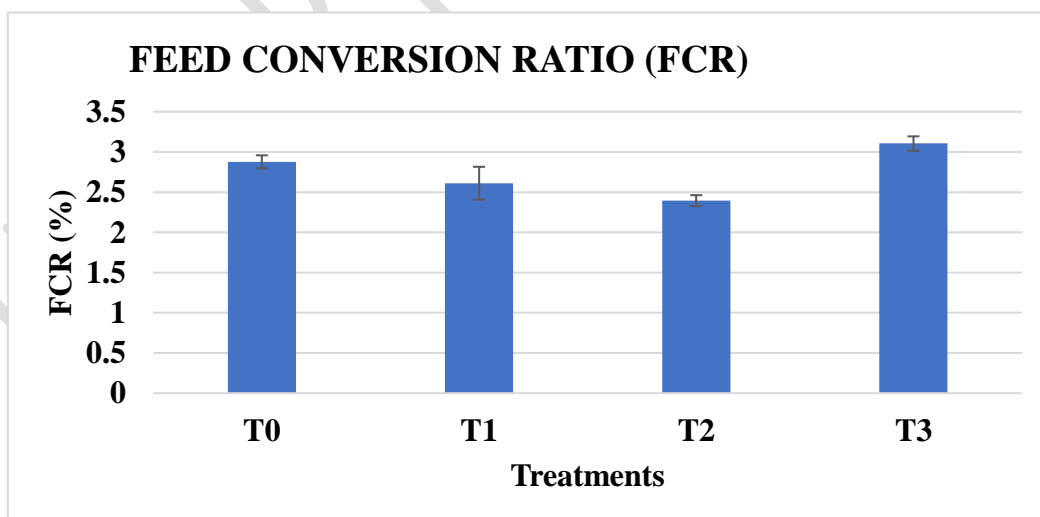


Figure 4: Impact of sodium alginate diet on *M. rosenbergii* feed conversion ratio (FCR) after the experiment

### 3.2.2 Protein Efficiency Ratio (PER) Analysis:

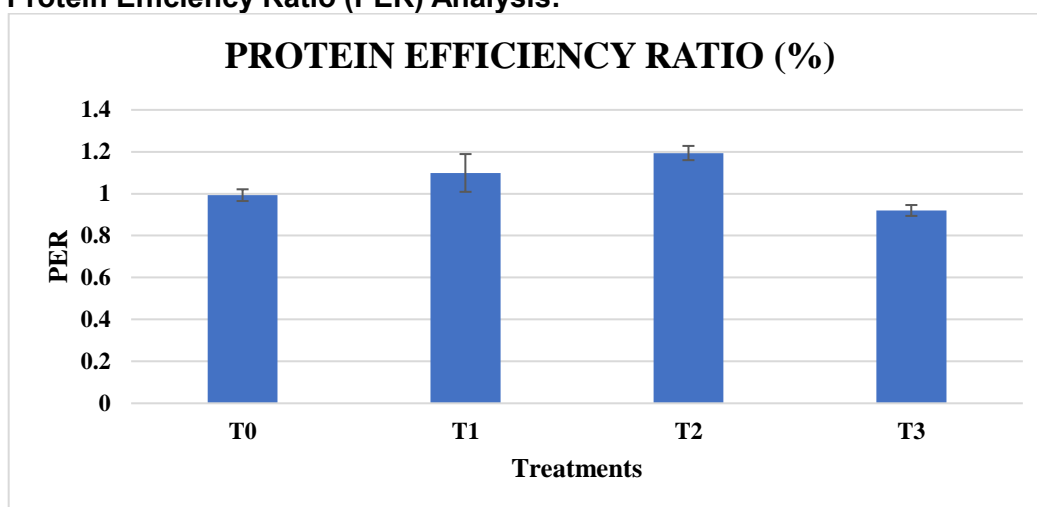
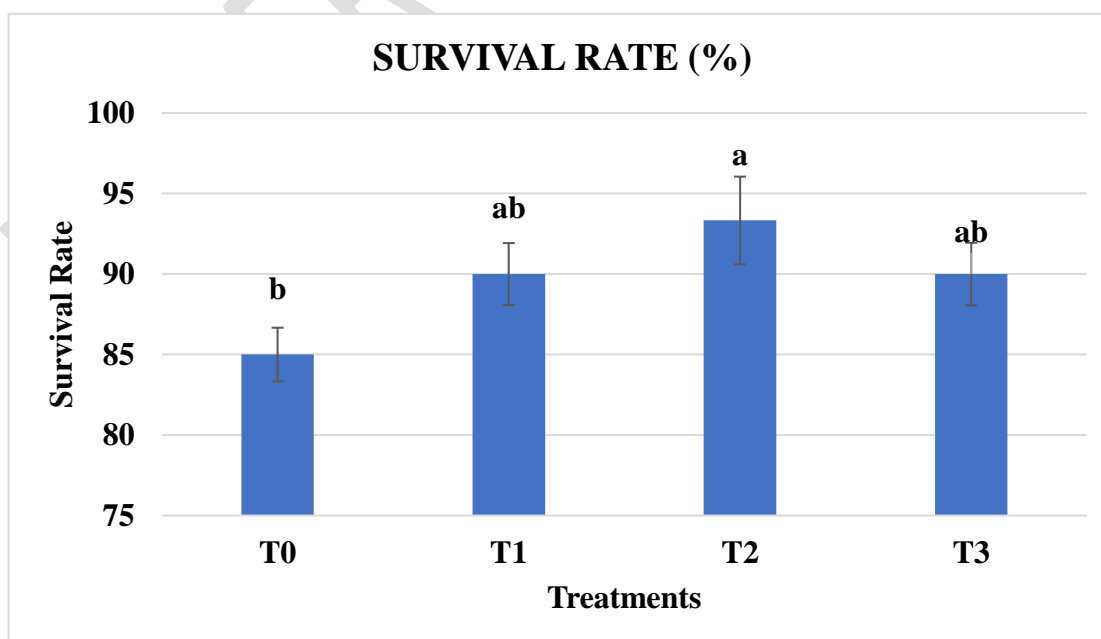


