

## Short Research Article

### Isolation and Partial characterization of 11-octadecenoic acid methyl ester and bis (2-ethylhexyl) phthalate from *Celtis integrifolia* Lam

#### ABSTRACT

**Aim:** The study was aimed at the isolation and characterization of phytochemicals from *Celtis integrifolia* crude extract. *Celtis integrifolia* is a traditional medicinal plant used as a remedy or cure for diarrhea, measles, bleeding, ebolic, sore throat, and as an antinociceptive agent.

**Methodology:** The stem bark sample was obtained, coarsely powdered and exhaustively extracted using maceration technique with methanol. The extract was filtered using cotton wool and clarified using Whatmann No.1 filter paper. The filtrate was concentrated on a rotavapor at 45°C. The crude extract was then reconstituted in water and partitioned successively with *n*-hexane, ethyl acetate, and *n*-butanol. The ethyl acetate and the *n*-hexane fractions were concentrated and subjected to purification on column chromatography.

**Results:** Gradient elution of extract fractions on open-column chromatography yielded two compounds coded C-3 and C-4 from the *n*-hexane and ethyl acetate fractions respectively. The isolated compounds were characterized using spectroscopic data from FT-IR and <sup>1</sup>H NMR, GC-MS and literature database. Compound C-3 with  $R_f = 0.43$  in hexane: Ethyl acetate (9:1) was identified as 11-Octadecenoic acid methyl ester while Compound C-4 isolated from ethyl acetate fraction as a yellow substance with  $R_f = 0.38$  in hexane: ethyl acetate (8:2) was identified as bis (2-ethylhexyl) phthalate.

**Conclusion:** The study had shown that *Celtis integrifolia* contains phytochemicals. To the best of our knowledge based on available literature, we report for the first time the isolation and characterization of 11-Octadecenoic acid methyl ester and bis (2-ethylhexyl) phthalate from *Celtis integrifolia*. Since the presence of phytochemical substances is believed to be the basis of any

*observed pharmacological activity, the study therefore lends credence to the traditional medicinal uses of Celtis integrifolia Lam.*

**Keywords:** *Celtis integrifolia*, 11-Octadecenoic acid methyl ester, bis(2-ethylhexyl) phthalate, Chromatography, Characterization

UNDER PEER REVIEW

## 1.0 INTRODUCTION

Medicinal plant exhibit a broad range of biological activities due to the presence diverse range of bioactive molecules with beneficial therapeutic properties [1]. Plants are considered as a reservoir of bioactive compounds which are useful in drug development and synthesis. These bioactive compounds had been exploited for the treatment of ailments and includes substances such as coumarins, terpenoids, carotenoid alkaloids, saponins, phenols, steroids, tannins and glycosides [2]. The *Celtis* genus is famous for the widespread use of its species in traditional medicine with several compounds isolated and characterized [3]. Specifically, Filali-Ansari *et al.* [4] reported the isolation and characterization of three antioxidant compounds from *Celtis australis* and were identified as  $\beta$ -sitosterol (1),  $\beta$ -sitosterol-3-O- $\beta$ -glucoside (2) and vanillic acid (3).

