

Original Research Article

Feeding behavior of imidacloprid resistant brown planthopper, *Nilaparvata lugens* (Stål)

ABSTRACT

A study was conducted to investigate the feeding behavior of imidacloprid resistant (RS) and laboratory (LS) strains of brown planthopper, *Nilaparvata lugens*, by using standard honeydew and probing tests. The honeydew production was higher in RS strain (7.12 and 14.63 cm²) as compared to LS strain (5.44 and 1.86 cm²) in both the imidacloprid treated and untreated conditions, respectively. The probing test revealed an increased probing activity by both the LS and RS strains on imidacloprid treated seedlings as compared to the untreated. The number of probing marks by LS and RS strains on the treated seedlings were 16 and 31, whereas in untreated seedlings 7 and 11 respectively. These results established the higher feeding capacity of imidacloprid resistant *N. lugens* strain as compared to the laboratory strain, measured in terms of probing behaviour and honeydew production.

Key words: Honeydew, probing, insecticide, imidacloprid, resistance, rice and brown planthopper.

1. INTRODUCTION

The brown planthopper, *Nilaparvata lugens* (Stål) (Hemiptera: Delphacidae), is one of the most notorious pests of rice. Due to the short life cycle and high fecundity, the population of *N. lugens* can increase rapidly under favourable conditions inflicting heavy crop losses. Besides, *N. lugens* also acts as a vector of grassy stunt, ragged stunt and wilted stunt viruses (1,2,3). Application of chemical insecticides has been a primary tool for managing this pest. However, due to extensive use, *N. lugens* has developed resistance to almost all the major classes of insecticides. Currently, *N. lugens* has developed resistance to 36 active ingredients of insecticides, with 432 reported cases across the globe, 11 posing serious challenges for the development of new insecticides and pest resistance management (4). Increased insecticide resistance has limited the efficacy of insecticides leading to a significant increase in production costs and control failure. Imidacloprid, the first commercialised neonicotinoid insecticide, was introduced for planthopper control in the early 1990s. It is a systemic

Commented [A1]: As per the IPM?

Commented [A2]: Extensive use of insecticides....

insecticide that translocate rapidly through the plant tissue and proved extremely effective against sucking pests. It disrupts the insect nervous system by competitive modulation of nicotinic acetylcholine receptors (nAChR) (5). Insecticide resistant insects often exhibit increased energy consumption or disturbances in their metabolic balance, resulting in certain fitness cost and change of feeding behaviour (6). Resistant insect populations show slower developmental rates, reduced survival rates and fecundity (7,8,9). Since, the feeding behaviour and dietary habits of an insect directly influence the fitness parameters such as developmental characteristics and fecundity (10), it is worthwhile to study the feeding behaviour of insecticide resistant populations of *N. lugens*. Resistant *N. lugens* populations exhibited distinct feeding patterns and increased reproduction rates when exposed to insecticides like deltamethrin, methyl parathion, cypermethrin, and fenvalerate (11). Several studies have examined the feeding behaviours of sap sucking insects such as aphids, planthoppers, and whiteflies, employing electrical penetration graph (EPG) (12,13,14). Insecticides also impact pest dispersal, locomotion, reproduction, feeding and host-finding behavior (15). Exposure of the potato psyllid, *Bactericera cockerelli* to imidacloprid resulted in reduced probing time, increased periods of rest and the ultimate abandonment of leaflets (16). The probing behavior of *Frankliniella fusca* was altered on imidacloprid and cyantraniliprole treated peppers (17). In this context the present study was conducted to understand the feeding behaviour of imidacloprid resistant *N. lugens* population.

2. MATERIAL AND METHODS

2.1 Insects

The laboratory strain (LS) of *N. lugens* was obtained from Bayer Biosciences, Hyderabad which was being maintained in glasshouse for about five years without exposure to any insecticides. Further, LS was maintained in the glass house at ICAR-Indian Institute of Rice Research, Hyderabad for 10 generations without exposure to any insecticides. The resistant strain (RS) was continuously selected from the LS for 10 generations by exposing to sub-lethal doses of imidacloprid at each generation. The third instar nymphs were sprayed with sub-lethal doses of imidacloprid and the surviving nymphs were advanced to the next generation. These strains were maintained in insect proof cages on 45-day old potted rice plants (TN-1) at 27 ±1 °C temperature, 70-80 per cent relative humidity and 16:8 h light:dark photoperiod.

