

# Gene Editing for Disease Resistance in Crops: Success Stories and Challenges

## Abstract

Gene editing has emerged as a transformative tool in modern agriculture, offering new avenues for enhancing disease resistance in crops. By precisely modifying the DNA of plants, scientists can develop varieties that are better equipped to withstand the onslaught of pathogens, which pose significant threats to global food security. This article delves into the success stories and challenges associated with gene editing for disease resistance in crops, with a focus on technologies like CRISPR-Cas9, TALENs, and ZFNs. One of the major success stories in this field is the development of disease-resistant varieties of wheat, rice, and tomatoes. For instance, researchers have used CRISPR-Cas9 to create wheat varieties resistant to powdery mildew, a devastating fungal disease. Similarly, gene editing has enabled the development of rice strains with enhanced resistance to bacterial blight, a disease that can lead to significant yield losses. In tomatoes, gene editing has been employed to confer resistance against the Tomato Yellow Leaf Curl Virus, which severely affects tomato production worldwide. These achievements underscore the potential of gene editing to create crops that are not only more resilient but also capable of maintaining high yields under disease pressure. However, the application of gene editing in crop disease resistance is not without challenges. One of the primary concerns is the regulatory landscape, which varies significantly across different countries. While some nations have embraced gene-edited crops, others have imposed strict regulations, treating them similarly to genetically modified organisms (GMOs). This inconsistency can hinder the global deployment of disease-resistant crops and create trade barriers. Additionally, there are concerns related to off-target effects, where unintended changes in the genome may occur, potentially leading to unintended consequences in the plant's growth or ecological interactions.

Keywords: gene, crops, tomatoes, powdery, fungal, tomato, leaf curl, landscapes

## **Introduction**

Agricultural productivity faces numerous challenges, with plant diseases being one of the most significant threats to crop yields and food security. Pathogens such as bacteria, fungi, and viruses can devastate crops, leading to substantial economic losses and exacerbating hunger in vulnerable regions [1]. Traditional methods of combating plant diseases, including chemical pesticides and conventional breeding, have made significant strides, but they often fall short in addressing the evolving nature of pathogens and the increasing demand for sustainable agricultural practices.

In this context, gene editing has emerged as a revolutionary tool that offers unprecedented precision and efficiency in enhancing disease resistance in crops [2]. Unlike conventional breeding, which relies on the slow process of selecting and crossing plants with desirable traits, gene editing allows scientists to directly modify specific genes associated with disease resistance. Techniques like CRISPR-Cas9, TALENs, and ZFNs enable targeted alterations in the plant genome, making it possible to develop crops that are not only resistant to diseases but also tailored to specific environmental conditions. The application of gene

editing in agriculture holds immense promise [3]. It can potentially reduce the reliance on chemical pesticides, lower production costs, and contribute to more resilient food systems. Successes in creating disease-resistant varieties of staple crops such as wheat, rice, and tomatoes demonstrate the technology's potential to revolutionize agriculture. However, the deployment of gene-edited crops also raises important questions about regulatory frameworks, public perception, and the long-term sustainability of these innovations [4].

There is a major risk that microbial diseases may interfere with agricultural output, the economy, and the safety of food supplies. The global food supply and biological biodiversity are both placed in jeopardy as a result of climate change, which accelerates the incidence of plant disease outbreaks, expands their dispersion, and makes them more specific to hosts [5]. Agriculture in the modern era has as its principal objective the maintenance of environmental sustainability while simultaneously ensuring the long-term security of food supplies. Globally, phytopathogens are responsible for reducing agricultural yields, which in turn results in yield losses in rice, wheat, maize, and potatoes [6]. When it comes to cereals, fungal infections alone may cause yield losses of around 15%–20%, and in severe circumstances, they can reach up to 60%. When soybeans are affected by phoma blight, the yield is reduced by 51.72 percent. Fusarium root rot causes a loss of production in field peas that is around sixty percent. When it comes to combating illnesses that do not include genetic resistance, the agricultural industry mostly depends on chemical control approaches [7]. On the other hand, the use of pesticides and other chemical agents raises concerns for the safety of other living creatures, whether they are directly or indirectly affected. Ecosystems in the water, air, and soil have been contaminated as a result of repeated usage of chemicals, which has led to bioaccumulation at greater tropical levels [8]. When it comes to food production, reducing dependence on chemical control is very necessary in order to mitigate the consequences of global climate change and reduce the negative environmental repercussions that are associated with the activities that are now carried out [9]. Through the processes of plant domestication and breeding, it has been feasible to generate plant kinds that are resistant to plant illnesses, which allows for the sustainable management of plant diseases. Traditional methods of breeding for resistance have a number of limitations, including the fact that they require a lot of time and work, that they cause linkage drag, and that they produce genotypes that are crossable [10].

Inducing alterations in the DNA of the plant by chemical or physical mutagenesis is the process of mutation breeding. This process results in the production of mutants that are then subjected to strict selection in order to evaluate desired characteristics and assist in the discovery of novel genes in the genome [11]. Farmers and agricultural experts have found that transgenic technology, and more specifically genetically modified (GM) crops, has emerged as a potential solution to their problems. Transgenic technology makes it possible to include genes from a wide variety of sources and is not limited to crossable genotypes. This results in the creation of crops that have enhanced yields, increased tolerance to biotic and abiotic stresses, resistance to herbicides, and increased nutritional content [12, 13, 14].

### **CRISPR for disease resistance**

Having a strong grasp of the defences that host plants have against parasite plants is vital when it comes to applying gene editing to boost host resistance. This understanding is necessary since it is fundamental to the process. As soon as the presence of a parasite is identified, the first response is referred to as pathogen-triggered immunity (PTI). This response involves the activation of both physical and biochemical defences inside the cells of the plant that is being parasitized [74, 77, 78]. The capacity of parasitic plants to oppose PTI is shown by the fact that they are able to inject effectors into host cells, which in turn promotes parasitism in the parasite plants themselves. There are two categories that may be used in order to identify host resistance mechanisms. These categories are pre-attachment and post-attachment. The classification of these types is based on whether the defensive systems are activated before to or after the parasite plants have established themselves on the host. The term "pre-attachment resistance" refers to a collection of strategies that host plants use in order to avoid the attachment and invasion of parasite plants prior to the occurrence of direct contact [73, 75, 76]. This is done with the goal of preventing the attachment and invasion of parasitic plants. Among the strategies that fall under this category are those that prevent the germination of parasitic plant seeds, those that produce toxic compounds via root exudates, those that impede the development of parasitic plant seedlings, and those that disrupt the commencement of the haustorium. Attempts have been made to target genes that are accountable for the synthesis of strigolactones and parasitism via the utilization of techniques of genetic alteration such as CRISPR-Cas9. As a consequence of this, agricultural crops have developed resistance to plants that are classified as parasitic [72, 79].