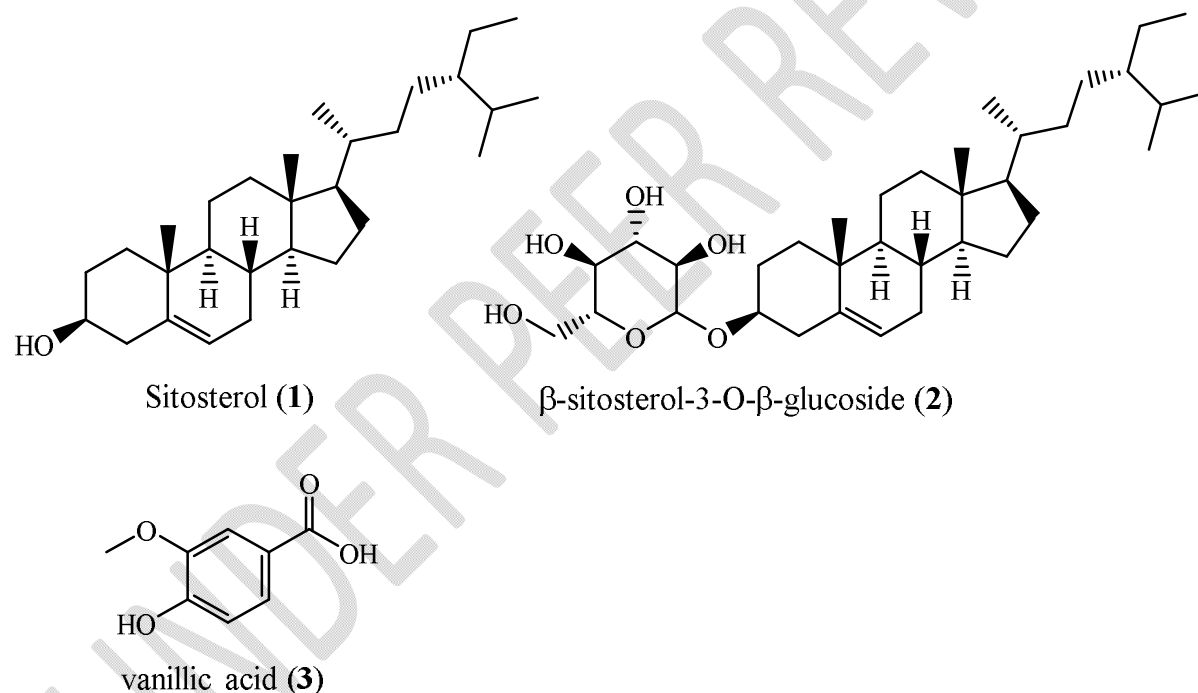


Fig.1: Some compounds isolated from *Celtis australis*

*Celtis integrifolia* (syn. *Celtis toka*) is a deciduous tree that grows up to about 30m tall usually with grey smooth stem bark. The plant is common to the temperate regions of the northern hemisphere and widely used as medicine in African countries like Nigeria. The leaves and the stem bark of the plant are reportedly the most used parts in traditional medicine for the treatment of epilepsy, mental disorders, cancer, wound healing, diarrhea, chicken pox, measles, bleeding, gout, eczema, sore throat and as an

antinociceptive[5,6]. Recent studies had proved that *Celtis integrifolia* leaf extract contains several classes of phytochemicals such as alkaloids, phenols, steroids and flavonoids[7]. Despite several reported pharmacological properties such as antimicrobial, cytotoxicity and antioxidants activities [8, 6] on *Celtis integrifolia* crude extract, there are no studies yet on the isolation and characterization of its phytochemicals based on available literature. Consequently, we report for the first time the isolation and partial characterization of some of the bioactive compounds of *C. integrifolia* Lam crude extract.

## **2.0 MATERIALS AND METHODS**

### **2.1 Collection and Sample Identification**

The stem bark sample of *Celtis integrifolia* stem was collected from NGalda, Yobe State Nigeria in December 2022 and identified by Dr. D. A. Zhigila Department of Botany, Gombe State University. The sample was compared with a previously deposited specimen and Voucher No. GSUH112 was allocated.

### **2.2 Preparation and Extraction Plant Material**

The collected sample of *C. integrifolia* stem bark was air dried and pulverized to powder. About 2.5 kg of the powdered sample was soaked in methanol for a period of seven days with occasional shaking. The extract was filtered using Whatmann No.1 filter paper. The marc was again re-soaked in methanol and then filtered off. The combined filtrate was concentrated on a rotavapor at about 45°C to yield a crude methanol extract [9]. The crude extract was reconstituted in methanol:water (1:9) and successively extracted with n-hexane, ethyl acetate and n-butanol.