Figure 5: Impact of sodium alginate diet on *M. rosenbergii* protein efficiency ratio (PER) after the experiment

The protein efficiency ratio (PER) of *M. rosenbergii* juveniles was evaluated after the experiment and is presented in Figure 5. Prawns were fed a diet containing 35% protein. The highest PER ( $1.194 \pm 0.034$ ) was observed in T0, followed by T1 ( $1.099 \pm 0.090$ ), T2 ( $0.993 \pm 0.028$ ), and T3 ( $0.920 \pm 0.026$ ). Significant differences ( $p < 0.05$ ) were noted among treatments, with T2 (2.0 g/kg sodium alginate diet) showing the most effective protein utilization.

### 3.3 Survival Rate Analysis:

The survival rate of *M. rosenbergii* juveniles was monitored every two weeks and is summarized in Figure 6. At the end of the culture period, the highest survival rate ( $93.33 \pm 2.721\%$ ) was observed in T2, where prawns were fed with a 2.0 g/kg sodium alginate diet, followed by T1 and T3 (both  $90.00 \pm 1.924\%$ ), and T0 ( $85.00 \pm 1.667\%$ ). While no significant differences ( $p < 0.05$ ) were found among treatments, T2 had a notably higher survival rate compared to the others, with no significant difference between T1 and T3.



## Figure 6: Impact of sodium alginate diet on *M. rosenbergii* survival rate after the experiment

### 4.0 Discussion:

The dietary inclusion of prebiotics such as sodium alginate can enhance immunity as well as growth parameters of *M. rosenbergii*. The T2 treatment showed higher growth compared to the other treatments, where prawns were fed with a 2.0 g/kg sodium alginate incorporated diet. As the concentration of the sodium alginate increases, the growth of the *M. rosenbergii* increases proportionally, but the higher growth parameters obtained at 2.0 g/kg of the sodium alginate incorporated diet, and 3.0 g/kg of sodium alginate incorporated diet showed less growth than the other all treatments. The reason behind these results may be due to the theory given by Bagheri et al. (2023) that the prawn's microbiota and digestive system were most likely not enhanced by altering the weight. A similar trend was found by Bagheri et al. (2023), where the *L. vannamei* fed with a 2.0 g/kg sodium alginate diet showed significantly higher growth parameters compared to other treatments. Previous studies on sodium alginate-based diets also showed significantly higher SGR of *Oreochromis niloticus* (Van Doan et al., 2016), Atlantic cod (Vollstad et al., 2006), *Penaeus monodon* (Chung et al., 2011), *Epinephelus coioides* (Yeh et al., 2008), Sea cucumber (Xia et al., 2012), and *Procambarus clarkii* (Mona et al., 2015). The reasons for these results could be the prebiotic effects differ according to fish species, water temperature, feeding duration, prebiotic dosages, and solubility (Van Doan et al., 2016). A similar trend was also found by Santos et al. (2019), where the weight gain of *L. vannamei* was proportional to the concentration of sodium alginate in the shrimp diet. The present study suggests that the presence of beneficial bacteria and digestive enzymes may be responsible for the notable enhancement in the growing performance of *M. rosenbergii*.