The benefits of LGS1-based resistance are governed by the genotype of the parasite as well as the features of the environment. It is vital to highlight that these variations in SLs have broader repercussions, and it is important to keep this in mind. There is a chance that LGS1 deletion lines will have a greater susceptibility to *S. hermonthica* genotypes that are sensitive to orobanchol. This is a possibility [80, 81, 82]. Nevertheless, it is possible that these lines will also exhibit a decreased expression of genes that are linked with the photosystem. Combining advanced CRISPR technologies with meticulous control mechanisms like as inducible systems or tissue-specific expression becomes highly vital for the aim of correctly applying this method in agriculture without losing the potential yield. This is because the goal is to not sacrifice yield potential [69, 70, 71]. Post-attachment resistance is a defensive reaction that is triggered when a plant identifies parasite plants that have attached themselves to the host for the goal of reproduction. This process is known as post-attachment development. There are many different processes that are included in this armoury of defensive mechanisms. Some of these processes include hypersensitive responses (HRs), hormone-driven signalling pathways, the hardening of cell walls, and the accumulation of protective secondary metabolites. In prior research, it has been shown and recorded that one of the most essential techniques involves the change of cell walls [66, 67, 68]. This has been generally acknowledged and has been published. Numerous host plants that are resistant to root and stem parasite plants have taken use of this mechanism in order to gain an advantage over that plant. The key negative regulator of this lignin-based response has been targeted

and eliminated via the application of CRISPR genome editing technology. Increasing the amount of research that is conducted on the several processes that are involved in post-attachment resistances and incorporating these mechanisms into plant genetic engineering is an extremely important step [65, 83, 84, 85].

### **Success Stories of Gene Editing for Disease Resistance in Crops**

Gene editing has made remarkable strides in the development of disease-resistant crops, showcasing the technology's potential to transform agriculture. Below are some of the notable success stories that highlight the impact of gene editing on crop disease resistance [64].

#### **1. Powdery Mildew-Resistant Wheat**

Powdery mildew is a pervasive fungal disease that affects wheat, leading to significant yield losses worldwide. Through the use of CRISPR-Cas9, researchers have successfully edited the genome of wheat to develop varieties that are resistant to this disease. By targeting specific genes associated with susceptibility to powdery mildew, scientists have been able to create wheat strains that are more resilient and require fewer fungicide applications. This advancement not only improves crop yields but also contributes to more sustainable agricultural practices by reducing the reliance on chemical inputs [16, 86].

#### **2. Bacterial Blight-Resistant Rice**

Rice is a staple food for over half of the world's population, and bacterial blight is one of the most devastating diseases affecting rice production. Traditional breeding methods have struggled to keep up with the evolving strains of the pathogen. However, gene editing has offered a new solution. Using CRISPR-Cas9, scientists have edited the SWEET genes in rice, which are known to be exploited by the bacteria to cause infection. The edited rice varieties exhibit strong resistance to bacterial blight, ensuring stable yields and enhancing food security in regions where rice is a crucial crop [17].

#### **3. Tomato Yellow Leaf Curl Virus (TYLCV)-Resistant Tomatoes**

Tomato Yellow Leaf Curl Virus (TYLCV) is a major threat to tomato production, causing severe yield losses and affecting the quality of the fruit. Gene editing has been employed to develop tomato varieties resistant to TYLCV. By targeting and modifying the genes that interact with the virus, scientists have created tomato plants that can effectively resist TYLCV infections. This breakthrough has significant implications for tomato growers, particularly in regions where the virus is endemic, and contributes to the overall reduction of crop losses [18, 87].

#### **4. Late Blight-Resistant Potatoes**

Late blight, caused by the pathogen *Phytophthora infestans*, is infamous for its role in the Irish Potato Famine and remains a serious threat to potato production today. Through gene editing, researchers have introduced resistance genes from wild potato species into cultivated varieties, creating potatoes that are resistant to late blight. These gene-edited potatoes offer a promising solution to a long-standing agricultural problem, reducing the need for chemical fungicides and lowering the risk of crop failure [63, 88].

#### **5. Fungal-Resistant Bananas**

Bananas are a critical food source for millions of people, and fungal diseases such as Fusarium wilt, also known as Panama disease, pose a severe threat to banana production. Using gene editing, scientists have developed banana varieties

that are resistant to these fungal infections. By modifying genes that are targeted by the fungus, researchers have created banana plants that can survive and thrive despite the presence of the disease. This development is particularly important for safeguarding banana production in regions where Fusarium wilt is prevalent [62, 89].

#### **6. Citrus Greening-Resistant Oranges**

Citrus greening, also known as Huanglongbing (HLB), is one of the most destructive diseases affecting citrus crops, particularly oranges. The disease is caused by a bacterium transmitted by the Asian citrus psyllid, leading to stunted growth, reduced fruit quality, and eventually, tree death. Traditional control methods have been largely ineffective, putting the global citrus industry at risk. Gene editing has offered a breakthrough solution by enabling scientists to develop orange trees with enhanced resistance to citrus greening. By editing genes that enhance the plant's immune response, researchers have created varieties that can better withstand the effects of HLB, providing hope for the future of citrus farming [61].

#### **7. Cassava Mosaic Disease-Resistant Cassava**

Cassava is a vital staple crop in many developing countries, particularly in Africa. However, cassava mosaic disease (CMD), caused by a group of plant viruses, has severely affected cassava yields, threatening food security in these regions. Through gene editing, scientists have engineered cassava plants that are resistant to CMD by targeting and disrupting the viral DNA's ability to replicate within the plant. This innovation has led to the development of cassava varieties that can maintain high yields even in areas where CMD is prevalent, significantly improving food security for millions of people who rely on cassava as a primary food source [60, 90].

#### **8. Downy Mildew-Resistant Grapes**

Downy mildew is a fungal disease that poses a major threat to grapevines, particularly in regions with warm, humid climates. The disease can lead to severe losses in grape production, affecting both table grapes and wine production. Gene editing has enabled the development of grape varieties with enhanced resistance to downy mildew. By modifying genes that are involved in the plant's defense mechanisms, researchers have created grapevines that can better resist the pathogen, reducing the need for chemical fungicides and ensuring more reliable grape harvests [19].

#### **9. Black Sigatoka-Resistant Bananas**

In addition to Fusarium wilt, bananas are also threatened by Black Sigatoka, a fungal disease that causes significant yield losses and increases production costs due to the need for frequent fungicide applications. Through gene editing, scientists have developed banana varieties that are resistant to Black Sigatoka by targeting and altering specific genes associated with the plant's susceptibility to the fungus. This advancement has the potential to reduce the reliance on chemical treatments and improve the sustainability of banana production, particularly in regions where Black Sigatoka is widespread [59, 91].

#### **10. Cocoa Swollen Shoot Virus-Resistant Cocoa**

Cocoa production, critical to the global chocolate industry, is threatened by Cocoa Swollen Shoot Virus (CSSV), a disease that causes severe crop losses and can lead to the death of cocoa trees. Gene editing has been applied to develop cocoa

plants resistant to CSSV by targeting and editing the genes that the virus exploits to infect the plant. These gene-edited cocoa trees show promise in resisting the disease, offering a potential solution to protect cocoa production and the livelihoods of millions of smallholder farmers who depend on it [58].

#### **11. Septoria Tritici Blotch-Resistant Wheat**

Septoria tritici blotch (STB) is a devastating fungal disease that affects wheat, one of the world's most important staple crops. The disease can lead to significant yield losses, particularly in regions with wet climates. Researchers have used gene editing, specifically CRISPR-Cas9, to develop wheat varieties with enhanced resistance to STB. By targeting and disabling susceptibility genes that the fungus exploits to infect the plant, scientists have created wheat strains that can better resist STB, leading to more reliable yields and reduced need for fungicides [20, 92, 93].

