

Original Research Article

NUTRITIONAL AND ANTINUTRITIONAL EVALUATION OF THE PULP AND SEED OF *Terminaliacatappa*L.(TROPICAL ALMOND) FRUIT AND PHYSICOCHEMICAL PROPERTIES OF ITS SEED OIL

ABSTRACT

Aims: This study aimed at determining the nutritional and antinutritional composition of the pulp and seeds as well as physicochemical characteristics of the seed oil of *Terminaliacatappa*L.(tropical almond).

Methodology: The analyses were done in the Agroecology and life science laboratory of the University of Buea between June 2023 and May 2024. The experimental design was employed. Oil was extracted using soxhlet method. UV-visible spectrophotometer was used for nutritional and antinutritional analyses; while gas chromatography and titrimetry were used to analyse the fatty acid profile and physicochemical properties of the seed oil respectively.

Results: Results revealed that the seeds recorded more concentrated energy (596.28 Kcal/100g) than the pulps (403.42 Kcal/100g). Total phenolic (133.70 ± 0.15 mg/100g), tannin (77.90 ± 0.01 mg/100g), oxalate (15.01 ± 0.01 mg/100g) and phytic acid (2.81 ± 0.06 mg/100g) contents were displayed by the pulp and 29.70 ± 0.25 , 18.00 ± 0.07 , 20.92 ± 0.07 and 1.88 ± 0.01 mg/100g, respectively for seeds. The abundant minerals in the pulp were K, Ca, Na, and P, those in the seeds were Mg, K, P and Fe. The Acid value, Saponification value, Iodine value, Peroxide value and P-anisidine value of the seed oil were 2.24 ± 1.16 mgKOH/g, 222.2 ± 3.11 mgKOH/g, 78.50 ± 0.54 gI₂/100g, 5.50 ± 0.71 mEq/kg and 3.65 ± 0.23 , respectively. Fatty acid profiling showed 66.68% unsaturated fatty acids of which 19.86% and 9.81% were linoleic and linolenic, respectively and 33.32% saturated.

Conclusion: The pulps and seeds of *T. catappa* have a significantly high content of macro and micronutrients that can meet up the nutritional needs of a population and the quality assessment of the oil suggests that it can be recommended as suitable for industrial usage.

Keywords: Pulp, Seed, oil, fatty acid profile, *Terminaliacatappa*, nutritional composition, physicochemical properties, fatty acid profile.

1. INTRODUCTION

Indigenous fruit trees are fruit-bearing tree native to a specific region or ecosystem and are sometimes traditionally cultivated or utilized by indigenous communities within the area [1]. These plant species are known for their leaves consumed as vegetables with high nutritive values as well medicinal properties. Their fruits are edible and sometimes contain seeds, classified as oilseeds [1, 2] of high nutritional and economic importance as fruits not only offer easily available energy, but also micronutrients necessary to sustain and support human growth and activity [2]. Ahmad and Pieroni [3], stipulated that edible indigenous fruit species have been used as an alternative to naturalized species, during dry periods and times of food crises and in this way bridge the 'hunger gap' during times of food shortage. However, in the last two decades, research efforts have been channeled toward harnessing the nutrient potentials of both conventional and unconventional fruits as a way of enhancing food security [4]. While a few studies have highlighted the nutritional potential of several of these fruits like in *Artocarpus heterophyllus* (jackfruit), *myrianthus arboreus* (giant yellow mulberry), *Anacardium occidentale* (cashew apple), *Elaeis guineensis* (palm nut); and found them naturally rich in phytochemicals compounds, likewise their by-products (peels and seeds), there is much still to be done when it comes to the nutritional awareness and valorization of others [5-10].

Terminalia catappa L. also known as country almond, Indian almond, Malabar almond, sea almond, tropical almond, and beach almond, is an endemic tropical fruit tree species from the Combretaceae family [11]. In Cameroon, it is known by a common appellation "Banga school". *T. catappa* tree has leaves and stem barks whose nutritional and medicinal importance has been revealed [12-13]. Although it also produces fruits and nuts which are edible, it is still among the non-conventional fruit trees that has been under-exploited, despite its wide distribution in some communities of Cameroon [14]. Contrary to leaves and stem bark, whose nutritional and medicinal importance has been revealed [15], very little has been done on the fruits and their seeds especially in Cameroon. Meanwhile previous studies conducted in Vietnam, revealed that *T. catappa* fruit had oil content of 56.38% which was significantly high when compared to the oil content reported for some commercial plant oils such as sunflower (44%), soybeans (18%) and corn 26.44% [16, 17].

Though several fruits are available in Cameroon, there is a lack of scientific information about them. A knowledge of their nutritional potential is important to their valorization and for better utilization by the population. The more specific contribution of food from native trees to nutrition, is poorly documented in science and often not acknowledged in poverty reduction strategies [2]. Therefore, several fruits trees may be considered for food uses, but their nutritional value is underestimated [14]. Information on the nutrient composition of food is essential to estimate adequate nutrient intake both at individual and group levels [3]. The consumption and utilization of the fruits of *T. catappa* in Cameroon is limited in that it is known only to the immediate localities where they are found. In order to encourage the increased cultivation and consumption and hence exploitation, it is essential to determine the nutrient composition of the pulp and seed as well as check if its seed has potential of producing oil of good quality for consumption. It is for the aforementioned reasons that this study sets out to evaluate the nutritional and antinutritional properties of the pulp and seed of *T. catappa* fruit as well as to determine the physicochemical properties of its seed oil.

