

# Comparative Analysis of Growth Responses in Indian Mustard to Azotobacter Priming and Vermicompost for Sustainable Cultivation

A study investigating the responses of Indian mustard (*Brassica juncea*) to biopriming with Azotobacter, incorporation of vermicompost and their comparative analysis was performed in 2024 Rabi season in new alluvial zone at Bidhan Chandra Krishi Viswavidyalaya, Nadia district, West Bengal. The research involved ten genotypes of Indian mustard and four treatments in completely randomized design. Treatments were designed as seeds sown in field soil, Azotobacter-primed seeds in field soil, seeds sown in vermicompost mixed with field soil, and Azotobacter-primed seeds in vermicompost with field soil. Eight key parameters were considered i.e. germination rate, seedling fresh and dry weight, seedling length, vigour index I and II, proline, and chlorophyll content. Results established improvements in germination and other growth parameters, particularly with the treatment combining Azotobacter priming and vermicompost, which presented the highest values across most genotypes. The performance of TM 306 - 1 and TBM 143 genotypes had produced the best results. The results emphasize the potential for utilizing biofertilizers and organic amendments in sustainable mustard cultivation, providing an effective substitute for chemical fertilizers.

## ABSTRACT

*Keywords: Azotobacter, Vermicompost, Seed Priming, Indian Mustard*

## 1. INTRODUCTION

Among the oilseeds, Indian mustard, from the *Brassicaceae* family, is a significant oilseed crop as it occupies a very vital place in the Indian agriculture scenario. It ranks second to groundnut in both area and production and is responsible for about 80% of the total rapeseed-mustard production. The mustard seeds are rich in nutrients, having an oil content that ranges from 38 to 50% and comprising erucic acid, linoleic acid, and oleic acid (Bater Dabi *et al.*, 2001; Gantait *et al.*, 2024; Janaki *et al.*, 2022; Kaushik *et al.*, 2024). The adverse effects of chemical fertilizers on Indian agriculture, both on soil and human health are manifold. The long-term effects of synthetic fertilizer usage have resulted in soil degradation, including reduced fertility and increased soil pH, so that at one point in time, it might turn into unproductive land (Bhokare P. R. & Wankhade R. R., 2024; Dube *et al.*, 2024). Excessive use of chemical fertilizers also leads to the contamination of the soil with metals, like cadmium and lead, which impose extensive environmental and health hazards (Dash *et al.*, 2022). Farmers are complaining that synthetic fertilizers not only detract nutritional quality from the crops but also taste bad, in addition to causing health problems such as hemoglobin disorders and chronic health issues because of high nitrate levels (Nichols, 2023). To solve the problem, biopriming with bio-fertilizers like Azotobacter and some organic amendments like vermicompost are good areas to explore as these provide sustainable and eco-friendly alternatives. Biopriming, a sort of seed treatment, refers to soaking seeds in a solution containing a beneficial microorganism. Microorganisms like bacteria or fungi colonizes and, in some cases, penetrate the seed coat (Gantait *et al.*, 2024; Govind *et al.*, 2024). Freelifving nitrogen fixing bacteria i.e. Azotobacter can convert atmospheric nitrogen into an available form from which plants can derive. Seed germination and seedling vigor are enhanced by growth-promoting chemical compounds produced by Azotobacter (Bater Dabi *et al.*, 2001; Janaki *et al.*, 2022; Kaushik *et al.*, 2024). Vermicompost refers to organic fertilizer formed from the digestion of organic waste materials by earthworms. It is a nutrient-rich semi-bulky organic fertilizer containing high concentrations of macro and micronutrients. Vermicompost is

Formatted: Font: Italic

31 a good additive to soil because it improves soil quality, soil fertility and microbial activity (M. Kumar *et al.*,  
32 2023; Singh *et al.*, 2018). The present experiment has been conducted to compare the germination,  
33 seedling vigor, chlorophyll and proline content of different varieties when primed and exposed to  
34 vermicompost.

## 35 2. MATERIALS AND METHODS

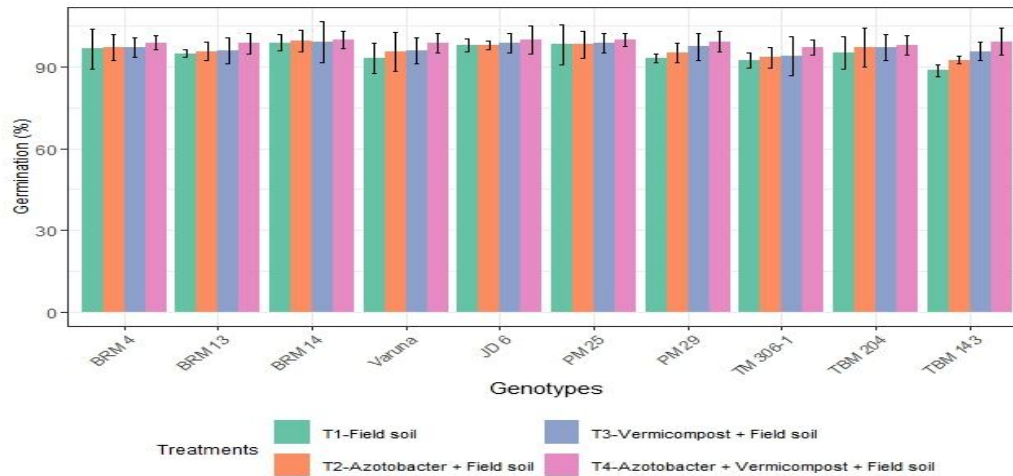
36 The research investigated the impacts of ten genotypes (G) and four treatments (T) on several  
37 seedling growth and physiological metrics in new alluvial zone at Bidhan Chandra Krishi Viswavidyalaya,  
38 Nadia district, West Bengal. The experiment was arranged in a completely randomized design with three  
39 replicates. Parameters including germination, seedling fresh and dry weight, seedling length, vigor index I  
40 and II, proline content, and chlorophyll content were evaluated to determine the growth potential and  
41 resilience of the crop under various treatments. The genotypes were BRM4(G<sub>1</sub>), BRM13(G<sub>2</sub>), BRM 14(G<sub>3</sub>),  
42 Varuna(G<sub>4</sub>), JD 6(G<sub>5</sub>), PM 25(G<sub>6</sub>), PM 29(G<sub>7</sub>), TM 306-1(G<sub>8</sub>), TBM 204(G<sub>9</sub>) and TBM 143(G<sub>10</sub>). The four  
43 treatments are as follows: Seeds sown in field soil (T<sub>1</sub>), Azotobacter primed seeds sown in field soil (T<sub>2</sub>),  
44 Seeds sown in vermicompost + field soil (T<sub>3</sub>), Azotobacter primed seeds sown in vermicompost + field  
45 soil (T<sub>4</sub>). 50 seeds were placed in each sterilized plastic container and left in open condition. In case of  
46 vermicompost treatment 50% of vermicompost and 50% of field soil was used. For Azotobacter seed  
47 priming a 5:1 ratio of Azotobacter to seed was maintained and seeds were soaked for one hour then  
48 dried. During the time of experiment maximum and minimum temperatures were 31.8°C and 12.4°C  
49 respectively, maximum, and minimum relative humidity were 78% and 54% respectively with 8.1 hours of  
50 average bright sunshine hours. After the seventh day seedlings from the container were counted and  
51 germination (%) was calculated by dividing the number of seeds germinated by the total seeds planted,  
52 then multiplied by 100. Ten seedlings were picked gently from container after 7<sup>th</sup> day and seedling length  
53 was measured using a centimeter scale. Average data was presented in centimeter (cm). For seedling  
54 fresh weight five random seedlings were taken out from each container and their weight was measured in  
55 a weighing balance and the average was calculated. To obtain seedling dry weight they were put in hot air  
56 oven till constant temperature was achieved. After that, weights of five dry seedlings were observed using  
57 a weighing balance and average was calculated. The vigor index I was assessed to evaluate the overall  
58 vigor and health of the seedlings under controlled conditions. The vigor index I was calculated using the  
59 formula given by Abdul-Baki and Anderson (1973) [Vigor Index I = Germination Percentage × Average  
60 seedling length (cm)]. The vigor index II is a critical parameter for evaluating seedling vigor, providing  
61 insights into the overall health and growth potential of plants. The vigor index II was calculated using the  
62 formula given by Abdul-Baki and Anderson (1973) [Vigor Index II = Germination Percentage ×  
63 Mean dry weight of seedlings (mg)]. Proline content was determined spectrophotometrically by adopting  
64 the ninhydrin method of Bates *et al.* (1973). Total chlorophyll was estimated following Arnon's method  
65 (Arnon, 1949). Statistical analysis was done using OPSTAT (Sheoran *et al.*, 1998).