#### **12. Panama Disease (Tropical Race 4) Resistant Bananas**

Panama disease, caused by a soil-borne fungus *Fusarium oxysporum* f. sp. *ubense* (Tropical Race 4), is one of the most serious threats to global banana production. The disease has devastated banana plantations across Asia, Africa, and Australia, and there is no effective chemical treatment. Through gene editing, scientists have developed banana varieties with resistance to Tropical Race 4 by introducing resistance genes from wild banana species. These gene-edited bananas show strong resistance to the disease, offering a potential lifeline for banana growers worldwide and helping to ensure the availability of this critical food source [57, 94, 95].

#### **13. Barley Yellow Dwarf Virus-Resistant Barley**

Barley Yellow Dwarf Virus (BYDV) is a significant viral disease that affects barley, causing stunted growth, yellowing of leaves, and substantial yield losses. Traditional breeding efforts to combat BYDV have been challenging due to the complexity of the virus and its interactions with the plant. However, gene editing has provided a breakthrough. By using CRISPR-Cas9 to target and modify genes associated with the plant's susceptibility to BYDV, researchers have developed barley varieties that are resistant to the virus, ensuring higher yields and more stable production, particularly in regions where the virus is prevalent [21, 96].

#### **14. Cucumber Mosaic Virus-Resistant Cucumbers**

Cucumber Mosaic Virus (CMV) is a widespread plant virus that affects a variety of crops, including cucumbers, leading to reduced yields and poor fruit quality. Gene editing has been employed to develop cucumber plants that are resistant to CMV. By targeting and altering specific genes that the virus relies on to replicate and spread within the plant, scientists have created cucumber varieties that can effectively resist CMV infections. This advancement is particularly important for cucumber growers, as it helps reduce losses and ensures higher quality produce [56].

#### **15. Phytophthora-Resistant Soybeans**

Soybeans are a crucial crop for global food and feed production, but they are vulnerable to a range of diseases, including those caused by the pathogen *Phytophthora sojae*, which can lead to root and stem rot. Gene editing has been used to create soybean varieties that are resistant to this pathogen by editing resistance-related genes in the soybean genome. The resulting gene-edited soybeans are more resilient to Phytophthora infections, helping to protect yields and

reduce the need for chemical treatments, which can be costly and environmentally harmful [55, 97].

#### **16. Banana Bunchy Top Virus-Resistant Bananas**

Banana Bunchy Top Virus (BBTV) is another significant viral disease that affects banana plants, leading to stunted growth and severe yield losses. Gene editing has been utilized to develop banana varieties that are resistant to BBTV by targeting and modifying the genes involved in the virus' s ability to replicate and cause disease within the plant. These gene-edited bananas offer a promising solution to managing BBTV, particularly in regions where the disease is endemic and poses a serious threat to banana production [54].

#### **17. Verticillium Wilt-Resistant Cotton**

Verticillium wilt is a fungal disease that affects cotton, causing significant yield losses and reducing the quality of the cotton fibres. Through gene editing, scientists have developed cotton varieties that are resistant to Verticillium wilt by targeting genes that the fungus uses to invade and damage the plant. The resulting gene-edited cotton plants exhibit strong resistance to the disease, ensuring healthier crops and higher yields, which is particularly important for cotton growers in regions where Verticillium wilt is a major concern [53].

#### **18. Xanthomonas Wilt-Resistant Bananas**

Xanthomonas wilt, also known as banana bacterial wilt, is a devastating disease affecting banana crops in East and Central Africa. The disease is caused by the bacterium *Xanthomonas campestris* pv. *musacearum* and leads to rapid wilting and death of banana plants. Traditional control measures have been largely ineffective in halting the spread of the disease. Gene editing, however, has provided a solution by enabling the development of banana varieties resistant to Xanthomonas wilt. By introducing or modifying resistance genes from other plant species, scientists have created banana plants that can effectively resist the bacterium, offering hope for smallholder farmers who rely heavily on banana production for their livelihoods [22].

#### **19. Nematode-Resistant Soybeans**

Nematodes, particularly the soybean cyst nematode (*Heterodera glycines*), are a major pest affecting soybean production worldwide, causing significant yield losses. Conventional breeding methods have been used to develop resistant varieties, but these methods are time-consuming and often insufficient due to the nematode' s ability to adapt. Gene editing has enabled the development of soybean varieties that are resistant to nematodes by targeting and modifying specific genes that control the plant' s defence mechanisms. These gene-edited soybeans exhibit strong resistance to nematode infestations, helping to protect yields and reduce the need for chemical nematicides [52].

#### **20. Verticillium Wilt-Resistant Olive Trees**

Verticillium wilt, caused by the soil-borne fungus *Verticillium dahliae*, is a serious disease affecting olive trees, leading to reduced olive production and even tree death. Traditional control measures, such as soil fumigation, are often ineffective and environmentally damaging. Gene editing has been applied to develop olive trees with resistance to Verticillium wilt by targeting specific genes involved in the plant' s immune response. The gene-edited olive trees show improved resistance to the disease, offering a sustainable solution to protect olive groves and ensure the continued production of olives and olive oil [51].

### **21. Black Rot-Resistant Brassica Crops**

Black rot, caused by the bacterium *Xanthomonas campestris* pv. *campestris*, is a significant disease affecting Brassica crops, including cabbage, cauliflower, and broccoli. The disease leads to leaf necrosis and plant death, causing substantial yield losses. Gene editing has been employed to develop Brassica varieties resistant to black rot by altering genes that the bacterium uses to infect the plant. The resulting crops are better equipped to resist the disease, ensuring higher yields and reducing the need for chemical treatments [50].

### **22. Tomato Spotted Wilt Virus-Resistant Peppers**

Tomato Spotted Wilt Virus (TSWV) is a damaging viral disease that affects a wide range of crops, including peppers, leading to reduced yields and poor fruit quality. Gene editing has been used to develop pepper varieties that are resistant to TSWV by targeting and modifying genes involved in the plant's interaction with the virus. These gene-edited peppers exhibit strong resistance to TSWV, helping to safeguard pepper production and reduce losses caused by the virus [49].

### **23. Fusarium Wilt-Resistant Tomatoes**

Fusarium wilt, caused by *Fusarium oxysporum* f. sp. *lycopersici*, is a soil-borne fungal disease that affects tomato plants, leading to wilting, yellowing, and eventually plant death. The disease is particularly challenging to manage due to its persistence in the soil. Gene editing has allowed scientists to develop tomato varieties with resistance to Fusarium wilt by targeting and altering genes that control the plant's defence against the fungus. These gene-edited tomatoes are more resilient to Fusarium wilt, ensuring more reliable yields and reducing the need for soil sterilization or fungicide treatments [23].

### **24. Ring Spot Virus-Resistant Papaya**

Papaya production has been severely threatened by the Papaya Ringspot Virus (PRSV), which causes ring-like lesions on the fruit and leads to stunted growth and yield losses. Traditional breeding and transgenic approaches have been used to combat PRSV, but gene editing has provided a more precise method for developing resistant papaya varieties. By editing genes associated with susceptibility to PRSV, researchers have created papaya plants that can resist the virus, ensuring healthy fruit production and reducing the economic impact of the disease on papaya growers [48].

