

# **A Study on the Freshwater Cladoceran *Daphnia magna* (Straus, 1820) and its uses in Fish Larviculture**

## **Abstract**

Over the last three decades, aquaculture has become increasingly important, leading to the commercial viability of various shellfish and finfish species. The production of live feed is gaining popularity as a way to enhance the nutritional quality of species raised in aquaculture. *Daphnia* is a type of small crustacean belonging to the Brachiopoda family, commonly found in freshwater environments. *Daphnia spp.* serves as an excellent natural food source for tropical fish fry and has emerged as an alternative to traditional fish feeds. Utilizing *Daphnia* in aquaculture offers benefits such as low cost and ease of cultivation. This study addresses the identification of *Daphnia*, its reproduction and life cycle, nutritional value, significance in aquaculture, and methods of cultivation.

## **Introduction**

In the past thirty years, aquaculture has gained significant importance, with numerous shellfish and finfish species becoming commercially viable. This growth has resulted in the establishment of effective methods for breeding, reproduction, and the management of broodstock in captivity. Live food organisms are crucial as they supply necessary nutrients such as carbohydrates, proteins, amino acids, lipids, fatty acids, vitamins, and minerals, earning them the title of "living capsules of nutrition" [1]. However, the high price of *Artemia nauplii* cysts has led aquaculturists to explore alternative, suitable zooplankton options like *Daphnia magna*, *Moina*, rotifers, infusoria, termites, and bloodworms, which can be produced in large quantities. While feeding preferences differ among species, many require high-protein live food to improve growth performance, reproductive success, and survival rates [1].

The nutrition of fish is essential in aquaculture production because it greatly impacts the survival and growth rates of cultured species, ultimately determining the industry's financial viability [2]. This aspect is particularly vital during the early larval stages, where mortality rates are elevated due to the significant nutritional requirements for growth and development, which are compounded by their underdeveloped digestive enzyme activity,

making it difficult for them to absorb nutrients from inert food sources. Moreover, research indicates that commercial diets often lack the necessary specificity and fail to meet the nutritional needs of all cultured species [3]. In response to this issue, the production of live feed is gaining popularity as a way to enhance the nutritional quality of species raised in aquaculture. Live food provides a natural source of proteins, lipids, carbohydrates, minerals, and vitamins. Additionally, it does not affect water quality, and its color, odor, and constant movement stimulate feeding behavior in predators [4].

Cladocerans are zooplankton that belong to the class Crustacea within the phylum Arthropoda. These organisms primarily inhabit freshwater habitats, though they can also be found in brackish and marine environments. Their soft, non-thorny bodies make them an important food source for the larvae and juvenile stages of commercially important fish and prawn species. Consequently, Cladocerans are commonly used as an economical live food source in aquaculture hatcheries [5]. Approximately 420 cladoceran species have been recognized globally, with *Daphnia* being the most commonly cultivated species for the initial stages of finfish and shellfish feeds. This organism thrives in a range of habitats, from tropical areas to the Arctic, particularly in smaller ponds and larger freshwater lakes. While 50 species of *Daphnia* have been reported worldwide, only six are typically found in tropical zones [6].

The genera *Daphnia* and *Moina* are closely related and can be found globally, and are collectively referred to as *Daphnia* [7]. *Daphnia* serves a vital role as a live feed in freshwater aquaculture. Its distinctive appearance and erratic swimming patterns have earned it the nickname "water flea." This small crustacean ranges in size from 0.2 to 3.0 mm in length, with *D. pulex* and *D. magna* being the two primary species farmed for aquaculture purposes [7]. The lack of commercial feed options for the early stages of larval development underscores the importance of *Daphnia* as a key live food source in many freshwater fish hatcheries. Thus, advancements in *Daphnia* cultivation and a better understanding of its nutritional profile could significantly improve hatchery operations moving forward. This study focuses on the identification of *Daphnia*, its reproductive and life cycle, nutritional value, role in aquaculture, and cultivation methods.

### **Identification of Daphnia:**

It has a large head with rostrum, thorax and abdomen covered by the carapace. The abdomen which is bent down ward is free of appendages. In cephalic appendages, second maxilla is absent. Paired eyes are fused in one. Five pairs of leaflike feet present on thorax [8]. Daphnia have a body consisting of a head and a trunk. The antennae are the main means of locomotion. Large compound eyes lie under the skin on the sides of the head. One of the major characteristics of daphnia is that the main part of the body, the trunk, is enclosed in an external skeleton (carapace). Periodically, they moult or shed their external shell. The brood pouch, where the eggs and embryos develop, is on the dorsal side of the female. In Daphnia, the brood pouch is completely closed, while Moina have an open pouch [7].