## 66 3. RESULTS AND DISCUSSION

### 67 3.1 Results

#### 68 3.1.1 Germination (%)

69 Among the genotypes highest mean germination was shown by G<sub>3</sub> (99.43%), G<sub>6</sub> (98.87%)  
70 followed by G<sub>5</sub> (98.68%), while the lowest mean germination was recorded for G<sub>8</sub> (94.31%). The  
71 treatment with the highest germination was T<sub>4</sub> (99.02%) followed by T<sub>3</sub> (97.07%), and the lowest was T<sub>1</sub>  
72 (95.00%). The interaction of genotypes and treatments showed the three highest germination rates for G<sub>3</sub>  
73 × T<sub>4</sub> (100.00%), G<sub>6</sub> × T<sub>4</sub> (100.00%), and G<sub>5</sub> × T<sub>4</sub> (100.00%). The difference between T<sub>4</sub> and T<sub>3</sub> was not  
74 statistically significant.



75

**Figure 1.** Comparative analysis of germination (%) across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEm(±)]. Values are means of three replicates.

76

### 77 3.1.2 Fresh weight of seedlings(mg)

78

79

80

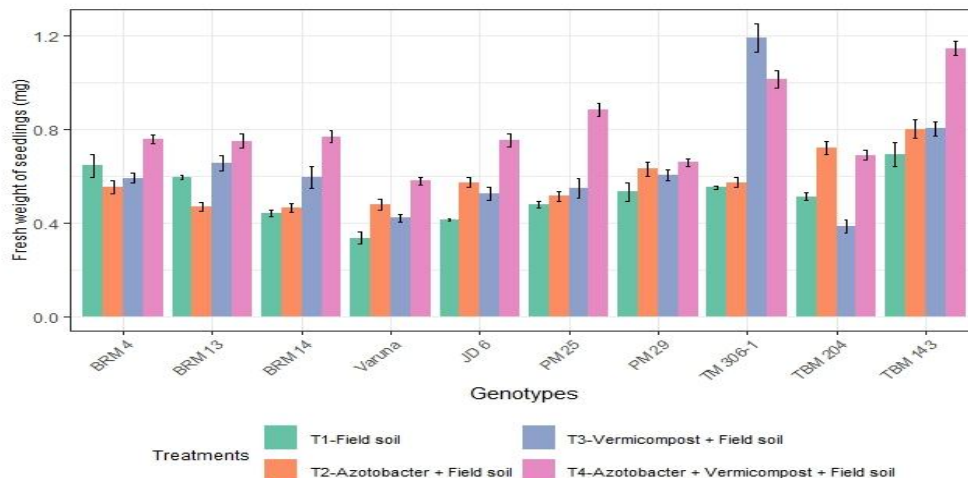
81

82

83

84

Genotypes G<sub>10</sub> (0.861mg), G<sub>8</sub> (0.833 mg), and G<sub>7</sub> (0.607 mg) had the highest seedling fresh weight while G<sub>4</sub> (0.453 mg) had the lowest. Among treatments, T<sub>4</sub> (0.801 mg) and T<sub>3</sub> (0.632 mg) had the highest fresh weight of seedlings with T<sub>1</sub> (0.520 mg) being the lowest. Among interaction effects G<sub>10</sub> × T<sub>4</sub> (1.147 mg), G<sub>8</sub> × T<sub>4</sub> (1.015 mg), and G<sub>8</sub> × T<sub>3</sub> (1.192 mg) had the most seedling fresh weight. G<sub>10</sub> and G<sub>8</sub> were significantly different from G<sub>7</sub>, but there was no significant difference between G<sub>10</sub> and G<sub>8</sub>. The difference between T<sub>4</sub> and T<sub>3</sub> was significant, suggesting that T<sub>4</sub> provides a notable improvement in fresh weight.



85

**Fig 2:** Comparative analysis of fresh weight of seedlings (mg) across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEm(±)]. Values are means of three replicates.

86

**Table 1:** Effect of Azotobacter priming and vermicompost treatments on germination (%) and fresh weight of seedlings (mg) in different Indian mustard genotypes

88

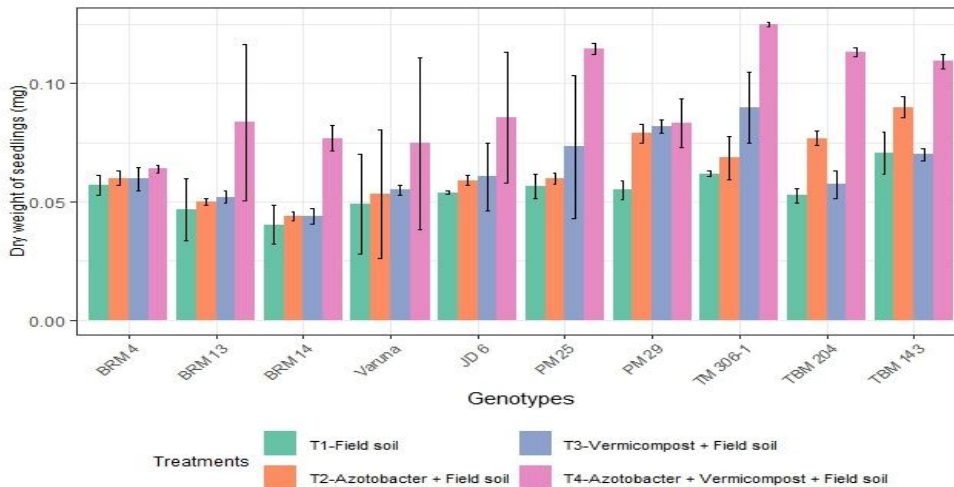
Table 1	Germination(%)					Fresh weight of seedlings(mg)				
	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	MeanG	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	MeanG
<b>G<sub>1</sub></b>	96.75	97.25	97.25	99.00	97.56	0.646	0.555	0.592	0.760	0.638
<b>G<sub>2</sub></b>	95.00	95.75	96.00	98.70	96.36	0.594	0.470	0.657	0.751	0.618
<b>G<sub>3</sub></b>	99.00	99.50	99.25	100.00	99.43	0.440	0.465	0.595	0.770	0.568
<b>G<sub>4</sub></b>	93.25	95.75	96.00	98.75	95.93	0.335	0.478	0.421	0.579	0.453
<b>G<sub>5</sub></b>	98.00	98.00	98.75	100.00	98.68	0.414	0.574	0.525	0.752	0.566
<b>G<sub>6</sub></b>	98.25	98.25	99.00	100.00	98.87	0.478	0.514	0.548	0.884	0.606
<b>G<sub>7</sub></b>	93.25	95.25	97.50	99.25	96.31	0.534	0.631	0.604	0.660	0.607
<b>G<sub>8</sub></b>	92.50	93.50	94.00	97.25	94.31	0.551	0.574	1.192	1.015	0.833
<b>G<sub>9</sub></b>	95.25	97.25	97.25	98.00	96.93	0.513	0.722	0.386	0.689	0.578
<b>G<sub>10</sub></b>	88.75	92.50	95.75	99.25	94.06	0.692	0.802	0.803	1.147	0.861