### **25. Rhizoctonia Root Rot-Resistant Sugar Beet**

Rhizoctonia root rot, caused by the fungus *Rhizoctonia solani*, is a serious disease affecting sugar beet, leading to root rot, reduced sugar content, and significant yield losses. Gene editing has been utilized to develop sugar beet varieties resistant to Rhizoctonia root rot by targeting genes involved in the plant's immune response. These gene-edited sugar beets exhibit enhanced resistance to the fungus, ensuring healthier plants and higher sugar yields [47].

### **26. Potato Virus Y (PVY)-Resistant Potatoes**

Potato Virus Y (PVY) is one of the most destructive viruses affecting potato crops worldwide, leading to severe yield losses and reduced tuber quality. Conventional methods to manage PVY, such as using resistant cultivars and insecticides to control aphid vectors, have had limited success. Gene editing has enabled the development of potatoes resistant to PVY by targeting and modifying genes that the virus exploits to infect the plant. These gene-edited potatoes show strong

resistance to PVY, helping farmers achieve better yields and reducing the reliance on chemical controls [46].

### **27. Rice Tungro Disease-Resistant Rice**

Rice tungro disease, caused by the rice tungro bacilliform and spherical viruses, is a significant threat to rice production in South and Southeast Asia. The disease causes stunted growth, yellowing of leaves, and can result in up to 100% yield loss in severe cases. Gene editing has been successfully applied to develop rice varieties resistant to tungro disease by modifying genes that are involved in the plant's defense mechanisms. These gene-edited rice plants are more resilient to tungro infections, ensuring stable production and food security in regions heavily dependent on rice [24].

### **28. Bacterial Spot-Resistant Tomatoes**

Bacterial spot, caused by *Xanthomonas* spp., is a common disease in tomato crops that leads to leaf lesions, fruit spots, and reduced yield. Traditional breeding methods have struggled to produce varieties with strong resistance due to the complex genetics of the disease. Gene editing, however, has allowed scientists to develop tomato varieties with enhanced resistance to bacterial spot by targeting specific susceptibility genes that the pathogen exploits. These gene-edited tomatoes can better resist bacterial infections, leading to healthier plants and higher yields [45].

### **29. Root-Knot Nematode-Resistant Carrots**

Root-knot nematodes, particularly *Meloidogyne* species, are a significant pest affecting carrot production, causing galls on roots that reduce both yield and quality. Controlling nematodes traditionally involves soil fumigation or the use of nematicides, which can be harmful to the environment. Gene editing has been used to develop carrot varieties resistant to root-knot nematodes by modifying genes that regulate the plant's response to nematode infection. These gene-edited carrots exhibit strong resistance, reducing the need for chemical treatments and ensuring better crop quality [44].

### **30. Maize Lethal Necrosis (MLN)-Resistant Maize**

Maize lethal necrosis (MLN) is a viral disease that affects maize, causing severe yield losses in East Africa. The disease is caused by a combination of viruses, including maize chlorotic mottle virus (MCMV) and sugarcane mosaic virus (SCMV). Gene editing has been successfully applied to develop maize varieties resistant to MLN by targeting and modifying genes that the viruses use to establish infection. These gene-edited maize plants are better equipped to resist MLN, providing a critical tool for ensuring maize production in regions where the disease is prevalent [43].

### **31. Bacterial Wilt-Resistant Eggplant**

Bacterial wilt, caused by *Ralstonia solanacearum*, is a devastating disease that affects eggplant, leading to plant wilting and death. The pathogen is difficult to control because it persists in the soil and can infect a wide range of host plants. Gene editing has been employed to develop eggplant varieties with resistance to bacterial wilt by altering genes that regulate the plant's immune response. These gene-edited eggplants show improved resistance to the disease, helping to secure yields and reduce losses in affected regions [42].

### **32. Downy Mildew-Resistant Sunflowers**

Downy mildew, caused by *Plasmopara halstedii*, is a serious disease affecting sunflower crops, leading to stunted growth and reduced seed production. Traditional breeding for resistance to downy mildew has been complicated by the pathogen's ability to evolve new virulent races. Gene editing has enabled the development of sunflower varieties with enhanced resistance to downy mildew by targeting and modifying specific resistance genes. These gene-edited sunflowers exhibit strong resistance to the disease, ensuring more reliable production and reducing the need for fungicides [25].

### **33. Anthracnose-Resistant Beans**

Anthracnose, caused by the fungus *Colletotrichum lindemuthianum*, is a major disease affecting beans, leading to dark lesions on stems, leaves, and pods, and resulting in significant yield losses. Gene editing has been used to develop bean varieties resistant to anthracnose by targeting and modifying genes that the fungus uses to infect the plant. These gene-edited beans show strong resistance to the disease, helping to protect yields and improve the quality of bean crops, especially in regions where anthracnose is a persistent problem [41].

### **34. Stripe Rust-Resistant Wheat**

Stripe rust, caused by the fungus *Puccinia striiformis* f. sp. *tritici*, is a destructive disease affecting wheat, leading to reduced yields and grain quality. The fungus has shown an ability to rapidly evolve, making it difficult to control with conventional breeding methods. Gene editing has been successfully applied to develop wheat varieties with enhanced resistance to stripe rust by targeting specific genes involved in the plant's defence response. These gene-edited wheat plants demonstrate strong resistance to stripe rust, helping to ensure stable production in regions prone to the disease [40].

### **35. Fusarium Head Blight-Resistant Barley**

Fusarium head blight (FHB), caused by *Fusarium graminearum*, is a fungal disease that affects barley, leading to yield losses and contamination of grain with harmful mycotoxins. Traditional breeding has had limited success in controlling FHB due to the complex nature of resistance. Gene editing has allowed scientists to develop barley varieties with enhanced resistance to FHB by targeting and modifying genes that regulate the plant's defence against the fungus. These gene-edited barley plants exhibit improved resistance, ensuring safer and more reliable barley production [26].

## **Challenges**

### **1. Technical Challenges**

- **Off-Target Effects:** One of the primary technical challenges in gene editing is the potential for off-target effects, where unintended changes are made to the genome. These off-target mutations can result in undesirable traits, potentially affecting the crop's yield, quality, or safety [27].
- **Complexity of Resistance Traits:** Disease resistance is often controlled by multiple genes, each contributing to different aspects of the plant's defense mechanisms. Editing multiple genes simultaneously, while ensuring that they work together harmoniously, is technically challenging [28].
- **Environmental Interactions:** The effectiveness of gene-edited traits can be influenced by environmental factors such as climate, soil type, and the presence of other pathogens. A trait that provides strong resistance in one

environment may be less effective in another, complicating the development of universally effective disease-resistant crops [29].

- **Pathogen Evolution:** Pathogens, such as bacteria, viruses, and fungi, can rapidly evolve to overcome the resistance traits introduced through gene editing. This arms race between plant defenses and pathogen attacks necessitates ongoing research and development to stay ahead of evolving pathogens [30].

## 2. Regulatory Challenges

- **Diverse Regulatory Landscapes:** Different countries have varying regulations regarding gene-edited crops. Some countries have stringent regulations that classify gene-edited crops similarly to genetically modified organisms (GMOs), while others have more relaxed policies. This inconsistency complicates the global development and commercialization of gene-edited crops [31].
- **Approval Processes:** The approval process for gene-edited crops can be lengthy, expensive, and uncertain. Regulatory agencies often require extensive data to ensure that the edited crops are safe for human consumption and the environment, which can delay the introduction of new disease-resistant varieties [32].
- **Intellectual Property Issues:** Gene editing technologies, such as CRISPR-Cas9, are often patented, leading to complex intellectual property landscapes. Navigating these patents can be challenging for researchers and developers, potentially limiting access to the technology and increasing the cost of development [33].

