

# EXPLORING THE ANTIMICROBIAL PROPERTIES OF LEMON: A COMPARATIVE ANALYSIS OF PEEL, SEED, AND PULP

## ABSTRACT

### Aim

Lemons are a treasure trove in nature, belonging to the Rutaceae family and rich in vitamin C, as well as various macro and micronutrients. They are widely known for boosting immunity and can potentially be used as a natural source of medication. D-limonene is one of the main bioactive compounds present in lemons, and it is responsible for the refreshing fragrance of lemons. The study aims to identify whether the waste from lemons can be used as potential nutraceuticals or functional foods

### Study design

We took five different species for comparative antibacterial studies from Citrus species those are ***C. aurantifolia***, ***C. limetta***, ***C. sinensis***, ***C. reticulata***, and ***C. maxima***.

### Place and Duration of Study:

Biotechnology lab, Techno India University, Kolkata. The duration of this study was 1 year.

### Methodology

In this comparative study, the major parts (pulp, seed, and peel) of a fruit were used for the experiments. It is important to reduce the amount of waste in the environment by creating creative and cost-effective, eco-friendly waste management techniques. The antibacterial potential against *E. coli* and *S. aureus* was thoroughly measured using the Kirby-Bauer disc diffusion method.

### Result

The results show that in the case of *E. coli* inhibition (water extractions), the peels of *C. reticulata*, the seeds of *C. aurantifolia*, and the pulp of *C. aurantifolia* perform well compared to other samples. In the case of *S. aureus* inhibition, the peels of *C. limetta*, the seeds of *C. sinensis*, and the pulp of *C. aurantifolia* perform well compared to other samples.

### Conclusion

Hence, the results indicate that PEELs can be potential antimicrobial agents and have discovered that various parts of the citrus fruit exhibit a wide range of antimicrobial effects against both gram-positive and gram-negative bacteria.

**Keywords:** *Citrus peel, seed, pulp, antimicrobial, Citrus fruits,*

## 1. INTRODUCTION

Citrus fruit belongs to the Rutaceae family. They are rich in citric acid and ascorbic acid, strengthening our immune system. Citrus fruits are one of the most widely grown fruits and are an important source of physiologically active substances and phytochemicals (Suri, Singh, and Nema 2022). They can potentially be employed as a natural medication source or a major component of functional foods since they are an excellent source of bioactive compounds and are high in ascorbic acid and citric acid, which boost our immune systems. These compounds are mostly recognized in their edible parts. On the other hand, limonoids—typical citrus fruit terpenoids with an intensely bitter taste and potentially anticarcinogenic and chemopreventive properties—can be extracted from seeds. (Sengupta et al. 2023).

Citrus fruits increase waste generation when consumed raw or juiced. Conventional rubbish disposal techniques contaminate land and waterways, potentially harming aquatic environments. Consequently, to lessen the quantity of waste that builds up

in the environment, it is essential to create creative processes and economical, ecologically friendly waste management techniques (Seid et al. 2023).

Bacterial infections worldwide are a leading cause of illnesses, physical impairments, and deaths. Medicinal plants are thought to offer a safer and more affordable alternative for treating bacterial infections because they contain a diverse array of phytochemicals.

Natural medicines made from medicinal plants have antibacterial properties that can be used to treat viral, fungal, and bacterial illnesses. Microorganisms are becoming more resistant to antibiotics, despite the pharmaceutical industry having developed several new ones during the past three decades. (Gupta and Sharma 2022) Many years of antibacterial study have been conducted on citrus peels, pulp, and seeds, and it has been demonstrated that seed wastes consist of a substantial number of essential oils and polyphenols with antimicrobial activity. Bioactive chemicals (Roquia et al. 2022) with potential medicinal applications can be found in abundance in citrus trash. Numerous advantageous characteristics of these compounds have been discovered, such as their anti-aging, anti-mutagenic, anti-carcinogenic, and anti-allergenic effects. One particularly interesting source of natural chemicals with potential for therapeutic use is *Citrus sinensis*. New medications may therefore be developed as a result of more research into the extraction and application of these substances from citrus trash. (Baker, Ibrahim, and Salama 2021)

In this current study the antibacterial properties of citrus seed, peel, and pulp extracts from *Citrus reticulata*, *Citrus limetta*, *Citrus aurantifolia*, *Citrus sinensis*, and *Citrus maxima* and  $\delta$ -limonene (essential oil rich in terpenes found in the rind of citrus fruits) were investigated using

the agar-well diffusion method against *Staphylococcus aureus* and *Escherichia coli* a gram-positive and a gram-negative bacteria respectively.

## LITERATURE STUDY

(Mohammed and Ayoub 2016) identified the presence of alkaloids, saponins, and other compounds in these seeds that contribute to their antimicrobial activity (Valarmathy et al. 2010) further highlighted the antimicrobial effects of Citrus limetta, attributing them to the presence of limonene and other compounds.

(Al-Aamri et al. 2018) According to the results of the antibacterial activity analysis, the essential oil exhibits dose-dependent activity against *S. aureus* more so than it does against *E. coli*. In contrast to ampicillin, the positive control, its antibacterial activity is significantly lower.

(Shakya et al. 2019) showed citrus reticulata extracts have antibacterial efficacy against specific strains of bacteria with a maximal zone of inhibition against *Bacillus* sp. citrus fruit peel extract was found to be more efficient than juice extract against gram-positive bacteria with a high inhibition zone against *Klebsiella pneumoniae* the extract of juice was shown to be more efficient than its peel extract among gram-negative bacteria although 10 DMSO showed no zone of inhibition Citrus limetta. antibacterial effectiveness against particular bacterial strains compared to its juice extract the peel extract shown a greater ability to inhibit gram positive bacteria with a maximal inhibition zone against *Bacillus* sp. with the exception of *Klebsiella pneumoniae* on which the peel extracts was more effective with a inhibition zone DMSO did not exhibit any zone of inhibition among the gram negative bacteria the juice extract of c2 was found to be more effective than its peel extract against the four other bacteria chosen with maximum zone of inhibition against *E. coli* Citrus maxima exhibit antibacterial action against specific strains of microorganisms the results indicate that peel extract was more efficient against *Bacillus* sp among gram positive bacteria whereas its juice extract was more effective against *S. aureus* and *S. aureus* with a maximum zone of inhibition against *E. coli* the juice extract of was shown to be more efficient against gram negative bacteria than its peel extract although 10 dms showed no inhibition zone.

According to (Garba et al. 2016) Caurantifolia is rich in phytochemical components such as flavonoids alkaloid tannins and phenols all of which have been shown to have antibacterial qualities.