2. MATERIAL AND METHODS

2.1. Sample collection

Ripe fruits of *T.catappa* were harvested from seaside and beaches in Limbe and Idenau, (coastal towns with several water sources and a tropical forest on the slope of Mount Cameroon with an equatorial climate and green vegetation throughout the year) localities found in the Southwest region of Cameroon, in June 2023. The fruits were transported in ice cold box to the life science laboratory of the University of Buea for preparation and analyses.

2.2. Samples Preparation

T.catappa fruits were washed, pulp removed and the seeds dehulled to obtain the kernel and dried in an oven at 50 °C for 48 h. The freshly pooled pulps were crushed using a mortar and pestle and dried in a vacuum oven at 50 °C for 48 h and later crushed into fine powder using an electric blender. Similarly, the dried seeds were de-husked manually cracking with a stone to remove the kernels which were further dried for another 48 hours until moisture content was less than 12%. The dry kernels were then ground using an electric grinder to obtain fine powder. The fine powder (Figure 1) obtained from the seed was divided into two portions, the first portion (1kg) was used for proximate and nutritional analysis while the second portion (4kg) was used for oil extraction.

2.3 Oil Extraction

Extraction of *T.catappa* seed oil was by the Soxhlet method. The oil was extracted from the seed powder in a Soxhlet extractor for 8 h to maximize the total oil yield using n-hexane as a solvent. The solvent was evaporated and recovered with the help of Rota vapor (Buchi, Switzerland) at 100 rpm, and dried in an oven at 45 °C for 2 h. The extracted oil (Figure 1) was then allowed to be collected, weighed and stored at 4 °C for further analysis. The total oil yield expressed as a percentage was calculated on dry weight basis as:

$$\% \text{ Oil yield} = \frac{\text{Weight in gram of extracted oil}}{\text{Weight in gram of extracted seed powder sample}}$$



Figure 1. The process of harvesting of seeds and oil extraction from *T. catappa*. A= a typical tropical almond tree with fruits, B=Fresh fruits with seeds, C= dried fruits with seeds, D = fresh dehulled seeds, E = oven dried seeds, F= powdered seeds prior to extraction of oil, G = Extracted oil, H= defatted flour

2.3. Nutritional Analyses of pulp and seed flour

2.3.1 Proximate analysis of pulp and seed flour

The methods used for sample treatment and analysis were carried out based on the standard procedures recommended by AOAC [18], The moisture content of both pulp and defatted seed powder was determined gravimetrically by heating at 105 °C for 24 h in an oven by the procedure described by AOAC [18], Proximate analysis (crude fat, ash, total

carbohydrates, fiber, and proteins) was determined according to AOAC [18]. Total oil was quantified gravimetrically and calculated as a percentage of oil using a soxhlet extractor. Protein ($N \times 6.25$) was determined by the Kjeldahl method. To determine the ash content of the sample, 5 g of the sample was incinerated at 550 °C in a muffle furnace. The crude fiber content of the samples was determined by treating the both samples with 1.25% sulfuric acid and 1.25% sodium hydroxide solutions under specific conditions and the dried residue remaining after digestion of the samples was considered as crude fiber. Digestive carbohydrate content was determined by difference method according to AOAC [18].

2.3.2. Determination of mineral composition of pulp and seed flour

Minerals were assayed by atomic absorption spectroscopy. The principle is based on the fact that when atoms of an element are in contact with a flame, they emit wavelengths of radiation whose intensity can be measured. The concentration of the cation to be determined was calculated from the absorbance values by linear regression equations. To determine the mineral content of both samples, 5 g of sample was incinerated in a furnace at 550 °C and the ashes dissolved in 50 ml of 2.5% HNO₃ solution. The concentrations of Na, Ca, Mg, Fe, P, K, Zn, and Fe were determined using atomic absorption spectrophotometer (Buck Science). A calibration curve was prepared using standard metal solutions.

2.3.3. Estimation of vitamin A content of pulp and seed flour

The vitamin A content was estimated after determining the β -carotene content of the samples. β -carotene content was determined as described by Hagos et al.[19]. A mass of 1 g of the sample was extracted in 5 ml of methanol for 2 h at room temperature under dark conditions. The β -carotene layer was separated using hexane through a separating funnel. The volume was made up to 10 ml with hexane and then this layer was again passed through sodium sulphonate through a funnel to remove any moisture from the layer. The absorbance of the β -carotene layer was measured at 436 nm using hexane as a blank. The beta carotene was calculated using the formula:

$$\text{Beta-carotene content } (\mu\text{g}/100\text{g}) = \text{Absorbance (436 nm)} \times V \times D \times 100 \times 100/W \times Y$$

Where: V = Total volume of extract; D = Dilution factor; W Sample weight; Y = Percentage dry matter content of the sample.

Vitamin A content was estimated using the 1 μg retinol = 1 RE method.

1 μg β -carotene = 0.167 μg RE as defined by Codex Alimentarius [20].

2.3.4. Determination of vitamin C content of pulp and seed flour

Vitamin C was determined by the titration method using the protocol described by AOAC [18]. A volume of 1 mL of both samples and 90% acetic acid was titrated with a solution of 50 μM DCIP. The endpoint titration was determined when the blue colour changes to a pale pink colour. A blank titration was performed against 90% acetic acid while a standard titration was performed against 40 mg/L pure L-ascorbic acid. The amount of Vitamin C was expressed in mg/100ml of sample.