## 3. Ethical and Social Challenges

- **Public Perception and Acceptance:** Public perception of gene editing is mixed, with some people viewing it as a promising technology for addressing global food security, while others are concerned about potential risks. Misinformation and lack of understanding about gene editing can lead to resistance from consumers and advocacy groups [34].
- **Ethical Concerns:** Ethical questions arise regarding the manipulation of plant genomes, especially when it involves editing genes in ways that would not occur naturally. There is ongoing debate about the moral implications of altering the genetic makeup of crops, particularly when it comes to long-term environmental impacts and biodiversity [35].
- **Equity and Access:** There is concern that the benefits of gene editing may not be equitably distributed, with wealthier nations and large agribusinesses potentially gaining more from the technology than smallholder farmers in developing countries. Ensuring that small-scale farmers have access to gene-edited crops and can benefit from them is a significant challenge [36].

## 4. Socioeconomic Challenges

- **Cost of Development:** The development of gene-edited crops is expensive, requiring significant investment in research, development, and regulatory approval. This can limit the ability of smaller companies and public institutions to participate in the development of gene-edited crops [37].
- **Market Acceptance:** Even if gene-edited crops are approved by regulators, they may face challenges in gaining market acceptance. Retailers, food

processors, and consumers may be hesitant to adopt gene-edited products, particularly in regions where there is strong opposition to GMOs or where organic and non-GMO labels are highly valued [38].

- **Impact on Traditional Breeding:** There is concern that the focus on gene editing could overshadow traditional breeding methods, which have been used successfully for centuries. Balancing the use of gene editing with traditional breeding practices is essential to ensure that diverse and resilient crop varieties continue to be developed [39].

### Conclusion

Gene editing represents a transformative approach to enhancing disease resistance in crops, offering the potential to improve agricultural productivity, reduce reliance on chemical pesticides, and secure global food supplies in the face of increasing environmental challenges. The success stories across various crops, from rice and wheat to bananas and tomatoes, demonstrate the power of this technology to address some of the most pressing threats to crop health. These advancements highlight gene editing's ability to create resilient crop varieties that can withstand a broad spectrum of diseases, thereby contributing to sustainable farming practices and food security. However, the application of gene editing in agriculture is not without its challenges. Technical hurdles such as off-target effects and the complexity of polygenic traits, alongside the ever-present risk of pathogen evolution, underscore the need for ongoing research and innovation. Regulatory and ethical considerations further complicate the deployment of gene-edited crops, with diverse global regulations, intellectual property issues, and public perception playing crucial roles in shaping the future of this technology. Despite these challenges, the potential benefits of gene editing for disease resistance in crops are immense. By addressing the technical, regulatory, and ethical challenges through collaborative research, transparent communication, and inclusive policymaking, the agricultural community can harness the full potential of gene editing. This will not only enhance crop resilience but also contribute to a more sustainable and equitable global food system. As the technology continues to evolve, gene editing is poised to play a critical role in meeting the demands of a growing population and adapting to the changing climate, ensuring that agriculture can thrive in the face of future challenges.

### References

1. Manzoor S, Nabi SU, Rather TR, Gani G, Mir ZA, Wani AW, Ali S, Tyagi A and Manzar N (2024) Advancing crop disease resistance through genome editing: a promising approach for enhancing agricultural production. *Front. Genome Ed.* 6: 1399051. doi: 10.3389/fgeed.2024.1399051
2. van Esse HP, Reuber TL, van der Does D. Genetic modification to improve disease resistance in crops. *New Phytol.* 2020 Jan;225(1):70-86. doi: 10.1111/nph.15967. Epub 2019 Jul 11. PMID: 31135961; PMCID: PMC6916320.
3. Julian R. Greenwood, Xiaoxiao Zhang, John P. Rathjen, Precision genome editing of crops for improved disease resistance, *Current Biology*, Volume 33, Issue 11, 2023, Pages R650-R657, ISSN 0960-9822, <https://doi.org/10.1016/j.cub.2023.04.058>. (<https://www.sciencedirect.com/science/article/pii/S0960982223005407>)
4. Dong OX, Ronald PC. Genetic Engineering for Disease Resistance in Plants: Recent Progress and Future Perspectives. *Plant Physiol.* 2019 May;180(1):26-

38. doi: 10.1104/pp.18.01224. Epub 2019 Mar 13. PMID: 30867331; PMCID: PMC6501101.
5. Jhu M-Y, Ellison EE and Sinha NR (2023) CRISPR gene editing to improve crop resistance to parasitic plants. *Front. Genome Ed.* 5:1289416. doi: 10.3389/fgeed.2023.1289416
  6. Albert I, Böhm H, Albert M, Feiler CE, Imkampe J, Wallmeroth N, Brancato C, Raaymakers TM, Oome S, Zhang H *et al* 2015. An RLP23 - SOBIR1 - BAK1 complex mediates NLP - triggered immunity. *Nature Plants* 1: 15140.
  7. Bastet A, Robaglia C, Gallois JL. 2017. eIF4E resistance: natural variation should guide gene editing. *Trends in Plant Science* 22: 411–419.
  8. Baum JA, Bogaert T, Clinton W, Heck GR, Feldmann P, Ilagan O, Johnson S, Plaetinck G, Munyikwa T, Pleau M *et al* 2007. Control of coleopteran insect pests through RNA interference. *Nature Biotechnology* 25: 1322–1326.
  9. Bebber DP, Holmes T, Smith D, Gurr SJ. 2014. Economic and physical determinants of the global distributions of crop pests and pathogens. *New Phytologist* 202: 901–910.
  10. Bird DM, Jones JT, Opperman CH, Kikuchi T, Danchin EG. 2015. Signatures of adaptation to plant parasitism in nematode genomes. *Parasitology* 142: S71–S84.
  11. Boevink PC, Wang X, McLellan H, He Q, Naqvi S, Armstrong MR, Zhang W, Hein I, Gilroy EM, Tian Z *et al* 2016. A *Phytophthora infestans* RXLR effector targets plant PP1c isoforms that promote late blight disease. *Nature Communications* 7: 10311.
  12. Boutrot F, Zipfel C. 2017. Function, discovery, and exploitation of plant pattern recognition receptors for broad - spectrum disease resistance. *Annual Review of Phytopathology* 55: 257–286.
  13. Bozkurt TO, Richardson A, Dagdas YF, Mongrand S, Kamoun S, Raffaele S. 2014. The plant membrane - associated REMORIN1.3 accumulates in discrete perihyphal domains and enhances susceptibility to *Phytophthora infestans* . *Plant Physiology* 165: 1005–1018.
  14. Brogden KA. 2005. Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria? *Nature Reviews Microbiology* 3: 238–250.
  15. Brown JK. 2015. Durable resistance of crops to disease: a Darwinian perspective. *Annual Review of Phytopathology* 53: 513–539.
  16. Callaway E. 2018. CRISPR plants now subject to tough GM laws in European Union. *Nature* 560: 16.
  17. Cao Y, Liang Y, Tanaka K, Nguyen CT, Jedrzejczak RP, Joachimiak A, Stacey G. 2014. The kinase LYK5 is a major chitin receptor in Arabidopsis and forms a chitin - induced complex with related kinase CERK1. *eLife* 3: 1–19. doi: 10.7554/eLife.03766