### **2.3 Isolation of Compounds from *C. integrifolia***

The column chromatography purification of n-hexane and ethyl acetate fractions was performed in accordance with the procedure previously reported by Kwaji *et al.*[10] with slight modifications. The n-hexane (15.8 g) and ethyl acetate (4.5 g) extract fractions were dissolved in minimum amounts of methanol and pre-adsorbed on to 20 g silica gel 60 (70-230 mesh) and allowed to dry. Column packing was done using the wet slurry method. Gradient elution was performed and consisted of n-hexane/ethyl acetate and ethyl acetate/methanol at 5% increase in volume of the selected eluting solvents (100:00-100:20 v/v). All eluent

fractions were combined based on their TLC profiles and subsequently concentrated. Fractions with single spot were washed and recrystallized from methanol. Compound C3 was isolated as a white substance while C4 was yellow in color. Characterization of the isolates were performed using IR, <sup>1</sup>H NMR and GC-MS data and in comparison with literature.

### 3.0 RESULTS AND DISCUSSIONS

The Table 1 below provides a summary of the result of column chromatography purification of the crude extract fractions.

**3.1 Table 1: Isolates from ethyl acetate and n-hexane fractions of *C. integrifolia*.**

SN	Isolate	R <sub>f</sub>	Colour	weight	Fraction
1	C-3	0.81	White	0.4g	n-Hexane
2	C-4	0.38	Yellow	0.5g	Ethyl acetate

### 3.2 Characterization of Compound C-3

**Table 2: FT-IR data of C-3**

SN	Freq. (cm <sup>-1</sup> )	Vibration Type
1	3469	O-H
2	2937	C-H of (CH <sub>3</sub> )
4	2869	C-H of (CH <sub>2</sub> )
5	1634	C=C stretching
6	1466	CH <sub>3</sub> bending
7	1383	CH <sub>2</sub> bending
8	1134	CH <sub>2</sub> bending
9	1049	C-O stretching

The peak at  $3469\text{ cm}^{-1}$  and  $1049\text{ cm}^{-1}$  suggests the presence of bonded hydroxyl (-OH) group of residual methanol solvent while the weak peak at  $1634\text{ cm}^{-1}$  indicates C=C olefinic stretching. The stretching and bending vibrations of methyl group was observed as an intense peak at  $2937\text{ cm}^{-1}$  and as medium intensity peak at  $1466\text{ cm}^{-1}$  while the peaks at  $2869\text{ cm}^{-1}$  and  $1383\text{ cm}^{-1}$  indicates the presence of methylene groups. The weak peak at  $1049\text{ cm}^{-1}$  is due to C-O vibration. These absorption frequencies are consistent with literature reports[11, 12].