(Sahlan et al. 2018) found that ethanolic extract of pomelo (*Citrus maxima*) seeds and pulp exhibited the largest zones of inhibition for *Staphylococcus aureus*. The ethanolic extract of pomelo (*Citrus maxima*) seeds and pulp showed low inhibition zone against *Escherichia coli* and *Bacillus subtilis* compared to ethanolic extract of grapefruit (*Citrus paradisi*) seeds and pulp. Ethanolic extract of grapefruit (*Citrus paradisi*) seeds and pulp showed inhibition zones more for *Staphylococcus aureus* compared to *Bacillus subtilis* *Escherichia coli*.

(Al-Âni, Tawfik, and Shehab 2011) stated that it is most likely that this beneficial antibacterial effect has contributed to the antimicrobial activity of citrus flavonoids, such as naringenin and hesperidine. The research also showed clear disparity between the antimicrobial activity of the pomelo (*Citrus maxima*) seeds and pulp ethanolic extract and the grapefruit (*Citrus paradisi*) seeds and pulp ethanolic extract

(Lahlou 2004)The inhibition zones for the *Bacillus subtilis* in the investigated pomelo (*Citrus maxima*) seeds and pulp ethanolic extract is higher than the grapefruit seeds extract sample.

(Arora and Kaur 2013) found pulp has more antioxidant activity and antimicrobial activity as compare with the orange peel.

(Han, Chen, and Sun 2021)found that limonene showed a significant antibacterial activity in the growth and reproduction of *S. aureus*. The number of colonies lowered gradually with the increased concentration of limonene.

## **2. MATERIAL AND METHODS**

### **2.1 MATERIAL**

#### **2.1.1 Collection of samples**

Sample collection was done from local vendors in Kolkata, West Bengal between August 2023 and October 2023 throughout the year. The collected sample of seeds, peels, and pulps were then powdered in a hot air oven. According to Sulaiman et al., It is best to limit the interval between sample harvest and experimental work, as dried samples are easier to work with than fresh samples, which are difficult to handle and tend to disintegrate easily. (Sulaiman et al. 2011). Oven drying that uses thermal energy to eradicate moisture from samples is considered an easy and rapid process that preserves phytochemicals and essential antioxidants as mentioned by(Mediani et al. 2013)\_D-limonene source from Sigma-Aldrich

#### **2.1.2 Microorganisms**

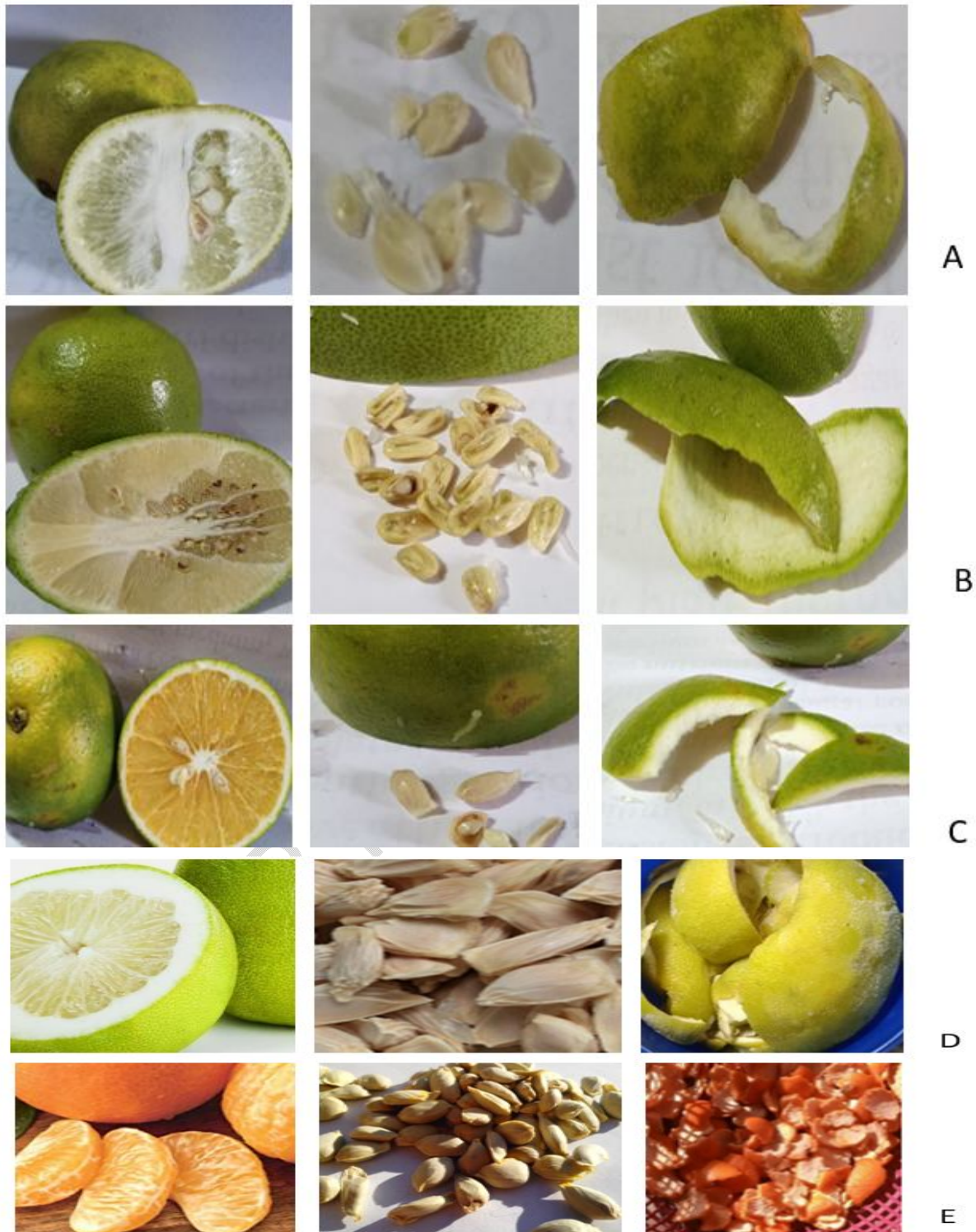
*Staphylococcus aureus* and *Escherichia coli* bacterial strains were obtained from the Department of Biotechnology, Techno India University, West Bengal, Salt Lake, Kolkata-700091. The bacteria strains were grown in Himedia M002-100G Nutrient broth at 37<sup>0</sup> Celsius or 98.6 Fahrenheit in a shaker incubator overnight.

#### **2.1.3 Sample preparation**

The dehydrated samples were then powdered finely using a household blender and 1 g of every sample was then added to 20 ml of double distilled water to make a final concentration of 50 mg/ml.