Mean T	95.00	96.30	97.07	99.02		0.520	0.579	0.632	0.801	
	Factor G	Factor T	Factor G X T			Factor G	Factor T	Factor G X T		
C.D(5%)	NS	NS	NS			0.040	0.025	0.079		
SEm(±)	2.112	1.336	4.224			0.014	0.009	0.028		
<sup>a</sup> G <sub>1</sub> : BRM 4; G <sub>2</sub> : BRM13; G <sub>3</sub> : BRM 14; G <sub>4</sub> : Varuna; G <sub>5</sub> : JD 6; G <sub>6</sub> : PM 25; G <sub>7</sub> : PM 29; G <sub>8</sub> : TM 306-1; G <sub>9</sub> : TBM 204; G <sub>10</sub> : TBM 143										
<sup>b</sup> T <sub>1</sub> : Seeds sown in field soil; T <sub>2</sub> : Azotobacter primed seeds in field soil; T <sub>3</sub> : Seeds in vermicompost + field soil; T <sub>4</sub> : Azotobacter primed seeds in vermicompost + field soil										
<sup>c</sup> CD: Critical Difference										
<sup>d</sup> SEm(±): Standard Error of Mean e NS: Non-significant										

89

### 3.1.3 Dry weight of seedlings(mg)

91 In the case of seedling dry weight, best genotypes were G<sub>8</sub> (0.086 mg), G<sub>10</sub> (0.085 mg), and G<sub>6</sub>  
 92 (0.076 mg), while the worst one was G<sub>3</sub> (0.051 mg). Among treatments, T<sub>4</sub> (0.093 mg) and T<sub>3</sub> (0.064 mg)  
 93 had the highest, with T<sub>1</sub> (0.054 mg) having the lowest value. The best three interactions were G<sub>8</sub> × T<sub>4</sub>  
 94 (0.125 mg), G<sub>6</sub> × T<sub>4</sub> (0.115 mg), and G<sub>10</sub> × T<sub>4</sub> (0.110 mg). Both genotypes and treatments showed  
 95 significant difference but G<sub>8</sub>, G<sub>10</sub>, and G<sub>6</sub> exhibited non-significant difference.



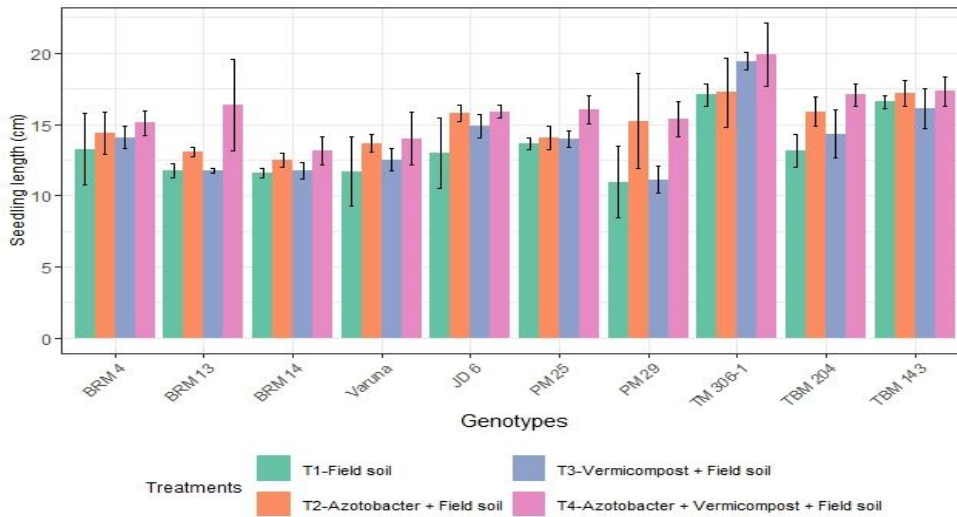
96

**Fig 3:** Comparative analysis of dry weight of seedlings (mg) across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEm(±)]. Values are means of three replicates.

97

98 **3.1.4 Seedling length(cm)**

99 G<sub>8</sub> (18.408 cm), G<sub>10</sub> (16.804 cm), and G<sub>5</sub> (14.892 cm) recorded maximum seedling length but G<sub>3</sub>  
 100 (12.233 cm) was the lowest. In the case of treatments T<sub>4</sub> (16.023 cm) and T<sub>2</sub> (14.900 cm) had the highest,  
 101 with T<sub>1</sub> (13.270 cm) having the lowest seedling length after 7 days. Both genotypes and treatments were  
 102 significant and G<sub>8</sub> was significantly different from G<sub>10</sub> and G<sub>5</sub>, while the latter two did not differ significantly  
 103 from each other. The difference between T<sub>4</sub> and T<sub>2</sub> is significant, highlighting that T<sub>4</sub> strongly enhances  
 104 seedling length.



105

**Fig 4:** Comparative analysis of seedling length (cm) across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEm(±)]. Values are means of three replicates.

106

107 **Table 2: Effect of Azotobacter priming and vermicompost treatments on dry weight of seedlings**

108 **(mg) and seedling length (cm) in different Indian mustard genotypes**

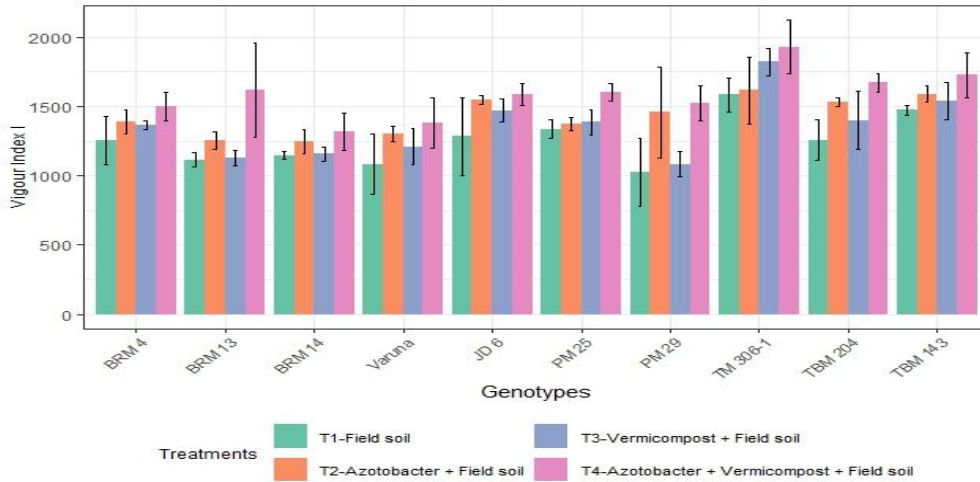
Table 2	Dry weight of seedlings(mg)					Seedling length(cm)				
	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	Mean G	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	Mean G
G <sub>1</sub>	0.057	0.060	0.060	0.064	0.060	13.26	14.40	14.06	15.10	14.20

