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2 **A nanotoxicological approach to the effects of**
3 **metformin and sodium metavanadate co-**
4 **encapsulated in polycaprolactone**
5 **nanoparticles under the biological parameters**
6 **of zebrafish (*Danio rerio*)**
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11
12 **ABSTRACT**
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The present study evaluated the sublethal and lethal effects, and heart rate of *Danio rerio* when exposed to antidiabetic drugs, metformin (M-10), and sodium metavanadate (V-10), their association (MV-10) and the polycaprolactone nanoparticles containing poloxamer 188 with (PPMV-1, PPMV-5, and PPMV-10) or without these antidiabetics (PP-10). Acute toxicity tests were carried out to evaluate these effects. Groups V-10, MV-10, PPMV-5, PPMV-10, and PP-10 had lethal and sublethal, such as effects as pericardial edema. Concerning the heart rate, the PPMV-1, PPMV-5, PPMV-10, and PP-10 groups had a reduction compared to the other groups, indicating toxicity of the constituents of the nanoparticles. PPMV-5, PPMV-10, and PP-10 groups had sublethal and lethal effects depending on the concentration. Antidiabetics were eliminated as a possible cause and the poloxamer 188 is non-toxic at the concentrations used. In the PP-10 group, there was a dynamic of sublethal and lethal effects like the PPMV-10 group. We conclude that the presence of PCL in the formulation of nanoparticles was harmful at the cardiovascular level in embryos and *eleuthero-embryos* of *D. rerio*, affecting their development and heart rate, regardless of their concentration. Because of the results obtained, we can conclude that the toxicological evaluation of a nanomaterial is important to anticipate problems in its veterinary application since such results show possible toxicity problems.

14
15 *Keywords: PCL, toxicology, animal model, nanomaterial, antidiabetics.*
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18 **1. INTRODUCTION**
19

20 Nanotechnology currently offers significant contributions with applications to animal
21 production and veterinary medicine, aiding in the prevention and treatment of diseases and
22 applying new therapies that increase longevity and improve the quality of life of animals (Bai
23 et al., 2018). About 25,000 patents and 2,000 articles on the application of nanotechnology
24 in veterinary medicine have been filed and published to date (Lens, 2021), indicating a
25 growing interest in the application of nanotechnology in this field. Among the areas of
26 nanotechnology activity, there is research in the development of nanomaterials that have
27 useful chemical and physical properties, which have attracted the attention of researchers.
28 These nanomaterials have been widely explored for the controlled release of drugs and can
29 be of different types such as metal-based, carbon-based, and polymeric. The latter are
30 colloidal particles with a size between 10-500 nm. They are considered to have high

31 biological safety and biodegradability, increasing the stability of antigens and drugs and
32 improving bioavailability. However, due to this small size, it is necessary to consider the risk
33 of accumulation of nanomaterials in different tissues (Han et al., 2018; Bueno, 2020).
34 Therefore, it is important to assess the toxicity of polymeric nanomaterials such as those
35 used in the production of nanoparticles (NPs), as well as the pharmaceutical excipients of
36 these formulations (Hering et al., 2020). The toxicity of nanoparticles is defined by the field of
37 nanotoxicology, which consists of understanding the interaction of nanoparticles with
38 organisms in various ways, such as the interrelationship with fluids and tissues that can
39 cause changes in cardiac functions or binding to mediators that can activate inflammatory
40 responses (Tiple et al., 2020).

41 In the field of veterinary medicine, polycaprolactone (PCL) is a biocompatible and
42 biodegradable polymer, approved by the *Food and Drug Administration*. It can be used in the
43 formulation of nanoparticles. Its applications are diverse, such as in the encapsulation of
44 antifungal, antiviral, and antiparasitic agents (Irache et al., 2011). Several surfactants are
45 used in the production of nanoparticles, such as poloxamer 188. This surfactant is a
46 hydrophilic triblock copolymer, approved by the *Food and Drug Administration* for oral and
47 intravenous administration, being used as a pharmacological membrane stabilizer and
48 rheological agent. It is efficient in repairing muscle injuries and reduces inflammation in
49 rabbits (Cadichon et al., 2007).