Commented [A3]: This reference is not related to this MS. Pl delete. The mentioned insecticides (organochlorine, synthetic pyrethroids etc.) are not recommended to manage *N. lugens* as well as they are not fall under the group of insecticide (imidacloprid) which was tented by the author(s).

Commented [A4]: Please mention the sub lethal dose of imidacloprid applied and how the insecticide was exposed i.e. sprayed to the tested LS insects.

2.2 Insecticide

Commercial imidacloprid 17.8SL formulation (Confidor, Bayer Crop Sciences) was used for the experiments.

2.3 Feeding test

The feeding behaviour of LS and RS strains of *N. lugens* was assessed by measuring honeydew excretion, which serves as an indicator of their feeding preferences and efficiency. Feeding capacity of *N. lugens* was determined by ninhydrin method (18) with suitable modifications. The lower 10 cm stems from 50-days old rice plant were cut, shade-dried and dipped in the imidacloprid solution (300 ppm) for 30s. Distilled water without any insecticide served as a control. Treated stem was inserted through the centre of a 15 cm dia. Whatman No.1 filter paper laid on a plastic card which is placed on a cup. A layer of water, touching the plant roots was maintained in the cup. Finally, each stem was enclosed within an inverted cup. Three newly formed brachypterous females were released per treatment. There were five replications per treatment. The insects were allowed to feed for 24 h. The filter papers with honeydew deposition were collected and treated with 0.001% ninhydrin in acetone solution followed by oven drying at 100°C for 5 minutes. Due to their amino acid content honeydew stains appeared violet or purple and these coloured areas were copied on a tracing paper and measured using Image J software.

2.4 Probing test

One seven-day-old rice seedling (variety TN-1) was placed in a test tube (25 mm X 150 mm) containing a 5 mm layer of water at the bottom. One newly formed brachypterous female was introduced into the tube and the tube opening was covered with a muslin cloth. After 24 hours, the insects were removed and the seedlings were stained with 1% aqueous solution of Erythrosine-B dye for an hour. Each treatment was replicated five times. Subsequently, the feeding punctures, also known as 'probing marks' were examined and counted under a stereo zoom microscope (Olympus, SZX 10).

2.5 Statistical analysis

Data were subjected to one-way ANOVA and treatment means were separated by LSD (P=0.05) (SAS Institute, 2008).

Commented [A5]: Please mention that how many test tubes were places for each treatment at a time.

3. RESULTS AND DISCUSSION

The area of honeydew production in LS strain was 5.44 and 1.86 cm² in imidacloprid treated and untreated conditions, respectively. Whereas, in RS strain it was 7.12 and 14.63 cm² respectively. Thus, the honeydew production was significantly higher in RS strain compared to LS strain in both the treated and untreated conditions. Homopteran insects feed on phloem sap that is rich in water and excrete excess water through the 'filtration chamber' mechanism in the form of honeydew containing sugars, amino acids, lipids, and waxes. Quantification of honeydew excretion serves as an indirect but precise measure of *N. lugens* feeding activity, offering a straightforward bioassay for *N. lugens* feeding behaviour. The energy requirements of RS insects might be higher due to diversion of energy to meet the demands of detoxification pathways (6). Similar effect on the feeding behaviour of was reported in other sap feeding insects. In *Myzus persicae* at low concentrations of imidacloprid, a reduction in honeydew excretion, loss of weight, restless behaviour, and movement from treated to untreated leaves was observed indicating antifeeding properties of this compound (19). *Bemesia tabaci* feeding on imidacloprid treated cotton leaf discs showed significantly low honeydew excretion and fecundity compared to the untreated control (20). In contrast, Chen et al. (2023) (21) observed that imidacloprid-susceptible *Diaphorina citri* feeding on citrus exhibited significantly more bouts associated with intercellular pathway, phloem penetration, phloem salivation, and non-probing activities than imidacloprid-resistant counterparts. However, there were no differences observed in the frequency or duration of phloem ingestion or xylem feeding between susceptible and resistant *D. citri*.

Table 1: Feeding behaviour of Laboratory (LS) and Resistant (RS) strains of *N. lugens*.

Treatment	Area of Honeydew (cm ²)	No. of probes
LS-Untreated	5.44 ^{ab}	7 ^a
LS-Treated	1.86 ^a	16 ^a
RS-Untreated	14.63 ^c	11 ^a
RS-Treated	7.12 ^b	31 ^b
LSD (P=0.05)	4.94	9.68

In a column means with same letters do not differ significantly, LSD (P=0.05).

