

Original Research Article

Plant Based Remedies Against the Microbial Maladies with A Focus on Bacterial Wilt Caused by *Ralstonia solanacearum* in Ginger (*Zingiber officinale* Roscoe)

Abstract

Bacterial wilt caused by *Ralstonia solanacearum* (Smith) Yabuuchi & al. is one of the worst disease-causing pathogens to ginger (*Zingiber officinale* Roscoe) that incur severe crop loss and affect the economic security of the farmers. Management of the disease involves both traditional and modern scientific techniques that necessitate or demand the use of synthetic chemicals. In this connection, a study was conducted with certain endemic plants of the Western Ghats to unravel the ability of these plants to control bacterial wilt that affecting the ginger crop. The plant species selected were *Hydnocarpus macrocarpus* (Bedd.) Warb., *Colubrina travancorica* Bedd., *Cynometra beddomei* Prain, *Prioria pinnata* (Roxb. ex DC.) Breteler and *Cynometra travancorica* Bedd. The plant samples were collected from different areas of Western Ghat, particularly the Wayanad district of Kerala. Phytochemical investigations, both qualitative and quantitative means revealed that all of them possess chemical compounds that have potential therapeutic and antimicrobial properties. Studies to check the ability of these plants against *R. solanacearum* revealed that the *Prioria pinnata* has the right ability to inhibit bacterial growth. Further investigation showed that the alkaloid fractions of the plant extract showed the highest antimicrobial properties compared to that of phenols and terpenoid fractions. The GC-MS and FTIR studies indicates the presence of good amount of chemical compounds in the tree species of *Prioria pinnata*, possessing antimicrobial and therapeutic potential. The study points out the conservation of plant wealth of the Western Ghats that hides many biomolecules of clinical and industrial potential. The threat caused by climate change to plant wealth should be minimized by appropriate public and private partnership-oriented conservation programs.

Introduction

Ginger (*Zingiber officinale* Roscoe) can be considered as the confluence of spice and medicine. William Roscoe described this plant and gave the name *Zingiber* derived from Sanskrit. In India, the area under cultivation of Ginger is 63,000 ha and the production is 3 tons/ha. India

is the world's highest producer of Ginger with 1/3 of the production, followed by China, Nigeria, Nepal, and Indonesia (Priyanka & Khanal, 2021). Madhya Pradesh is the state with the largest production with 31.18% of the share (2021–22). Kerala ranks 9th with 2924 MT during 2021–22 and Wayanad is the district with the largest area under cultivation 1232ha (2021-22). Soft rot in ginger is caused by *Pythium aphanidermatum* (Edson) Fitzpn and it causes a crop loss of up to 50%. The Bacterial wilt is Caused by *Ralstonia solanacearum* (Smith) Yabuuchi & al. and it is called 'Mahali disease' and makes a yield drop of nearly 40%, however, in conducive conditions, the yield reduces up to 100% (Mathew et al., 1979; Dohroo, 1991). Ginger farming become a nightmare in Kerala state because of microbial diseases and the decline in the area under cultivation of ginger is a usual trend that is prevailing in districts like Wayanad. While looking into the area under cultivation of Ginger in the district, during the year 2001-2002, Ginger was cultivated in 10706 ha (2012-13), however, in 2021-22 it considerably come down to 2924 ha (Agriculture Statistics 2022–23). This indicates the percentage of reduction in area under cultivation was nearly 73%. In this context, an eco-friendly approach that can best control the microbial disease in need of the hour can better save the lives and livelihood of the Ginger farmers of the state and nation as well.

The present study focuses on plant-based remedies in the control of microbial diseases with a focus on bacterial wilt or Mahali disease caused by *Ralstonia solanacearum*. Five selected plants that are endemic to the Western Ghats were tested for their ability to control the bacterial wilt with an assumption that the phytochemicals present in the plants would have the right potential to check the growth of microorganisms that are pathogenic to crops. A detailed study could unravel the chemical constituents' quantity and mode of action in controlling the pathogen would be of greater use for farmers and ginger production as well.