G <sub>2</sub>	0.047	0.050	0.052	0.084	0.058	11.73	13.06	11.76	16.36	13.23
G <sub>3</sub>	0.040	0.044	0.044	0.077	0.051	11.60	12.46	11.73	13.13	12.23
G <sub>4</sub>	0.049	0.053	0.055	0.075	0.058	11.70	13.66	12.53	14.00	12.97
G <sub>5</sub>	0.054	0.059	0.061	0.086	0.065	12.96	15.80	14.90	15.90	14.89
G <sub>6</sub>	0.057	0.060	0.073	0.115	0.076	13.63	14.03	13.96	16.06	14.42
G <sub>7</sub>	0.055	0.079	0.082	0.083	0.075	10.96	15.23	11.13	15.36	13.17
G <sub>8</sub>	0.062	0.069	0.090	0.125	0.086	17.06	17.23	19.43	19.90	18.40
G <sub>9</sub>	0.053	0.077	0.057	0.113	0.075	13.16	15.90	14.33	17.06	15.11
G <sub>10</sub>	0.071	0.090	0.070	0.110	0.085	16.60	17.20	16.08	17.33	16.80
Mean T	0.054	0.064	0.064	0.093		13.27	14.90	13.99	16.02	
	Factor G	Factor T	Factor G X T			Factor G	Factor T	Factor G X T		
C.D(5%)	0.018	0.011	NS			2.058	1.302	NS		
SEm(±)	0.006	0.004	0.012			0.730	0.461	1.459		
<sup>a</sup> G <sub>1</sub> : BRM 4; G <sub>2</sub> : BRM13; G <sub>3</sub> : BRM 14; G <sub>4</sub> : Varuna; G <sub>5</sub> : JD 6; G <sub>6</sub> : PM 25; G <sub>7</sub> : PM 29; G <sub>8</sub> : TM 306-1; G <sub>9</sub> : TBM 204; G <sub>10</sub> : TBM 143										
<sup>b</sup> T <sub>1</sub> : Seeds sown in field soil; T <sub>2</sub> : Azotobacter primed seeds in field soil; T <sub>3</sub> : Seeds in vermicompost + field soil; T <sub>4</sub> : Azotobacter primed seeds in vermicompost + field soil										
<sup>c</sup> CD: Critical Difference										
<sup>d</sup> SEm(±): Standard Error of Mean e NS: Non-significant										

109

### 110 3.1.5 Vigour Index I

111 The best vigour index I was presented by T<sub>4</sub> (1,585) and T<sub>2</sub> (1,429), and T<sub>1</sub> (1,255) was the  
112 lowest. The best genotypes for high vigour index I was G<sub>8</sub> (1,736), G<sub>10</sub> (1,580), and G<sub>5</sub> (1,471), while the  
113 worst was G<sub>7</sub> (1,270). The highest interactions were G<sub>8</sub> × T<sub>4</sub> (1,927), G<sub>8</sub> × T<sub>3</sub> (1,819), and G<sub>10</sub> × T<sub>4</sub>  
114 (1,724). both treatments and genotypes were significant and G<sub>8</sub>, G<sub>10</sub>, and G<sub>5</sub> also showed significant  
115 differences among each other. The difference between T<sub>4</sub> and T<sub>2</sub> is highly significant, indicating a  
116 substantial effect of T<sub>4</sub> on vigour index I.



**Fig 5:** Comparative analysis of vigour index I across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEM(±)]. Values are means of three replicates.

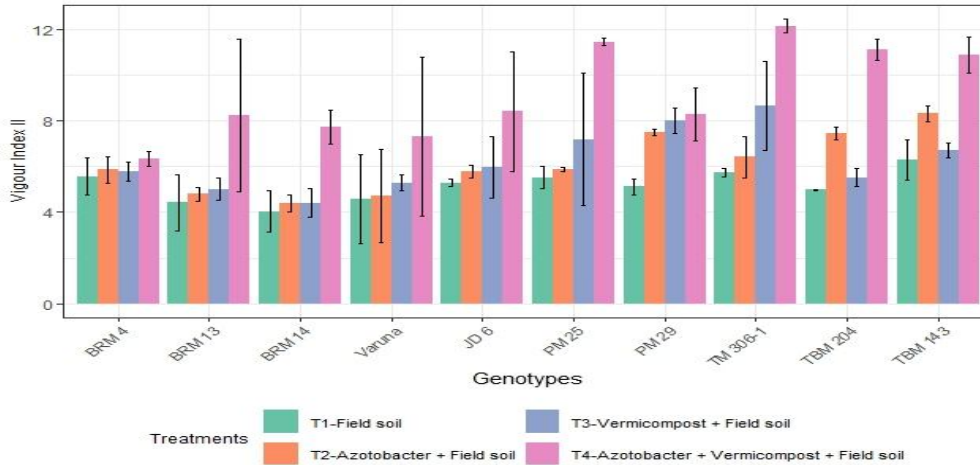
117

118

### 3.1.6 Vigour Index II

119

120 The genotypes for vigour index II were G<sub>8</sub> (8.247), G<sub>10</sub> (8.053), and G<sub>6</sub> (7.514), with G<sub>3</sub> (5.146)  
 121 being the lowest. For treatments T<sub>4</sub> (9.198) and T<sub>3</sub> (6.257) were the highest, and T<sub>1</sub> (5.156) was the  
 122 lowest. The three highest interactions were G<sub>8</sub> × T<sub>4</sub> (12.152), G<sub>6</sub> × T<sub>4</sub> (11.457), and G<sub>10</sub> × T<sub>4</sub> (10.896).  
 123 Both genotypes and treatments were significant, with G<sub>8</sub> being significantly different from G<sub>10</sub> and G<sub>6</sub>,  
 124 while G<sub>10</sub> and G<sub>6</sub> did not differ significantly. The difference between T<sub>4</sub> and T<sub>3</sub> was significant, reinforcing  
 125 T<sub>4</sub>'s superior performance in vigor improvement.



126

127  
128  
129

**Fig 6:** Comparative analysis of vigour index II across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEm(±)]. Values are means of three replicates.