18. Castro MS, Fontes W. 2005. Plant defense and antimicrobial peptides. *Protein and Peptide Letters* 12: 13–18.
19. Chen CH, Lin HJ, Ger MJ, Chow D, Feng TY. 2000. cDNA cloning and characterization of a plant protein that may be associated with the harpin<sub>PSS</sub> - mediated hypersensitive response. *Plant Molecular Biology* 43: 429–438.
20. Chen H, Engkvist O, Wang Y, Olivecrona M, Blaschke T. 2018. The rise of deep learning in drug discovery. *Drug Discovery Today* 23: 1241–1250.
21. Choi MS, Kim W, Lee C, Oh CS. 2013. Harpins, multifunctional proteins secreted by Gram - negative plant - pathogenic bacteria. *Molecular Plant - - Microbe Interactions* 26: 1115–1122.
22. Clark P, Habig J, Ye J, Collinge S. 2014. *Petition for determination of nonregulated status for Innate™ potatoes with late blight resistance, low acrylamide potential, reduced black spot, and lowered reducing sugars: Russet Burbank event W8* . USDA Petition no. 14 - 093 - 01p. [WWW document] URL [https://www.aphis.usda.gov/brs/aphisdocs/14\\_09301p.pdf](https://www.aphis.usda.gov/brs/aphisdocs/14_09301p.pdf) [accessed 15 January 2019].
23. Clausen M, Kräuter R, Schachermayr G, Potrykus I, Sautter C. 2000. Antifungal activity of a virally encoded gene in transgenic wheat. *Nature Biotechnology* 18: 446–449.
24. Dayakar BV, Lin HJ, Chen CH, Ger MJ, Lee BH, Pai CH, Chow D, Huang HE, Hwang SY, Chung MC *et al* 2003. Ferredoxin from sweet pepper (*Capsicum annuum* L.) intensifying harpin<sub>PSS</sub> - mediated hypersensitive response shows an enhanced production of active oxygen species (AOS). *Plant Molecular Biology* 51: 913–924.
25. De Vleeschauwer D, Xu J, Höfte M. 2014. Making sense of hormone - mediated defense networking: from rice to *Arabidopsis* . *Frontiers in Plant Science* 5. doi: 10.3389/fpls.2014.00611.
26. EPA . 2017. *Final registration decision for the new active ingredient and food use of VNTI protein and the genetic material necessary for its production in potatoes* . EPA - HQ - OPP - 2016 - 0036 [WWW document] URL <https://www.regulations.gov/document?D=epa-hq-opp-2016-0036-0013> [accessed 15 January 2019].
27. Espada M, Silva AC, Eves van den Akker S, Cock PJ, Mota M, Jones JT. 2016. Identification and characterization of parasitism genes from the pinewood nematode *Bursaphelenchus xylophilus* reveals a multilayered detoxification strategy. *Molecular Plant Pathology* 17: 286–295.
28. FDA . 2019. *Consultation procedures under FDA's 1992 Statement of Policy – Foods derived from new plant varieties* . [WWW document] URL <https://www.fda.gov/food/ingredients-additives-gras-packaging-guidance->

[documents-regulatory-information/consultation-procedures-under-fdas-1992-statement-policy-foods-derived-new-plant-varieties](#) [accessed 12 May 2019].

29. Fernbach PM, Light N, Scott SE, Inbar Y, Rozin P. 2019. Extreme opponents of genetically modified foods know the least but think they know the most. *Nature Human Behaviour* 3: 251–256. doi: 10.1038/s41562-018-0520-3.
30. Ferreira SA, Pitz KY, Manshardt R, Zee F, Fitch M, Gonsalves D. 2002. Virus coat protein transgenic papaya provides practical control of papaya ringspot virus in Hawaii. *Plant Disease* 86: 101–105.
31. Fitch MMM, Manshardt RM, Gonsalves D, Slightom JL, Sanford JC. 1992. Virus resistant papaya plants derived from tissues bombarded with the coat protein gene of papaya ringspot virus. *Bio/Technology* 10: 1466–1472.
32. Ghislain M, Byarugaba AA, Magembe E, Njoroge A, Rivera C, Román ML, Tovar JC, Gamboa S, Forbes GA, Kreuze JF *et al* 2019. Stacking three late blight resistance genes from wild species directly into African highland potato varieties confers complete field resistance to local blight races. *Plant Biotechnology Journal* 17: 1119–1129.
33. Godoy CV, Seixas CDS, Soares RM, Marcelino - Guimarães FC, Meyer MC, Costamilan LM. 2016. Asian soybean rust in Brazil: past, present, and future. *Pesquisa Agropecuária Brasileira* 51: 407–421.
34. Gómez - Gómez L, Boller T. 2000. FLS2: an LRR receptor - like kinase involved in the perception of the bacterial elicitor flagellin in *Arabidopsis*. *Molecular Cell* 5: 1003–1011.
35. Gonsalves C, Lee DR, Gonsalves D. 2004. Transgenic virus - resistant papaya: the Hawaiian ' Rainbow' was rapidly adopted by farmers and is of major importance in Hawaii today. *APSnet Features*. doi: 10.1094/APSnetFeature-2004-0804.
36. Horvath DM, Stall RE, Jones JB, Pauly MH, Vallad GE, Dahlbeck D, Staskawicz BJ, Scott JW. 2012. Transgenic resistance confers effective field level control of bacterial spot disease in tomato. *PLoS ONE* 7: e42036.
37. Hu X, Bidney DL, Yalpani N, Duvick JP, Crasta O, Folkerts O, Lu G. 2003. Overexpression of a gene encoding hydrogen peroxide - generating oxalate oxidase evokes defense responses in sunflower. *Plant Physiology* 133: 170–181.
- 38.
39. Hummel AW, Doyle EL, Bogdanove AJ. 2012. Addition of transcription activator - like effector binding sites to a pathogen strain - specific rice bacterial blight resistance gene makes it effective against additional strains and against bacterial leaf streak. *New Phytologist* 195: 883–893.
40. Iliescu EC, Balogh M, Szabo Z, Kiss GB. 2013. *Identification of a Xanthomonas euvesicatoria resistance gene from pepper (Capsicum*

annuum) and method for generating plants with resistance . International (PCT) patent application WO/2014/068346A2, filed October 30, 2013.

