

Impact of *Toxoplasma gondii* on Serum Protein Levels and Mineral Content in Sheep and Goats at a Small Dairy Farm in White Nile State, Sudan

Abstract

Aim: This study aims to investigate the effects of *Toxoplasma gondii* infection on the serum proteins and macro elements (Sodium, Potassium, Calcium, Magnesium and Phosphate) for sheep and goat. **Introduction:** In sheep and goats, *Toxoplasma* infection is a major cause of abortion and stillbirth. Subclinical infections are also quite common in adult animals of affected flocks and herds. Serum proteins and macro elements play an essential role in animal metabolism. **Material and Methods:** Serological survey of *T. gondii* was conducted using Latex Agglutination Test (LAT) for 20 sera collected from sheep and goats (8 and 12 respectively) in White Nile State, Sudan. **Results:** Out of twelve goats' serum and eight sheep serum samples, seven samples were found to be positive for *Toxoplasma* (58.33%) in goats and four samples were found to be positive for *Toxoplasma* (50%) in sheep, the remaining samples were negative. It was observed that serum Total protein (Tb) level was increased significantly ($p < 0.05$) in both species, and albumin (Alb) level decreased significantly ($p < 0.05$) in goats, while Globulin (Glob) was significantly ($p < 0.05$) low in sheep compared with control animals. The results showed that some minerals (Na and K) concentrations in the serum were higher in both sheep and goats compared to the controls. **Conclusion:** In conclusion infestation of goats with *Toxoplasma gondii* increases the total protein, globulins and significantly decreases the albumin concentrations. For sheep the infestation increases the total protein, albumin and significantly decreases the globulin concentrations. Sheep and goats infected with *Toxoplasma gondii* showed increased serum sodium, potassium and magnesium levels affecting most of the enzyme systems.

Key Word: Macro elements, Total proteins, *Toxoplasma*, sheep and goat.

Introduction

The global consumption of livestock, particularly sheep and goats, is particularly prevalent in developing nations (Mazinani & Rude, 2020). Among the numerous pathogens affecting these animals, *Toxoplasma gondii* stands out as an obligate intracellular protozoan belonging to the phylum Apicomplexa, subclass Coccidia (Bandyopadhyay, and Chattopadhyay, 2022.)

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Toxoplasmosis, caused by *T. gondii*, is a common parasitic zoonosis affecting a wide range of warm-blooded animals, including humans and domestic livestock (Stelzer *et al.*, 2019; Stanić, and Fureš, 2020)

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Sheep and goats, in particular, exhibit high rates of chronic *T. gondii* infection, often resulting in abortion and stillbirth (Nayeri *et al.*, 2021). Subclinical infections are also prevalent in adult animals within affected flocks and herds. The severity of toxoplasmosis in these animals correlates with the gestational stage; infections during early gestation can lead to fetal death, resorption, and abortion, while infections later in gestation may have minimal clinical effects, resulting in the birth of normal but infected and immune lambs (Dubey and Beattie, 1988; Buxton *et al.*, 2007).

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Beyond reproductive and economic losses, *T. gondii* infection in animals poses significant public health concerns. Consumption of infected meat or milk can facilitate zoonotic transmission, with the parasite being transmitted through various routes such as direct contact with animals, contaminated faeces, soil, herbage, or contaminated food and water (Jittapalapon *et al.*, 2005). The transmission cycle of *T. gondii* involves both definitive and intermediate hosts, as well as inter-host transmission (Calero-Bernal *et al.*, 2022; Lymbery, and Thompson, 2012).