**Table 3 :** Effect of Azotobacter priming and vermicompost treatments on Vigour Index I and Vigour Index II in different Indian mustard genotypes

Table 3	Vigour Index I					Vigour Index II				
	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	Mean G	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	Mean G
G <sub>1</sub>	1,253	1,386	1,362	1,496	1,374	5.577	5.865	5.784	6.344	5.892
G <sub>2</sub>	1,114	1,252.0	1,129	1,618	1,278	4.423	4.797	5.017	8.258	5.624
G <sub>3</sub>	1,146	1,242	1,157	1,319	1,216	4.041	4.392	4.416	7.733	5.146
G <sub>4</sub>	1,083	1,300	1,209	1,381	1,243	4.571	4.714	5.283	7.335	5.476
G <sub>5</sub>	1,282	1,546	1,469	1,588	1,471	5.293	5.785	5.966	8.412	6.364
G <sub>6</sub>	1,333	1,371	1,384	1,601	1,423	5.528	5.872	7.199	11.457	7.514
G <sub>7</sub>	1,024	1,455	1,081	1,522	1,270	5.121	7.498	8.003	8.281	7.226
G <sub>8</sub>	1,583	1,614	1,819	1,927	1,736	5.734	6.429	8.672	12.152	8.247
G <sub>9</sub>	1,257	1,531	1,399	1,669	1,464	4.980	7.443	5.530	11.109	7.266
G <sub>10</sub>	1,472	1,588	1,538	1,724	1,580	6.295	8.318	6.702	10.896	8.053
Mean T	1,255	1,429	1,355	1,585		5.156	6.111	6.257	9.198	
	Factor G	Factor T	Factor G X T			Factor G	Factor T	Factor G X T		
C.D(5%)	206.239	130.437	NS			1.747	1.105	NS		
SEm(±)	73.110	46.239	146.220			0.619	0.392	1.239		

<sup>a</sup> G<sub>1</sub>: BRM 4; G<sub>2</sub>: BRM13; G<sub>3</sub>: BRM 14; G<sub>4</sub>: Varuna; G<sub>5</sub>: JD 6; G<sub>6</sub>: PM 25; G<sub>7</sub>: PM 29; G<sub>8</sub>: TM 306-1; G<sub>9</sub>: TBM 204; G<sub>10</sub>: TBM 143

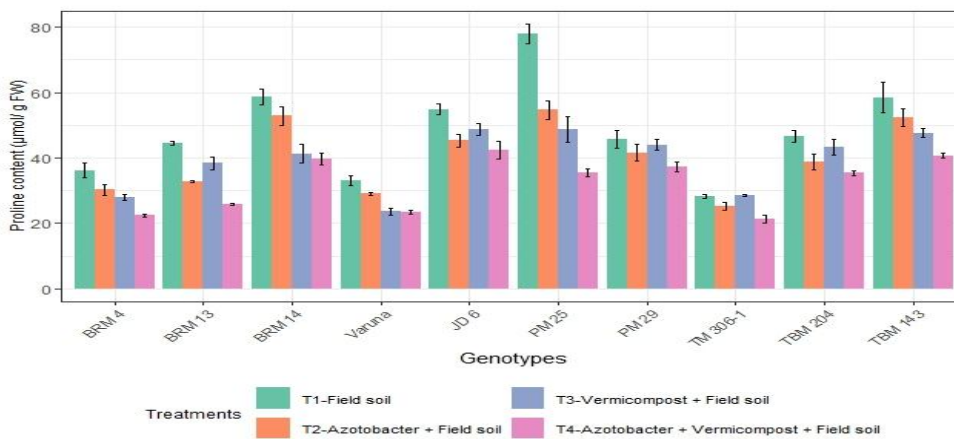
<sup>b</sup> T<sub>1</sub>: Seeds sown in field soil; T<sub>2</sub>: Azotobacter primed seeds in field soil; T<sub>3</sub>: Seeds in vermicompost + field soil; T<sub>4</sub>: Azotobacter primed seeds in vermicompost + field soil

<sup>c</sup> CD: Critical Difference

<sup>a</sup> SEM(±): Standard Error of Mean e NS: Non-significant

130 **3.1.7 Proline content(μmol/ g FW)**

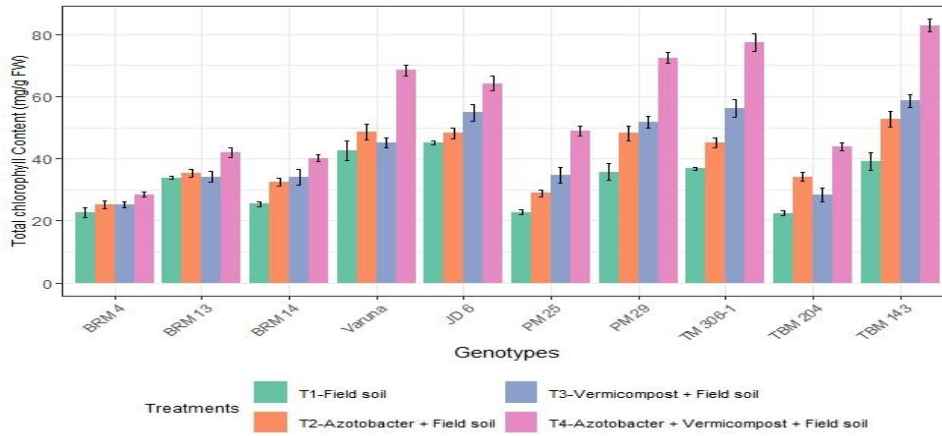
131 In the case of proline content of seedlings, highest values were observed in G<sub>7</sub> (56.311 μmol/ g  
 132 FW), G<sub>8</sub> (46.200 μmol/ g FW), and G<sub>4</sub> (44.600 μmol/ g FW), with G<sub>5</sub> (28.567 μmol/ g FW) being the  
 133 lowest. Also, in treatments, T<sub>4</sub> (40.801 μmol/ g FW) and T<sub>3</sub> (40.071 μmol/ g FW) were the highest, and T<sub>2</sub>  
 134 (38.511 μmol/ g FW) was the lowest. The three highest interactions were G<sub>7</sub> × T<sub>4</sub> (77.656 μmol/ g FW), G<sub>6</sub>  
 135 × T<sub>4</sub> (45.744 μmol/ g FW), and G<sub>8</sub> × T<sub>4</sub> (34.878 μmol/ g FW). The results showed significant differences  
 136 between G<sub>7</sub> and the other groups (G<sub>8</sub> and G<sub>4</sub>), while G<sub>8</sub> and G<sub>4</sub> were not significantly different. However,  
 137 treatments were not significant.



138 **Fig 7: Comparative analysis of proline content (μmol/ g FW) across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEM(±)]. Values are means of three replicates.**

140 **3.1.8 Total chlorophyll Content(mg/g FW)**

141 T<sub>4</sub> (56.840 mg/g FW) and T<sub>3</sub> (42.290 mg/g FW) showed the highest chlorophyll content with T<sub>1</sub>  
 142 (32.640 mg/g FW) being the lowest. For genotypes, G<sub>10</sub> (58.300 mg/g FW), G<sub>8</sub> (53.900 mg/g FW), and G<sub>7</sub>  
 143 (52.000 mg/g FW) had the highest total chlorophyll content values, while G<sub>1</sub> (25.375 mg/g FW) was the  
 144 lowest. The three highest interactions were G<sub>10</sub> × T<sub>4</sub> (82.800 mg/g FW), G<sub>8</sub> × T<sub>4</sub> (77.400 mg/g FW), and  
 145 G<sub>7</sub> × T<sub>4</sub> (72.400 mg/g FW). Factors Genotype, treatment, and their interaction were significant. Significant  
 146 differences were observed among G<sub>10</sub>, G<sub>8</sub>, and G<sub>7</sub>. The difference between T<sub>4</sub> and T<sub>3</sub> was significant,  
 147 indicating that T<sub>4</sub> greatly enhances chlorophyll content.