Materials and Methods

Collection of the plant samples

The study was conducted at Wayanad district of Kerala and the plant samples were collected from evergreen patches of the Western Ghats that spread over the district. There were five plants were selected based on the survey in which the plants were used by the farmers of the

district to protect the Ginger from the attack of pests and diseases. The selected plants were identified after consulting the authentic specimens deposited at MH and the plants are *Hydnocarpus macrocarpus* (Bedd.) Warb., *Colubrina travancorica* Bedd., *Cynometra beddomei* Prain., *Cynometra travancorica* Bedd., and *Prioria pinnata* (Roxb. ex DC.) Breteler (\equiv *Kingiodendron pinnatum* (DC.) Harms). The samples are collected in a sterile polythene bag and taken into the laboratory. The samples were surface sterilized under tap water several times to remove residual contaminants followed by distilled water (Ahmad et al., 2009).

Microorganism Culture

Ralstonia solanacearum cultures were collected from the Indian Institute of Spices Research (IISR), Calicut. The microbial culture collection number was *Ralstonia solanacearum* IISR-GRS-SPR. The cultures were kept in nutrient agar plates and incubator at 28-32°C. The cultures were sub-cultured for the experiments in nutrient agar plates (Nnamdi et al., 2011; Sreena et al., 2013).

Qualitative and Quantitative Determination of Phytochemicals

Preliminary phytochemical analysis was carried out for the extract as per standard methods. The methanol extract of the sample was prepared by following the procedure mentioned by Pareksh et al., (2006) in which 10g of shade-dried powder was taken in 100ml methanol in a conical flask, plugged with cotton wool and then kept on a rotary shaker for 24 hours. After 24 hours, the methanol extract was centrifuged at 2000rpm for 10 minutes and the supernatant was collected and filtered by using Whatman no.1 filter paper. The solvent was evaporated to make the final volume which is one-fourth of the original volume and stored at 4°C in an airtight bottle.

The extracts were subjected to a qualitative chemical test for various phytoconstituents like carbohydrates (Sadasivam & Manickam, 1991); reducing sugars, phenols, tannins, phlobatannins, sterols and saponins (Raaman, 2006); flavonoids (Shamaki et al., 2012); alkaloids (Mayer' test & Wagner' test as described by Raaman 2006); glucosides (De et al., 2006) and terpenoids (Shamaki et al., 2010). The quantitative determination of alkaloids and phenolic compounds was taken up by adopting the methods of Dattatreya et al. (2020).

Antimicrobial Assay

A disc diffusion assay (Jaishree & Kumar, 2017) was performed to determine the antimicrobial activity of the selected five plant species. The test organism was grown in SDA plates diluted in distilled water (1×10^8) and seeded into the petri plates. The methanol extract of the selected five plant samples at varying concentrations was impregnated in Whatman no.1 filter paper in 6mm diameter. These discs were kept at 4°C for one hour to effectively diffuse the extracts to the media and thereafter the plates were incubated at 37°C for 24 hours. The inhibition zone (IZ) that formed around the disc after 24 hours was measured. The Activity Index (AI) was calculated by the formula

$$\text{Activity Index} = \frac{\text{Inhibition Zone of the Test}}{\text{Inhibition Zone of the Control}}$$

The experiment was repeated three times to reduce the errors and values were statistically assessed.

GC MS and FTIR

Gas Chromatography-Mass Spectroscopy (GC-MS)

The phytochemical analysis was performed using a combined 7890A gas chromatograph system (Agilent 19091-433HP, USA) and a mass spectrophotometer. The system was equipped with an HP-5 MS fused silica column (30.0 m \times 250 μ m, film thickness 0.25 μ m), and interfaced with a 5675C Inert MSD with a Triple-Axis detector. Helium gas was used as the carrier gas, adjusted to a column velocity flow of 1.0 ml/min. Plant extract that dissolved in Dichloroethane was taken for the GC-MS studies (Olivia et al., 2021).