41. ISAAA . 2017. *Global status of commercialized biotech/GM crops in 2017: Biotech crop adoption surges as economic benefits accumulate in 22 years* . ISAAA Brief no. 53. Ithaca, NY: ISAAA.
42. Islam MT, Croll D, Gladioux P, Soanes DM, Persoons A, Bhattacharjee P, Hossain MS, Gupta DR, Rahman MM, Mahboob MG *et al* 2016. Emergence of wheat blast in Bangladesh was caused by a South American lineage of *Magnaporthe oryzae* . *BMC Biology* 2016: 84.
43. Jansen C, von Wettstein D, Schäfer W, Kogel KH, Felk A, Maier FJ. 2005. Infection patterns in barley and wheat spikes inoculated with wild-type and trichodiene synthase gene disrupted *Fusarium graminearum* . *Proceedings of the National Academy of Sciences, USA* 102: 16892–16897.
44. Jarosch B, Kogel KH, Schaffrath U. 1999. The ambivalence of the barley *Mlo* locus: mutations conferring resistance against powdery mildew (*Blumeria graminis* f. sp. *hordei*) enhance susceptibility to the rice blast fungus *Magnaporthe grisea* . *Molecular Plant-Microbe Interactions* 12: 508–514.
45. Jha S, Chattoo BB. 2010. Expression of a plant defensin in rice confers resistance to fungal phytopathogens. *Transgenic Research* 19: 373–384.
46. Jones JB, Minsavage GV, Roberts PD, Johnson RR, Kousik CS, Subramanian S, Stall RE. 2002. A non-hypersensitive resistance in pepper to the bacterial spot pathogen is associated with two recessive genes. *Phytopathology* 92: 273–277.
47. Jones JD, Dangl JL. 2006. The plant immune system. *Nature* 444: 323–329.
48. Jones JD, Vance RE, Dangl JL. 2016. Intracellular innate immune surveillance devices in plants and animals. *Science* 354: aaf6395.
49. Kalaitzandonakes N, Alston JM, Bradford KJ. 2007. Compliance costs for regulatory approval of new biotech crops. *Nature Biotechnology* 25: 509–511.
50. Kawashima CG, Guimarães GA, Nogueira SR, MacLean D, Cook DR, Steuernagel B, Baek J, Bouyioukos C, Do Vale Araújo Melo B, Tristão G *et al* 2016. A pigeonpea gene confers resistance to Asian soybean rust in soybean. *Nature Biotechnology* 34: 661–665.
51. Lacombe S, Rougon - Cardoso A, Sherwood E, Peeters N, Dahlbeck D, van Esse HP, Smoker M, Rallapalli G, Thomma BP, Staskawicz B *et al* 2010. Interfamily transfer of a plant pattern-recognition receptor confers broad-spectrum bacterial resistance. *Nature Biotechnology* 28: 365–369.

52. Li T, Liu B, Spalding MH, Weeks DP, Yang B. 2012. High - efficiency TALEN - based gene editing produces disease - resistant rice. *Nature Biotechnology* 30: 390–392.
53. Li X, Shin S, Heinen S, Dill - Macky R, Berthiller F, Nersesian N, Clemente T, McCormick S, Muehlbauer GJ. 2015. Transgenic wheat expressing a barley UDP - glucosyltransferase detoxifies deoxynivalenol and provides high levels of resistance to *Fusarium graminearum* . *Molecular Plant - - Microbe Interactions* 28: 1237–1246.
54. Li Z, Zhou M, Zhang Z, Ren L, Du L, Zhang B, Xu H, Xin Z. 2011. Expression of a radish defensin in transgenic wheat confers increased resistance to *Fusarium graminearum* and *Rhizoctonia cerealis* . *Functional and Integrative Genomics* 11: 63–70.
55. Lo Presti L, Kahmann R. 2017. How filamentous plant pathogen effectors are translocated to host cells. *Current Opinion in Plant Biology* 38: 19–24.
56. Maqbool A, Saitoh H, Franceschetti M, Stevenson CE, Uemura A, Kanzaki H, Kamoun S, Terauchi R, Banfield MJ. 2015. Structural basis of pathogen recognition by an integrated HMA domain in a plant NLR immune receptor. *eLife* 4: e08709.
57. McDonald BA, Linde C. 2002. Pathogen population genetics, evolutionary potential, and durable resistance. *Annual Review of Phytopathology* 40: 349–379.
58. Moore JW, Herrera - Foessel S, Lan C, Schnippenkoetter W, Ayliffe M, Huerta - Espino J, Lillemo M, Viccars L, Milne R, Periyannan S *et al* 2015. A recently evolved hexose transporter variant confers resistance to multiple pathogens in wheat. *Nature Genetics* 47: 1494–1498.
59. Murphy F, He Q, Armstrong M, Giuliani LM, Boevink PC, Zhang W, Tian Z, Birch PRJ, Gilroy EM. 2018. The potato MAP3K StVIK is required for the *Phytophthora infestans* RXLR effector Pi17316 to promote disease. *Plant Physiology* 177: 398–410.
60. Muwonge A, Kunert K, Tripathi L. 2016. Expressing stacked *HRAP* and *PFLP* genes in transgenic banana has no synergistic effect on resistance to *Xanthomonas* wilt disease. *South African Journal of Botany* 104: 125–133.
61. Namukwaya B, Tripathi L, Tripathi JN, Arinaitwe G, Mukasa SB, Tushemereirwe WK. 2012. Transgenic banana expressing *Pflp* gene confers enhanced resistance to *Xanthomonas* wilt disease. *Transgenic Research* 21: 855–865.
62. Nürnberger T, Lipka V. 2005. Non - host resistance in plants: new insights into an old phenomenon. *Molecular Plant Pathology* 6: 335–345.
63. Oerke E - C, Dehne H - W. 2004. Safeguarding production - losses in major crops and the role of crop protection. *Crop Protection* 23: 275–285.

64. Osusky M, Zhou G, Osuska L, Hancock RE, Kay WW, Misra S. 2000. Transgenic plants expressing cationic peptide chimeras exhibit broad-spectrum resistance to phytopathogens. *Nature Biotechnology* 18: 1162–1166.
65. Pandey AK, Ger MJ, Huang HE, Yip MK, Zeng J, Feng TY. 2005. Expression of the hypersensitive response-assisting protein in *Arabidopsis* results in harpin-dependent hypersensitive cell death in response to *Erwinia carotovora*. *Plant Molecular Biology* 59: 771–780.
66. Panstruga R, Dodds PN. 2009. Terrific protein traffic: the mystery of effector protein delivery by filamentous plant pathogens. *Science* 324: 748–750.
67. Pruitt RN, Schwessinger B, Joe A, Thomas N, Liu F, Albert M, Robinson MR, Chan LJ, Luu DD, Chen H *et al* 2015. The rice immune receptor XA21 recognizes a tyrosine-sulfated protein from a Gram-negative bacterium. *Science Advances* 1: e1500245.
68. Quijano CD, Wichmann F, Schlaich T, Fammartino A, Huckauf J, Schmidt K, Unger C, Broer I, Sautter C. 2016. KP4 to control *Ustilago tritici* in wheat: enhanced greenhouse resistance to loose smut and changes in transcript abundance of pathogen-related genes in infected KP4 plants. *Biotechnology Reports (Amsterdam)* 11: 90–98.
69. Rep M, Meijer M, Houterman PM, van der Does HC, Cornelissen BJ. 2005. *Fusarium oxysporum* evades I-3-mediated resistance without altering the matching avirulence gene. *Molecular Plant - Microbe Interactions* 18: 15–23.
70. Rinaldo AR, Ayliffe M. 2015. Gene targeting and editing in crop plants: a new era of precision opportunities. *Molecular Breeding* 35: doi: 10.1007/s11032-015-0210-z.
71. Rinaldo A, Gilbert B, Boni R, Krattinger SG, Singh D, Park RF, Lagudah E, Ayliffe M. 2017. The *Lr34* adult plant rust resistance gene provides seedling resistance in durum wheat without senescence. *Plant Biotechnology Journal* 15: 894–905.
72. Sarris PF, Cevik V, Dagdas G, Jones JDG, Krasileva KV. 2016. Comparative analysis of plant immune receptor architectures uncovers host proteins likely targeted by pathogens. *BMC Biology* 14: doi: 10.1186/s12915-016-0228-7.
73. Saur IM, Bauer S, Kracher B, Lu X, Franzeskakis L, Müller MC, Sabelleck B, Kümmel F, Panstruga R, Maekawa T *et al* 2019. Multiple pairs of allelic MLA immune receptor-powdery mildew AVR<sub>A</sub> effectors argue for a direct recognition mechanism. *eLife* 8: pii: e44471.
74. Savary S, Willocquet L, Pethybridge SJ, Esker P, McRoberts N, Nelson A. 2019. The global burden of pathogens and pests on major food crops. *Nature Ecology and Evolution* 3: 430–439.