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Minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iodine (I), manganese (Mn), copper (Cu), selenium (Se), and zinc (Zn) play crucial roles in governing successful reproductive processes (Fadlalla, 2022). These minerals, originating mainly from external sources, constitute approximately 4% of an animal's body weight and serve structural, physiological, catalytic, and regulatory functions (Bednarek and Bik, 1994; Suttle, 2000). Imbalances in mineral levels, whether due to excess or deficiency, can lead to alterations in serum levels and contribute to various pathological conditions, including infectious diseases (Asín *et al.*, 2021)

Therefore, the aim of this study is to investigate the effects of *Toxoplasma gondii* infection on serum proteins and macro elements in sheep and goats. Specifically, we aim to examine the impact of *T. gondii* infection on the content of selected minerals and total proteins in these animals.

Material and methods

This study was conducted on a farm located in White Nile State,, Sudan. The farm comprised a mix of eight male sheep and twelve female goats. The aim was to investigate the impact of Toxoplasmosis on serum total proteins, and mineral levels. The study was conducted on the local breeds. The age of more than three years, with the history of weakness, apportion, ticks infestation, loss of appetite, low milk production and reduced fertility were selected.

Sampling: Blood samples were collected from the jugular vein of both sheep and goats. The samples were transported to the Parasitology Laboratory at the College of Veterinary Medicine, University of Bahri. Upon arrival, serum was separated from the clotted blood and stored at - 20°C until further analysis. Initially, the sera were qualitatively examined for anti-*T. gondii* antibodies using the Latex Agglutination Test (LAT) according to the manufacturer's instructions (Spinreact, S.A./S.A.U, Spain). Sera showing positive or doubtful reactions were subsequently diluted in twofold serial increments (1:2 up to 1:128) for further testing.

All sera were subjected to appropriate analytical procedure to determinate different biochemical parameters. Serum calcium was determined by the Photometric Test CPC method according to the procedure described by (Gitelman, 1967). The concentration of the phosphate PO₄ in serum sample was determined by (Shervedani and Bagherzadeh 2008). Serum sodium and potassium was determined colormetrically according to the method of Trender(Trender, 1951). Serum magnesium was determined by Howard method (Howard, 1964).

Serum total protein levels were assessed in each serum sample using the methodology outlined by Lowry *et al.*, (1951). Serum albumin was quantified colorimetrically employing the dye binding technique with Bromocresol Green (BCG), following the protocol outlined by Young (1975). Serum globulin concentrations were determined using Enzyme-Linked Immunosorbent Assay (ELISA), as per the procedure outlined by Piomelliet *al.*, (1998).

Statistical analysis:Statistical analysis was performed using SPSS-version 20 software (IBM, Armonk, NY, USA.).An independent sample T-test was used to analyze the effects of toxoplasmosis. Parametric data were expressed in tables as means ± SDs.

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Results

As depicted in Table 1, examination of eight blood samples collected from sheep showed that four samples (50%) tested positive, whereas the remaining four samples tested negative.

Similarly, analysis of twelve blood samples obtained from goats revealed that seven samples (58.33%) tested positive for the presence of the target analyte, while the remaining five samples (41.66%) yielded negative results.

Table (1): percentage of *Toxoplasma gondii* in the farm

Bacteria	species	Tested samples	Positive	Chi-square value	P-value
Toxoplasma	Sheep	8	4 (50%)	0.13	0.71
	Goats	12	7 (58.33%)		

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Form serum protein concentrations results were illustrated in table (2). In sheep, toxoplasmosis infection lead to increase in total protein and albumin and significant ($p < 0.05$) decreased in globulins concentrations in positive sheep compared with negative sheep. For goats, toxoplasmosis infection lead to increase in total protein and globulins and significant ($p < 0.05$) decreased in albumin concentrations in positive goats compared with negative goats.

Table (2): Serum protein concentrations in goats and sheep with Toxoplasmosis

Species	Sheep		Goats	
Disease	Toxoplasma		Toxoplasma	
Parameters	Control (negative)	Positive	Control (negative)	Positive
TP g/l	65.36±5.73	77.57±2.94	70.16±13.03	92.75±5.33
Alb g/l	48.74±3.69	64.77±3.99	58.82±11.19	58.31±4.72*
Glob g/l	16.61±2.04	12.79±6.12*	11.34±2.79	34.43±4.98

TP= Total proteins, Alb= Albumin, Glob = Globulin, *Significantly different from control at ($P < 0.05$). *Significantly different from control at ($P < 0.01$).