The Fourier Transform Infrared Spectrometer (FTIR) assessment is one of the most important tools to assess the functional groups and bodies present in the given chemical samples. It relies on the principle that the light absorbed would be related to the characteristic of the chemical compound present in the sample. Hence, by interpreting the wavelength that was absorbed the chemical compound can be identified. The present study used the methodology described by Ashokkumar and Ramaswamy (2014). The instrument used was Shimadzu, IRAffinity1, Japan, with a Scan range from 400 to 4000 cm^{-1} with a resolution of 4 cm^{-1} .

Results

Phytochemical screening

The results of the phytochemical screening revealed that the selected plants from the Western Ghats possess different phytochemical components that may have therapeutic and antimicrobial properties. The outcomes of the qualitative experiments conducted indicate that all the selected plants showed positive results for most of the tests, however, negative results were shown in the case of sterols and saponins as given in Table 1. Phlobotannins showed positive results in *Colubrina travancorica* only and the rest of them showed negative results.

Table 1. Qualitative determination of the phytochemicals present in the selected five endemic plant species from the Western Ghats of Kerala

Test	Plant species Selected				
	<i>Hydnocarpus macrocarpus</i>	<i>Colubrina travancorica</i>	<i>Cynometra beddomei</i>	<i>Prioria pinnata</i>	<i>Cynometra travancorica</i>
Carbohydrates	+	+	+	+	+
Reducing sugar	+	+	+	+	+
Phenols	+	+	+	+	+
Tannins	+	+	+	+	+
Flavonoids	+	+	+	+	+
Alkaloids	+	+	+	+	+
Phlobotannins	-	+	-	-	-
Glycosides	+	+	+	+	+
Sterols	-	-	-	-	-
Saponins	-	-	-	-	-
Terpenoids	+	+	+	+	+

+ Present; - Absent

Determination of Phenolic Compounds

All the selected plants possess varying amounts of phenolic compounds (Table 2). The *Colubrina travancorica* 0.80 ± 0.00 showed the highest amounts of Tannic acid followed by *Prioria pinnata* with 0.57 ± 0.01 . The composition of flavonoids and phenols was found to be varying in all the selected plants.

Table 2. Determination of the Total Phenolic Compounds present in the selected plant species

Samples (mg/ml)	<i>Hydnocarpus macrocarpus</i>	<i>Colubrina travancorica</i>	<i>Cynometra beddomei</i>	<i>Prioria pinnata</i>	<i>Cynometra travancorica</i>
Flavonoids	0.14±0.04	0.15±0.005	0.10±0.006	0.15±0.02	0.14±0.02
Tanic Acids	0.07±0.009	0.80±0.00	0.075±00	0.57±0.01	0.08±00
Phenols	0.13±0.04	0.141±0.01	0.12±0.05	0.15±0	0.15±0

$n=3\pm$ standard deviation

Determination of Alkaloid Content

The result of the determination of Alkaloid content is provided in Table 3. *Colubrina travancorica* possesses the highest amount of alkaloid content with 0.95 ± 0.03 mg/ml followed by *Hydnocarpus macrocarpus* with 0.92 ± 0.02 . However, all the plants show the presence of alkaloids in varying concentrations.

Table 3: Determination of the total alkaloid composition in selected plant species.

Samples	<i>Hydnocarpus macrocarpus</i>	<i>Colubrina travancorica</i>	<i>Cynometra beddomei</i>	<i>Prioria pinnata</i>	<i>Cynometra travancorica</i>
Alkaloid Content (mg/ml)	0.92±0.02	0.95±0.03	0.89±0.02	0.90±0.01	0.87±0.04

$n=3\pm$ standard deviation

Antimicrobial Assay

Varying concentrations of the plant extract such as 5%, 10%, and 15% were taken for the antimicrobial activity assessment against the pathogen *R. solanacearum* and the results are given in Table 4. However, as the table indicates, none of the plants in any of the selected concentrations showed any impact on the growth of *R. solanacearum*. From the Table 4 it is clear that only *Prioria pinnata* in all the selected concentrations of 20%, 40% and 60% showed inhibitory effects against the selected organism *R. solanacearum*. None of the other plants showed any significant inhibition against the pathogen.