50 In the veterinary field, nanoparticles are being used to increase the quality of treatment of
51 patients. Several drugs, such as praziquantel, used in the treatment of fungi infection, and
52 plant compounds, such as 4-nerolidylcatechol (Cheng et al., 2010; Greatti et al., 2020), have
53 already been encapsulated in PCL nanoparticles as examples of veterinary products.
54 Diabetes mellitus is a common disease in cats and dogs with an incidence that is increasing
55 over the years (Hoening, 2002). Vanadium and metformin, drugs used to treat diabetes, were
56 not yet co-encapsulated in the proposition of nanotechnological products for veterinary use.
57 The first is a transitional element with several valence states (-3, -1, 0, and +1 to +5) and in
58 recent decades this trace element has been reported to have insulin-mimetic/enhancer
59 effects in orally administered diabetic patients. Due to this characteristic, vanadium was
60 used in the stabilization of metabolic parameters of periparturient dairy cows (Heidari et al.,
61 2016). The latter, metformin, on the other hand, is a drug of the biguanide class that acts to
62 reduce hepatic glucose production and insulin resistance of peripheral tissues, thus being
63 considered an insulin-sensitizing drug. It is used in the treatment of feline diabetes when
64 these animals have a functional reserve of β -pancreatic cells, as well as in dogs affected by
65 hyperadrenocorticism (Miceli et al., 2018). Evaluating the toxicity of metformin, the main drug
66 in the treatment of diabetes, and vanadium, a new therapeutic alternative, is essential to
67 ensure safety and efficacy in clinical use.

68 The zebrafish (*Danio rerio*) stands out as a widely used model organism for the rapid and
69 economical evaluation of the safety and efficacy of new compounds for animal health,
70 including nanopharmaceuticals. This model gains even more importance in a context in
71 which more than a thousand new substances, such as vaccines, medicines, food additives,
72 and agrochemicals, are introduced to the market annually (Böhme et al., 2017; Fukushima et
73 al., 2020). Due to attributes such as genetic, anatomical (kidneys, brain, liver, intestine,
74 heart, spine, eyes, mouth, ears, etc.), and physiological homology to mammals, external
75 fertilization, high number of descendants through reproduction, embryo transparency, small
76 size, and rapid development facilitate large-scale phenotypic approaches while maintaining
77 the capacity to respond to the 3Rs (Macrae; Peterson, 2015). Therefore, in this work we
78 propose the evaluation of the toxicity of PCL nanoparticles containing co-encapsulated
79 metformin and vanadium and the constituents used in the production of these nanoparticles,
80 evaluating their effects on the development of embryos and eleuthero-embryos of *D. rerio*.

81

82 2. MATERIAL AND METHODS

83

84 2.1 Zebrafish husbandry and crossing

85 The experiments were carried out at the *Laboratório de Ecofisiologia e Comportamento*
86 *Animal* – LECA of the *Universidade Federal Rural de Pernambuco* – UFRPE, a vivarium
87 registered in the CIUCA-CONCEA Platform with a certificate of regularity issued by the
88 regional council of veterinary medicine to perform tests with aquatic animals. All tests
89 involving animals were previously approved by the Ethics Committee for Animal Use
90 (License nº 071/2019). Adult wild-type fish (1 year) were bred and housed in 80 L
91 aquariums, where they were quarantined to detect or confirm the absence of pathogens or
92 diseases. They were housed under the following laboratory conditions: artificial aeration of
93 11 mg/L DO, temperature of 25 ± 1 °C, pH 7.5 ± 0.5 , and 14/10 h cycle (light/dark). The
94 water was partially renewed once a week. Abiotic parameters such as dissolved oxygen,
95 ammonia, nitrite, and nitrate were also measured and maintained within ideal ranges
96 (Gomes *et al.*, 2024). The animals were fed three times a day, 2x with Fort Color® fish food
97 (30% crude protein) and 1x with live brine shrimp nauplii (*Artemia* ssp). To obtain the
98 embryos, the adult animals were separated in a 2:1 male-to-female ratio (OECD 236, 2013)
99 and placed in spawning tanks (Alesco® Zebclean, Monte Mor, Brazil) for reproduction. Thirty
100 minutes after the start of spawning, the eggs were collected, with the removal of unfertilized
101 ones. Fertilized eggs (with normal blastula development) (OECD 236, 2013) were washed
102 with distilled water and randomly transferred for exposure in sterile polystyrene pots (80 mL)
103 and kept in an incubator at a controlled temperature (27 ± 1 °C) and photoperiod (14/10h
104 light/dark).