After thoroughly mixing the mixture, it was left in the dark for an entire night on a vibration table set to a low setting. After passing the mixture through the Whatman No. 1 filter paper, the aqueous extract was obtained and used without any additives.

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**Pic 1.** The samples are A – *Citrus aurantifolia*, B- *Citrus reticulata*, C – *Citrus limetta*, D- *Citrus maxima*, E- *Citrus sinensis*. 1 denotes pulp, 2 denotes seed and 3 denotes peel of the above-mentioned samples.

## 2.2 Methodology

The Kirby-Bauer test was used to assess antibiotic susceptibility. Extracts and d-limonene were tested using the disc diffusion method on agar plates. Samples were added to filter paper discs and placed on bacterial plates, which were then incubated to observe the zones of inhibition.

## 3. RESULTS AND DISCUSSION

The seeds, pulp, and peel of a total of 5 commonly found citrus species samples were taken namely *Citrus reticulata*, *Citrus limetta*, *Citrus aurantifolia*, *Citrus sinensis*, and *Citrus maxima* along with d-limonene, a vital essential oil found in every citrus species. Water extracts of the samples were subjected to the Kirby-Bauer test for antibiotic susceptibility. We chose two different bacteria, *Staphylococcus aureus*, and *Escherichia coli* which are gram-positive and gram-negative respectively. Different concentrations of ampicillin were also subjected to the test to compare the efficiency of the samples.

### 3.1 Antimicrobial efficacy of samples against gram-negative bacteria *Escherichia coli*

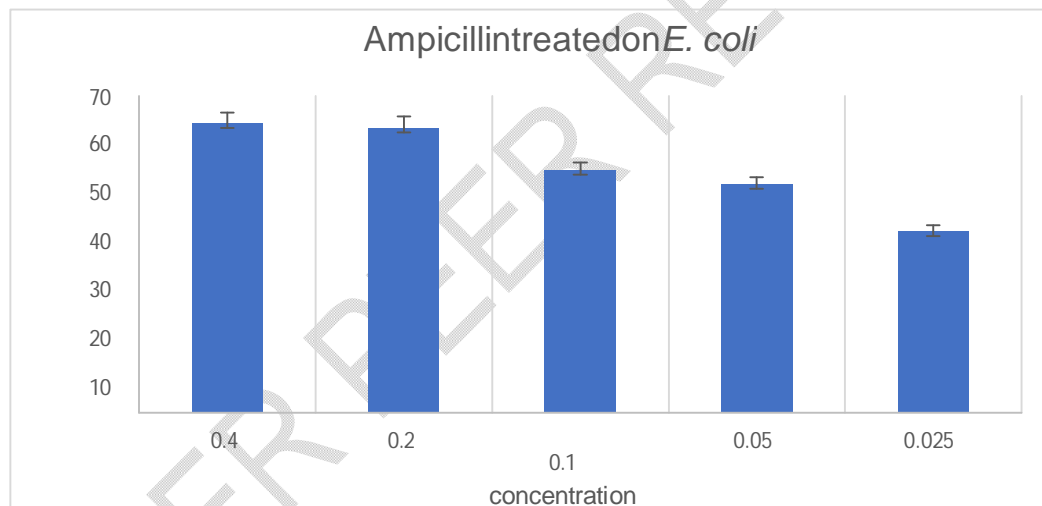
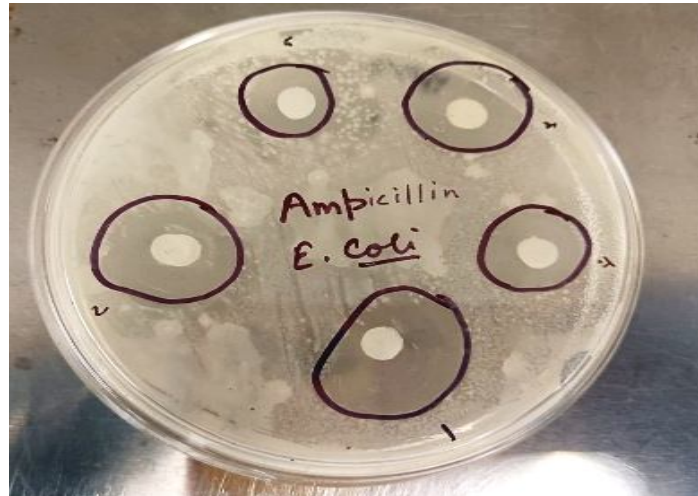
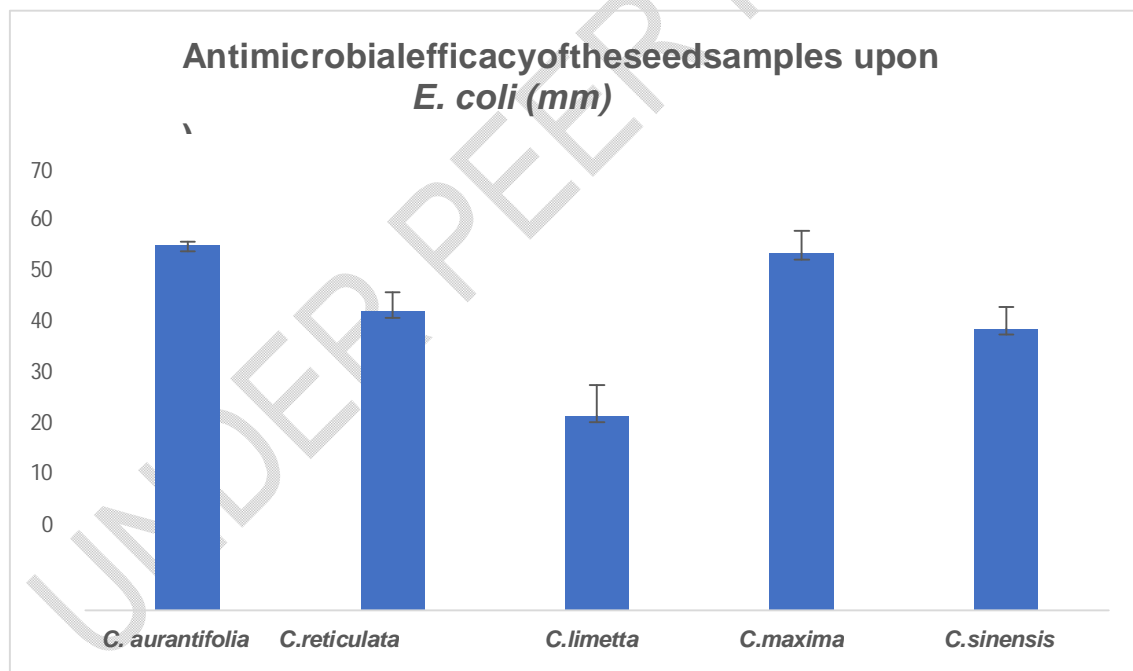