Table 4 Antimicrobial activity of selected five plant species against *Ralstonia solanacearum*

Sl. No	Plant extract	Concentration (%)	Inhibition (mm)
--------	---------------	-------------------	-----------------

1.	<i>Hydnocarpus macrocarpus</i>	5	No zone
		10	No zone
		15	No zone
		20	No zone
		40	3
		60	10
2.	<i>Prioria pinnata</i>	5	No zone
		10	No zone
		15	No zone
		20	12
		40	17
		60	20
3.	<i>Colubrina travancorica</i>	5	No zone
		10	No zone
		15	No zone
		20	No zone
		40	No zone
		60	No zone
4.	<i>Cynometra travancorica</i>	5	No zone
		10	No zone
		15	No zone
		20	No zone
		40	No zone
		60	7
5.	<i>Cynometra beddomei</i>	5	No zone
		10	No zone
		15	No zone
		20	6
		40	7
		60	9

Antimicrobial activity index

The antimicrobial activity index of phenolics, alkaloid and terpenoid fractions of *P. pinnata* against *R. solanacearum* is given in Table 5. From Table 5 it is clear that the alkaloid fractions of the *P. pinnata* showed the highest inhibitory actions against *R. solanacearum*. All samples were taken with 20% concentration in which the highest activity index was shown by the alkaloid fraction followed by phenolics and terpenoids.

Table 5 Antimicrobial activity index of phenolics, alkaloid and terpenoid fractions of *Prioria pinnata* against *Ralstonia solanacearum*

Sl. No.	Extracts	Concentration (%)	Diameter of zone of inhibition (mm)	Activity index
1.	Alkaloids	20	5	0.15

			6	0.18
			5	0.15
2.	Phenolic Compounds	20	4	0.125
			3	0.09
			4	0.125
3.	Terpenoids	20	3	0.09
			2	0.06
			2	0.06

The minimum inhibitory concentration (MIC) of alkaloid extracts against *R. solanacearum* provided in Table 6 indicates that the increasing concentration of the alkaloid fraction of the plant *P. pinnata* is having a positive impact on the growth of the selected pathogen *Ralstonia solanacearum*. The highest concentration taken was 30% and that showed an inhibition of 6mm in the growth of the pathogen followed by 25% with 4mm and 20% with 3.5mm.

Table 6 Minimum inhibitory concentration (MIC) of alkaloid extracts against *Ralstonia solanacearum*

Extract	Concentration in %	Diameter of zone of inhibition (mm)
Alkaloids	5	No zone
	10	2
	15	3
	20	3.5
	25	4
	30	6

Table 7 shows that the components that have a high retention time were 2-Tert-Butyl-4,6-Bis(3,5-Ditert-Butyl-4-HydroxyBenzyl) Phenol with a value of 45.21% followed by Vitamin E with a retention time of 39.6%. Other compounds such as Tetrakis (2,3- Ditert- Butylphenyl)-4,4'Biphenylene Diphosphonate and Beta- Sitosterol also showed good retention values. Figure 1 is the GCMS spectrum of the methanolic extract of the *P. pinnata*.