### **Reproduction and Life Cycle of Daphnia**

Being a cladocera, it has the capability to reproduce in both sexually and asexually. However, parthenogenesis reproduction is very common among cladocerans. In this, female produces a amictic eggs-a clutch of parthenogenetic eggs-and keep them in brood pouch. The developmental process takes place in brood pouch and after completion of development in 3 days the smaller young ones directly released from the mother brood pouch. The produced daphnia looks much similar to their mother, like a clone but without a brood pouch [9]. After six moltings, the young daphnia attain maturity and produce the egg for first time – primipare stage. In general, it takes 5-10 days to reach the primipare stage by the younger daphnia; however, the duration is highly temperature dependent. Once in 3-4 days interval, the adult daphnia releases the eggs which continue till her death. Life span of daphnia under captive conditions is about 2 months [9].

Factors, such as change in water temperature or food deprivation as a result of population increase, may induce the production of males. These males have one or two gonopores, which open near the anus and may be modified into a copulatory organ. The male clasps the female with the first antennae and inserts the copulatory processes into the single, median female gonopore [9]. The fertilized eggs are large, and only two are produced in a single clutch (one from each ovary), and are thick-shelled: these resting or dormant eggs being enclosed by several protective membranes, the ephippium. In this form, they are resistant to desiccation, freezing and digestive enzymes, and as such play an important role in

colonizing new habitats or in the re-establishment of an extinguished population after unfavourable seasonal conditions.

### **Nutritional quality of Daphnia**

Among zooplankton species, *D. magna* has excellent nutritional content that is recommended for use in feeding of fish larvae[10].*Daphnia sp.* is highly nutritious live food containing high protein content ranging from 45-72 % and the fat ranges from 6.5-8 % of its dry weight. Furthermore, its linoleic and linolenic fatty acids contents are 7.5 % and 6.7 %, respectively [11,12, 13].

### **Importance of Daphnia in Aquaculture**

*Daphnia* is one of the most popular live feeds for aquaculture fishes. It is a frequently used food source in the fresh water larviculture (ie, for different carp spp). They have been used extensively to rear larvae and fry [14,15]. *Daphnia* includes several species, the largest of which is *D. magna*, shows high reproductively. *D. magna* is mainly used, alive or preserved as food for fish in aquaculture [16,17].

*Daphnia magna* comprises several digestive enzymes like proteases, peptidases, amylase, lipase, and cellulase which act as exoenzymes in the gut of fish. Being larger in size than *Moina*, it serves as live food for advanced stages of fish [18]. *D. magna* contains added protein and consequently, is a good replacement for *Artemia nauplii* in aqua hatcheries. *D. magna* has also been extensively utilized as a live-food source in various hatcheries and in the care and culture of aquarium fishes of commercial importance [19]. *D. magna* has the advantage of high reproduction rates, wide temperature tolerance, and the ability to thrive on phytoplankton and organic wastes. This enrichment of food with *D. magna* is accomplished with a basis of DHA, which helps the fish to make immune resistance against gill and water fouling problems [18, 20]. Recently, *Daphnia* meal is successfully used as a fish meal replacer in the diet of *Pelteobagrus fulvidraco*[21]. Moreover, *D. magna* can be used as a bioencapsulation of probiotics during fish larvae feeding, as indicated by [22] who reported that Persian sturgeon (*Acipenser persicus*) larvae fed with bioencapsulated *D. magna* with *Saccharomyces cerevisiae* had higher growth performance and better feed utilization.