Fig 1: Diagrammatic representation of the antimicrobial activity of Ampicillin on *E. coli* bacteria



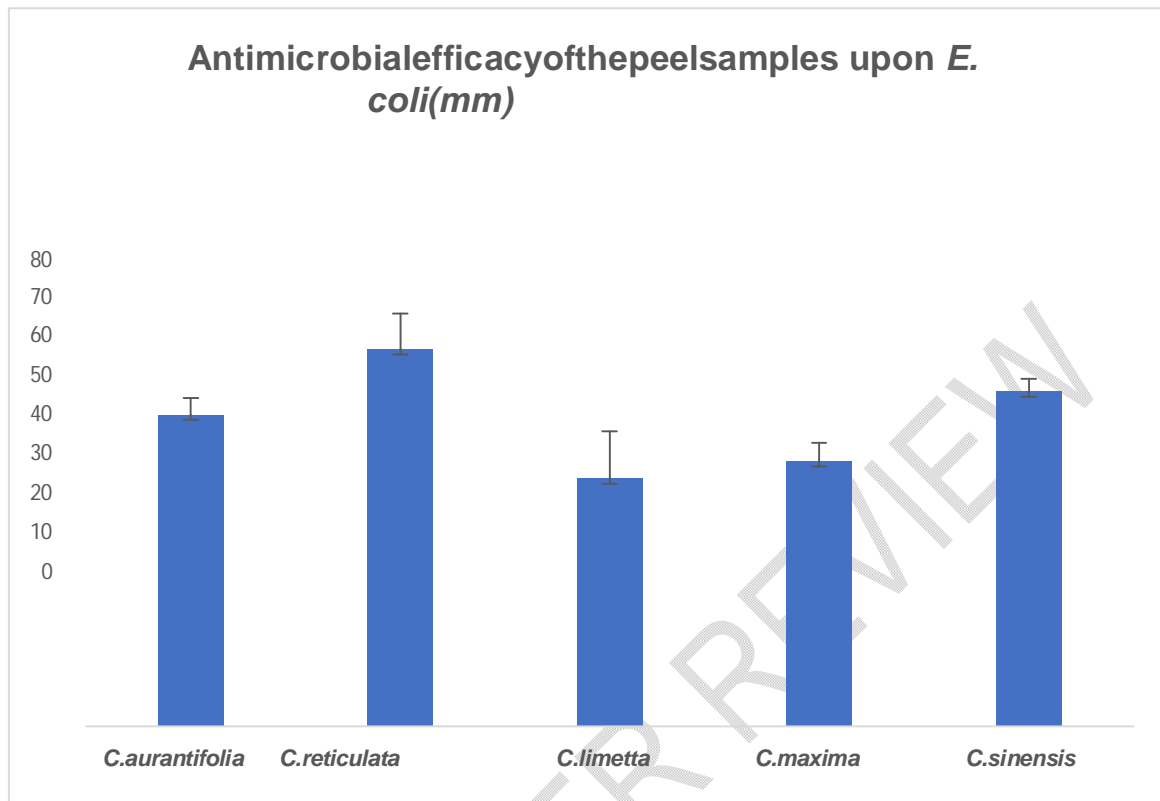
**Fig 2 : a pictorial representation of the inhibitory zones of ampicillin**

Different concentrations of **Ampicillin** have shown different inhibitory zones on *E. coli*. Every time 40  $\mu$ l of the sample was used for the treatment 0.4 mg  $64 \pm 2.44$  mm, 0.2 mg -  $63 \pm 2.5$  mm, 0.1 -  $53.66 \pm 1.6$  mm, 0.05 mg -  $50 \pm 1.5$  mm, 0.025mg -  $40 \pm 1.5$  mm [fig 2]



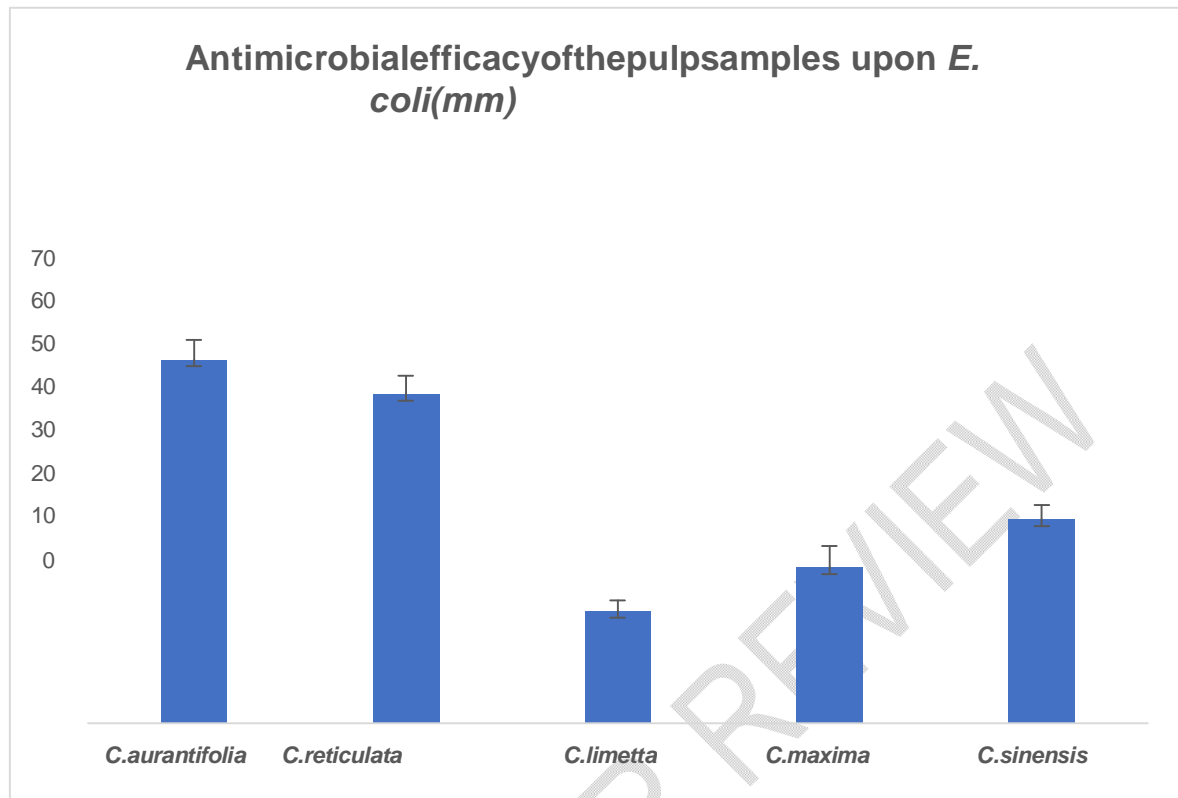
**Fig. 3: Diagrammatic representation of the antimicrobial activity of the seed samples on *E. coli* bacteria.**