For minerals the results were shown in table (3): In sheep, toxoplasmosis infection lead to increase in Na, K, Ca, Mg and PO₄ in positive sheep compared to the negatives. While in goats, toxoplasmosis infection lead to significant (p<0.05) increased in Na⁺ and Mg concentration and significant (p<0.001) increased in K. Also, increased in both Ca and PO₄ in the positive goats compared with negatives.

Table (3): Serum minerals concentrations in m.mol/L in sheep and goats with toxoplasmosis

Species	Sheep		Goats	
Disease	Toxoplasma		Toxoplasma	
Parameters	Control (negative)	Control (negative)	Positive	Positive
Na m.mol/L	125.47±4.13	126.61±4.02	137.18±1.83*	139.62±1.91
K m.mol/L	3.55±0.26	3.79±0.30	4.44±0.09***	4.44±0.06
Cam.mol/L	2.26±0.04	2.30±0.07	2.44±0.03	2.36±0.02
Mg m.mol/L	0.68±0.06	0.92±0.22	0.95±0.01*	0.96±0.01
PO ₄ m.mol/L	1.16±0.03	1.18±0.05	1.24±0.05	1.29±0.01

*Significantly different from control at (P<0.05).

*Significantly different from control at (P<0.01).

Discussion

Small ruminants, such as sheep and goats, are particularly vulnerable to *T. gondii*, resulting in various problems in these animals (Holec-Gąsior, and Sołowińska, 2023).

In the present study Latex agglutination test was used to detect the sero-prevalence of *Toxoplasma gondii* in sheep and goats in a small farm in western Omdurman Sudan.

The current investigation revealed that over 50% of the goats (seven out of twelve) and sheep (four out of eight) tested positive for *Toxoplasma* infection, signifying a significant prevalence among the livestock population at the farm. These findings are consistent with previous research (Dubey, 2009b), which underscores the high susceptibility of sheep and goats to *T. gondii* infections, making this protozoan parasite a leading cause of reproductive losses in small

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ruminants globally. While much of the research has focused on sheep, toxoplasmosis holds similar, if not greater, significance as a cause of abortion in goats (Dubey, 2010b). A recent meta-analysis conducted in Africa, spanning data from 1969 to 2016, estimated an overall prevalence of 26.1% (with a range of 17.0–37.0%) for sheep and 22.9% (with a range of 12.3–36.0%) for goats (Tonouhewa et al., 2017).

For sheep with toxoplasmosis there was an increase in total protein and albumin concentrations and decreased a significant ($p < 0.05$) in globulin concentrations. On the other hand goats with toxoplasmosis infection there was an increase in total protein and globulins concentrations and significant ($p < 0.05$) decreased in albumin compared with negative ones. These results are in agreement with that recorded by Roland *et al.*, (2022) who reported that, a significant increase in albumin and ALT ($p = 0.04$) was observed between small ruminants infested with toxoplasmosis compared to the control group. These results are also similar to those obtained by Nora *et al.*, (2018), who indicated that ALT and albumin activities were increased in animals infected with *T. gondii*. These results could be explained by the fact that during toxoplasmic infestation there is a visceral hypertrophy, especially in the liver (El-Sayed *et al.*, 2016). Also Kaneko *et al.*, (2008) and Roussel, *et al.*, (1997) reported that some of the parameters were significantly altered due to protozoan infections. The reduction of serum total protein and albumin concentrations were observed in both Toxoplasma, Eimeria, and Cryptosporidium infections. It has been reported that protein requirement increase in the presence of any lesions on the body and chronic organ disease (El-Manyawee *et al.*, 2010). A significant decrease of total serum proteins in goats with internal parasitic infections was recorded by Moudgil *et al.*, (2017). This was attributed to protein-losing gastroenteropathy and malabsorption of proteins from damaged intestinal mucosa (Jesse *et al.*, 2019). The increased concentrations of total serum protein in goats with internal parasites was also observed by Hassan *et al.*, (2012).