105 2.2 Preparation and characterization of nanoparticles

106 The concentrations of the compounds PCL, poloxamer 188, metformin, and sodium
107 metavanadate used in the production of nanoparticles whose toxicity was evaluated, are
108 presented in Table 1. To obtain the NPs, the technique of deposition of performed polymers
109 was used, where the PCL was dissolved in acetone and dichloromethane and heated at 30°
110 C for 5 minutes for total dissolution, forming the organic phase. This solution was then
111 poured into an aqueous phase containing phosphate buffer (0.1 M, pH 7.4), metformin,
112 sodium methavanadate, and poloxamer 188. The mixture was kept under magnetic stirring
113 with a relative centrifugal force of 21 *g* for total evaporation of organic solvents for 24 hours
114 (patent BR 102020020499). To produce the white nanoparticle, there was no addition of
115 metformin and/or sodium metavanadate. The waste generated during the experiment
116 underwent treatment in an advanced oxidative process in a reactor using UV photo-oxidation
117 and H₂O₂ before disposal (Wolset *et al.*, 2013). The average size (nm), polydispersity index
118 (PDI), and zeta potential (mV) of the nanoparticles were evaluated using the standard
119 photon correlation spectroscopy (PCS) technique fixed at 90° to 25 °C using a Zetasizer
120 Nano ZS (Malvern, UK) (Cadena *et al.*, 2013; dos Santos Magnabosco *et al.*, 2022). The
121 analysis was performed at the *Laboratórios Associados em Rede de Nanotecnologia*
122 (LARnano) of the *Universidade Federal de Pernambuco* (UFPE). The data were measured
123 in triplicate.

124 Table 1. Concentrations of the compounds used in the experiments. Acronyms: C – Control;
 125 V – Sodium metavanadate; M – Metformin; MV – Sodium metavanadate associated with
 126 Metformin; PP – Polycaprolactone Nanoparticles containing Poloxamer 188; PPMV -
 127 Polycaprolactone nanoparticles containing Poloxamer 188 with association of co-
 128 encapsulated sodium methavanadate and metformin. Legends: * - Xu *et al.*, 2003; # - Kumar
 129 *et al.*, 2017; ## - Mazzarino *et al.*, 2012.

Group Compound	C	V-10	M-10	MV-10	PP-10	PPMV-1	PPVM-5	PPMV-10
Sodium Metavanadate*	0	0.025 mg/mL	0	0.025 mg/mL	0	0.0025 mg/mL	0.0125 mg/mL	0.025 mg/mL
Metformin#	0	0	1,875 mg/mL	1,875 mg/mL	0	0.1875 mg/mL	0.9375 mg/mL	1,875 mg/mL
Polycaprolactone##	0	0	0	0	0.313 mg/mL	0.031 mg/mL	0.156 mg/mL	0.313 mg/mL
Poloxamer 188##	0	0	0	0	0.0125 mg/mL	0.000125 mg/mL	0.00625 mg/mL	0.0125 mg/mL

130

131 2.3 Toxicity test

132 For the evaluation of the toxicity of the nanoparticles, the development of embryos and
 133 *eleuthero-embryos* of *D. rerio* was evaluated at 24, 48, 72, and 96hpf. With the aid of an
 134 optical microscope (400x, 1000x), pictures were taken of the embryos and *eleuthero-*
 135 *embryos* for the identification of possible lethal and sublethal effects (Lammer *et al.*, 2009;
 136 Bittencourt *et al.*, 2018; Cadena *et al.*, 2020). Also, for lethal effects, mortality was observed
 137 daily. Mortality was determined as coagulation, tail not detached, no somite formation,
 138 absence of heartbeat, and/or lack of hatching (OECD 236, 2013). In addition to the
 139 assessment of sublethal effects, heart rate was measured by manually counting heartbeats
 140 (Bittencourt *et al.*, 2018). Pigmentation Reduction (ReP), Pericardial Edema (PE), Yolk Sac
 141 Edema (YSE), and Spine Deformation (SDef) were also evaluated (Lammer *et al.*, 2009;
 142 Bittencourt *et al.*, 2018). The lethal and sublethal effects observed were first analyzed by
 143 dichotomous response (presence and absence), considering the affected animal when it had
 144 at least one lethal or sublethal effect (Cadena *et al.*, 2020). This response was analyzed by
 145 its relative frequency, and its data was presented in the frequency of affected animals (%).