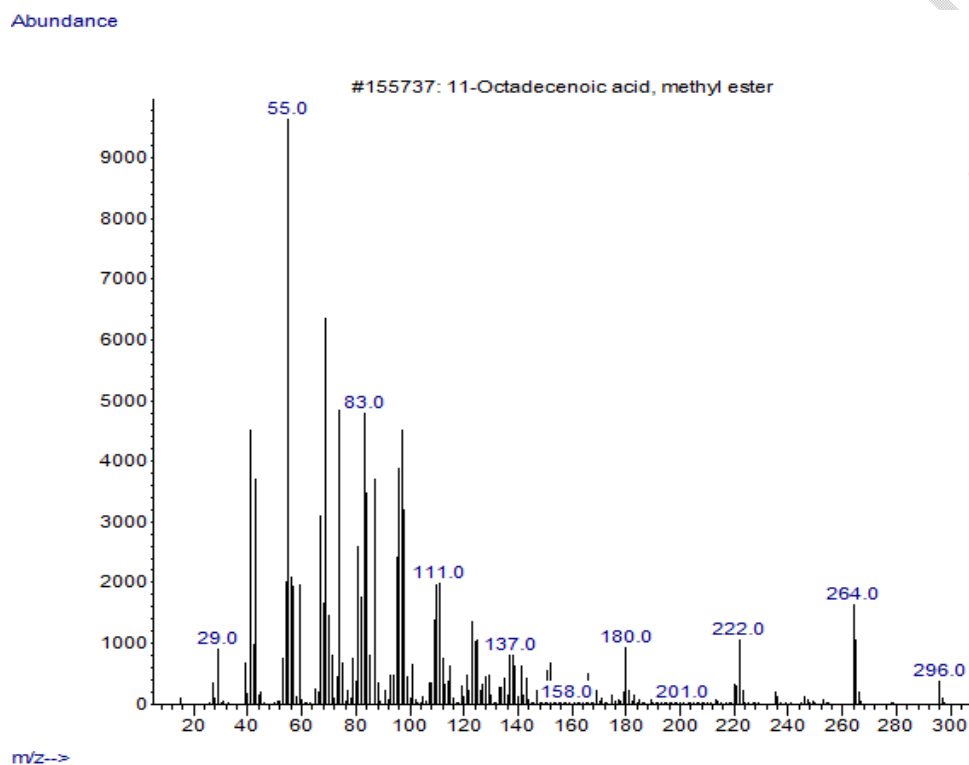


Fig. 2: GC-MS spectrum of C-3

The GC-MS library suggests that the compound C-3 is 11-octadecenoic acid methyl ester. The molecular formula of 11-octadecenoic acid methyl ester as suggested by GC-MS library is  $\text{C}_{19}\text{H}_{36}\text{O}_2$ . The molecular ion  $m/z$  ( $M^+$ ) is 296.0 as obtained in the spectrum (Fig. 2). Other daughter ions such as  $m/z$  222 is due to  $\text{C}_{16}\text{H}_{30}$  after removal of  $\text{C}_3\text{H}_6\text{O}_2$ . The peak at  $m/z$  180 is due to  $\text{C}_{13}\text{H}_{24}$ . The removal of  $\text{CH}_3\text{OH}$  from the molecular ion gave rise to the peak at  $m/z$  264. The peak at  $m/z$  111 is due to the fragment

C<sub>7</sub>H<sub>11</sub>O, while the base peak is due to the fragment C<sub>4</sub>H<sub>7</sub>O. The observed fragmentation pattern is consistent with literature [11, 12].

The <sup>1</sup>H NMR analysis result is in agreement with that of the mass spectrometry. The peak signals at δ<sub>H</sub>5.34 indicates geminal ethylene protons, δ<sub>H</sub>3.52 signifies the presence of methoxy protons, δ<sub>H</sub>2.62 indicates signals for α-methylene group to –C=C–, δ<sub>H</sub>1.84 ppm indicates β-methylene groups to –C(=O)–O–C, δ<sub>H</sub>1.49ppm indicates signals of β-protons to –C–C–C= and δ<sub>H</sub>0.69 ppm indicates signal for terminal methyl group of the fatty acid ester. All the above observations are consistent with the molecular structure of 11-Octadecenoic acid methyl ester (Fig.3).

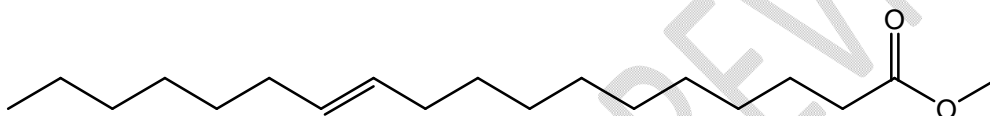


Fig. 3: 11-octadecenoic acid methyl ester

### 3.3 Characterization of Compound C-4

The compound C-4 was isolated as a yellow substance and Table 3 below shows vibrational peaks from its FT-IR spectrum.

Table 3: FT-IR data of C-4

S/No	Frequency (cm <sup>-1</sup> )	Type of Vibration
1	3492	OH band
2	1660	C=C stretching
3	1451	CH <sub>3</sub> bending
4	1374	CH <sub>2</sub> bending
5	1292	CH <sub>2</sub> bending
6	1174	C–O–C stretching
7	832	CH bending