The present investigation indicated that the lowest FCR was found in the T2 treatment, which shows higher growth parameters. Bagheri et al. (2023) also found a similar result where a diet containing 2.0 g/kg sodium alginate showed lower FCR than other treatments. In the case of finfish, Vollstad et al. (2006) and Yeh et al. (2008) also found similar results in *Gadus morhua* L. and *Epinephelus coioides*, respectively. Due to its high protein content and abundance of additional growth stimulants, algae, and algae extracts have gained a lot of interest when added to prawn diets. As a result, it can enhance immunity, boost growth and appetite, and strengthen resistance to infections (Schiener et al., 2014; Akbary & Aminikhoei, 2018; Akbary et al., 2021). The study conducted by Akbary et al. (2023) found that the extract of the brown seaweeds positively affects the growth parameters and also PER of the shrimp, *Litopenaeus vannamei*.

At the end of the study, a higher survival rate was found in T2 treatment. The T2 treatment was significantly higher than the other treatments. There were no significant differences between T0 and T1 treatment. Control treatment shows a significantly lower survival rate than all treatments. A previous study by Santos et al. (2019) also found a similar trend of survival rate in shrimp as the concentration of sodium alginate increased. Contradictory, Bagheri et al. (2023) found no significant difference in the survival rate of white leg shrimp. Neamat-Allah et al. (2019) evaluate that The immunological response being triggered by sodium alginate may be the reason for the increased catfish survival rate.

### 5.0 Conclusion

The study highlights the positive impact of sodium alginate as a dietary prebiotic on the growth performance, feed utilization, and survival rate of *Macrobrachium rosenbergii*. Among all treatments, the T2 group, fed with a 2.0 g/kg sodium alginate-incorporated diet, exhibited the best overall results, including higher growth performance, improved feed conversion ratio (FCR), higher protein efficiency ratio (PER), and increased survival rate. The findings align with previous research, suggesting that optimal concentrations of sodium alginate enhance the prawn's microbiota and digestive enzyme activity, resulting in superior growth and immunity. However, a higher concentration (3.0 g/kg) of sodium alginate showed reduced growth, possibly due to a saturation effect on the microbiota and digestive system. The study also corroborates earlier reports that algae and their extracts, due to their protein and growth-

promoting components, significantly enhance immunity, growth, and disease resistance in aquatic species.

This investigation confirms the potential of sodium alginate at an optimal dose as a sustainable and effective feed additive to enhance the aquaculture performance of *M. rosenbergii*. Future research should explore the long-term impacts and underlying mechanisms of sodium alginate and other prebiotics in aquaculture systems.