The bioencapsulation of *D. magna* using the commercial probiotic *Bacillus* bacteria (Protexin Aquatic) has been shown to enhance the resilience of *Acipenser persicus* larvae to various stressors[23]. Munirasu[24] investigated the bio enrichment of live *D. magna* as a

means to improve the survival and growth of the freshwater fish *Catlacatla*. Vivi [25] reported on the mass cultivation of *Daphnia magna* Straus, 1820 in a fermented medium, which served as feed to enhance the nutrient quality and growth performance of Nile tilapia larvae (*Oreochromis niloticus*, Linnaeus, 1758). Rameshwar [26] explored the production of *Daphnia* for the nursery rearing of cultivable freshwater fish species. Abo [27] examined the impact of using *Daphnia magna* (Straus, 1820) as a substitute for fish meal, assessing its effects on the growth, feed efficiency, histological condition, and economic viability of grey mullet (*Mugilcephalus*, Linnaeus 1758). Manjit [1] conducted experiments with *D. magna* (Straus, 1820) as an alternative live feed for *Pterophyllumsalare*, focusing on its potential for commercialization. Kethavath[8] studied *Daphnia* culture and evaluated its role in the growth and development of molly fish.

### **Culture of Daphnia**

**This can be segregated as**

#### **Culture Methods and Applications:**

##### **1. Culture techniques: *Daphnia* can be cultured using various techniques, including batch culture, continuous culture, and recirculating systems.**

*Daphnia* is a very good source of live feed in freshwater aquaculture industry. Cultivation of this species is not difficult as it can feed wide varieties of unused food residues. Herawati [12] mentioned that rice bran has high nutritional value and enhances the growth of *D. magna*. Additionally, it has widespread overall freshwater bodies. It is characterized as a suitable size for the mouth opening of many fish larvae and fry, and it has a high nutritional composition [28]. Its nutrient content varies according to the culture medium and the degree of availability of phytoplankton [12]. These are herbivores which mainly feed on micro algae. The herbivorous zooplankton growth rates are sometimes strongly correlated with the mineral and biochemical composition of the phytoplankton they consume reported that food nutrient content influenced the growth rate of *D. magna* even at very low food levels [29]. The best foods for culturing *Daphnia* are algae and yeast. For culturing, the algal feeds such as *A. pinnata*, *C. vulgaris* and the yeast (*S. cerevisiae*) are used. The advantages of algae as a food are that, algal feeds are easy to culture and it is an excellent feed for the growth of *Daphnia*. Yeast is easy to acquire, and there is a minimum of fuss while preparing it for the culture [29, 30]. The most critical environmental factor to successfully culture *Daphnia* is temperature, which should remain close to 20 °C (68 °F). Higher temperatures may be fatal

to *Daphnia* and lower temperatures slow reproduction. It has a short generation time (9-11 days at 20°C), but total life span is longer (> 60 days at 20°C, up to one year at colder temperatures[30].

Freshwater zooplankton, specifically *Daphnia magna*, can be cultured using different combinations of feed. Castro [31] investigated the laboratory cultivation of *D. magna* (Straus 1820) using microalgae and active dry yeast as feed. In a similar vein, Munirasu[5] explored how various feeds, including *Chlorella vulgaris*, *Azolla pinnata*, and yeast, influenced the population growth of *D. magna*, which is commonly found in freshwater environments. Herman[32] examined the culture of *Daphnia sp.* and analyzed the impact of different manure types on growth, fertility, and mortality rates. Furthermore, Monowar[33] assessed the culture of freshwater zooplankton *D. magna* in Bangladesh, focusing on the effects of various feeding combinations. *Daphnia* can be easily enriched with various compounds via a bio-enrichment process that enhances the nutritional quality of these live feed organisms. The materials that can be used for enriching *Daphnia* include microdiets, microencapsulated diets, baker's yeast, and vitamins, which can be applied through feeding or incorporation methods [9].

## Conclusion

*Daphnia* is a very important cladoceran group crustacean and it is considered as healthy and highly nutritional diet in aquaculture. Smaller size of *daphnia* and their fast-moving nature attract the fish larvae to hunt them. Additionally, the simple rearing techniques and smaller rearing unit making them as important live feed in freshwater hatcheries. *Daphnia* have huge amount of exoenzyme like proteinases, amylases, lipases and cellulase these are helpful in the digestion and growth of the developing fish larvae which exoenzymes are not that much available in the other live feed.

## References

1. Monjit Paul, Mukti Chanda, Joydeep Das, Priyanka Maity, Sudip Mondal, Asim Kumar Giri. *Daphnia magna* (Straus, 1820)– an alternative live fish food for *Pterophyllumsalare* and its commercialization. *TIJER - International Research Journal*. 2023;10(7): 770-776.
2. Lazo JP. Conocimiento Actual y Nuevas Perspectivas en el Desarrollo de Dietas para Larvas de Peces Marinos. Cruz-Suárez LE, Ricque-Marie D, Tapia-Salazar M, Golvera-Novoa MA, Civera-Cerecedo R. (Eds). *Avances en Nutrición Acuicola* V. 2000;300-312.