The result shows *C. aurantiifolia* and *C. maxima* have the highest antimicrobial activity among other samples. Sd measured (n=3) for each sample [fig 3] Pictures show the zones in which the samples have inhibited bacterial growth. Seed extracts of *C. aurantiifolia* ( $58.33 \pm 0.4$  mm) and *C. maxima* ( $57 \pm 3.5$  mm) showed good antibacterial efficacy [fig 6]



**Fig. 4: Diagrammatic representation of the antimicrobial activity of the peel samples on *E. coli* bacteria**

*C. reticulata* and *C.aurantifolia* have the highest antimicrobial activity among other samples. Sd measured (n=3) for each sample [fig 4] Pictures show the zones in which the samples have inhibited bacterial growth. Seed extracts of *C. aurantiifolia* ( $58.33 \pm 0.4$  mm) and *C. maxima* ( $57 \pm 3.5$  mm) showed good antibacterial efficacy



**Fig. 5: Diagrammatic representation of the antimicrobial activity of the pulp samples on *E. coli* bacteria**

The result shows *C. aurantiifolia* and *C. maxima* have the highest antimicrobial activity among other samples. Sd measured (n=3) for each sample [ fig 5] Pictures show the zones in which the samples have inhibited bacterial growth. Pulp extracts of *C. aurantiifolia* (55.33 ±2.94 mm) and *C. reticulata* (50±2.6 mm) showed good antibacterial efficacy [fig 6]

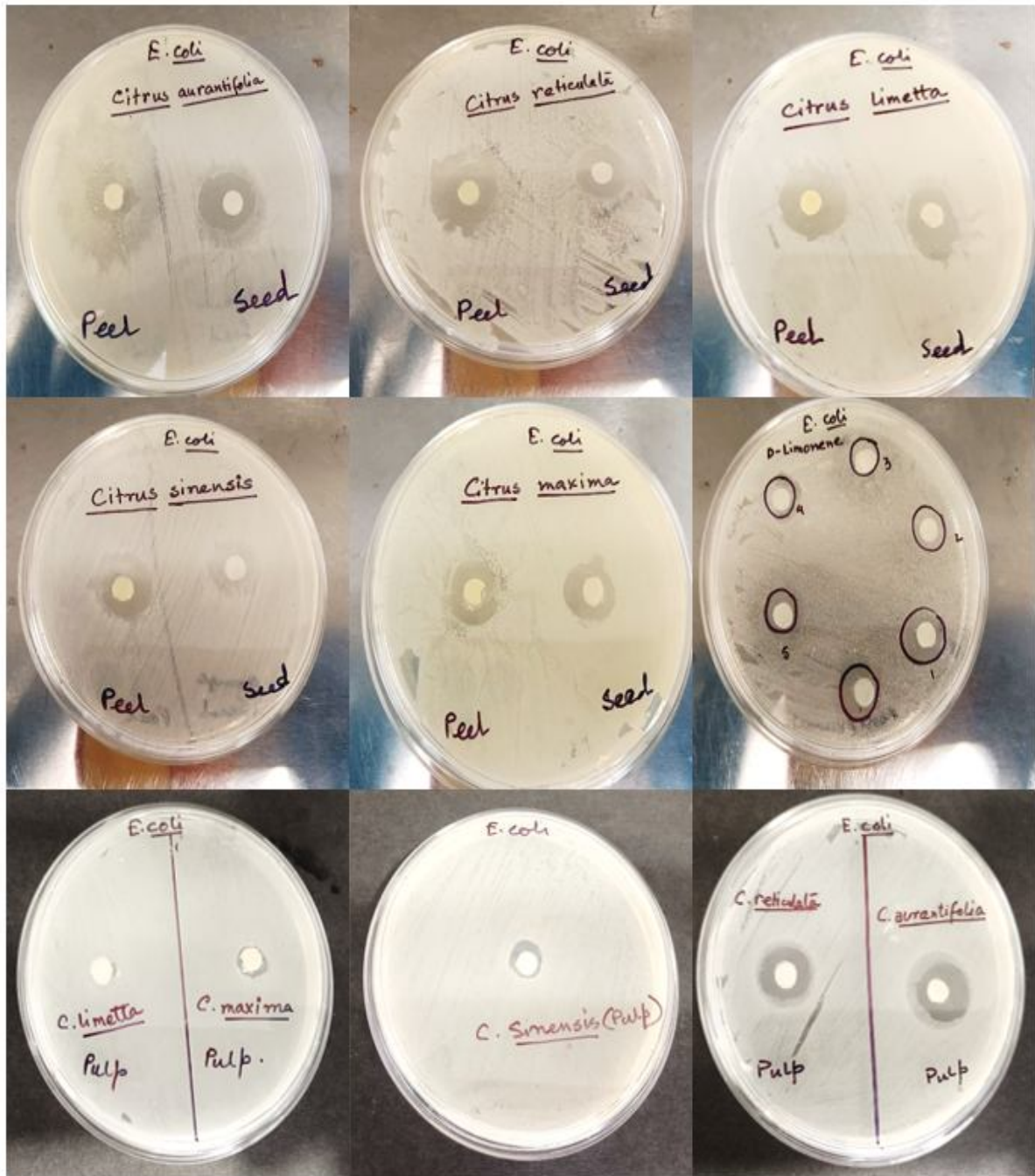


Fig. 6: Pictorial representation of the antimicrobial activity [zone of inhibition] of the seed peel and pulp samples on *E. coli* bacteria.

Part	Samples	Zone of Inhibition(mm)
Seed	<i>C.aurantifolia</i>	58.33±0.471
	<i>C.reticulata</i>	47.66±3.09
	<i>C.limetta</i>	31±4.89
	<i>C.maxima</i>	57±3.55
	<i>C.sinensis</i>	45±3.45
Peel	<i>C.aurantifolia</i>	54±2.82
	<i>C.reticulata</i>	65.33±6.16
	<i>C.limetta</i>	43±8.06
	<i>C.maxima</i>	46±3.09
	<i>C.sinensis</i>	58±2.16
Pulp	<i>C.aurantifolia</i>	55.33±2.94
	<i>C.reticulata</i>	50±2.82
	<i>C.limetta</i>	17±1.63
	<i>C.maxima</i>	23.66±3.29
	<i>C.sinensis</i>	31±2.16

Table 1. Inhibitory zones(mm) of three different parts of citrus fruits of five different species.