148

**Fig 8:** Comparative analysis of total chlorophyll Content (mg/g FW) across different Indian mustard genotypes (G<sub>1</sub>-G<sub>10</sub>) under four treatments (T<sub>1</sub>-T<sub>4</sub>). Error bars represent Standard Error of Means [SEm(±)]. Values are means of three replicates.

149

150

151

**Table 4 :** Effect of Azotobacter priming and vermicompost treatments on proline content (µmol/ g FW) and total chlorophyll Content (mg/g FW) in different Indian mustard genotypes

Table 4	Proline content(µmol/ g FW)					Total chlorophyll Content(mg/g FW)				
	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	Mean G	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	Mean G
G <sub>1</sub>	36.20	31.66	30.53	27.28	31.42	22.70	25.20	25.10	28.50	25.37
G <sub>2</sub>	22.36	36.92	41.11	32.62	33.25	33.80	35.40	34.20	41.80	36.30
G <sub>3</sub>	38.33	30.11	36.97	58.35	40.94	25.40	32.40	34.10	40.20	33.02
G <sub>4</sub>	52.80	44.00	41.97	39.62	44.60	42.50	48.60	45.10	68.40	51.15
G <sub>5</sub>	33.06	30.15	28.06	22.97	28.56	45.10	48.20	54.80	64.20	53.07
G <sub>6</sub>	23.43	44.54	51.16	45.74	41.22	22.80	28.90	34.70	48.90	33.82
G <sub>7</sub>	48.66	44.82	54.10	77.65	56.31	35.70	48.20	51.70	72.40	52.00
G <sub>8</sub>	54.63	49.57	45.71	34.87	46.20	36.80	45.20	56.20	77.40	53.90
G <sub>9</sub>	45.63	42.01	44.07	43.30	43.75	22.50	34.20	28.40	43.800	32.22
G <sub>10</sub>	37.30	31.30	26.98	25.56	30.28	39.10	52.70	58.60	82.80	58.30
Mean T	39.24	38.51	40.07	40.80		32.64	39.90	42.29	56.84	

	Factor G	Factor T	Factor G X T	Factor G	Factor T	Factor G X T
<b>C.D(5%)</b>	5.773	NS	11.545	2.627	1.661	5.253
<b>SEm(±)</b>	2.046	1.294	4.093	0.931	0.589	1.862
<sup>a</sup> G <sub>1</sub> : BRM 4; G <sub>2</sub> : BRM13; G <sub>3</sub> : BRM 14; G <sub>4</sub> : Varuna; G <sub>5</sub> : JD 6; G <sub>6</sub> : PM 25; G <sub>7</sub> : PM 29; G <sub>8</sub> : TM 306-1; G <sub>9</sub> : TBM 204; G <sub>10</sub> : TBM 143 <sup>b</sup> T <sub>1</sub> : Seeds sown in field soil; T <sub>2</sub> : Azotobacter primed seeds in field soil; T <sub>3</sub> : Seeds in vermicompost + field soil; T <sub>4</sub> : Azotobacter primed seeds in vermicompost + field soil <sup>c</sup> CD: Critical Difference <sup>d</sup> SEm(±): Standard Error of Mean e NS: Non-significant						

152

### 153 3.2 Discussion

154 The germination study indicated higher mean values for treatments G<sub>3</sub>, G<sub>6</sub>, and G<sub>5</sub>, especially under  
155 T<sub>4</sub>, which consistently demonstrated greater germination rates. However, the interactions between  
156 genotype and treatment were non-significant which means the treatments did have an effect but their  
157 influence was largely consistent among genotypes. Many studies reported the positive effect of  
158 vermicompost on the germination and growth of mustard seedlings and plants (Merta, 2023; Haque & Ali,  
159 2020; Reza, 2023; Reza *et al.*, 2022). The improved germination upon vermicompost application could be  
160 attributed to several factors. Vermicompost has many available mineral nutrients, humic substances, and  
161 plant growth-promoting agents such as auxins, which are known to improve seed germination and  
162 seedling growth (Bhattacharya *et al.*, 2019; Pathma & Sakthivel, 2012). Vermicompost helps in increasing  
163 the porosity, aeration, and water retention capabilities which enhance the germination and growth of  
164 mustard plants (Sarma & Gogoi, 2015; Merta, 2023). Azotobacter further increases seed germination in  
165 crops by ensuring plant health through nitrogen fixation, phosphate solubilization, and growth hormone  
166 production. Together, these factors lead to optimum growth, increased vigor, and effective germination  
167 (Abbas *et al.*, 2024). Azotobacter has been found effective in promoting seed germination in several crops  
168 of paddy (Chennappa *et al.*, 2017a), wheat (Silini *et al.*, 2012), buckwheat, winter wheat (Roi *et al.*, 2022),  
169 and beetroot (Kurdish *et al.*, 2008).

170 The fresh and dry weight of the seedlings indicated that genotypes G<sub>10</sub>, G<sub>8</sub>, and G<sub>7</sub> performed to  
171 the best under T<sub>4</sub> conditions. The genotypes that showed a significant improvement in growth, based on  
172 fresh weight measurements under T<sub>4</sub> conditions were G<sub>10</sub> and G<sub>8</sub>. In addition, the treatments together with  
173 the genotypes significantly affected seedling length; under T<sub>4</sub> conditions, G<sub>8</sub> was the best combination.  
174 Vermicompost significantly increases the fresh weight and dry weight of seedlings, especially for  
175 tomatoes and pepper plants (Brace, 2017). Riwardi *et al.* (2023) showed that vermicomposting had a  
176 great influence on both fresh and dry weights in maize seedlings. Shoot fresh weights were increased  
177 by over 23% for wheat inoculated with Azotobacter strain Azo-8, and increases by over 23% in shoot dry  
178 weights along with marked improvements in root biomass have also been reported (Singh *et al.*, 2013). In  
179 addition to the above, Vigna radiata seedlings have shown a 20.07% increase in fresh weight and a little  
180 over 62% increase in dry weight through Azotobacter inoculation (Munnaza *et al.*, 2012). Scientists show  
181 that the addition of vermicompost to growth medium can bring a change in seedling height to cucumbers

182 from 1.9% to about 18.6%, related to leaf area and fresh weight increase (Jankauskienė and others,  
183 2022).

184 The vigour indices (I and II) reconfirmed the superiority of G<sub>8</sub>, G<sub>10</sub>, and G<sub>6</sub>. Most importantly, G<sub>8</sub> under  
185 T<sub>4</sub> recorded the maximum values. By enhancing seed germination, promoting disease resistance, and  
186 enhancing the overall health of the plants, vermicompost improves vigour index of crops (Mohite *et al.*,  
187 2024). Research conducted by Bajaj (2023) disclosed that the tomato plants' vigour index has increased  
188 with altered levels of vermicompost, indicating a positive effect of vermicompost on the crop's growth. The  
189 culture filtrate of *Azotobacter salinestris* (GVT-1) has improved the vigour index of paddy seeds, thus  
190 enhancing growth and seedling germination rates in crops (Chennappa *et al.*, 2017b).