3. Sanchez-Estudillo L. Alimentonutritivo, colorido y enmovimiento: Los Cultivos de apoyoenAcuicultura. Ciencia. 2011;15(43):55-60.
4. Castro BT, De Lara AR, Castro MG, Castro MJ, Malpica SA. Alimento vivo en la Acuicultura. Contactos. 2003;48:27-33.
5. Munirasu S, Uthayakumar V, Arunkumar P Ramasubramanian V. The effect of different feeds such as *Chlorella vulgaris*, *Azolla pinnata* and yeast on the population growth of *Daphnia magna* commonly found in freshwater systems. International Journal of Fisheries and Aquatic Studies. 2016;4(6):05-10.
6. Delbare D, Dert P. Cladocerans, nematodes and trochophora larvae. Manual on the Production and Use of Live Food for Aquaculture. 1996;283-295.
7. Rottmann RW, Scott Graves J, Craig Watson, Roy PE, Yanong. Culture Techniques of Moina: The Ideal *Daphnia* for Feeding Freshwater Fish Fry. UF/IFAS Extension. University of Florida. CIR. 2017;1054.
8. KethavathMadhudeepika, Dharavath Ram kumar, Jagadeeshwarachari T. Daphnia culture and evaluating the role of *Daphnia* in Molly fish growth and development. International Journal of Research Publication and Reviews. 2024;5,(9):230-236.
9. Bhosle et al. Production of Daphnia for Freshwater Nursery Rearing of Cultivable Fishes. Biotica Research Today. 2020;2(11):1218-1221.
10. El-feky, MM, Abo-Taleb H. Effect of feeding with different types of nutrients on intensive culture of the water flea, *Daphnia magna* Straus, 1820. Egypt. J. Aquat. Biol. Fish. 2020;24,655–666.
11. Herawati VE, Hutabarat J, Radjasa OK. Growth and survival rate of tilapia (*Oreochromis niloticus*) larvae fed by *Daphnia magna* cultured with organic fertilizer resulted from probiotic bacteria fermentation. HAYATI J. Biosci. 2015;22,169-173.
12. Herawati VE, Nugroho RA, Pinandoyo, Darmanto YS, Hutabarat J. The effect of fermentation time with probiotic bacteria on organic fertilizer as *Daphnia magna* cultured medium towards nutrient quality, biomass production and growth performance enhancement. IOP Conference Series: Science. 2018;116:012089.
13. Macedo CF, Pinto-Coelho RM. Nutritional Status Response of *Daphnia laevis* and *Moinamicrura* from a tropical reservoir to different algal diets: *Scenedesmusquadricauda* and *Ankistrodesmusgracilis*. Brazilian J. Biol., 2001. 61: 555-562.
14. De Pauw N, Laureys P, Morales J. Mass cultivation of *Daphnia magna* (Straus) on ricebran. Aquaculture. 1981;25:141-152.