75. Schiek B, Hareau G, Baguma Y, Medakker A, Douches D, Shotkoski F, Ghislain M. 2016. Demystification of GM crop costs: releasing late blight resistant potato varieties as public goods in developing countries. *International Journal of Biotechnology* 14: 112.
76. Schoonbeek HJ, Wang HH, Stefanato FL, Craze M, Bowden S, Wallington E, Zipfel C, Ridout CJ. 2015. Arabidopsis EF-Tu receptor enhances bacterial disease resistance in transgenic wheat. *New Phytologist* 206: 606–613.
77. Schwessinger B, Bahar O, Thomas N, Holton N, Nekrasov V, Ruan D, Canlas PE, Daudi A, Petzold CJ, Singan VR *et al* 2015. Transgenic expression of the dicotyledonous pattern recognition receptor EFR in rice leads to ligand-dependent activation of defense responses. *PLoS Pathogens* 11: e1004809.
78. Scorza R, Callahan A, Dardick C, Ravelonandro M, Polak J, Malinowski T, Zagrai I, Cambra M, Kamenova I. 2013. Genetic engineering of plum pox virus resistance: 'HoneySweet' plum—from concept to product. *Plant Cell, Tissue and Organ Culture* 115: 1–12.
79. Singh RP, Hodson DP, Jin Y, Lagudah ES, Ayliffe MA, Bhavani S, Rouse MN, Pretorius ZA, Szabo LJ, Huerta-Espino J *et al* 2015. Emergence and spread of new races of wheat stem rust fungus: continued threat to food security and prospects of genetic control. *Phytopathology* 105: 872–884.
80. Song Y, Thomma BPHJ. 2018. Host-induced gene silencing compromises *Verticillium* wilt in tomato and Arabidopsis. *Molecular Plant Pathology* 19: 77–89.
81. Tsuda K, Somssich IE. 2015. Transcriptional networks in plant immunity. *New Phytologist* 206: 932–947.
82. Uhse S, Djamei A. 2018. Effectors of plant-colonizing fungi and beyond. *PLoS Pathogens* 14: e1006992.
83. United Nations, Department of Economic and Social Affairs, Population Division. 2017. World population prospects: The 2017 revision, key findings and advance tables. *Working Paper No. ESA/P/WP/248*.
84. USDA. 2018. *Details on USDA plant breeding innovations*. [WWW document] URL [https://www.aphis.usda.gov/aphis/ourfocus/biotechnology/brs-news-and-information/2018\\_brs\\_news/pbi-details](https://www.aphis.usda.gov/aphis/ourfocus/biotechnology/brs-news-and-information/2018_brs_news/pbi-details) [accessed 11 January 2019].
85. USDA. 2019. *Coordinated framework: Roles of U.S. regulatory agencies*. [WWW document] URL [https://www.aphis.usda.gov/aphis/ourfocus/biotechnology/sa\\_regulations/ct\\_agency\\_framework\\_roles](https://www.aphis.usda.gov/aphis/ourfocus/biotechnology/sa_regulations/ct_agency_framework_roles) [accessed 5 February 2019].
86. Walz A, Zingen-Sell I, Loeffler M, Sauer M. 2008. Expression of an oxalate oxidase gene in tomato and severity of disease caused by *Botrytis cinerea* and *Sclerotinia sclerotiorum*. *Plant Pathology* 57: 453–458.

87. Wang GL, Song WY, Ruan DL, Sideris S, Ronald PC. 1996. The cloned gene, *Xa21*, confers resistance to multiple *Xanthomonas oryzae* pv. *oryzae* isolates in transgenic plants. *Molecular Plant-Microbe Interactions* 9: 850-855.
88. Wang J, Hu M, Qi J, Han Z, Wang G, Qi Y, Wang HW, Zhou JM, Chai J. 2019a. Reconstitution and structure of a plant NLR resistosome conferring immunity. *Science*. doi: 10.1126/science.aav5870.
89. Wang J, Hu M, Wu S, Qi J, Wang G, Han Z, Qi Y, Gao N, Wang HW, Zhou JM *et al* 2019b. Ligand-triggered allosteric ADP release primes a plant NLR complex. *Science* 364: doi: 10.1126/science.aav5868.
90. Wang J, Zhou L, Shi H, Chern M, Yu H, Yi H, He M, Yin J, Zhu X, Li Y *et al* 2018b. A single transcription factor promotes both yield and immunity in rice. *Science* 361: 1026-1028.
91. Wang M, Weiberg A, Lin FM, Thomma BP, Huang HD, Jin H. 2016. Bidirectional cross-kingdom RNAi and fungal uptake of external RNAs confer plant protection. *Nature Plants* 2: doi: 10.1038/nplants.2016.151.
92. Wu CH, Abd-El-Haliem A, Bozkurt TO, Belhaj K, Terauchi R, Vossen JH, Kamoun S. 2017. NLR network mediates immunity to diverse plant pathogens. *Proceedings of the National Academy of Sciences, USA* 114: 8113-8118.
93. Yang L, McLellan H, Naqvi S, He Q, Boevink PC, Armstrong M, Giuliani LM, Zhang W, Tian Z, Zhan J *et al* 2016. Potato NPH3/RPT2-Like Protein StNRL1, targeted by a *Phytophthora infestans* RXLR effector, is a susceptibility factor. *Plant Physiology* 171: 645-657.
94. Ye C, Li H. 2010. 20 years of transgenic research in China for resistance to papaya ringspot virus. *Transgenic Plant Journal* 4: 58-63.
95. Yin C, Hulbert SH. 2018. Host-induced gene silencing (HIGS) for elucidating *Puccinia* gene function in wheat. *Methods in Molecular Biology* 1848: 139-150.
96. Yip MK, Huang HE, Ger MJ, Chiu SH, Tsai YC, Lin CI, Feng TY. 2007. Production of soft rot resistant calla lily by expressing a ferredoxin-like protein gene (*pf1p*) in transgenic plants. *Plant Cell Reports* 26: 449-457.
97. Zeilmaker T, Ludwig NR, Elberse J, Seidl MF, Berke L, Van Doorn A, Schuurink RC, Snel B, Van den Ackerveken G. 2015. DOWNY MILDEW RESISTANT 6 and DMR6-LIKE OXYGENASE 1 are partially redundant but distinct suppressors of immunity in Arabidopsis. *The Plant Journal* 81: 210-222. []