191 The proline content was variable from one genotype to another, with a maximum content of G<sub>7</sub> and G<sub>8</sub>  
192 met under T<sub>4</sub> treatment. This indicates an increased possibility for genotypes to develop physiological  
193 resistance toward such stresses. In high correlation, some genotypes recorded very high chlorophyll  
194 contents which are vital for photosynthesis, that is, G<sub>10</sub>, G<sub>8</sub>, G<sub>7</sub> under T<sub>4</sub> conditions. These results  
195 confirmed those genotypes as most suitable for maximizing treatment benefits towards better  
196 physiological output. Mixed inoculation with different *Azotobacter* strains on wheat seedlings has been  
197 studied, which increased proline level and growth parameters under osmotic stress, shown to be  
198 significantly related to drought resistance (Liu *et al.*, 2013). Various *Azotobacter* strains have  
199 been reported to improve the physiological attributes such as proline synthesis in maize grown on saline  
200 soils and hence its usefulness toward osmotic adjustment and alleviation of stress in plants (Abdel Latef  
201 *et al.*, 2020). Research suggests that the addition of vermicompost resulted in an increase in proline  
202 concentration, which is the major osmotic regulator that helps plants in overcoming abiotic problems like  
203 drought and salinity (Bokobana *et al.*, 2020; Hosseinzadeh *et al.*, 2017). During water stress conditions,  
204 30% vermicompost induced a 39% increase in the proline content of chickpea seedlings (Hosseinzadeh *et al.*,  
205 2017). In fact, when tomato seedlings are subjected to vermicompost-leachate, especially during heat  
206 and moisture stresses, there are increased proline levels (Chinsamy *et al.*, 2014). Researchers have well  
207 put vermicompost as an important source of macro- and micronutrients which henceforth augments plant  
208 nutrition and at the same time improves chlorophyll concentration, as commonly exhibited by *Capsicum*  
209 *annuum* and other vegetable crop seedlings (Kamalkant Yadav *et al.*, 2014; Theunissen, 2010). Kumar *et al.*  
210 (2016) observed improvement in the photosynthetic pigments of *Jatropha* by *Azotobacter* and  
211 arbuscular mycorrhizal fungus. A treatment of *Azotobacter* in wheat plants indicated a very high increase  
212 in the total chlorophyll content (mg g<sup>-1</sup>) (El-zawawy *et al.*, 2023). The chlorophyll content is increased with  
213 the inoculation of *Azotobacter*, either alone or with *Rhizobium*, in black gram (*Vigna mungo*) as compared  
214 to the control (Tiwari *et al.*, 2017).

215 The combination of vermicompost with *Azotobacter*, produced positive improvements in crop growth,  
216 yield, and nutrient uptake in many crops. This is a blend of the benefits of vermicompost from organic  
217 matter and nitrogen-fixing *Azotobacter*, which enhances plant growth parameters in crops such as chili,  
218 strawberry, and maize, more than that by either one of the components, or chemical fertilizers alone  
219 (Kalpana, 2019; Shirkhani & Nasrolahzadeh, 2016; Tripathi *et al.*, 2015). In *Amaranthus*, this combination  
220 of vermicompost and *Azotobacter* was conducive to early emergence and higher germination  
221 percentages, which led to the development of more vigorous seedlings (Yadav *et al.*, 2024). Furthermore,  
222 the combined application of *Azotobacter* and plant-based composts, such as *Moringa*, has been  
223 demonstrated to enhance growth parameters and nutrient levels in various crops, further supporting the  
224 synergistic benefits of *Azotobacter* when utilized in conjunction with organic soil amendments (Albureikan,  
225 2024). It also helps in increasing the availability and uptake of essential nutrients such as nitrogen,  
226 phosphorus, and potassium in plants and much better nutrient content in crops such as rice and wheat  
227 (Ghadimi *et al.*, 2021; V. Kumar & Singh, 2001; Rather & Sharma, 2009) reducing the use of chemical  
228 fertilizer, being among the sustainable agricultural practices towards environmental sustainability (Rather  
229 & Sharma, 2009; Shirkhani & Nasrolahzadeh, 2016). The presence of *Azotobacter* in vermicompost,  
230 therefore, helps improve the microbial activity of the soil, which translates to a better soil structure and  
231 health, thus benefitting long-term crop productivity (Ghadimi *et al.*, 2021; Mal *et al.*, 2021).

232 **4. LIMITATIONS**

233 Although the study establishes the benefits of Azotobacter seed priming and vermicompost  
234 integration in Indian mustard cultivation, it is limited because it focused only on seedling stages in a  
235 controlled setup. Exactly similar results may not be replicated in a field trial also without exploring long  
236 term effects of treatments on seed yield and plant health. So further field trials and cost-benefit analysis  
237 are needed.

238 **5. CONCLUSION**

239 The study highlights the importance of combining genotype selection with advanced treatment  
240 techniques for improving mustard cultivation. Genotypes TM 306-1(G<sub>8</sub>) and TBM 143(G<sub>10</sub>) are the top  
241 performers, especially when paired with treatment Azotobacter primed seeds sown in vermicompost +  
242 field soil(T<sub>4</sub>), which provides favorable conditions for nutrient uptake, growth, and stress resilience.  
243 Farmers can improve germination rates, seedling vigor, and stress tolerance by selecting these  
244 genotypes and applying T<sub>4</sub>. These genotypes are adaptable to varying conditions, making them ideal for  
245 cultivation in diverse agro-climatic regions.

246 **COMPETING INTERESTS**

247 Authors have declared that no competing interests exist.

248 **AUTHORS' CONTRIBUTIONS**

249 Aritra Mukherjee designed the study, performed the statistical analysis, wrote the protocol, and wrote the  
250 first draft of the manuscript. Ananya Baidya and Achyuta Basak managed the analyses of the study.  
251 Md.Sabir Ahmed Mondol managed the literature search. All authors read and approved the final  
252 manuscript.

253

254 **Disclaimer (Artificial intelligence)**

255 **Option 1:**

256 Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT,  
257 COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this  
258 manuscript.

259 **Option 2:**

260 Author(s) hereby declare that generative AI technologies such as Large Language Models, etc. have been  
261 used during the writing or editing of manuscripts. This explanation will include the name, version,  
262 model, and source of the generative AI technology and as well as all input prompts provided to the  
263 generative AI technology