FT-IR result showed a broad peak at 3492 cm<sup>-1</sup> indicating O–H bond vibrations of hydroxyl group possibly due to residual methanol solvent. The weak vibrations of methyl (CH<sub>3</sub>) and methylene (CH<sub>2</sub>)

groups were observed at  $2900\text{cm}^{-1}$  and  $2800\text{cm}^{-1}$  and as medium intensity peaks at  $1451\text{cm}^{-1}$  and  $1384\text{cm}^{-1}$  respectively. The out of plane C-H vibrations of the unsaturation was observed at  $832\text{cm}^{-1}$ . The corresponding C=C bond vibrations was observed at  $1651\text{cm}^{-1}$  as a weak peak of olefinic bond stretching. The C-O-C bond vibration was observed as a weak intense peak at  $1174\text{cm}^{-1}$ . These data summarize the functional groups present in the molecule and suggests that compound (C4) is an aromatic ester. These absorption frequencies are consistent with those of bis (2-ethylhexyl) phthalate [13, 14, 15, 16].

The GC-MS library of the compound C4 suggested that the compound is bis (2-ethylhexyl) phthalate with 64% abundance and retention time (33.96 minutes). The isolated compound C4 parent molecular ion  $[M^+]$ ,  $m/z$ , = 390 amu. The base peak  $m/z$  = 149 (100). The percentage fragment of other peaks relative to the base peak are  $m/z$  - 390(0), 279(12) and 167(37). The parent molecular ion  $m/z$  ratio correspondsto the molecular formula  $C_{24}H_{38}O_4$  suggesting that the compound is bis (2-ethylhexyl) phthalate (Fig. 4).

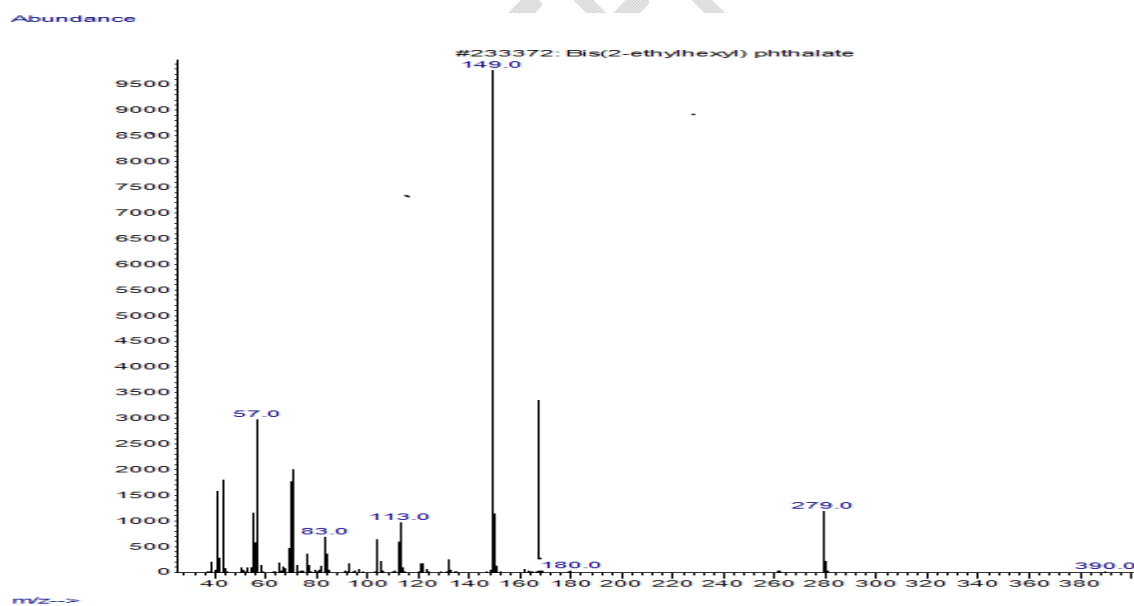


Fig. 4: GC-MS spectrum of C-4

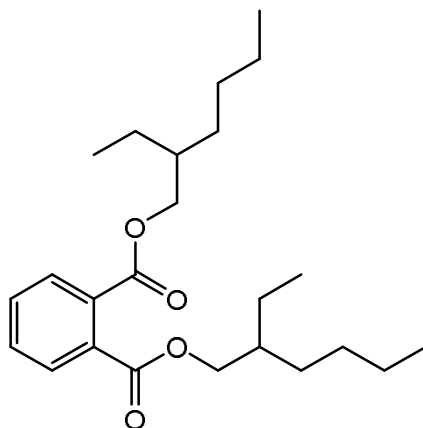


Fig. 5: bis (2-ethylhexyl) phthalate

The parent molecular ion  $[M]^+$  with  $m/z$  390amu undergoes several cleavages and radical-site rearrangement to give ion fragments with  $m/z$  279, 261, 167, and 149.