2.3.5 Anti-nutritional factors of pulp and seed flour

2.3.5.1 **Determination of Total Phenolics Content (TPC).**

The total phenolics content of the pulp and seed was determined using the Folin-Ciocalteu colorimetric method described by Chlopicka et al. [21]. The calibration curve for the standard is shown on figure 2 below.

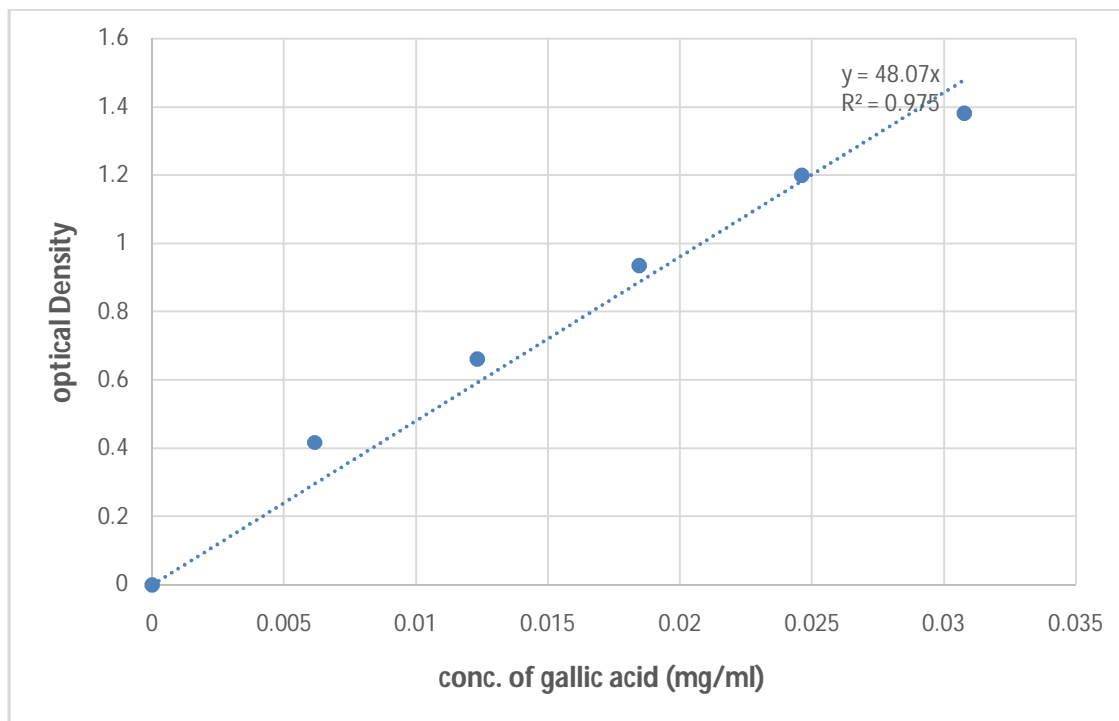


Figure 2. Calibration curve for determination of Total phenolic content

2.3.5.2. **Estimation of tannins content**

The content of Tannins was estimated by the Vanillin-HCl method. Defatted *T. catappa* flour and pulp flour (5 g) were treated with acidic methanol for extraction of tannins. From the diluted extract, 1 ml was mixed with 5 ml of freshly prepared vanillin-HCl reagent, and the optical density was determined at 500 nm by using a spectrophotometer. The results were expressed mg/100 gdm using catechin as standard.

2.3.5.3. **Determination of phytates content**

Phytates content was determined according to Andreia et al. [22]. The amount of phytate phosphorus content was calculated from the standard curve by assuming that 4:6 iron-to-phosphorus molar ratio.

2.3.5.4 **Determination of oxalates content**

To determine oxalates content in *T. catappa* seed flour and pulp, 2 g of flour were extracted with 100 ml of boiling distilled water for 30 min, filtered, and adjusted to 200 ml. The hot

water extract residue was further extracted with 150 ml of boiling 1 M HCl for 30 min, adjusted to 200 ml, and filtered. The two filtrates were combined and titrated with potassium permanganate.

2.4 Seed oil analysis

2.4.1 Determination of oil quality indices

After extraction of the *T. catappa* seed oil, it was filtered to remove non-oil materials. Sodium sulfate crystals were added to the crude oil to remove any trace water. The dry agent was separated by decanting and filtration. The physicochemical analyses of the oil for free fatty acid, iodine, peroxide, saponification, and P-anisidine values were carried out according to the methods of AOAC [18]. Total Oxidation (TOTOX) value was determined based on the obtained peroxide and P-Anisidine values using the formula;

$$\text{TOTOX} = 2\text{PV} + \text{AV}$$

2.4.2. Determination of Fatty acids composition

The fatty acids were analyzed using gas chromatography-mass spectrophotometry (GC-MS) (Agilent Technologies, 7890A GC-System, 5975C insert XL EI/CI MSD) with Triple-Axis Detector equipped with a VF-5ms MS capillary column (30 m x 0.25 mm id, 0.25 μm). Injector (G4513A) and detector temperatures were set at 220 and 250 $^{\circ}\text{C}$, respectively. One micro-liter (1 μL) of the filtered sample diluted with hexane was injected and analyzed with the column held initially at 50 $^{\circ}\text{C}$ for 1 min and then increased by 5 $^{\circ}\text{C}/\text{min}$ up to 280 $^{\circ}\text{C}$. Helium was employed as carrier gas (1 mL/min). The identification of the different fatty acids was performed by comparison of their relative retention times and mass spectra (Figure 3), with those of authentic reference compounds using NIST (National Institute of Standards and Technology) library database. Identification of different fatty acids present in samples was investigated using GC-MS.