264 Details of the AI usage are given below:

265 **1.**

266 **2.**

267 **3.**

268

269

270 **References**

- 271 1. Abdul-Baki, A. A., & Anderson, J. D. (1973). Vigor determination in soybean seed by multiple  
272 criteria 1. *Crop science*, 13(6):630-633.
- 273 2. Albureikan, M. O. I. (2024). Enhancement of Plant Growth with Plant-Based Compost and the  
274 Heterotrophic Azotobacter and Streptomyces Inoculation under Greenhouse Conditions. *Journal*  
275 *of Pure and Applied Microbiology*, 18(3), 1632–1647. <https://doi.org/10.22207/jpam.18.3.13>
- 276 3. Bater Dabi, J.K. Singh, Rajesh Kumar Singh, & Akhilesh Vishwakarma. (2001). Quality and  
277 profitability of Indian mustard (*Brassica juncea*) as affected by nutrient-management practices  
278 under irrigated condition. *Indian Journal of Agronomy*, 60(1), 168–171.  
279 <https://doi.org/10.59797/ija.v60i1.4435>
- 280 4. Bates, L. S., Waldren, R. P. A., & Teare, I. D. (1973). Rapid determination of free proline for water-  
281 stress studies. *Plant and soil*, 39, 205-207.
- 282 5. Bhokare P. R. & Wankhade R. R. (2024). Impacts of chemical fertilizer on agricultural soil of  
283 digras region, yavatmal district, maharashtra (india): a case study. *International Education and*  
284 *Research Journal*, 10(5). <https://doi.org/10.21276/IERJ24050062972803>
- 285 6. Dash, G., Mohanty, K. G. R., Sahoo, D., Jali, P., B. Jyotirmayee, Parida, S., Deo, B., & Mahalik,  
286 G. (2022). Studies on the Impact of Agrochemicals used on the Croplands of Jagatsinghpur  
287 District of Odisha, India. *Ecology, Environment and Conservation*, 28, 43–50.  
288 <https://doi.org/10.53550/EEC.2022.v28i07s.008>
- 289 7. Dube, A., Lal, K., Laik, R., & Jaiswal, S. (2024). Effects of 38-year continuous manure and  
290 fertilizer application on soil Physico-chemical characteristics at various depths in rice-wheat  
291 cropping system in Indo-Gangetic plain. *International Journal of Research in Agronomy*, 7(1),  
292 250–254. <https://doi.org/10.33545/2618060X.2024.v7.i1d.217>
- 293 8. Gantait, A., Masih, S. A., & Maxton, A. (2024). Effect of Biological Priming on Metabolomic and  
294 Molecular Changes in Response to Drought Stress in Brassica juncea. *Journal of Advances in*  
295 *Biology & Biotechnology*, 27(8), 1325–1338. <https://doi.org/10.9734/jabb/2024/v27i81256>
- 296 9. Ghadimi, M., Sirousmehr, A., Ansari, M. H., & Ghanbari, A. (2021). Organic soil amendments  
297 using vermicomposts under inoculation of N<sub>2</sub>-fixing bacteria for sustainable rice production.  
298 *PeerJ*, 9. <https://doi.org/10.7717/peerj.10833>
- 299 10. Govind, Kumar, M., Kumar, M., Hardeep, & Sangwan, D. (2024). Effect of seed priming on  
300 germination parameters of Bael (*Aegle marmelos* Corr.) under laboratory conditions. *Environment*  
301 *Conservation Journal*, 25(1), 199–205. <https://doi.org/10.36953/ECJ.24392668>
- 302 11. Janaki, B., Singh, R., & Tripathi, P. (2022). Effect of Biofertilizers and Potassium on Yield and  
303 Economics of Yellow Mustard (*Brassica campestris* L.). *International Journal of Environment and*  
304 *Climate Change*, 1282–1287. <https://doi.org/10.9734/ijeccl/2022/v12i1131106>

- 305 12. Kalpana, J. C. B. (2019). Co-inoculation effect of vermicompost and plant growth promoting  
306 rhizobacteria (*Azotobacter* Sp.) on the growth of chilli (*Capsicum annuum* L.). *Journal of*  
307 *Pharmacognosy and Phytochemistry*, 8, 2340–2348.
- 308 13. Kaushik, S., Yadav, K. G., Kumar, P., Kumar, A., Kumar, P., Qidwai, S., & Yadav, V. (2024). Effect  
309 of Liquid Biofertilizer and Variable Source of Nutrients on Growth and Yield of Indian Mustard  
310 (*Brassica juncea* L.) in Western U.P., India. *Journal of Advances in Biology & Biotechnology*,  
311 27(5), 721–729. <https://doi.org/10.9734/jabb/2024/v27i5834>
- 312 14. Kumar, M., Yadav, D. D., Singh, S., Verma, V. K., Prasad, J., Singh, U., & Sachan, D. S. (2023).  
313 Effect of FYM, Vermicompost and Fertility Levels on Yield Attributes of Indian Mustard (*Brassica*  
314 *juncea* L.). *International Journal of Plant & Soil Science*, 35(21), 604–612.  
315 <https://doi.org/10.9734/ijpss/2023/v35i214015>
- 316 15. Kumar, V., & Singh, K. (2001). Enriching vermicompost by nitrogen fixing and phosphate  
317 solubilizing bacteria. *Bioresource Technology*, 76 2, 173–175. [https://doi.org/10.1016/S0960-](https://doi.org/10.1016/S0960-8524(00)00061-4)  
318 [8524\(00\)00061-4](https://doi.org/10.1016/S0960-8524(00)00061-4)
- 319 16. Mal, S. V. V., Chattopadhyay, G., & Chakrabarti, K. (2021). Microbiological integration for  
320 qualitative improvement of vermicompost. *International Journal of Recycling of Organic Waste in*  
321 *Agriculture*. <https://doi.org/10.30486/IJROWA.2021.1902019.1087>
- 322 17. Nichols, C. E. (2023). Inflammatory agriculture: Political ecologies of health and fertilizers in India.  
323 *Environment and Planning E: Nature and Space*, 6(2), 1030–1053.  
324 <https://doi.org/10.1177/25148486221113557>
- 325 18. Rather, S. A., & Sharma, N. (2009). The effect of integrated use of vermicompost, biofertilizer  
326 (*Azotobacter chroococcum*) and inorganic fertilizers (N, P, K and Zn) on yield and nutrient content  
327 and their uptake by wheat. *International Journal of Agricultural Sciences*, 5, 371–373.
- 328 19. Sheoran, O. P., Tonk, D. S., Kaushik, L. S., Hasija, R. C., & Pannu, R. S. (1998). Statistical  
329 software package for agricultural research workers. *Recent advances in information theory,*  
330 *statistics & computer applications* by DS Hooda & RC Hasija Department of Mathematics  
331 Statistics, CCS HAU, Hisar, 8(12):139-143.
- 332 20. Shirkhani, A., & Nasrolahzadeh, S. (2016). Vermicompost and Azotobacter as an ecological  
333 pathway to decrease chemical fertilizers in the maize, *Zea mays*. *Biochemical and Biophysical*  
334 *Research Communications*, 9, 382–390. <https://doi.org/10.21786/BBRC/9.3/7>
- 335 21. Singh, R., Babu, S., Avasthe, R., Yadav, G. S., Chettri, T. K., & Singh, A. (2018). Effect of organic  
336 mulches and vermicompost on productivity, profitability and energetic of mustard (*Brassica*  
337 *campestris*) in popcorn (*Zea mays everta*)- mustard cropping system in rainfed Sikkim Himalaya.  
338 *The Indian Journal of Agricultural Sciences*, 88(11), 1735–1739.  
339 <https://doi.org/10.56093/ijas.v88i11.84916>
- 340 22. Singh, N. K., Chaudhary, F. K., & Patel, D. B. (2013). Effectiveness of Azotobacter bio-inoculant  
341 for wheat grown under dryland condition. *Journal of Environmental Biology*, 34(5), 927.

342 23. Tripathi, V., Kumar, S., & Gupta, A. (2015). Influence of Azotobacter and vermicompost on growth,  
343 flowering, yield and quality of strawberry cv. Chandler. *Indian Journal of Horticulture*, 72, 201–  
344 205. <https://doi.org/10.5958/0974-0112.2015.00039.0>

345 24. Yadav, S., Jangu, R., Malviya, S., Prajapati, J., Das, K., & Asati, K. (2024). Integrated Nutrient  
346 Management: A Pathway to Enhanced Growth and Yield in *Amaranthus tricolor* L. *Journal of*  
347 *Advances in Biology & Biotechnology*, 27(7), 885–892.  
348 <https://doi.org/10.9734/jabb/2024/v27i71048>

349

350