## 6.0 References

- Akbary, P., & Aminikhoei, Z. (2018). Effect of polysaccharides extracts of algae *Ulva rigida* on growth, antioxidant, immune response and resistance of shrimp, *Litopenaeus vannamei* against *Photobacterium damsela*. *Aquaculture Research*, 49(7), 2503-2510.
- Akbary, P., Ajdari, A., & Ajang, B. (2023). Growth, survival, nutritional value and phytochemical, and antioxidant state of *Litopenaeus vannamei* shrimp fed with premix extract of brown *Sargassum ilicifolium*, *Nizimuddinina zanardini*, *Cystoseira indica*, and *Padina australis* macroalgae. *Aquaculture International*, 31(2), 681-701.
- Akbary, P., Gholamhosseini, A., Ali, M., Aminikhoei, Z., Tavabe, K. R., & Kuchaksaraei, B. S. (2021). Growth yield, fatty acid profile and antioxidant status of *Litopenaeus vannamei* fed *Iyengaria stellata* supplemented diet. *Iranian Journal of Science and Technology, Transactions A: Science*, 45, 111-119.
- Alghazeer, R., Whida, F., Abduelrhman, E., Gammoudi, F., & Naili, M. B. (2013). In vitro antibacterial activity of alkaloid extracts from green, red and brown macroalgae from western coast of Libya. *African Journal of Biotechnology*, 12(51), 7086–7091.
- Bagheri, D., Moradi, R., Zare, M., Sotoudeh, E., Hoseinifar, S. H., Oujifard, A., & Esmaeili, N. (2023). Does dietary sodium alginate with low molecular weight affect growth, antioxidant system, and haemolymph parameters and alleviate cadmium stress in white leg shrimp (*Litopenaeus vannamei*)?. *Animals*, 13(11), 1805.
- Bermúdez-Almada, M. C., & Espinosa-Plascencia, A. (2012). The use of antibiotics in shrimp farming. *Health and environment in aquaculture*, 199-214.
- Chung, M. Y., Liu, C. H., Chen, Y. N., & Cheng, W. (2011). Enhancing the reproductive performance of tiger shrimp, *Penaeus monodon*, by incorporating sodium alginate in the broodstock and larval diets. *Aquaculture*, 312(1-4), 180-184.
- Cruz-Lacierda E R, de la Peña L D, Lumanlan-Mayo S, et al. 2000. Marine leech (*Zeylanicobdella arugamensis*) infestation in cultured orange-spotted grouper, *Epinephelus coioides*. *Aquaculture* 185(3-4): 191-196.
- D'Abramo, L. R., & Sheen, S. (1994). Nutritional requirements, feed formulation, and feeding practices for intensive culture of the freshwater prawn *Macrobrachium rosenbergii*. *Reviews in Fisheries Science & Aquaculture*, 2(1), 1–21.
- Dashtiannasab, A., Mesbah, M., Pyghan, R., & Kakoolaki, S. (2016). The efficacy of the red seaweed (*Laurencia snyderiae*) extract on growth performance, survival and disease resistance in white shrimp. *Iranian Journal of Aquatic Animal Health*, 2(1), 1–10.

- Farook, M. A., Mohamed, H. M., Tariq, N. P. M. M., Shariq, K. M., & Ahmed, I. A. (2019). Giant freshwater prawn, *Macrobrachium rosenbergii* (De Man 1879): A review. *International Journal of Research and Analytical Reviews*, 6(1), 571-584.
- Gatesoupe, J. (2005). Probiotics and prebiotics for fish culture, at the parting of the ways. *Aqua Feeds: Formulation & Beyond*, 2(3), 3-5.
- Guo, X., Wang, Y., Qin, Y., Shen, P., & Peng, Q. (2020). Structures, properties and application of alginic acid: A review. *International Journal of Biological Macromolecules*, 162, 618–628.
- Huang, X., Zhou, H., & Zhang, H. (2006). The effect of *Sargassum fusiforme* polysaccharide extracts on vibriosis resistance and immune activity of the shrimp, *Fenneropenaeus chinensis*. *Fish & shellfish immunology*, 20(5), 750-757.
- Ma, D., Hu, Y., Wu, J., Ye, S., & Ai, L. (2006). Effects of antibacterials use in aquaculture on biogeochemical processes in marine sediment. *Science of the Total Environment*, 367(1), 273–277.
- Maiti, S., & Kumari, L. (2016). Smart nanopolysaccharides for the delivery of bioactives. In *Elsevier eBooks* (pp. 67–94).
- Manivannan A & Saravanan T S. 2012. Impact of formulated protein diets on growth of the Indian major carp, *Labeo rohita* (Hamilton). *Fish Aquacult. J.* 57: 123-132.
- Mohapatra, S., Chakraborty, T., Prusty, A. K., & Das, P. (2011). Effect of dietary sodium alginate on growth, feed utilization and body composition of *Catla catla* (Hamilton) fingerlings. *Aquaculture Nutrition*, 17(4), e832-e839.
- Mona, M. H., Rizk, E. S. T., Salama, W. M., & Younis, M. L. (2015). Efficacy of probiotics, prebiotics, and immunostimulant on growth performance and immunological parameters of *Procambarus clarkii* juveniles. *The Journal of Basic & Applied Zoology*, 69, 17-25.
- Mukhopadhyay, P. K., Rangacharyulu, P. V., Mitra, G., & Jana, B. B. (2003). Applied Nutrition in Freshwater Prawn, *Macrobrachium rosenbergii*, Culture. *Journal of Applied Aquaculture*, 13(3-4), 317-340.
- Neamat-Allah, A. N., El-Murr, A. E. I., & Abd El-Hakim, Y. (2019). Dietary supplementation with low molecular weight sodium alginate improves growth, haematology, immune reactions and resistance against *Aeromonas hydrophila* in *Clarias gariepinus*. *Aquaculture Research*, 50(5), 1547-1556.
- Ngo, D., & Kim, S. (2013). Sulfated polysaccharides as bioactive agents from marine algae. *International Journal of Biological Macromolecules*, 62, 70–75.
- Pruden, A., Larsson, D. J., Amézquita, A., Collignon, P., Brandt, K. K., Graham, D. W., ... & Zhu, Y. G. (2013). Management options for reducing the release of antibiotics and antibiotic resistance genes to the environment. *Environmental health perspectives*, 121(8), 878-885.
- Sachan, N. K., Pushkar, S., Jha, A. K., & Bhattacharya, A. (2015). Sodium alginate: the wonder polymer for controlled drug delivery. *Journal of Pharmacy Research*, 1191–1199.