Fatty acid profiling (ΣSFA , ΣMUFA , ΣUFA , ΣPUFA , $\Sigma\text{n-3/n-6}$ ratio, Σ omega-3, Σ omega-6, $\Sigma\text{PUFA}/\Sigma\text{SFA}$ ratio) was done by summing different classes of fatty acids.

The atherogenicity index (AI) and the thrombogenicity index (TI) were obtained using the formulas of Ulbricht & Southgate [23].

$$\text{AI} = \frac{[(\text{C12:0} + (4 \times \text{C14:0}) + \text{C16:0})]}{(\Sigma \text{MUFA} + \Sigma \omega 6 + \Sigma \omega 3)}$$

$$\text{TI} = \frac{(\text{C14:0} + \text{C16:0} + \text{C18:0})}{[(0.5 \times \Sigma \text{AGMI}) + (0.5 \times \Sigma \omega 6 + (3 \times \Sigma \omega 3) + (\Sigma \omega 3 / \Sigma \omega 6))]}$$

The ratio of hypocholesterolemic and hypercholesterolemic (H/H) was obtained using the formula:

$$\text{H/H} = \frac{(\text{C18:1cis9} + \text{C18:2}\omega 6 + 20:4\omega 6 + \text{C18:3}\omega 3 + \text{C20:5}\omega 3 + \text{C22:5}\omega 3 + \text{C22:6}\omega 3)}{(\text{C14:0} + \text{C16:0})}$$

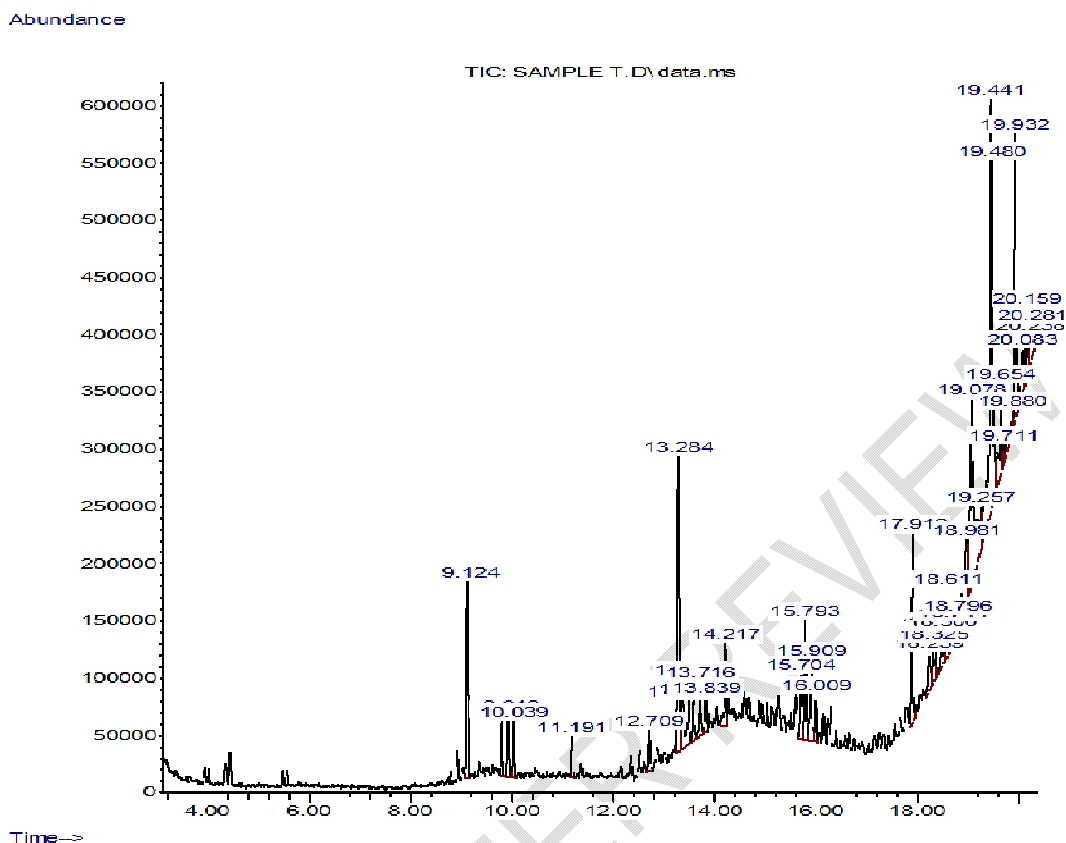


Figure 3. Calibration curve showing abundance and retention of fatty acids from *T. catappa* seed oil

2.5. Statistical Analysis

The data were subjected to one-way analysis of variance (ANOVA) to test if the fruit part had an effect on the measured parameters. Since there were more than two levels of the factor for the proximate composition, the significant ANOVA result was followed by Student-Newman-Keuls test. The analyses were conducted in Statgraphics Centurion version XVI and Graphpad-InStat version 3.05 at $p < 0.05$.

3. RESULTS

3.1 Nutritional of *T. catappa* pulp and seed

3.1.1. Proximate composition

Table 1 presents the results of the chemical composition of *T. catappa* seed and fruit pulp. The moisture content of dried seed in this study was 10.21% while the moisture content for the pulp was 86.53%. The fruit pulp on a dry matter basis recorded a high carbohydrate content (51.81%) as the predominant component, followed by proteins (23.96%) and fat (15.58%). While for the seeds, the predominant was crude fats (49.79%), followed by crude proteins (31.47%).