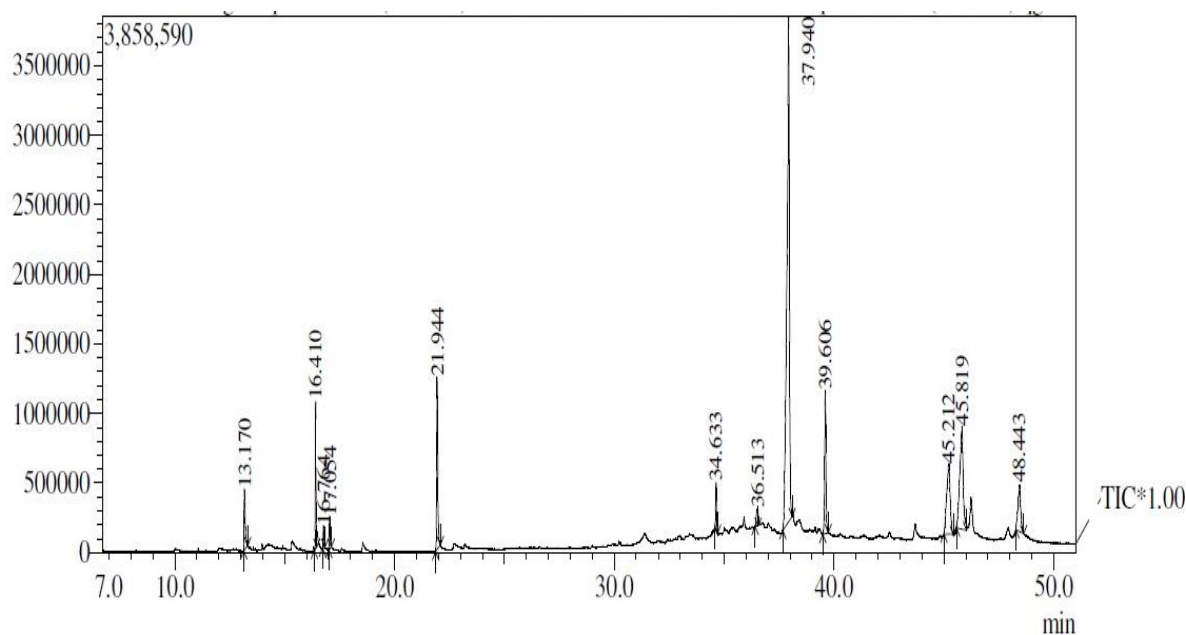


Fig 1 GCMS Spectrum of the methanol extract of *Prioria pinnata*

Table 7 Major Compounds Present in *Prioria pinnata*

Peak	R. Time	Area	Area%	Height	Height%	Name	Base m/z
1	13.170	1499705	2.48	438333	4.46	3,5'-Dimethoxyacetophenone	180.10
2	16.410	2421680	4.00	1057913	10.77	Neophytadiene	68.05
3	16.764	396647	0.65	159747	1.63	3,7,11,15- Tetramethyl -2-Hexadecen-1-ol	82.05
4	17.054	597666	0.99	228600	2.33	Phytol, Acetate	71.05
5	21.944	4454073	7.35	1235878	12.58	Isophytol, Acetate	71.05
6	34.633	919434	1.52	342050	3.48	Squalene	69.05
7	36.513	660935	1.09	126938	1.29	Beta- Sitosterol	55.05
8	37.940	28790903	47.54	3633482	36.98	Tetrakis (2,3- Ditert-Butylphenyl)-4,4'Biphenylene Diphosphonate	57.10

9	39.606	4349973	7.18	1022509	10.41	Vitamin E	165.10
10	45.212	5504834	9.09	501934	5.11	2-Tert-Butyl-4,6-Bis(3,5-Ditert-Butyl-4-HydroxyBenzyl) Phenol	57.10