146 2.4 Statistical analysis

147 These results were analyzed by *one-way* ANOVA followed by Tukey's test ($p < 0.05$). Also,
 148 as validation criteria, the animals in the control group did not present any of the sublethal
 149 effects analyzed, and the mortality of the group did not exceed 10% (OECD 236, 2013). For
 150 heart rate, the mean and standard deviation of each group were determined, and the results
 151 were analyzed by *two-way* ANOVA followed by Tukey's test ($p < 0.05$). Statistical analyses
 152 were performed using the *Origin Pro Academic* 2015 software (Origin Lab. Northampton,
 153 MA, USA).

154 **3. RESULTS AND DISCUSSION**

155

156 **3.1 Results**

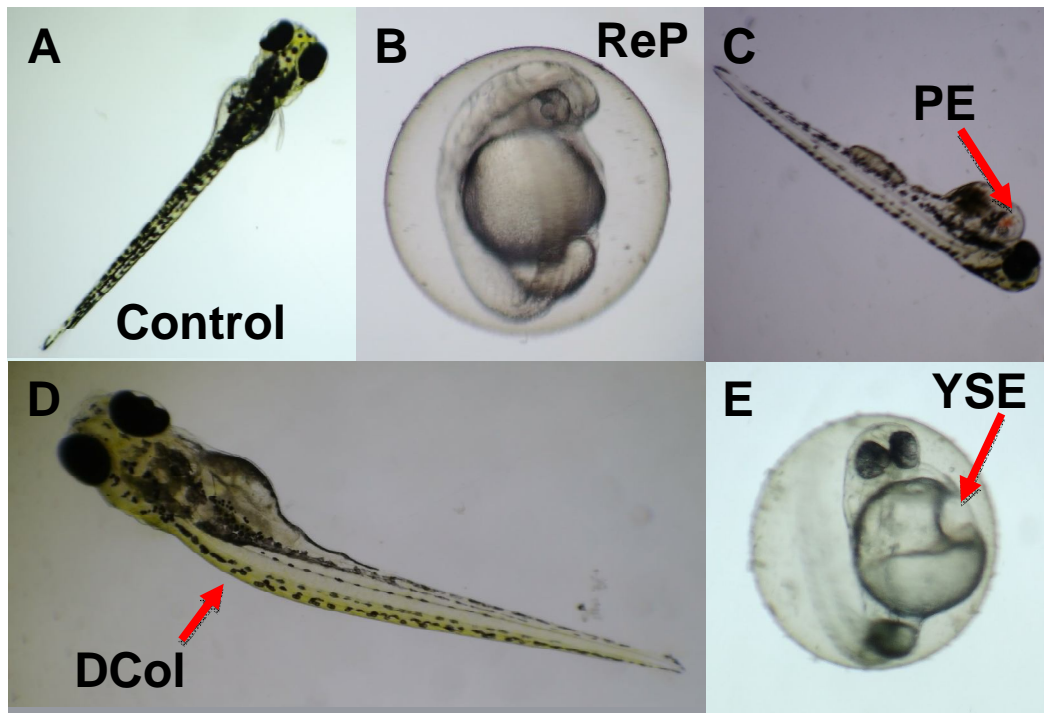
157

158 **3.1.1 Physicochemical characterization of nanoparticles**

159 The results of the physicochemical characterization show nanoparticles with an average size
160 between 165.3 and 341.07 nm, polydispersity index (PDI) between 0.025 and 0.198, and
161 zeta potential (ζ) between -7.83 to -3.03 mV.

162 **3.1.2 Effects of metformin, sodium metavanadate, white PCL nanoparticles, and**
163 **different concentrations of antidiabetic drugs co-encapsulated in PCL on zebrafish**
164 **embryonic development**

165 Examples of typical sublethal effects presented in all experimental groups can be seen in
166 Fig. 1. V-10 and MV-10 induced pericardial and yolk sac edema and spinal deformation. PP
167 induced a reduction of pigmentation and pericardial edema. PPMV-5 induced spine
168 deformation. PPMV-10 induced yolk sac edema.