The fragment of C-4at  $m/z$  279 displayed (Fig.4) is due to the loss of  $C_8H_{15}$  (111 amu). Subsequent fragments at  $m/z$  261 might be due to the loss of a  $C_8H_{17}$  (113 amu) and loss of  $C_8H_{16}$  (112amu)yields a fragment ion at  $m/z$  149. The signal at  $m/z$  279 might also be due to loss of  $C_8H_{16}$  (112amu)to give another fragment at  $m/z$  167 followed by loss of  $H_2O$  (18amu) to give a fragment ion at  $m/z$  149 again. The mass fragmentation pattern above is in accordance with previous literature report[17]. Based on  $^1H$ -NMR spectrum data of C-4 ( $CDCl_3$ ; 600Hz,). The two proton signals at  $\delta_H$  7.46 ppm and  $\delta_H$  7.06 ppm indicates the presence of aromatic ring protons, while the proton signal  $\delta_H$  4.86 ppm region indicates the presence of oxygenated methylene protons characteristic of esters. The methylene protons also appears in the region  $\delta_H$  3.32-3.21 ppm region and  $\delta_H$  1.40-1.30 ppm indicates the presence of aliphatic methyl ( $CH_3$ ) groups. The methylene protons at the  $\delta_H$  4.86 ppm region are more deshielded than those at  $\delta_H$  2.78-2.15 ppm as a result of being directly bonded to the electronegative oxygen heteroatom [17].Consequently, data from IR,  $^1H$  NMR, GC-MS and literature strongly suggests that C-4 is bis (2-ethylhexyl) phthalate.

#### 4.0CONCLUSION

The isolation and identification of 11-octadecenoic acid methyl ester and bis (2-ethylhexyl) phthalate in *Celtis integrifolia* Lam crude extract confirms the presence of bioactive compounds. The two compounds

are known to possess broad spectrum of antibacterial activity especially against *Escherichia coli* and *Shigella dysenteriae* which are known causative agents of diarrhea. The presence of these compounds in *Celtis integrifolia* is consistent with its traditional medicinal application; also the reported cytotoxic nature of these compounds lends credence to the use of the plant in cancer treatment. These substances may act independently or in synergy. The study also showed that the traditional medicinal use of a plant may serve as a guide for the isolation of bioactive compounds.