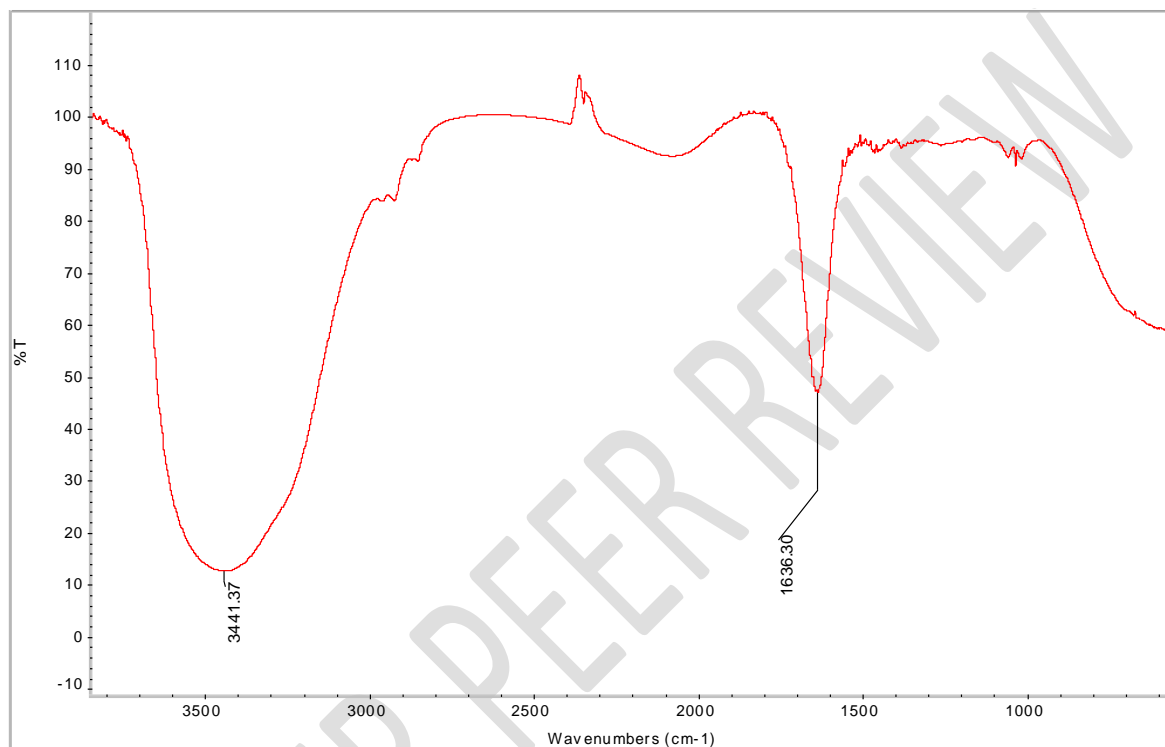


Fig 2 FTIR- Spectrum of an alkaloid fraction of *Prioria pinnata*

Table 8 indicates the presence of O-H stretch, free hydroxy bonds that may be indicative of the alcohols and phenolic compounds present in the sample and -C=C- stretch bonds that are indicative of Alkenes in the samples. Figure 2 shows the peaks of the representing functional groups present in the sample.

Table 8 FTIR spectral peak values and functional groups obtained for the leaf extract in Methanol solvent

Extract Prepared In	Peal Values	Bonds	Functional Groups

Methanol (ME)	3441.37	O-H Stretch, free hydroxy	Alcohols and Phenols
	1636.30	-C=C- stretch	Alkenes

Discussion

Ralstonia solanacearum is one of the dreadful pathogens that affect crops around the world and is considered as one of the major threats to ginger cultivation (Nair, 2013). *R. solanacearum* naturally inhabits soil, plants, and animals and has wide host ranges. Control *R. solanacearum* is a difficult task since it has multiple host ranges, long survival in the soil, a wide mechanism for spreading, ability to associate with weeds in the cropland and grow endophytically (Sarkar & Choudhari, 2016). Attempts to control the pathogen include cultural, biological, chemical or even physical means (Cordoso et al., 2006). The present study revealed that among the five selected plant species only *P. pinnata* showed inhibitory effects that start with a concentration of 20%. While looking at the chemical components that inhibit the growth it can be noticed that the alkaloid fractions of the leaf extract showed maximum inhibitory effects compared to the phenols and terpenoids. Values of the minimum inhibitory concentrations (MIC) of the alkaloid fractions further revealed that a concentration of above 5% only showed inhibitory properties. However, the treatment with alkaloid fractions with a concentration of more than 10% showed positive results with 2mm and a further increase in inhibition showed with an escalation in concentrations. Earlier research attempts with *P. pinnata* also showed similar results as reported by earlier research attempts (Sheik & Chandrashekar, 2014; Baheti et al., 2020). The dreadful pathogen can be controlled with leaf extracts of *P. pinnata* and a focused attempt to reveal what are the chemical constituents present in the alkaloid fractions is essential.