Table 1. Proximate composition of *T. catappa* fruit pulp, seed and defatted seed

Proximate Composition	Pulps	Seeds	Defatted Seed
Moisture (%)	86.53 ± 0.11 ^c	10.21 ± 0.98 ^a	8.05 ± 2.98 ^a
Crude protein (%)	23.96 ± 2.61 ^b	31.47 ± 0.02 ^b	70.14 ± 5.02 ^a
Crude fat (%)	15.58 ± 2.69 ^a	49.79 ± 0.27 ^c	5.84 ± 0.27 ^a
Total ash (%)	9.66 ± 1.83 ^b	0.47 ± .014 ^a	0.48 ± .014 ^a
Total fiber (%)	9.98 ± 0.45 ^c	2.66 ± 0.09 ^b	2.66 ± 0.09 ^b
Carbohydrate (%)	41.83±0.90 ^a	5.60 ± 0.01 ^b	5.76 ± 0.01 ^b
Total carbohydrate (%)	51.81± 4.01 ^a	8.26 ± 0.01 ^b	8.48 ± 0.01 ^b
Energy value (Kcal/100g)	403.42± 0.01 ^a	596.28±4.01 ^b	356.16±4.01 ^c

*Moisture content on oven dry weight basis

Values are mean ± standard deviation, n=3, values with different letters across the rows are significantly different ($P \leq 0.05$)

3.1.2 Mineral Composition

Table 2 shows the mineral content of *T. catappa* pulps and seeds. The abundant minerals in the pulps were potassium (1989.36 mg/100g), calcium (336 mg/100g) sodium (253.15 mg/100g), and phosphorus (178.46 mg/100g); while in the seeds, abundant minerals were magnesium (1085 ± 0.04mg/100 g), potassium (1035 ± 4.24 mg/100 g), phosphorus (960.5 ± 4.95), iron (166 ± 1.41 mg/100 g).

Table 2. Mineral composition of *T. catappa* pulp and seed

Mineral content (mg/100g)	Pulps	Seeds
Phosphorus	178.46 ± 0.001 ^b	960.5 ± 4.95 ^a
Potassium	1989.36 ± 2.38 ^a	1035 ± 4.24 ^b
Magnesium	58.32 ± 1.37 ^a	1085 ± 0.04 ^b
Calcium	336 ± 0.74 ^a	36 ± 1.41 ^b
Zinc	79.33 ± 0.001 ^a	6.96 ± 0.05 ^b
Sodium	253.15 ± 0.82 ^a	53 ± 1.41 ^b
Iron	51.24 ± 0.78 ^b	166 ± 1.41 ^a

*Values are mean ± standard deviation, n=3, values with different letters within rows are significantly different ($P \leq 0.05$)

3.1.3 Vitamin Composition of *T. catappa* pulp and seed

Figure 4 demonstrates the results for the vitamin composition of the pulps and seeds of *T. catappa*. From the results obtained in this study, the Vitamin A content of the pulp is 178.46 mg/100g while that for the seed is 960.5 mg/100g. The vitamin C content of the pulp is 1989.36 mg/100g while that for the seeds is 1035 mg/100g.

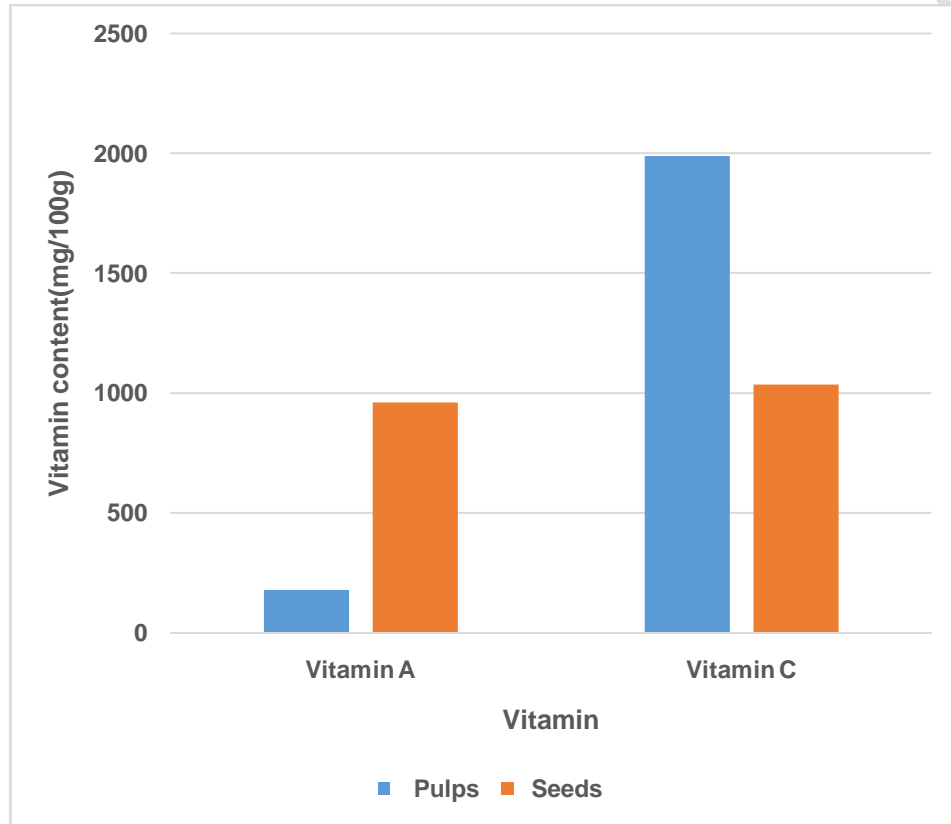


Figure 4. Vitamin Composition of *T. catappa* pulp and seed

3.1.4. Antinutrients Composition of pulp and seed of *T. catappa*

Antinutritional components of *T. catappa* fruit pulp and seed are summarized in Table 3. The results show Total phenolic content of pulp and seed to be 133.70 mg/100g and 29.70 mg/100g respectively. Phytate, oxalates and tannins contents are 2.81 mg/100g, 15.01 mg/100g and 77.90 mg/100g respectively for pulps while for seeds the values were 1.88 mg/100g, 20.92 mg/100g and 18.00 mg/100g respectively.