## REFERENCES

- [1]. Egamberdieva, D., Mamedov, N., Ovidi, E., Tiezzi, A. and Craker L. Phytochemical and Pharmacological properties of Medicinal plant from Uzbekistan: A Review. University of Massachusetts Amherst, 2015, <https://doi.org/10.7275/R5571969>.
- [2]. Mahre, MB., Umaru, B., Ojo, NA., Saidu, AS., Yahi, D., Ibrahim, RM. and Mshelbwala, PP. Acute Toxicity, Phytochemistry and Anti-diarrheal Effects of *Celtis integrifolia* Lam. Aqueous Leaf Extract in Wistar Albino Rats. *British Journal of Pharmaceutical Research*, 2016, **14**(5): 1-7.
- [3]. Qi, Z., Fang, X., Xie, Y., Wang, L. and Zhang, Y. and Zhao, L. Bioassay-guided Isolation of anti-inflammatory Constituents from *Celtis sinensis* leaves, *Journal of Food Biochemistry*, 2020, Doi:10.1111/fbc.13580.
- [4]. Filali-Ansari, N., El-Abbouyi, A., Kijjoa, A., El-Maliki, S. and El-Khyari, S. Antioxidant and Antimicrobial Activity of Chemical Constituents from *Celtis australis*. *Der Pharma Chemica*, 2016, **8**(4):338-347.
- [5]. Abah, KO., Mahre, MB. and Mshelbwala, PP. Elemental Analysis and Antimicrobial assay of aqueous leaf extract of *Celtis integrifolia* Lam. *Sokoto Journal of Veterinary Sciences*, 2018, **16**(4): 79-82.
- [6]. Ezekiel, JS. Phytochemical Analysis, Antioxidant Assay and Cytotoxicity of Stem Bark Extracts of *Celtis integrifolia* L. (Cannabaceae). *J. Chem. Pharm. Res.*, 2018, **10**(8):137-142.
- [7]. Idris, SA. and Muhammed, AH. Phytochemical screening, antimicrobial and antioxidant activity of *Celtis intergrifolia* (Lam.) leaves. *International Journal of Scientific Research and Engineering Development*, 2021, **4**(2):13-21.
- [8]. Fall, AD., Dieng, SIM., Diatta-Badji, K., Diatta, W. and Bassène, E. Phytochemical screening, phenol content and antioxidant studies of ethanol leaf extract of *Celtis toka*(Forssk.) Hepper & J.R.I. Wood. In *Journal of Pharmacognosy and Phytochemistry*. 2017, **6**(1). 488–492.
- [9]. Shakeri, A., Hazeri, N., Vlizadeh, J., Ghasemi, A. and Tavallaei, FZ. Phytochemical Screening, Antimicrobial and Antioxidant Activities of *Anabasis aphylla* L. *Kragujevac Journal of Science*, 2012, **34**: 71-78.
- [10]. Kwaji, A., Adamu, HM. and Chindo, IY. Isolation, characterization and biological properties of betulin from *Entada africana* Gull. And Perrot (Mimosaceae). *Journal of Applied and Advanced Research*, 2018, **2**(3): 28-31.

- [11]. Shoge, M. and Amusan, T. Phytochemical, Antidiarrheal activity, Isolation and Characterization of 11-Octadecenoic acid methyl ester Isolated from the seeds of *Acacia nilotica* Linn. *Journal of Biotechnology and Immunology*,2020, **2**(1):1-12.
- [12]. Bitire, SO., Jen, T. and Belaid, M. Production and Optimization of Biodiesel from Parsley Seed oil using KOH as a catalyst for Automobiles Technology, 2021, Doi:10.1007/s00170-021-07415-6.
- [13]. Namikoshi, M., Fujiwara, T., Nishikawa, T and Ukai, K. Natural abundance  $^{14}\text{C}$  content of Dibutyl phthalate (DBP) from three marine algae. Tokyo University of marine Science and Technology. *Marine Drugs*,2006, **4**: 290-297
- [14]. Olawumi, OO. and Koma, SO. Stigmasterol and Stigmasterol Glycoside: Isolated Compounds from *Tetrapleura Tetraptera* extracts. *Journal of Biochemistry International*,2019, **6**(1): 21-48
- [15]. Jiang, K., Gachumi, G., Poudel, A., Shurmer, B., Bashi, Z. and El-Aneed, A. The establishment of tandem mass spectrometric fingerprints of phytosterols and tocopherols and the development of targeted profiling strategies in vegetable oils. *American Society for Mass Spectrometry*,2019, **30** (1): 1700-1712.
- [16]. Dawa, SI., Shagal, MH., Abdulrahman, FI., Sodipo, OA., Yakubu, J. and Mamza, UT. Isolation and Characterization of bis (2-ethylhexyl) benzene-1,2-dicarboxylate from the roots of *Detariummicrocarpum*. *European Journal of Medical and Health Research*,2023, **1**(2):38-44.
- [17]. Rajamanikyam, M., Vadlapudi, V., Parvathaneni, SP., Koude, D., Sripadi, P., Misra, S. *et al.* Isolation and characterization of phthalates from *Brevibacteriummbrellneri* that cause cytotoxicity and cell cycle arrest. *EXCLI Journal*, 2017, **16**: 375-387.