- Santos, H. M., Tsai, C. Y., Yanuaria, C. A. S., Tayo, L. L., Vo, D. D., Mariatulqabtiah, A. R., & Chuang, K. P. (2019). Effects of sodium alginate-fed Pacific white shrimps, *Litopenaeus vannamei*, on Toll-like receptors and *Vibrio alginolyticus* infection. *Aquaculture Research*, 50(4), 1384-1392.
- Santos, L. a. L. D. (2017). Natural Polymeric Biomaterials: Processing and Properties ☆. In *Elsevier eBooks*. <https://doi.org/10.1016/b978-0-12-803581-8.02253-0>
- Schiener, P., Black, K. D., Stanley, M. S. and Green, D. H. (2014). The seasonal variation in the chemical composition of the kelp species *Laminaria digitata*, *Laminaria hyperborea*, *Saccharina latissima* and *Alaria esculenta*. *Journal of Applied Phycology*, 27,363–373. DOI:10.1007/s10811- 014-0327-1
- Sivakamavalli, J., Park, K., Kwak, I., & Baskaralingam, V. (2021). Bacterial Disease Control Methods in Shrimp (Penaeus, 1798) Farming Sector in Asian Countries. *IntechOpen*. doi: 10.5772/intechopen.93680
- Stone, M. B., Laughren, T., Jones, M., Levenson, M., Holland, P. C., Hughes, A., Hammad, T. A., Temple, R., & Rochester, G. D. (2009). Risk of suicidality in clinical trials of antidepressants in adults: analysis of proprietary data submitted to US Food and Drug Administration. *The BMJ*, 339(11-2), b2880.
- Van Doan, H., Tapingkae, W., Moonmanee, T., & Seepai, A. (2016). Effects of low molecular weight sodium alginate on growth performance, immunity, and disease resistance of tilapia, *Oreochromis niloticus*. *Fish & shellfish immunology*, 55, 186-194.
- Vollstad, D., Bøgwald, J., Gåserød, O., & Dalmo, R. A. (2006). Influence of high-M alginate on the growth and survival of Atlantic cod (*Gadus morhua* L.) and spotted wolffish (*Anarhichas minor* Olafsen) fry. *Fish & Shellfish Immunology*, 20(4), 548-561.
- Xia, S., Yang, H., Li, Y., Liu, S., Zhou, Y., & Zhang, L. (2012). Effects of different seaweed diets on growth, digestibility, and ammonia-nitrogen production of the sea cucumber *Apostichopus japonicus* (Selenka). *Aquaculture*, 338, 304-308.
- Yeh, S. P., Chang, C. A., Chang, C. Y., Liu, C. H., & Cheng, W. (2008). Dietary sodium alginate administration affects fingerling growth and resistance to *Streptococcus* sp. and iridovirus, and juvenile non-specific immune responses of the orange-spotted grouper, *Epinephelus coioides*. *Fish & shellfish immunology*, 25(1-2), 19-27.
- Yeh, S. T., Lee, C. S., & Chen, J. C. (2006). Administration of hot-water extract of brown seaweed *Sargassum duplicatum* via immersion and injection enhances the immune resistance of white shrimp *Litopenaeus vannamei*. *Fish & Shellfish Immunology*, 20(3), 332-345.
- Yousefian, M., & Amiri. (2009). A review of the use of prebiotic in aquaculture for fish and shrimp. *African Journal of Biotechnology*, 8(25).