Table 3. Antinutrients Composition of pulp and seed of *T. catappa*

Antinutrients (mg/100g)	Pulp	Seed
TPC	133.70 ± 0.15 ^a	29.70 ± 0.25 ^b
Phytate	2.81 ± 0.06 ^b	1.88 ± 0.01 ^b
Oxalate	15.01 ± 0.01 ^a	20.92 ± 0.07 ^a
Tannin	77.90 ± 0.01 ^a	18.00 ± 0.07 ^b

Values are mean ± standard deviation, n=3, values with different letters within rows are significantly different ($P \leq 0.05$)

3.2 Seed Oil Analysis

3.2.1 Physicochemical characterization of oil

In order to know the safety of the oil for consumption, the oil quality indices were analyzed and presented on the table 4. The acid value, free fatty acid, saponification value, iodine value, peroxide value, P-anisidine value and total oxidation value obtained all fell within the recommended Codex Alimentarius commission range for safety. The physicochemical properties of *T. catappa* seed oil are shown on Table 4.

Table 4. Oil quality indices of *T. catappa* seed oil

Quality indices	<i>T. catappa</i> oil	Codex safety level (WHO/FAO)
Acid index (mgKOH/g)	2.24 ± 1.16	≤ 4.0
FFA (%)	1.54 ± 0.59	≤ 2.5
Saponification value(mgKOH/g)	222.2 ± 3.11	188-194
Iodine value (gl ₂ /100g)	78.50 ± 0.54	104-120
Peroxide value (mEq/kg)	5.50 ± 0.71	≤ 15
P-anisidine value	3.65 ± 0.23	≤ 20
Totox value	14.65 ± 1.18	≤ 40

Values are mean ± standard deviation, n= 3

3.2.2 Fatty acid composition of *T. cattapaseeds*

The fatty acid composition of *T. catappa* seed oil was determined by gas chromatography (Table 5). The major unsaturated fatty acids present in the oil were Linoleic acid, Oleic acid, and Linolenic acid, while the major saturated fatty acids were Lauric, capric, margaric, and arachidic acids. Total saturated fatty acids account for 33.32% of the fat represented mainly by lauric acid (7.32%). Unsaturated fatty acids constitute 66.68% of the total seed fatty acids and 32.47% of these are monounsaturated.

Table 5. Fatty acid profile of *T.catappa* oil

Fatty Acids	Common Name	IUPAC Name (Systematic)	(%)
C10:0	Capric acid	Decanoic acid	5.26
C12:0	Lauric (Dodecanoic) acid	Dodecanoic acid	7.32
C14:0	Myristic (Tetradecanoic) acid	Tetradecanoic acid	2.06
C15:0	Pentadecanoic acid	Pentadecanoic acid	5.26
C15:1	Pentadecenoic acid	14-Pentadecenoic acid	9.09
C16:0	Palmitic acid	Hexadecanoic acid	0.83
C16:1 (9)	Palmitoleic acid	Hexadecenoic acid	9.09
C17:0	Margaric acid	Heptadecanoic acid	5.26
C18:0	Stearic acid	Octadecanoic acid	2.06
C18:1(9)	Oleic acid	Octadecenoic acid	14.30
C18:2(9,12)	Linoleic acid	Octadecadienoic acid	19.86
C18:3(9,12,15)	Linolenic acid	Octadecadienoic acid	9.81
C20:0	Arachidic acid	Eicosanoic acid	5.26
NI	Non Identified		4.54
ΣSFA	Saturated Fatty acids		33.32
ΣMUFA	Monounsaturated fatty acids		32.47
ΣUFA	Unsaturated fatty acids		66.68
ΣPUFA	Polyunsaturated Fatty acids		29.67
Σ (ω-3)	Omega-3		9.81
Σ(ω-6)	Omega-6		19.86
PUFA/SFA	polyunsaturated/saturated ratio		0.89
ω-3/ω-6	omega-3/omega-6 ratio		0.49
AI	Atherogenicity Index		0.26
TI	Thrombogenicity index		0.09
H/H	Hypocholesterolemic and hypercholesterolemic ratio		15.21

Values are mean ± standard deviation, n= 3

4. DISCUSSION

The moisture contents were found to be higher in the edible pulp compared to the seeds. High moisture content reduces the shelf life of fruits since moisture increases microbial activity, implying that both fresh seeds and pulps, cannot be stored for long in the fresh state and therefore need to be preserved by drying techniques, to increase their shelf life as reported by Ojwang et al.[5].