The probing test revealed an increased probing activity in both the LS and RS strains in imidacloprid treated seedlings compared to the untreated. The number of probing marks by LS and RS strains in treated seedlings were 16 and 31, whereas in untreated seedlings were 7 and 11, respectively. In addition, the number of probes by RS under treated conditions was significantly higher as compared to the rest of the treatments. In homopterans, the piercing organ is the stylet that secretes a sheath after penetration and it remains within the plant tissues even after withdrawal of the stylets and could be easily visualized by histological staining. Before the insect stylet reaches the phloem, multiple attempts are made to find a suitable site for feeding. Insect feeding is modulated by the complex mechanisms that respond to internal and external signals (22). Imidacloprid acts on several types of post-synaptic nicotinic acetylcholine receptors and it binds irreversibly to these receptors leading to spontaneous discharge of the nerve impulses at first, followed by failure of the neuron to propagate any signal (23,24). Wang *et al.* 2020 (25) reported that imidacloprid resistant aphids showed increased activity in searching for a suitable feeding site. Imidacloprid resistant aphids showed a higher frequency of apoplastic stylet probing compared to the susceptible aphids. The duration of phloem ingestion was notably increased in resistant aphids on imidacloprid treated plants in contrast to control plants. Whereas, imidacloprid significantly reduced the capacity of susceptible aphids to locate and feed from the phloem. Zhu *et al.* 2020 (26) revealed that *Sogatella furcifera* when exposed to plants treated with triflumezopyrim concentrations of LC₁₀, LC₅₀ and LC₉₀ through direct contact method revealed reduction of 27.5, 33.5 and 34.3 per cent probing frequencies, respectively compared to the untreated control.

4. CONCLUSION

Our results revealed that RS strain of *N. lugens* has higher feeding capacity in terms of probing behaviour and honeydew production as compared to the LS strain. The higher feeding capacity might be due to higher energy requirements in resistant strains to meet the demands of detoxification pathways. Higher metabolic energy demand in resistant populations influence the developmental parameters negatively, imposing fitness cost. Thus provide an opportunity for the reversal of the insecticide resistance in the crop ecosystem on withdrawal of the selection pressure.

Commented [A6]: This reference is not related to this MS. PI delete.

REFERENCES

1. Chen CC, Ko WH, Chiu RJ. Rice wilted stunt and its transmission by the brown planthopper *Nilaparvata lugens*. Plant Protection Bulletin Taiwan. 1978; 20, 376-380.
2. Hibino H. Rice ragged stunt, a new virus disease occurring in tropical Asia. Review of Plant Protection Research. 1979; 12:98-110.
3. Chiu SC, Chien CC, Chow LY, Chen BH. Biological control of insect pests on Cruciferous vegetables. IN C N Chen, W Y Su and W F Hsiao (eds.). Proceedings of the symposium on production and insect control of Cruciferous vegetables in Taiwan. Taichung, Taiwan: Plant Protection Center .1981.
4. APRD. Arthropod pesticide resistance database (2024). Available: <http://www.pesticideresistance.org>.
5. IRAC. 2024. <http://www.irac-online.org/>. Based on Insecticide MoA Classification Scheme, Version 11.1.
6. Ullah F, Gul H, Desneux N, Said F, Gao X, Song D. Fitness costs in chlorfenapyr-resistant populations of the chive maggot, *Bradysia odoriphaga*. Ecotoxicology. 2020; 29(4):407-416.
7. Zhang X, Mao K, Liao X, He B, Jin R, Tang T, Wan H, Li J. Fitness cost of nitenpyram resistance in the brown planthopper *Nilaparvata lugens*. Journal of Pest Science. 2018; 91:1145-1151.
8. Liao X, Mao K, Ali E, Jin R, Li Z, Li W, Li J, Wan H. Inheritance and fitness costs of sulfoxaflor resistance in *Nilaparvata lugens* (Stål). Pest Management Science. 2019; 75(11):2981-2988.
9. Li W, Mao K, Liu C, Gong P, Xu P, Wu G, Le W, Wan H, You H, Li, J. Resistance monitoring and assessment of the control failure likelihood of insecticides in field populations of the white-backed planthopper *Sogatella furcifera* (Horváth). Crop Protection. 2020; 127:104973.
10. Leather SR. Factors Affecting Fecundity, Fertility, Oviposition, and Larviposition in Insects, In S.R. Leather & J. Hardie (eds), Insect Reproduction. CRC Press, Boca Raton. 1995; p. 143-174.
11. Chelliah S, Heinrichs EA, Smith WH. Factors contributing to brown planthopper resurgence, Judicious and Efficient Use of Insecticides on Rice, IRRI, Philippines. 1984; 107-115.
12. Zhang Z, Cui B, Zhang Y. Electrical penetration graphs indicate that triclin is a key secondary metabolite of rice, inhibiting phloem feeding of brown planthopper, *Nilaparvata lugens*. Entomologia Experimentalis et Applicata. 2015; 156(1):14-27.
13. Hao ZP, Hou SM, Hu BC, Huang F, Dang XL. Assessment of probing behavior of the cabbage aphid, *Brevicoryne brassicae* (Hemiptera: Aphididae), on three *Brassica napus* cultivars at three developmental stages using Electropenetrography (EPG). Journal of the Kansas Entomological Society. 2017; 90(1):11-23.