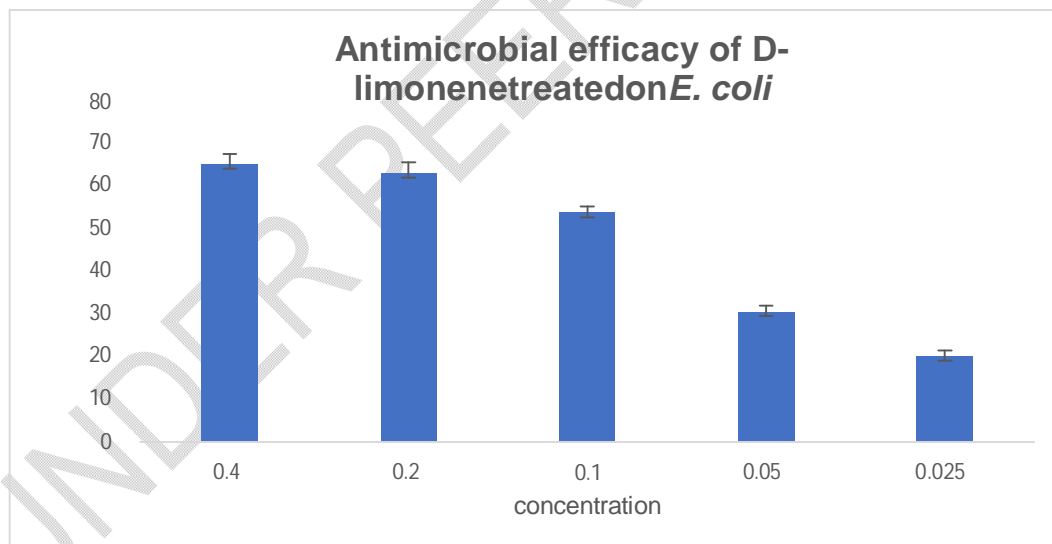
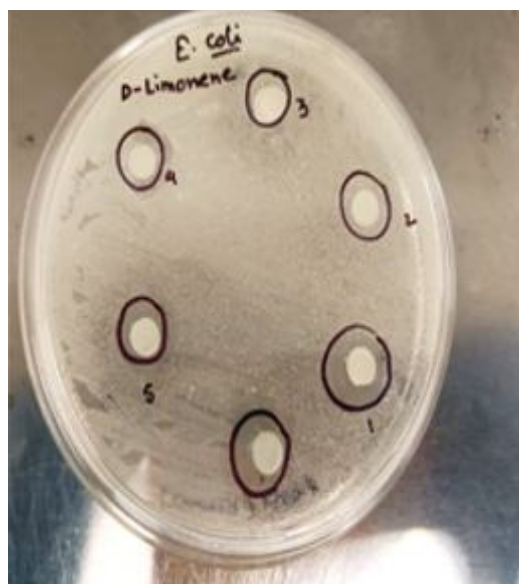


Fig 7: Different concentrations of D limonene have shown different inhibitory zones on *E.coli*.



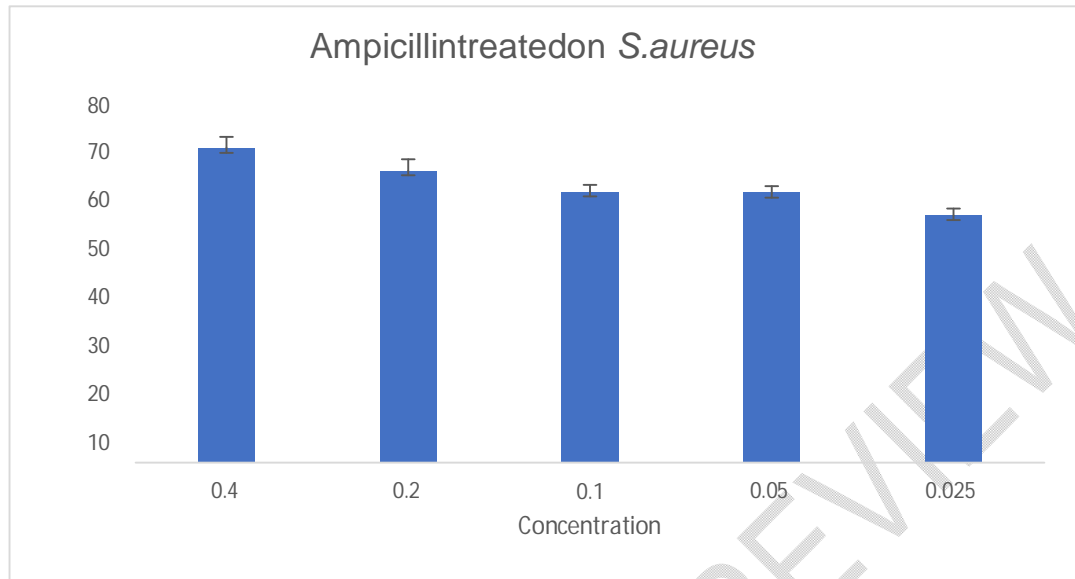
**Fig 8: Pictorial representation of the antimicrobial activity of the bioactive compounds D-Limonene**

Treatment	Inhibitory zones (mm)
0.4	65±2.44
0.2	63±2.5
0.1	53.66±1.6
0.05	30.5±1.5
0.025	20±1.5

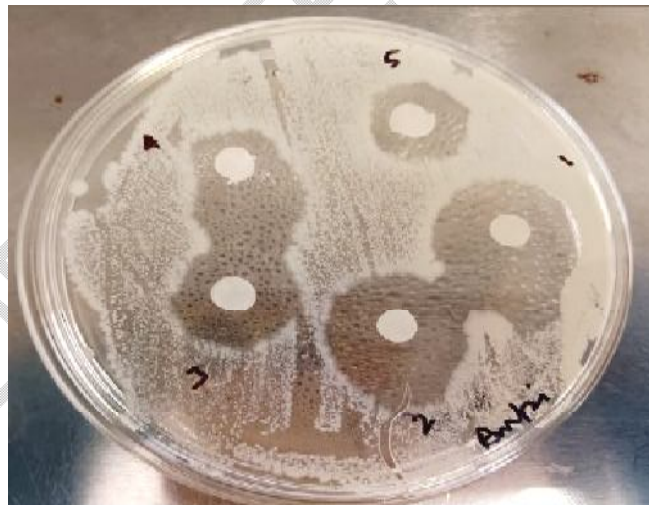
**Table 2. Range of concentration of d-limonene treatment along with their Inhibitory zones(mm)**

Every time 40 µl of the sample has been used for the treatment 0.4 mg 65± 2.44 mm, 0.2 mg - 63 ± 2.5 mm, 0.1 – 53.66 ±1.6 mm, 0.05 mg - 30.5 ±1.5 mm, 0.025mg – 20 ±1.5 mm [ fig 7, 8]