Proteins are important nutrients in the body, as they help in repairing worn-out tissues and are a source of amino acids required for protein synthesis. The findings of this study showed that the protein content of the pulps and seeds were 23.96% and 31.47% respectively, indicating that tropical almond seeds are good sources of protein. When defatted, the seeds had 70.14% protein indicating that it can be used to supplement a protein-deficient diet and as a complementary food to solve protein-energy deficiency [24]. These results are similar to results obtained for *T. catappa* seeds (28.70%) but higher than the value obtained for the pulps (8.75%) from Nigeria [25]. The protein content obtained for the seeds in this study is very similar to those harvested from Cote D'Ivoire (31.21%) [26], and higher than 20.14% from Benin [27]. However, the value for the protein content in seeds was slightly lower than 36.3% in cashew nut [7].

Lipids are known to have numerous health benefits in the body such as the synthesis of cell membranes, steroid hormones, and eicosanoids which ensure the integrity of the cell and enable it to carry out several vital body processes. Consumption of lipids is therefore important to ensure the proper functioning of the body [5]. *T. catappa* seeds reported a significantly higher values than those of most conventional seeds such as soybean (14%), palm fruit (20%), palm kernel (36%), groundnut (42%), sunflower (32%), cotton (35%) and unconventional seeds such as mango (12%) and sesame seeds (26%) [27-29]. The high amount of *T. catappa* lipids in the seeds makes it a distinct potential for the oil industry [30]. Generally, the results reveal *T. catappa* seed as an oilseed with the potential of high oil and protein content to satisfy the calorie and protein demands of a population.

For ash content, the pulps recorded the highest ash content (9.66%) relative to the seeds (0.47%). The ash content is a measure of the inorganic materials present in food, which are minerals. *T. catappa* fruits are therefore rich in minerals. The fiber content of the pulp (9.98%) is significantly higher than that of the seeds (2.66%). Fiber is an essential component of a healthy diet as it aids in digestion, promotes satiety and helps maintain bowel movement. A high fiber content in the pulp could be beneficial for overall gut health and may contribute to a feeling of fullness, which can support weight management [31].

The pulp contains significantly higher quantities of carbohydrate (51.81%) than the seed (8.26%). Carbohydrates, like proteins contribute to more than 10% of the total bio-available energy of the seeds which will make these seeds good sources of energy (Table 1). The result shows that the seed has a higher caloric value than the pulp and this was in line with results obtained for pulps (370 Kcal/100g) and seeds (587 Kcal/100g) of *T. catappa* fruits from Nigeria [25].

Mineral analyses revealed that for all the minerals analyzed, the values obtained for the pulp was significantly different ($P \leq 0.05$) from that of the seed. The seed contain more proportion of most minerals compared to the pulp of the fruit and magnesium was the most abundant mineral in the seeds while potassium was most abundant in the pulp. This result was in line with Vibha et al. [15] who also recorded that Magnesium (1687.5 mg/kg) was the most abundant mineral among the minerals analyzed in *T. catappa* seeds. Therefore; these seeds

have the potential to supply a sufficient amount of minerals for consumers as well as to balance human electrolytes.

Vitamin A is important for normal vision, gene expression, growth, and immune function through its maintenance of epithelial cell functions [32]. Vitamin C is a potent antioxidant that facilitates the transport and uptake of non-heme iron at the mucosa. Vitamin A was higher in the seed than in the pulp, while vitamin C was higher in the fruit pulp than in the seed. Vitamin C is known to be water-soluble and is found more in fruits than in seeds. Carotene generally is higher in fruits [33] and fat-soluble vitamins are lower than their water-soluble vitamins in fruits. This could also be the reason for the increased pro-vitamin A content in the dry seed.

Antinutrients are natural or synthetic compounds that interfere with the absorption of nutrients and their presence in any food sample is of significant importance since antinutrients may pose some deleterious effects, depending on the dose present [34]. Phenolic compounds inhibit the activity of digestive enzymes like α -amylase, trypsin, chymotrypsin, and lipase and decrease the digestibility of proteins, carbohydrates, and availability of vitamins and minerals [35]. The pulps exhibit a lower level of phenolics compared to the seeds. The levels of both phenolics and tannins in *T. catappa* appear to be lower than an earlier report for *T. catappa* seeds (35 mg/100g) harvested in Benin [27].

Phytic acid is an antinutrient that inhibits the absorption and utilization of iron and calcium in the body by forming insoluble phytates [36]. Therefore, when present in large amounts, they are known to act as a major inhibition to the absorption of iron. The phytate content for *T. catappa* pulp (2.81 mg/100g) is higher than that for the seeds (1.88 mg/100g). The low phytate content is an indication that they may not pose any hindrance to the metabolism of some nutrients in the body when consumed.

Oxalate is a chelating agent, which binds calcium very effectively. Foods with high oxalate content may produce acute metabolic calcium deficiency [37]. The concentration of oxalate (15.01 mg/100g for pulp and 20.92 mg/100g for seed) in this study seems to be low. Tannins are a class of phenolic compounds. High amounts of tannins are well known to form complexes with proteins reducing the solubility of proteins [38]. The tannin level (77.9 and 18.00 mg/100g for pulp and seed respectively) obtained in this study was found to be relatively high in comparison with tannins content found in previous study on *T. catappa* fruits in Nigeria which reported 44 mg/100g for pulp and 35 mg/100g for seeds [25]. Therefore the tannin level in tropical almonds may not be as harmful as expected for consumption.