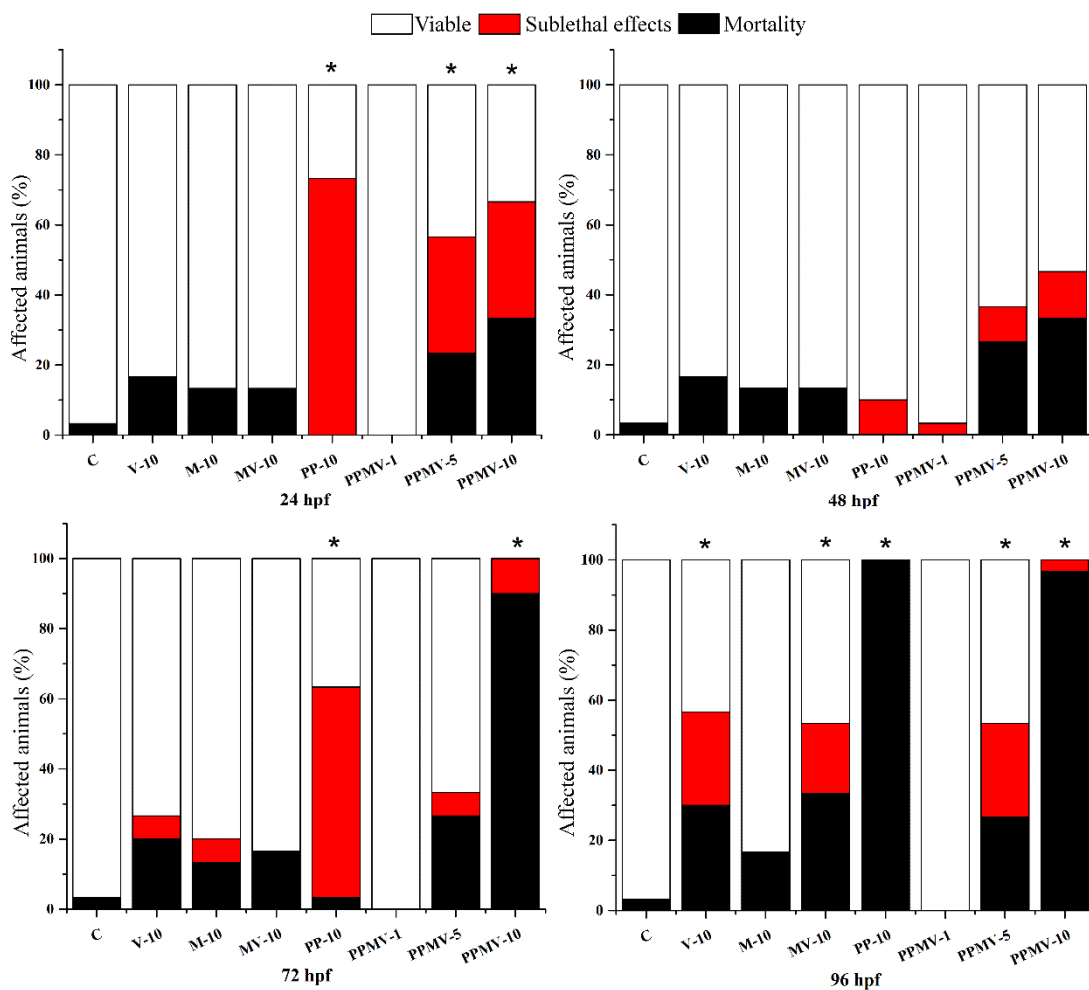


169

170 **Fig. 1. Typical sublethal effects were observed by exposure to white polycaprolactone**
171 **(PCL) nanoparticles and the different concentrations of metformin and sodium**
172 **metavanadate co-encapsulated in PCL nanoparticles in the period between 24-96 hpf.**
173 **Legend: A – Group C in dorsal view; B and C – Sublethal effects in the PP group; D –**
174 **Sublethal effect in PPMV-5; E – Sublethal effect in PPMV-10. Acronyms: ReP –**
175 **reduction of pigmentation; PE – pericardial edema; SDef – spine deformation; YSE –**
176 **yolk sac edema.**

177

178 Embryos and *eleuthero-embryos* were exposed to free antidiabetics, white PCL
 179 nanoparticles, and different concentrations of antidiabetic drugs co-encapsulated in PCL
 180 nanoparticles to evaluate the lethal and sublethal effects. The results are presented in Fig. 2.
 181 V-10 and MV-10 caused sublethal and lethal effects at 96hpf. PP-10 caused sublethal
 182 effects at 24 and 72hpf and lethal effects at 96hpf. PPMV-5 caused sublethal effects at
 183 48hpf and lethal effects at 96hpf. PPMV-10 caused sublethal and lethal effects at 24, 72,
 184 and 96hpf, with mostly lethal effects at 96hpf.