Minerals play a vital role in maintaining homeostasis by being involved in all living processes, either in the capacity of structural elements or as regulators of almost all metabolic processes (Bauman and Currie, 1980). Significant increase ($P > 0.05$) of sodium, potassium and Phosphate in small ruminants reported in this study table (3) was on contradiction to that obtained by Soheiret *et al.*, (2010) who reported low levels of sodium and potassium in serum of infected goats compared to controls. This low level may be attributed to diarrhea and reduced absorption in the infected

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tissues of intestine (Ciraket *et al.*, 2004). For calcium concentrations our results showed that there is no significant change in serum calcium of goat and significant increase ($P < 0.01$) in magnesium concentration regarding the negative controls. This was in disagreement with Genget *et al.*, (2001) who reported that Toxoplasma infected goat serums showed a reduction in magnesium, calcium and potassium concentration. But in our study the mean positive serum Mg concentration of sheep are not significantly increased compared to the negative controls, no significant differences could be observed with that reported by Seyreket *et al.*, (2004) who reported that the mean concentration of Mg in serum was higher but not significant ($P > 0.05$) in sero-positive sheep than in their controls. Magnesium (Mg) is most abundant divalent cation in intracellular fluid. It is needed in many enzymes such as alkaline phosphatase, ATPase (Maji *et al.*, 2023). Sheep and goats are highly susceptible to the infections with *T. gondii* and this protozoan parasite is considered as a major cause of reproductive losses in small ruminants worldwide (Ali *et al.*, 2019).

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Conclusion

Infestation with Toxoplasma in sheep elicits a pronounced elevation in total protein and albumin levels, concomitant with a significant reduction in globulin concentrations. Conversely, Toxoplasma gondii infestation in goats demonstrates an augmentation in total protein and globulin levels, coupled with a noteworthy decline in albumin levels, potentially impacting various enzymatic pathways. Furthermore, sheep and goats afflicted with toxoplasmosis exhibit heightened serum concentrations of sodium, potassium, and magnesium. These findings underscore the imperative for stringent protozoan disease management strategies in small-scale farming operations to mitigate substantial economic repercussions on animal productivity.

Availability of Data and Material

Data are available within the submitted article.

References

Ali, S., Zhao, Z., Zhen, G., Kang, J.Z. and Yi, P.Z., 2019. Reproductive problems in small ruminants (Sheep and goats): A substantial economic loss in the world. *Large Animal Review*, 25(6), pp.215-223.

Anand, C.V., 1994. Statistics for analytical chemistry second edition by JC Miller and JN Miller. pp 227. Prentice Hall, Englewood Cliffs, NJ. 1992 ISBN 0-13-845421-3.

Asín, J., Ramírez, G.A., Navarro, M.A., Nyaoke, A.C., Henderson, E.E., Mendonça, F.S., Molín, J. and Uzal, F.A., 2021. Nutritional wasting disorders in sheep. *Animals*, 11(2), p.501.

Bandyopadhyay, P.K., Das, N.R. and Chattopadhyay, A., 2022. Protozoan Parasites. In *Biochemical, Immunological and Epidemiological Analysis of Parasitic Diseases* (pp. 9-90). Singapore: Springer Nature Singapore.

Buxton, D., Maley, S.W., Wright, S.E., Rodger, S., Bartley, P. and Innes, E.A., 2007. *Toxoplasma gondii* and ovine toxoplasmosis: new aspects of an old story. *Veterinary parasitology*, 149(1-2), pp.25-28.

Calero-Bernal, R., Fernández-Escobar, M., Katzer, F., Su, C. and Ortega-Mora, L.M., 2022. Unifying virulence evaluation in *Toxoplasma gondii*: a timely task. *Frontiers in Cellular and Infection Microbiology*, 12, p.868727.