T. catappa seed oil has a very low acid value when compared with the safety level of Codex Alimentarius of ≤ 4.0 . The acid value obtained for *T. catappa* seed oil in this study is in line with values obtained for *T. catappa* seed oil from Nigeria (2.42) and Benin (2.24) [11,27]. The low acid value of *T. catappa* seed suggests that it can be stored for long since oil with high acid value cannot be stored for long because they are more prone to oxidation than their esterified ones. The free fatty acid (FFA) content of *T. catappa* seed oil (1.54 \pm 0.59%) fell within the safety range of $\leq 2.5\%$ as reported by Leung *et al.* [39]. The low FFA value in this study suggests that *T. catappa* seed oil may have a long shelf life since oils with low FFA tend to have a long shelf life.

The saponification value of the oil in this study was 222.25 mg KOH/g. This value is closer to values for some common oils like castor seed oil (185.83), palm oil (190-209 mgKOH/g), groundnut oil (187-196 mgKOH/g), and corn oil (187-196 mgKOH/g) [40]. The saponification value of *T. catappa* is within the range of (175–287) specification for oils by the American

Society for Testing and Materials [41]. Oils with lower saponification values contain high amount of long chain fatty acids. Therefore, the value obtained for *T. catappa* seed oil contained medium quantity of higher fatty acids, fatty acids ≥ 16 carbon (Table 5).

The iodine value indicates the number of reactive double bonds present in an oil. A high iodine value indicates more double bonds (unsaturation). The iodine value of *T. catappa* seed oil was 78.50 $\text{gI}_2/100\text{g}$ which was below the 107-135 $\text{gI}_2/100\text{g}$ range recorded for *T. catappa* oil from Malaysia [30], but within the range of 77–94 $\text{gI}_2/100\text{g}$ olive oil, above 8–10 $\text{gI}_2/100\text{g}$ coconut oil, 12–18 $\text{gI}_2/100\text{g}$, palm oil and 44.4 $\text{gI}_2/100\text{g}$ cashew nut oil [42]. Based on the iodine for *T. catappa* seed oil, the oil can be classified as a non-drying oil since such oils display $\text{I} < 100$. The seed oil can therefore be recommended in the edible cream industries and as raw vegetable oil.

The peroxide value obtained ($5.50 \pm 0.71 \text{ meqO}_2/\text{kg}$) was within the recommended safety range of $\leq 10 \text{ meq}$ of active oxygen/kg of oil by the Codex Alimentarius Committee for all edible oils [20]. The low peroxide value of *T. catappa* oil reported in this study indicates a low level of primary oxidation products, which is characterized by the presence of hydroperoxides in oil. The P-anisidine value (18.57 ± 0.30) was within the recommended safety range by the Codex Alimentarius Commission (P-anisidine value ≤ 20). This value suggests that oil may contain low secondary oxidation products such as aldehydes of α - and β -unsaturation. The total oxidation value (34.24 ± 0.8) was within the recommended safety range ($\text{TOTOX} \leq 40$) for edible oils indicating, high primary hydroperoxides and secondary (aldehydes) oxidative stability for storage [20]. Overall, these results suggest that *T. catappa* seed oil is good for human consumption but must not be kept exposed on sun.

Table 5 shows that the tropical almonds oil is greater in unsaturated fatty acid than that for palm oil (50%) but lower than the 85% mentioned for soya oil by Sarwar et al. [9]. Among the unsaturated fatty acids present, polyunsaturated fatty acids, had as values 19.86% and 9.81% for linoleic and linolenic acid respectively. The high level of linoleic acid, which is one of the most important polyunsaturated fatty acids in human food, makes the consumption of this oil beneficial for health. PUFA/SFA is an indicator used to evaluate lipid quality. A PUFA/SFA ratio of 0.2 has been associated with high cholesterol levels and with high risk of coronary heart disorders, while a ratio as high as 0.8 is associated with desirable levels of cholesterol and reduced coronary heart diseases [43].

The omega-3/omega-6 (ω -3/ ω -6) ratio (0.49) was within the safe limits, since the maximum recommended value of 4. WHO recommends that the n-6 PUFA/n-3 PUFA ratio should not exceed 10, while the European Nutritional Societies suggest that this ratio should not exceed 5, for the prevention of inflammatory, cardiovascular, and neurological disorders [44]. The oil examined exhibited a ω -6/ ω -3 ratio below 5. The monounsaturated fatty acid (MUFA) content reported in this study was 32.47%. In human nutrition, the MUFAs play an important role, due to their hypocholesterolemic action, reducing the risk of arteriosclerosis. The rich content in MUFAs indicates that *T. catappa* seed oil can be good for cooking processes and seasoning like olive oil if fractionated into oleic fractions.

5. CONCLUSION

From the result of the analysis, it has been shown that the pulp and seed flour of *T. catappa* have higher nutrient composition and calorie value compared to some nuts, especially in terms of crude oil and protein. The amount of protein in the seed is high for un-defatted and defatted seed in comparison with that of most protein-rich crops. *T. catappa* pulp and defatted kernels contained significant amounts of minerals as well as Vitamin A and C and

also chemical compounds like Phenolic compounds, tannins, oxalates and phytates, which were within permissible limits. The percentage oil content of the seed was found to be much more than some conventional oils. The physicochemical properties of the oil produced in this study were within the permissive value of edible oils according to codex standards. The oil was rich in linoleic acids, oleic, linolenic and palmitoleic, composed of unsaturated fatty acids. The high UFA/SFA and PUFA/SFA ratios of this oil indicates its cholesterol-lowering potential and suggests that the seeds of tropical almond fruit have a potential to be used in the dietetic management of certain coronary heart diseases. The fatty acid composition of the seed oil makes it suitable for consumption.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

CONFLICTING INTERESTS

Authors have declared no conflicting interests exist.

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