Eco-Friendly Management of Powdery Mildew Pathogen Infecting Cucumber Using Botanicals and Biostimulants

ABSTRACT:

Powdery mildew, caused by *Podosphaera xanthii*, is a major disease affecting cucumber (*Cucumis sativus L.*) crops, leading to significant yield losses. The study evaluates the efficacy of botanicals and biostimulants in managing powdery mildew under *in vitro* conditions. Treatments were tested at three concentrations (5, 7.5 and 10 %) against *P. xanthii*. Results revealed that the efficacy of treatments increased with concentration, with *Ascophyllum nodosum* (seaweed extract), *Mimosa pudica* (touch-me-not) and *Osmium sanctum* (tulasi) demonstrating the highest inhibition rates at 10 per cent. These findings suggest the potential of eco-friendly alternatives for managing powdery mildew in cucumber.

Keywords: Powdery mildew, cucumber, *Podosphaera xanthii*, botanicals, biostimulants, eco-friendly management.

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1. INTRODUCTION:

Powdery mildew is a destructive fungal disease caused by *Podosphaera xanthii* that affects cucumber and other cucurbits, reducing photosynthetic activity and causing significant yield losses (Perez-Garcia et al., 2009, El-Naggar et al., 2012 and Rur et al., 2018)). When resistant sources are unavailable or new virulent pathogen races emerge, fungicides are an alternative method for managing plant diseases. Evaluating new-generation fungicides in both *in vitro* and *in vivo* conditions help to determine their efficacy for field recommendations (Urban and Lebeda 2006). However, continuous use of the same fungicides can lead to pathogen resistance and negatively impact the ecosystem. Thus, screening plant products (botanicals), bio-stimulants, biosynthesized nanoparticles and bio-agents for antifungal activity is necessary to reduce fungicide use.

Conventional management strategies often rely on synthetic fungicides, raising concerns about environmental impact and resistance development. Eco-friendly alternatives, such as botanicals and biostimulants, have gained attention as sustainable disease management solutions (Marzani et al., 2021). This study evaluates the *in vitro* efficacy of selected botanicals and biostimulants against *P. xanthii*.

2. MATERIALS AND METHODS

2.1 Collection of Pathogen and Preparation of Botanicals

P. xanthii isolates were collected from infected cucumber leaves grown under greenhouse conditions at the Zonal Agricultural Research Station, Bengaluru. Fully expanded, powdery mildew-infected leaves were used for pathogen isolation and subsequent experiments. Botanicals and biostimulants were prepared as aqueous extracts at concentrations of 5, 7.5 and 10 per cent (McGrath, 2001) (Table 1).

Twenty-five grams of leaves of corresponding plant material were rinsed in water and cut into small pieces and macerated using a sterilized pestle and mortar with 25 ml of distilled water. The contents were filtered through a clean double-layered muslin cloth. From this solution 5 ml, 7.5 ml and 10 ml and added to 95 ml, 92.5 and 90 ml of distilled water to get 5 per cent, 7.5 per cent and 10 per cent concentrations respectively. These extracts were centrifuged for five minutes at 3000 rpm to get a clear plant extract. Four drops of plant extract solution were added on to the 2cm cut detached leaves with the help of a dropper/ camel hair brush. In each treatment three replications were maintained. These Petri plates containing slides were lined with moist blotting paper and were incubated at room temperature (28 ± 1°C) for 72 hours under dark condition. The observation on per cent leaf area inhibition of powdery mildew was recorded at 72 hours after incubation under Leica microscope with 10X magnification. The average of three replications was calculated and the per cent leaf area inhibition was calculated with the following formula given by Vincent (1927).

$$I = (C – T /C) \times 100$$

Where, I = Per cent inhibition of spore germination

C = Germination of conidia in control

T = Germination of conidia in treatment

Table 1: List of botanicals and biostimulants used for *in vitro* evaluation against *P. xanthii*

Sl. No.	Treatment code	Treatments

1	T1	*Simarouba glauca* (Simarouba)
2	T2	*Cymbopogon ambiguous* (Lemon grass)
3	T3	*Azadirachta indica* (Neem)
4	T4	*Bougainvillea spectabilis* (Bougainvillea)
5	T5	*Pongamia pinnata* (Pongamia)
6	T6	*Eucalyptus obliqua* (Eucalyptus)
7	T7	*Osmium sanctum* (Tulsi)
8	T8	*Mimosa pudica* (Touch-me-not)
9	T9	*Ascophyllum nodosum* (Seaweed extract)
10	T10	*Sargassum sp.* (Seaweed extract)

2.2 Experimental Design The experiment was conducted in a completely randomized design (CRD) with 10 treatments and three replications and control. Disease severity was assessed using a 0-9 scale (Jenkins and Wehner 1983) and converted to Percentage Disease Index (PDI) proposed by Wheeler (1969) (Table 2). Pathogen inhibition was calculated based on disease severity reduction relative to an untreated control and within treatment.

$$\text{PDI (\%)} = \frac{\text{Sum of individual disease ratings}}{\text{Total no. of leaves observed x Maximum disease grade}} \text{ X 100}$$