Cirak, V.Y., Kowalik, S., Bürger, H.J., Zahner, H. and Clauss, W., 2004. Effects of *Eimeria separata* infections on Na⁺ and Cl⁻ transport in the rat large intestine. *Parasitology research*, 92, pp.490-495.

Dubey, J.P., 2010b. *Toxoplasmosis of Animals and Humans*. 2nd edition. CRC Press, Boca Raton.

El-Manyawe, S.M., Abdel Rahman, M.A.M., Abd El Aal, A.M.I., Kamal, A.M. and Snousi, S.A., 2010. Prevalence of some protozoa and its effects on biochemical changes in goats in Cairo, Marsa Matrouh, and El-Wadi El-Gadid provinces. *Egypt J Comp Path & Clinic Path*, 23, pp.102-115.

Fadlalla, I.M., 2022. The Interactions of Some Minerals Elements in Health and Reproductive Performance of Dairy Cows. *New Advances in the Dairy Industry*.

Geng, Z.H., Fang, Y.Q., Liu, L., Shi, Y. and Li, S.H., 2001. Determination of T lymphocytes and trace elements in spleen from rats infected with *Toxoplasma gondii*. *Zhongguoji sheng chongxue yuji sheng chongbing zazhi* = Chinese journal of parasitology & parasitic diseases, 19(6), pp.357-359.

Gitelman, H.J., 1967. An improved automated procedure for the determination of calcium in biological specimens. *Analytical Biochemistry*, 18(3), pp.521-531.

- Hassan, M., Hoque, M.A., Islam, S.A., Khan, S.A., Hossain, M.B. and Banu, Q., 2012. Efficacy of anthelmintics against parasitic infections and their treatment effect on the production and blood indices in Black Bengal goats in Bangladesh. *Turkish Journal of Veterinary & Animal Sciences*, 36(4), pp.400-408.
- Holec-Gąsior, L. and Sołowińska, K., 2023. Detection of *Toxoplasma gondii* Infection in Small Ruminants: Old Problems, and Current Solutions. *Animals*, 13(17), p.2696.
- Howard H Sky-Peck (1964). *Clinical Chemistry*, Volume 10, Issue 5, 1 May 1964, Pages 391–398.
- Jesse, F.F.A., Bitrus, A.A., Chung, E.L.T., Peter, I.D., Mohd, M.A. and Salleh, L.A., 2019. Severe parasitic gastroenteritis (PGE) in a goat: a veterinary case report and way forward. *The Thai Journal of Veterinary Medicine*, 49(3), pp.295-299.
- Jittapalapong, S., Sangvaranond, A., Pinyopanuwat, N., Chimnoi, W., Khachaeram, W., Koizumi, S. and Maruyama, S., 2005. Seroprevalence of *Toxoplasma gondii* infection in domestic goats in Satun Province, Thailand. *Veterinary Parasitology*, 127(1), pp.17-22.
- Kaneko, J.J., Harvey, J.W. and Bruss, M.L. eds., 2008. *Clinical biochemistry of domestic animals*. Academic press. 6th ed., Academic Press, Inc., San Diego, London, Boston, New York, Sydney, Tokyo, Toronto, pp. 882-888.
- Lowry O. H., Rosebrough N. J., Farr A. L., Randall RJ (1951). Protein Measurement with the Folin Phenol Reagent. *J Biol Chem* 193: 265-275.
- Lymbery, A.J. and Thompson, R.C.A., 2012. The molecular epidemiology of parasite infections: tools and applications. *Molecular and biochemical parasitology*, 181(2), pp.102-116.
- Maji, C., Biswas, S. and Kaur, J., 2023. Nutritional Deficiency Diseases in Goats. *Principles of Goat Disease and Prevention*, pp.221-236.
- Mazinani, M. and Rude, B., 2020. Population, world production and quality of sheep and goat products. *American Journal of Animal and Veterinary Sciences*, 15(4), pp.291-299.
- Moudgil A.D., Sharma, A. Verma, M.S. Kumar, R. Dogra, P.K. and Moudgil, P. "Gastrointestinal parasitic infections in Indian Gaddi (goat) breed bucks: clinical, hemato-biochemical, parasitological and chemotherapeutic studies," *Journal of Parasitic Diseases*, vol. 41, no. 4, pp. 1059–1065, 2017.
- Nayeri, T., Sarvi, S., Moosazadeh, M. and Daryani, A., 2021. Global prevalence of *Toxoplasma gondii* infection in the aborted fetuses and ruminants that had an abortion: A systematic review and meta-analysis. *Veterinary parasitology*, 290, p.109370.