15. Tay SH, Rajbanshi VK, Ho WH, Chew J, Yap EA. Culture of Cladoceran *Moinamicrurakurz* using agro industrial wastes. In Proceedings of the Fourth Asian fish Nutrition Workshop (de silva, S.S., ed.), 1991;135-141.
16. Edwards P, Pullin RSV. Waste water-fed aquaculture. Proceedings of the International seminar on Waste water Reclamation and Reuse for Aquaculture, Calcutta, India, 6-9 December 1988. Asian Institute of Technology Bangkok, Thailand. 1990;1:295-299.
17. Sevrin-Reyssac J, Prolux D, Watteau S, Chemillier J. Eaux usees et lagunage. Plus qu'un exutoire. Aquac. Rev. 1994;54:14-32.
18. Reza R, Hadi J, Mahdih JH. A Comparative Study on the Growth Rate of Persian Sturgeon, *Acipenser persicus*, Larvae Fed with *Artemia nauplii* and *Daphnia magna*, Global Veterinaria. 2013;10(2):116-120.
19. Martin L, Fajardo J, Arenal A, Pimental E, Pacheco M, Garcia C, Hidalgo L, Santiesteban D. Complete and partial replacement of *Artemia nauplii* by *Moinamicrura* during early post-larval culture of white shrimp *Litopenaeus schmitti*. Aquacult. Nutr. 2006;12(2):89-96.
20. Jafaryan HM, Makhtomii, Mahdavi M. The effect of baker's yeast for better utilization of nutrient compositions of *Daphnia magna* larviculture of Persian sturgeon (*Acipenser persicus*). Aquaculture Europe. Trondheim, Norway. 2009;282-283.
21. Zhang H, Wang Y, Wu X, Zeng X, Yang G, Zhang J, Zhu G, Bai D. Effects of *Daphnia magna* meal replacing fish meal on growth, biochemical indexes of *Pelteobagrus fulvidraco* and water quality indexes. Feed Ind. 2019;2,212-221.
22. Soltani M. Effects of bioencapsulated *Daphnia magna* with *Saccharomyces cerevisiae* on the growth and feeding performance of Persian sturgeon (*Acipenser persicus*) larvae. Iran. J. Vet. Med. 2012;6,13-18.
23. Faramarzi M, Jafaryan H, Roozbehfar R, Jafari M, Rashidi Y, Biria M. Influences of probiotic bacilli via bioencapsulated *Daphnia magna* on resistance of Persian sturgeon larvae against challenge tests. Glob. Vet. 2012;8,421-425.
24. Munirasu S, Ramasubramanian V, Uthayakumar V, Muthukumar S. Bio enrichment of live feed *Daphnia magna* for the survival and growth of freshwater fish *Catla catla*. Int J Cur Res Rev. 2014;05(08):20-26.
25. Vivi Endar Herawati, Johannes Hutabarat, Pinandoyo Pinandoyo, Nurmanita Rismaningsih, Ocky Karnaradjasa. Mass culture of *Daphnia magna* Straus, 1820 in Fermented Medium as Feed to Enhance Nutrient Quality and Growth Performance of Nile Tilapia *Oreochromis niloticus* (Linnaeus, 1758) Larvae 2019;2,42-49.

26. Rameshwar V, Bhosle1, Stephen Sampath Kumar J, Somu Sunder Lingam. Production of *Daphnia* for Freshwater Nursery Rearing of Cultivable Fishes. *Biotica Research Today*. 2020;2(11):1218-1221.
27. Abo-Taleb HA, Ashour M, Elokaby MA, Mabrouk MM, El-feky MMM, Abdelzaher OF, Gaber A, Alsanie WF, Mansour AT. Effect of a New Feed *Daphnia magna* (Straus, 1820), as a Fish Meal Substitute on Growth, Feed Utilization, Histological Status, and Economic Revenue of Grey Mullet, *Mugilcephalus* (Linnaeus 1758). *Sustainability* 2021;13,7093. <https://doi.org/10.3390/su13137093>
28. Bogut I, Adamek Z, Puskadija Z, Galovic, D. Nutritional value of planktonic cladoceran *Daphnia magna* for common carp (*Cyprinus carpio*) fry feeding. *Croat. J. Fish. Ribar.* 2010;68,1-10.
29. Sterner RW, Schulz KL. Zooplankton nutrition: recent progress and a reality check. *Aquat. Ecol.* 1998;32:261-279.
30. Stones CS, Mills DV. The use of live yeast and yeast culture products in aquaculture. *International Aquaculture feed.* 2004; (5):28-34.
31. Castro-Mejía Jorge, Ocampo-Cervantes Jose Antonio, Castro-Mejía German, Cruz-Cruz Irina, Monroy-Dosta Ma del Carmen, Becerril Cortes Dan. Laboratory production of *Daphnia magna* (Straus 1820) fed with microalgae and active dry yeast. *Journal of Entomology and Zoology Studies* 2016;4(2):548-553.
32. Herman H, Andriani Y, Sahidin A, Hidayat T, Herawati T. Culture of *Daphnia* sp. (crustacean-cladocera): the effect of manure variation on the growth, natality, and mortality. *IOP Conf. Series: Earth and Environmental Science.* 2018;2, 32-40. doi :10.1088/1755-1315/137/1/012018.
33. Monowar Asif Khan, Md. Mohibul Hasan, Kizar Ahmed Sumon Harunur Rashid. Culture of freshwater zooplankton *Daphnia magna* fed with different feed combination. *Bangladesh J. Fish.* 2020;32(1):55-59.

Good kind of research. The manuscript contributes to the scientific knowledge. Therefore, it should be **accepted**.