Table 2: Disease Severity Scale for Powdery Mildew

Score	% Disease severity (PDI)	Description
0	0	No disease
1	0–3	Few small leaf lesions
2	3–6	Few lesions on few leaves with no stem lesions
3	6–12	Few lesions on few leaves or with superficial stem lesions
4	12–25	Few well-formed leaf lesions or superficial stem lesions
5	25–50	Few well-formed leaf lesions or enlarging stem lesions
6	50–75	Many large leaf lesions or deep stem lesions with abundant sporulation, or plant more than 50 per cent defoliated
7	75–87	Many large coalescing leaf or stem lesions, over 75 per cent of plant area affected or defoliated

8	87–100	Plants largely defoliated, leaf or stem with abundant sporulating lesions
9	100	Plants dead

2.3 Statistical Analysis Data were analyzed using ANOVA, and significant differences were determined using LSD at a 1 per cent significance level. Arc-sine transformations were applied to percent values for normalization.

3. Results and Discussion

3.1 *In Vitro* Evaluation of Botanicals and Biostimulants

3.1.1 Effect of Botanicals and Biostimulants at 5 per cent Concentration

Among the treatments, *Ascophyllum nodosum* (T9) and *Osmium sanctum* (T7) recorded the lowest PDI of 27.06 and 29.63 per cent, with inhibition rates of 51.21 and 54.25 per cent, respectively. *Eucalyptus obliqua* (T6) showed a PDI of 30.82 per cent with an inhibition rate of 52.41 per cent. Moderate efficacy was observed for *Pongamia pinnata* (T5) and *Azadirachta indica* (T3), with inhibition rates of 44.83 and 41.75 per cent (Table 3).

Table 3: *In vitro* efficacy of botanicals/ biostimulants against *P. xanthi* infecting cucumber

Treatment	Botanicals/ Biostimulants	Per cent inhibition over control				Efficacy within treatment			
		5 %	7.5 %	10 %	Mean	5%	7.5%	10%	Mean
T1	*Simarouba glauca*	27.03 (31.39)	29.10 (32.63)	32.03 (34.45)	29.38 (32.08)	11.03 (19.39)	13.89 (21.88)	15.85 (23.45)	13.59 (21.57)
T2	*Cymbopogan ambiguous*	28.82 (32.45)	36.83 (37.35)	38.88 (38.56)	34.84 (36.12)	11.48 (19.80)	16.83 (24.21)	20.99 (27.26)	16.44 (23.76)
T3	*Azadirachta indica* A. Juss	41.75 (40.24)	41.75 (43.65)	47.67 (49.39)	49.03 (44.42)	32.42 (34.69)	35.90 (36.79)	37.67 (37.85)	35.33 (36.44)
T4	*Bougainvillea spectabilis*	35.89 (36.79)	43.11 (41.03)	60.16 (50.84)	46.39 (42.88)	34.89 (36.19)	39.03 (38.65)	49.82 (44.88)	41.25 (39.91)
T5	*Pongamia pinnata L*	44.83 (42.02)	46.44 (42.94)	47.78 (43.71)	46.35 (42.89)	14.17 (22.10)	23.28 (28.84)	27.78 (31.79)	21.74 (27.58)
T6	*Eucalyptus obliqua*	52.41 (46.36)	54.29 (47.44)	55.86 (48.34)	54.18 (47.38)	23.68 (29.10)	35.62 (36.63)	46.52 (42.99)	35.27 (36.24)
T7	*Osmium sanctum*	54.25 (47.42)	55.42 (48.09)	56.89 (48.94)	55.52 (48.15)	34.25 (35.25)	33.33 (35.25)	48.06 (44.35)	38.82 (38.47)
T8	*Mimosa pudica*	48.15 (43.92)	58.21 (49.71)	58.93 (50.12)	55.10 (47.92)	28.15 (32.02)	34.75 (36.11)	38.93 (38.59)	33.94 (35.57)
T9	*Ascophyllum nodusum*	51.21 (45.67)	58.41 (49.82)	63.37 (52.73)	57.66 (49.41)	23.87 (29.24)	37.41 (37.69)	43.79 (41.42)	35.02 (36.11)
T10	*Sargassum sp.*	43.32 (41.14)	54.49 (47.56)	58.24 (49.72)	52.02 (46.14)	21.56 (27.65)	25.39 (30.24)	38.39 (38.27)	28.45 (32.06)
T11	Control	64.76 (53.56)	-	-	-	-	-	-	-

* Arc sin transformed values

	S. Em ±	C. D. at 1%
Botanicals (B)	0.36	1.39
Concentration (C)	0.27	1.08
B x C	0.62	2.41

3.1.2 Effect at 7.5 per cent Concentration

At 7.5 per cent, disease suppression was highest with *Mimosa pudica* (T8) and *Ascophyllum nodosum* (T9), achieving inhibition rates of 58.21 and 58.41 per cent. *Osmium sanctum* (T7) and *Eucalyptus obliqua* (T6) showed high effectiveness, with inhibition rates of 55.42 and 54.29 per cent, respectively. Moderate reductions were noted for *Azadirachta indica* (T3) and *Bougainvillea spectabilis* (T4) (Table 3) (Fig. 1a).