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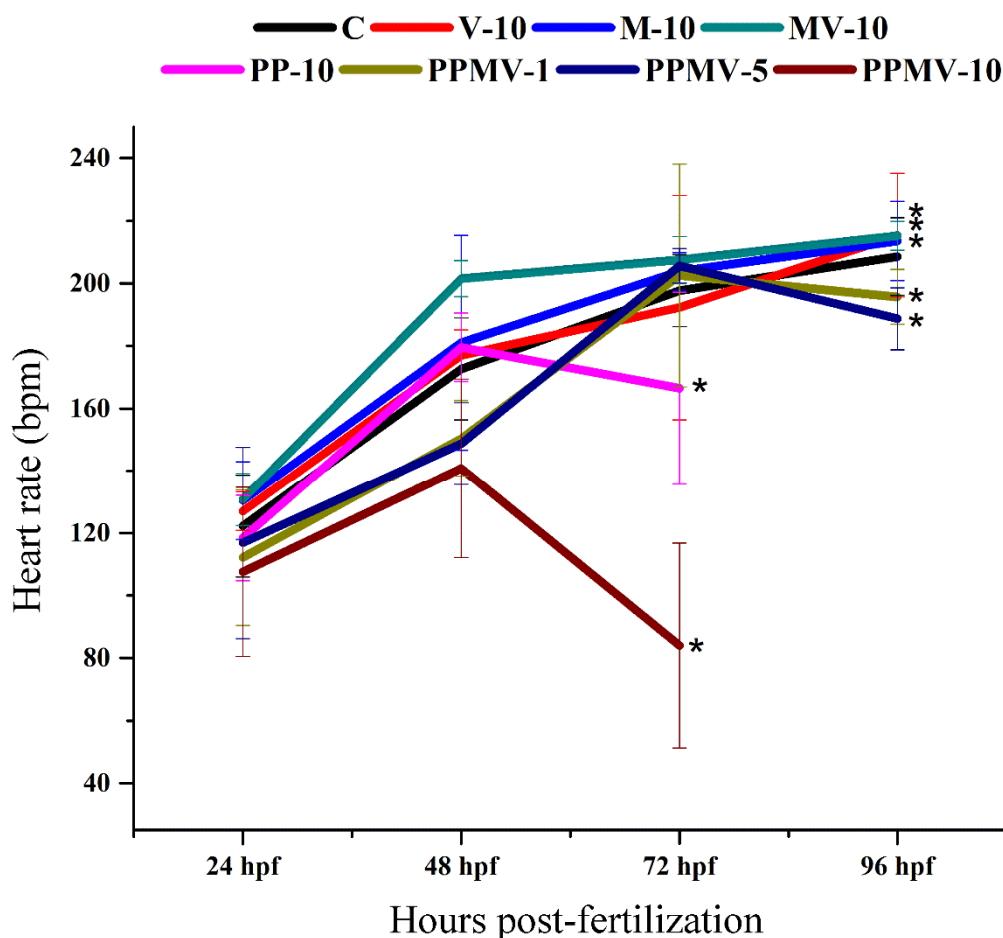
186 **Fig. 2.** Frequency of affected animals in the period of 24-96 hpf after exposure to free
 187 antidiabetic drugs metformin and sodium metavanadate, white polycaprolactone
 188 (PCL) nanoparticles, and the different concentrations of metformin and sodium
 189 metavanadate co-encapsulated in PCL nanoparticles. The increase in the frequency of
 190 affected animals was considered significant when * = $p < 0.05$ by Tukey's test in
 191 relation to the Control group. Each experimental group was compared with the control
 192 group by *one-way* ANOVA in the periods of 24 hpf ($F(7.23) = 12.41$, $p < 0.001$), 48 hpf
 193 ($F(7.23) = 2.62$, $p = 0.05$), 72 hpf ($F(7.23) = 18.36$, $p < 0.001$) and 96 hpf ($F(7.23) = 23.68$,
 194 $p < 0.001$). Acronyms: C – Control; V – Sodium metavanadate; M – Metformin; MV –
 195 Sodium metavanadate associated with Metformin; PP – Polycaprolactone
 196 Nanoparticles containing Poloxamer 188; PPMV - Polycaprolactone nanoparticles