### 3.2 Antimicrobial efficacy of samples against gram-positive bacteria *Staphylococcus aureus*

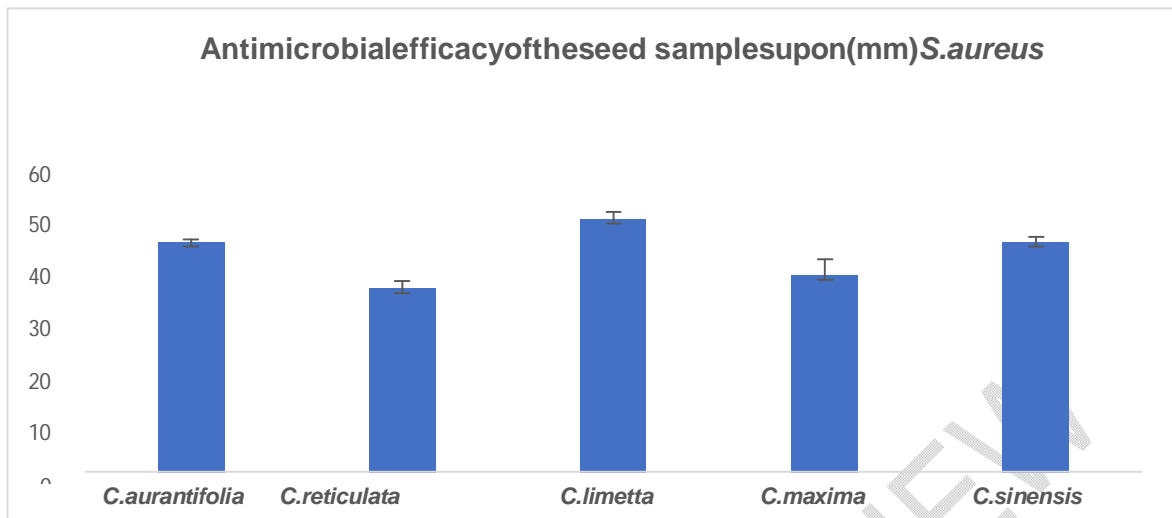


**Fig. 9: Diagrammatic representation of the antimicrobial activity of Ampicillin on *S. aureus* bacteria**



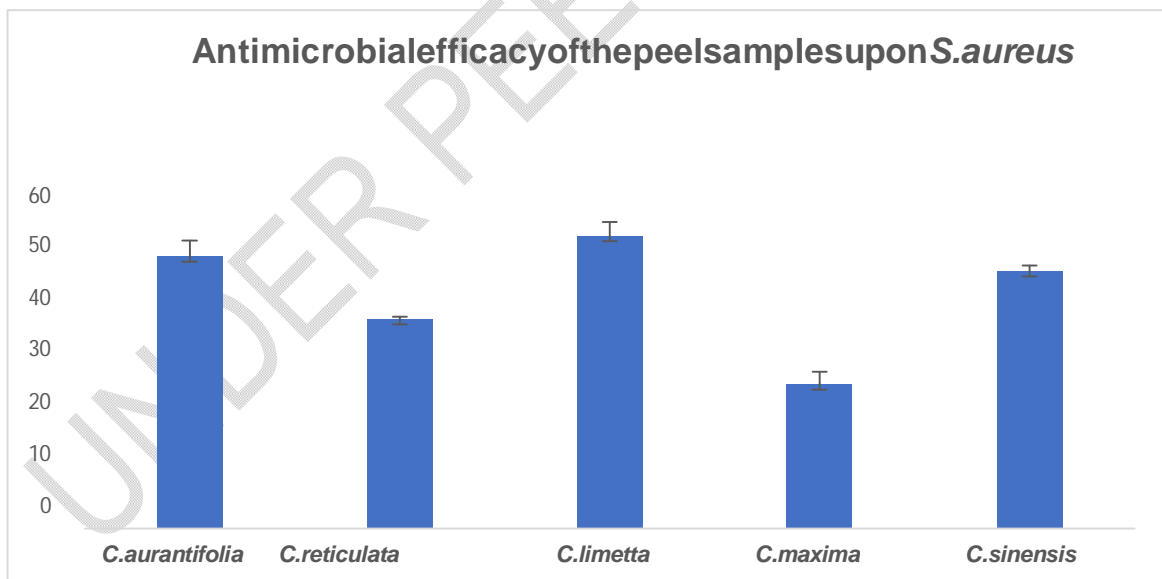
**Fig. 10: Pictorial representation of the antimicrobial activity of Ampicillin on *S. aureus* bacteria**

Different concentrations of **Ampicillin** have shown different inhibitory zones on *S. aureus*. Every time 40  $\mu$ l of the sample was used for the treatment 0.4 mg 70  $\pm$  2.44 mm, 0.2 mg - 65  $\pm$  2.5 mm, 0.1 - 60.2  $\pm$  1.6 mm, 0.05 mg - 60  $\pm$  1.5 mm, 0.025mg - 55  $\pm$  1.5 mm [ fig 9 and 10]

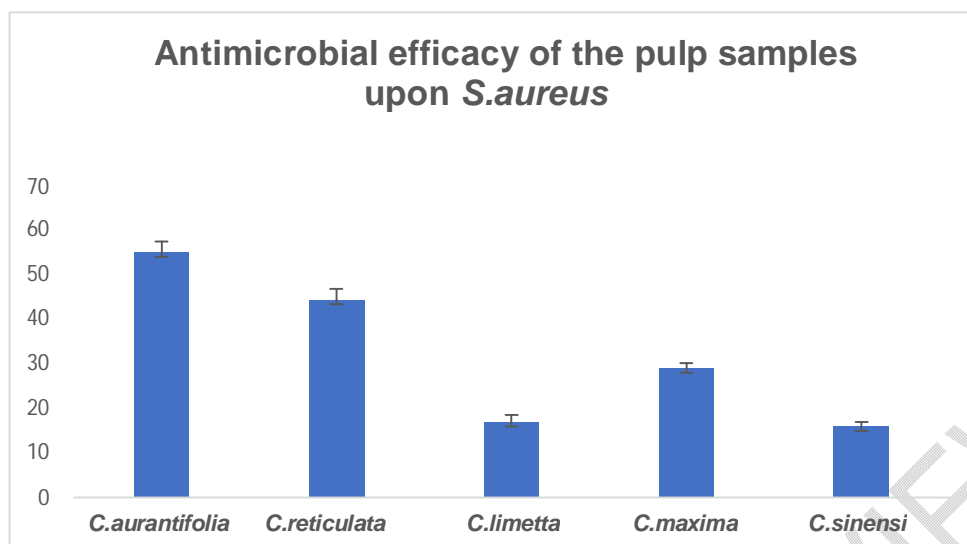


**Fig 11 : Diagrammatic representation of the antimicrobial activity of the seed samples on *E. coli* bacteria.**

The result shows *C. aurantiifolia* and *C. maxima* have the highest antimicrobial activity among other samples. Sd measured (n=3) for each of the samples Pictorial representations show the zones in which the samples have inhibited bacterial growth. Seed extracts of *C. aurantiifolia* ( $58.33 \pm 0.4$  mm) and *C. maxima* ( $57 \pm 3.5$  mm) showed good antibacterial efficacy