Antimicrobial compounds derived from plants have got wider attention since they can be used against drug-resistant microorganisms and other resistant varieties of microbes. Alkaloids are naturally found in plant kingdoms that proved to have anticancer, antiviral, anti-inflammatory, and antimicrobial properties (Liu et al., 2020). Many drugs have developed out of alkaloids and one of the best examples to cite is morphine- the narcotic analgesic (Rodrigues et al., 2020). The present study also corroborates that the alkaloids from the plants have antimicrobial properties and could best inhibit the most dreadful pathogen *R. solanacearum* that affecting ginger cultivation. An alkaloid fraction of 10% can rightly check the growth of tested microorganisms and also reveal that an increase in the testing concentration could control further microbial growth. The importance of the plant wealth in the Western Ghats is one of the areas of concern and it was reported that out of the 4000 species of flowering plants reported

1500 species belong to the endemic category. These plant species should be studied in detail to unravel the chemical compounds present in them, which may lead to the development of novel biomolecules that have therapeutic values and disease-prevention abilities both in plant and animal kingdoms.

Conclusion

Bacterial wilt caused by *Ralstonia solanacearum* is one of the dreadful diseases that cause significant crop loss to the farmers of the Kerala state especially the Wayanad district. Farmers are managing to control this disease with cultural, physical, chemical, and biological means. The ability of five endemic plant species from the Western Ghats has been tested in the control of *R. solanacearum*. Out of the five plant species, the *Prioria pinnata* found to be control the growth of the tested microorganism *R. Solanacearum*. Further investigation revealed that among the alkaloid, phenolics and terpenoid fractions of plant extract, an alkaloid portion could rightly control the microorganism. Detailed studies using GC-MS and FTIR further revealed that the plant *P. pinnata* possesses a series of chemical compounds that have antimicrobial properties and therapeutic values. The FTIR study revealed that the plant possesses functional groups that belong to alkaloids, phenols and alkenes. The study throws open light into the importance of chemical compounds that may hidden in the plant kingdom of Western Ghats the unique mega biodiverse hot spot of the world.

References

- Agriculture Statistics (2022-23). Department of Economics and Statistics, Government of Kerala. 50-105.
- Ahmad M., Noreen A., Abdul M., Tanveer S., Muhammad Z.C. Alamgeer and A. Bashir (2009). Effects of aqueous methanol extract of *Albizia lebbek* Benth. Seeds on various biochemical parameters in Alloxan-induced diabetic rabbits. *Pharmacology online*, 1: 134-143.
- Ashokkumar R. and Ramaswamy M. (2014). Phytochemical screening by FTIR spectroscopic analysis of leaf extracts of selected Indian medicinal plants. *International Journal of Current Microbiology and Applied Sciences*, 3(1): 395-406.