14. Chesnais Q, Mauck KE. Choice of tethering material influences the magnitude and significance of treatment effects in whitefly electrical penetration graph recordings. *Journal of Insect Behavior*. 2018; 31:656-671.
15. Haynes KF. Sublethal effects of neurotoxic insecticides on insect behavior. *Annual Reviews of Entomology*. 1988; 33:149-168.
16. Butler CD, Walker GP, Trumble JT. Feeding disruption of potato psyllid, *Bactericera cockerelli*, by imidacloprid as measured by electrical penetration graphs. *Entomologia Experimentalis et Applicata*. 2012; 142:247-257.
17. Jacobson AL, Kennedy GG. Effect of cyantraniliprole on feeding behavior and virus transmission of *Frankliniella fusca* and *Frankliniella occidentalis* (Thysanoptera: Thripidae) on *Capsicum annuum*. *Crop Protection*. 2013; 54:251-258.
18. Heinrichs EA, Chelliah S, Valenciana SL, Arceo MB, Fabellar LT, Aquino GB, Pickin S. *Manual for Testing Insecticides on Rice*. IRRRI, Philippines. 1981; 134p.
19. Nauen R. Behavior modifying effects of low systemic concentrations of imidacloprid on *Myzus persicae* with special reference to an antifeeding response. *Pesticide Science*. 1995; 44:145-153.
20. He Y, Zhao J, Zheng Y, Weng Q, Biondi A, Nicolas D, Wu, K. Assessment of Potential Sublethal Effects of Various Insecticides on Key Biological Traits of The Tobacco Whitefly, *Bemisia tabaci*. *International Journal of Biological Sciences*. 2013; 9(3):246-255.
21. Chen XD, Justin G, Lauren MD, Hunter G, Liu G, Jawwad AQ, Stelinski LL. Feeding behavior and hormoligosis associated with imidacloprid resistance in Asian citrus psyllid, *Diaphorina citri*. *Insect Science*. 2023; 0:1-11.
22. Tjallingii WF. Regulation of phloem sap feeding by aphids. In *Regulatory mechanisms in insect feeding*, Boston, MA: Springer US. 1995; 190-209.
23. Buckingham SD, Lapied B, Corronc HL, Grolleau F, Sattelle DB. Imidacloprid actions on insect neuronal acetylcholine receptors. *Journal of Experimental Biology*. 1997; 200: 2685-92.
24. Sheets LP. Imidacloprid: a neonicotinoid insecticide. In: Krieger RI, editor. *Handbook of pesticide toxicology*. 2nd ed. San Diego, CA: Academic Press. 2001; p. 1123-30.
25. Wang L, Wang Q, Wang Q, Rui C, Cui L. The feeding behavior and life history changes in imidacloprid-resistant *Aphis gossypii* glover (Homoptera: Aphididae). *Pest Management Science*. 2020; 76:1402-1412.
26. Zhu J, Sun W, Li Y, Ge L, Yang GQ, Xu JX, Fang L. Effects of a novel mesoionic insecticide, triflumezopyrim, on the feeding behavior of rice planthoppers, *Nilaparvata lugens* and *Sogatella furcifera* (Hemiptera: Delphacidae). *Journal of Integrative Agriculture*. 2020; 19(10):2488-2499.