3.1.3 Effect at 10 per cent Concentration

At 10 per cent, *Ascophyllum nodosum* (T9) demonstrated the highest inhibition rate of 63.37 per cent, followed by *Mimosa pudica* (T8) and *Osmium sanctum* (T7) with 58.93 and 56.89 per cent, respectively. *Eucalyptus obliqua* (T6) continued to perform well, with a PDI of 17.40 per cent and an inhibition rate of 55.86 per cent (Table 3). Moderate efficacy was observed for *Pongamia pinnata* (T5) and *Sargassum sp.* (T10). *In vitro* evaluation of botanicals and biostimulants revealed that as the concentration of the extracts increased the effectiveness was also increased. The results are in agreement with several workers Dinesh *et al.* 2015.

3.1.4 Treatment efficacy of botanicals and biostimulants for managing powdery mildew under *in vitro* conditions

The efficacy of different botanicals and biostimulants in inhibiting *P. xanthi* at three concentrations: 5, 7.5 and 10 per cent within treatment. Among the treatments, *B. spectabilis* showed the highest disease inhibition, particularly at 10 per cent concentration, reducing disease severity by 49.82 per cent with an overall mean inhibition of 41.25 per cent. Similarly, *E. obliqua* was highly effective, with 46.52 per cent inhibition at 10 per cent, demonstrating a notable reduction in disease severity from 39.42 to 25.52 per cent. *A. indica* and *O. sanctum* also performed well, with mean inhibitions of 35.33 and 38.82 per cent, respectively. Neem reduced the disease severity from 52.01 to 33.63 per cent, and Tulasi exhibited its highest inhibition (48.06 %) at 10 per cent (Table 3) (Fig. 1b).

Moderate efficacy was observed with *C. ambiguous*, *S. glauca* and the seaweed extracts *A. nodosum* and *Sargassum* sp. Lemon grass had a mean inhibition of 16.44 per cent, with the highest inhibition of 20.99 per cent at 10 per cent. Simarouba had a mean inhibition of 13.59 per cent, with its peak inhibition at 15.85 per cent at 10 per cent. The seaweed extracts had lower mean inhibitions 35.02 per cent for *Ascophyllum* and 28.45 per cent for *Sargassum*, although their effectiveness increased at higher concentrations. *P. pinnata* and *M. pudica* exhibited moderate disease inhibition, reducing the severity by up to 27.78 and 38.93 per cent at 10 per cent, respectively. In contrast, the control treatment showed no inhibition, maintaining a constant disease severity of 64.76 per cent.

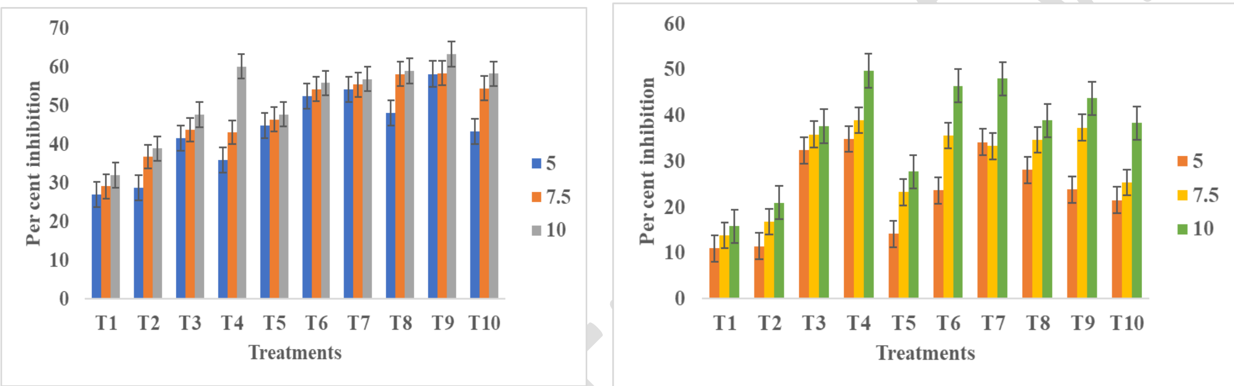

Fig. 1: Per cent powdery mildew leaf area inhibition a) botanicals/biostimulants over control; b) botanicals/biostimulants within treatment;

Overall, higher concentrations (10 %) resulted in the most significant disease suppression. *Ascophyllum nodosum*, *Osmium sanctum*, and *Eucalyptus obliqua* were consistently effective across all concentrations. These findings align with studies by Marzani *et al*. (2021) and Kumar and Chandel (2018), which demonstrated the potential of plant-based extracts for managing powdery mildew.

4. Conclusion

The study highlights the potential of botanicals and biostimulants as eco-friendly alternatives to synthetic fungicides for managing powdery mildew in cucumber. Among the tested treatments, *Ascophyllum nodosum*, *Mimosa pudica* and *Osmium sanctum* emerged as the most effective, particularly at higher concentrations. Future research should focus on field-scale validation and the development of integrated disease management strategies.

5. References

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