197 containing Poloxamer 188 with association of co-encapsulated sodium
198 methavanadate and metformin.
199

200 **3.1.3 Effects of metformin, sodium metavanadate, white PCL nanoparticles, and**
201 **different concentrations of antidiabetic drugs co-encapsulated in PCL on zebrafish's**
202 **heart rate**

203

204 In relation to the effect of nanoparticles on heart rate, the results can be seen in Fig. 3. An
205 increase in heart rate is expected over time, as it can be seen in the Group C. This was also
206 observed in the V-10 and M-10 groups, indicating that these isolated compounds did not
207 affect the heart rate of the animals. However, in the MV-10 group, there was an increase in
208 heart rate, indicating that the interaction between the compounds can cause some toxicity. In
209 the PPMV-1 and PPMV-5 groups, there was an increase in heart rate in relation to time up to
210 72 hpf, with a decrease of 96 hpf. In the PPMV-10 and PP-10 groups, a reduction in heart
211 rate occurred from 72 hpf. When compared with each other, the reduction in heart rate in the
212 PPMV-10 group is significant in relation to the PP-10 group ($p = 0.0001$) at 48 hpf (Fig. 3),
213 with PPMV10 being the most toxic among all.
214



215

216 Fig. 3. Comparison of the heart rate of the groups in the period of 24-96 hpf after
217 exposure to the antidiabetic drugs metformin and sodium metavanadate, the white
218 polycaprolactone (PCL) nanoparticles and the different concentrations of metformin

219 and sodium metavanadate co-encapsulated in PCL nanoparticles by two-way ANOVA
220 in the periods of 24 hpf ($F(7.26) = 5.37, p < 0.001$), 48 hpf ($F(7.26) = 32.45, p < 0.001$),
221 72 hpf ($F(7.24) = 22.32, p < 0.001$) and 96 hpf ($F(6.20) = 21.98, p < 0.001$) followed by
222 Tukey's test ($p < 0.05$).

223

224 3.2 Discussion

225 Toxicity assessment using *D. rerio* is a simple, effective, and ethical method to analyze
226 the acute effects of exposure to nanoparticles and their long-term impact during
227 embryonic development, providing important information on sublethal and lethal effects
228 that may occur in the organism (da Silva et al., 2023). However, few studies address the
229 effects of pharmaceutical excipients used for the formulation of nanoparticles for
230 veterinary use. In this study, we provide evidence that PCL nanoparticles can increase
231 mortality and morbidity in *D. rerio* embryos and eleuthero-embryos, causing
232 malformations such as spine deviation and pericardial and yolk sac edema.

233 All exposed groups presented sublethal and/or lethal effects. The mortality presented in
234 the group exposed to metformin (M-10) may be related to the concentration of the drug
235 used since Elizalde-Velázquez *et al.* (2021) used a concentration of ≤ 0.1 mg/mL, which
236 is lower than those used in our study, and they did not see such effects. The sublethal
237 effects observed in the group exposed to sodium metavanadate and the association of
238 this drug with metformin, such as pericardial and yolk sac edema and spinal
239 deformation, may be associated with the presence of the vanadium compound in
240 question, since in Bittencourt *et al.* (2018) these effects were the most evident in relation
241 to this drug and were not observed in the group exposed to metformin only. Additionally,
242 Santos *et al.* (2021) found that nanoparticles of vanadium at 0.010 mg/mL caused
243 malformation in embryos after 96hpf. Their results were found at a concentration of
244 almost half of ours and at the same time of exposure. The mortality presented in the
245 group exposed to the association of these drugs may be related to a possible synergistic
246 effect between these two compounds, since there was a higher lethality than in the
247 groups exposed to metformin or sodium metavanadate alone.

248 In the groups exposed to PCL-nanoencapsulated drugs, the sublethal effects, such as
249 pericardial and yolk sac edema, and lethal effects were dependent on the concentration
250 used, indicating that some of the excipients used in the formulation caused such effects.
251 Metformin and sodium methavanadate can be discarded as possible agents that have
252 caused such effects, since in our study there is a group of interactions of these drugs
253 that caused sublethal and lethal effects in the animals, but not to such a high degree. An
254 excipient that can be ruled out as a possible agent of the effects presented is the
255 surfactant known as poloxamer 188 since Hering *et al.* (2020) indicate that only
256 concentrations above those used in our study showed sublethal effects and no lethality.