- Baheti D., Kumbhar and Prasad (2020). Exploration of Ayurveda potential in tuberculosis: Current scenario and Future prospects. *International Journal of Ayurveda and Pharma Research*, 8(5): 19-32.
- Cardoso S. C, Soares A.C. F, Brito A.D.S., Laranjeira F.F, Ledo C.A.S. and A.P. Dos Santos (2006). Control of tomato bacterial wilt through the incorporation of aerial part of pigeon pea and crotalaria to the soil. *Summary of Phytopathology*, 32:27-33.
- Dattatray S., Gamit Rakesh., Shukla V. J., and Rabinarayan Acharya. (2020). Quantification of total alkaloid, tannin, flavonoid, phenolic, and chlorogenic acid contents of *Leea macrophylla* Roxb. ex Hornem. *International Journal of Green Pharmacy*. 14 (2) 138.
- De S., Dey Y.N and Ghosh A. K. (2010). Phytochemical investigation and chromatographic evaluation of different extracts of tuber of *Amorphaphallus paeoniifolius* (Araceae). *International Journal on Pharmaceutical and Biomedical Research*, 1(5):150-157.
- Dohroo N.P. (1991). New record of bacterial wilt of ginger in Himachal Pradesh. IPS North Zone Meet. April 29-30, pp 16 (Abstract)
- Jaishree S. and P. Kumar (2017). Comparative study of antimicrobial activity and phytochemical screening of serial extracts from leaves and fruit of *Aegle marmelos* and *Carica papaya*. *International Journal of Pharmacology and Pharmacological Sciences* 9 (12), 119-123
- Liu Y., Cui Y., Lu L., Gong Y., Han W., Piao G. (2020). Natural Indole-containing alkaloids and their antibacterial activities. *Archives of Pharmacology*. 353-361
- Mathew, J., Koshy A. Indrasenan, and M. Samuel (1979). A new record of bacterial wilt of ginger infected by *Pseudomonas solanacearum* E. F. Smith from India. *Current Science*. 48: 213-214.
- Nair K. P. (2013). The agronomy and economy of turmeric and ginger: The invaluable medicinal spice crops. Elsevier, London.61-77.
- Nnamdi L. Obasi, Madus., Ejikeme P., Cemaluk A., and C. Egbuonu (2011). Antimicrobial and phytochemical activity of methanolic extract and its fractions of *Jatropha curcas* Linn. (Euphorbiaceae) stem bark. *African Journal of Pure and Applied Chemistry*, 5(5): 92-96.

Olivia N.U., Goodness, U.C. and O.M. Obinna (2021). Phytochemical profiling and GC-MS analysis of aqueous methanol fraction of *Hibiscus asper* leaves. *Future Journal of Pharmaceutical Sciences* 7: 59-64.

Parekh J., Nehal K. and S. Chandra (2006). Evaluation of antibacterial activity and phytochemical analysis of *Bauhinia variegata* L. bark., *African Journal Biomedical Research*, 9: 53-56.

Priyanka J. and S. Khanal (2021). Production status, export analysis, and future prospects of ginger in Nepal. *Archives of Agriculture and Environmental Science*, 6(2): 202-209.

Raaman N. (2006). Phytochemical Techniques. In. New Indian Publishing- Botanical Chemistry New Delhi. 19-24

Rodrigues S., Shin D., Conway M., Smulski S., Trenker E., Shanthanna H., Vanniyasingam T., Thabane L., and Paul J. (2020) Hydromorphone versus morphine: A historical cohort study to evaluate the quality of postoperative analgesia. *Canadian Journal of Anaesthesia*, 68:226–234.

Sadasivam S. and Manickam A. (1991). Biochemical Methods 2nd edition, Scientific publishers. 6-194.

Sarkar S., and S. Chaudhuri (2016). Bacterial wilt and its management. *Current Science*.110(8):1439-1445.

Shamaki B.U., Geidam Y.A., Abdulrahma F., Ogbe A.O. and U.K. Aandabe (2012). Evaluation of phytochemical constituents and *in vitro* antibacterial activity of organic solvent fractions *Ganoderma lucidum* methanolic extract. *International Journal of Medicinal Plant Research* 1(3). 26-31.

Sheik and Chandrashekar (2014). Antimicrobial and antioxidant activities of *Kingiodendron pinnatum* (DC.) Harms and *Humboldtia brunonis* Wallich: endemic plants of the Western Ghats of India. *Journal of National Science Foundation of Sri Lanka*. 42(4): 307-313.

Sreena R., Manon B., Merlene A. B., Karthikeyan S., and K M. Gothandam (2013). Antioxidant, antimicrobial and antiproliferative activity and phytochemical analysis of selected medicinal plant Dasapushpam of Kerala. *International Journal of Pharmaceutical Science Review and Research* 23(1):172-179.