Nora, M.B., Adel, A.H., AhMed, M.A., HaNy, M. I., 2018. Biochemical Effects of Toxoplasma gondii and Neosporacanium Infection on Diary Bovine Models in Menoufia Province, Egypt. *Advances in Adv. Anim. Vet. Sci. Biochem.*, 4(4): 527–534.

Piomelli, D., Beltramo, M., Giuffrida, A. and Stella, N., 1998. Endogenous cannabinoid signaling. *Neurobiology of disease*, 5(6), pp.462-473.

Roland, N.C., Kouamo, J., Prudence, K.K.A., Tchizze, G.J.T. and Ngoula, F., 2022. Metabolic Profiles Associated with Toxoplasma gondii Infestation in Goats and Sheep in Cameroon. *PSM Veterinary Research*, 7(1), pp.20-30.

Roussel, A.J., Whitney, M.S. and Cole, D.J. (1997): "Interpreting a bovine serum chemistry profile", part 1. *Vet.Med.*, 92:553-558.

Sayed, N.M., Ramadan, M.E. and Ramadan, M.E., 2016. Toxoplasma gondii infection and chronic liver diseases: evidence of an association. *Tropical medicine and infectious disease*, 1(1), p.7.

Shervedani, R.K. and Bagherzadeh, M., (2008). Hydroxamation of gold surface via in-situ layer-by-layer functionalization of cysteamine self-assembled monolayer: Preparation and electrochemical characterization. *Electrochimica Acta*, 53(22), pp.6293-6303.

Soheir M. El Manyaw*, M.A.M. AbdelRahman*, A.M.I. Abd El Aal***, Azza M. Kamal** and Samira A. Snousi (2010). Prevalence of some protozoa and its effects on biochemical changes in goats in Cairo, Marsa Matrouh, and El-Wadi El-Gadid provinces. *Egypt. J. Comp. Path. & Clinic. Path.* Vol.23 No.1 (February) 2010; 102 – 115.

Stanić, Ž. and Fureš, R., 2020. Toxoplasmosis: a global zoonosis. *Veterinaria*, 69(1).

Stelzer, S., Basso, W., Silván, J.B., Ortega-Mora, L.M., Maksimov, P., Gethmann, J., Conraths, F.J. and Schares, G., 2019. Toxoplasma gondii infection and toxoplasmosis in farm animals: Risk factors and economic impact. *Food and waterborne parasitology*, 15, p.e00037.

Suttle, N.F., 2000. Minerals in livestock production. *Asian-Australasian Journal of Animal Sciences*, 13, pp.1-9.

Tonouhewa, A.B., Akpo, Y., Sessou, P., Adoligbe, C., Yessinou, E., Hounmanou, Y.G., Assogba, M.N., Youssao, I., Farougou, S., 2017. Toxoplasma gondii infection in meat animals from Africa: systematic review and meta-analysis of sero-epidemiological studies. *Vet World* 10, 194–208.

Trinder, P., 1951. A rapid method for the determination of sodium in serum. *Analyst*, 76(907), pp.596-599.

Young DS (1975).Quantitative colorometric determination of albumin in serum or plasma.ClinChem 21: 244.

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