257 Poly(ethyleneglycol) (PEG)-b-poly(ϵ -caprolactone) (PCL), namely PEG-b-PCL, a nano-
258 micelle similar to the PCL nanoparticles used in our study, on the other hand, presented
259 sublethal and lethal effects from 0.06 mg/mL in a dose-dependent manner in an animal
260 model (Zhou *et al.*, 2016), which corroborates our study, since in the group exposed to
261 the highest concentration of the association of nanoencapsulated metformin and sodium
262 metavanadate, all animals observed presented sublethal effects, with a lethality of about
263 96%. The group exposed to the white nanoparticle has the same concentration of PCL
264 as the group exposed to the highest concentration of the association of
265 nanoencapsulated metformin and sodium metavanadate, presenting very similar

266 dynamics of sublethal and lethal effects, demonstrating that the increase in the
267 concentration of PCL is responsible for the lethal and sublethal effects presented in
268 these groups.

269 The nature of the lethality presented in the animals at 24 hpf may be related to the
270 process of embryonic angiogenesis, as PCL downregulates the expression of the
271 vascular endothelial growth factor (VEGF) after 24h. Furthermore, it was proved that
272 PEG-b-PCL nano-micelles upregulated the expression of apoptotic genes, p53 and AIF,
273 increasing endothelial cell apoptosis and thus inhibiting the process of angiogenesis
274 (Zhou *et al.*, 2016). Also, *in vitro*, these micelles showed cell toxicity by increasing the
275 level of tumor necrosis factor (TNF), an inflammatory factor, in cells in contact with PEG-
276 b-PCL nano-micelles after 24h (Zhao *et al.*, 2013). At 72 hpf, there is an increase in
277 lethality, notably in groups exposed to PCL nanoparticles. Since these animals are now
278 in the *eleuthero-embryo* stage and do not have chorion, they are more exposed to the
279 compounds without a protective envelope (Duan *et al.*, 2020). These results
280 demonstrate that the toxicity of nanoparticles is more related to the *eleuthero-embryo*
281 and the constituents of the nanoparticles than to the metformin and sodium
282 metavanadate.

283 Regarding heart rate, the association of free drugs caused an increase in the exposed
284 animals, indicating that there is a synergy between the two drugs, which is in line with
285 the administration of these drugs alone, since metformin and sodium metavanadate
286 decrease heart rate (Bittencourt *et al.*, 2018; Borg *et al.*, 2020). In the case of PCL, the
287 heart rate in the animals decreased, regardless of the concentration used. This change
288 may be a result of the toxic effect caused by this polymer on angiogenesis (Zhou *et al.*
289 2016).

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292

4. CONCLUSION

293 The presence of PCL in the formulation of nanoparticles was harmful at the cardiovascular
294 level in embryos and *eleuthero-embryos* of *D. rerio*, affecting their development and heart
295 rate, regardless of their concentration, and may cause sublethal and lethal effects in these
296 animals. This results in greater care regarding the amount of nanomaterial used for the
297 development of nanoparticles. This implies that although they convey the idea that
298 nanomaterials are more bioavailable for veterinary use, they need to undergo careful
299 evaluation of their toxicity in animal models. Therefore, the toxicological evaluation of the
300 nanomaterial used is as important as its therapeutic efficacy, since such results can
301 anticipate possible toxicity problems, which may compromise its veterinary application in the
302 future. Finally, because polycaprolactone can alter nanoparticle toxicity, we recommend that
303 complexes are not treated as free molecule enhancements but rather as novel products that
304 should be evaluated individually regarding their toxicity.

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315

316 **AUTHORS' CONTRIBUTIONS**

317

318 This work was carried out in collaboration among all authors. All authors read and approved
319 the final manuscript.

320

321 **COMPETING INTERESTS**

322

323 The authors declare that no competing interests exist.

324

325

326 **ETHICAL APPROVAL**

327

328 All tests involving animals were previously approved by the Ethics Committee for Animal Use
329 of the *Universidade Federal Rural de Pernambuco*, under License nº 071/2019.

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