**Fig 12: Diagrammatic representation of the antimicrobial activity of the peel samples on *E. coli* bacteria.**



**Fig 13: Diagrammatic representation of the antimicrobial activity of the pulp samples on *E. coli* bacteria.**

The seeds of citrus species *Citrus limetta* and *Citrus sinensis* both gave high zone of inhibition of  $51 \pm 1.41$  mm and  $46.33 \pm 0.94$  mm respectively against *Staphylococcus aureus* followed by *Citrus aurantifolia* with a zone of  $46.33 \pm 0.47$  mm. In the case of peels, *Citrus limetta* gave the highest zone of inhibition of  $52.66 \pm 2.49$  mm followed by *Citrus aurantifolia* with a zone of  $49 \pm 2.82$  mm. In the case of the pulp portion of the fruit, *Citrus aurantifolia* gave the highest zone of inhibition of  $55 \pm 2.44$  mm followed by *Citrus reticulata* with an inhibition zone of  $44.33 \pm 2.49$  mm. [fig 14]

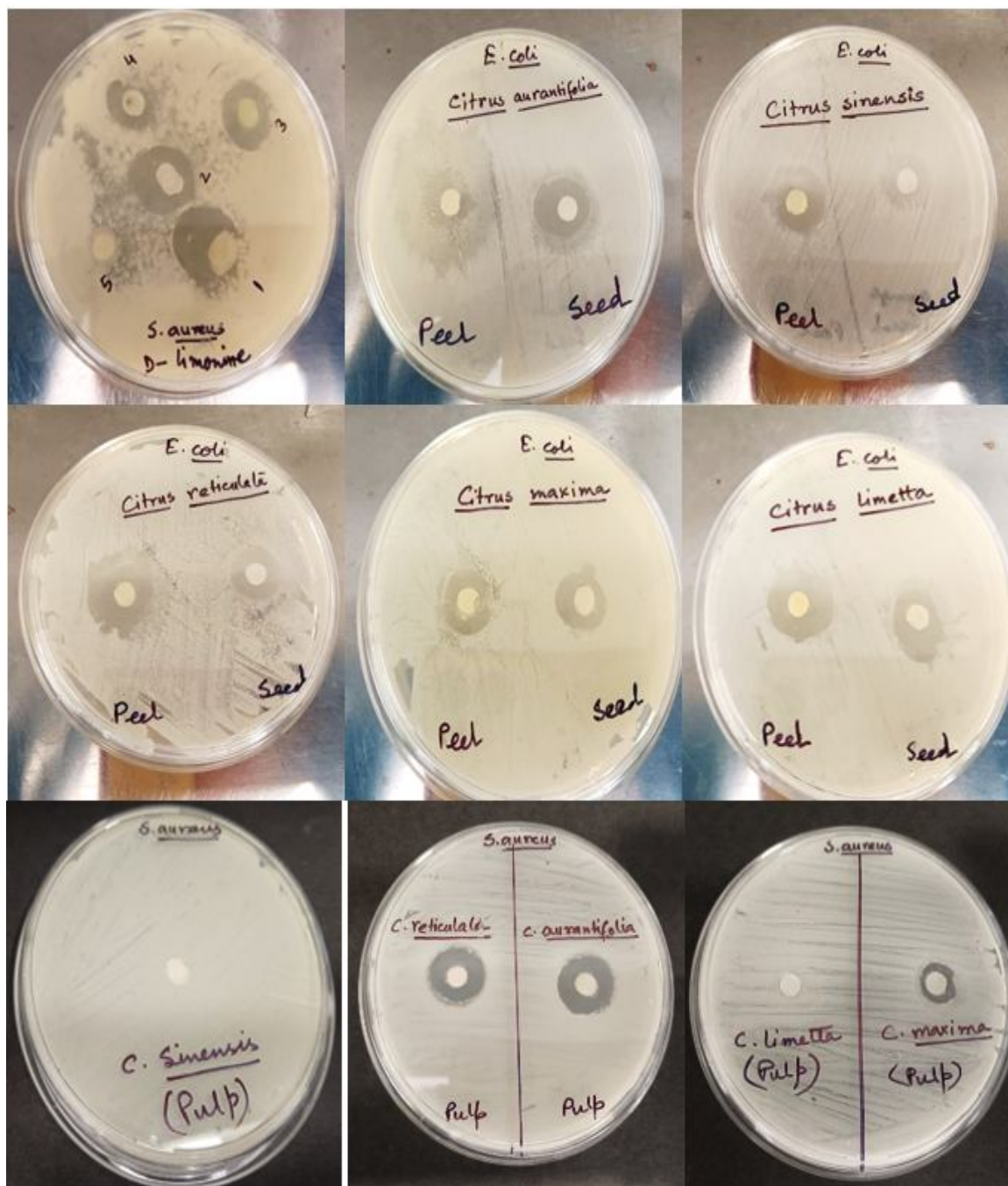


Fig. 14. Zone of inhibition of seeds, peels, and pulps of all five citrus species along with d-limonene against gram-positive bacteria *Staphylococcus aureus*

Part	Samples	Zone of Inhibition(mm)
Seed	<i>C.aurantifolia</i>	46.33±0.47
	<i>C.reticulata</i>	37±1.41

	<i>C.limetta</i>	51±1.41
	<i>C.maxima</i>	39.66±3.09
	<i>C.sinensis</i>	46.33±0.94
Peel	<i>C.aurantifolia</i>	49±2.82
	<i>C.reticulata</i>	37.66±0.4
	<i>C.limetta</i>	52.66±2.49
	<i>C.maxima</i>	26±2.16
	<i>C.sinensis</i>	46.33±0.943
Pulp	<i>C.aurantifolia</i>	55±2.44
	<i>C.reticulata</i>	44.33±2.49
	<i>C.limetta</i>	17±1.63
	<i>C.maxima</i>	29±1.24
	<i>C.sinensis</i>	16±0.94

Table 3. Inhibitory zones(mm) of three different parts of citrus fruits of five different species.

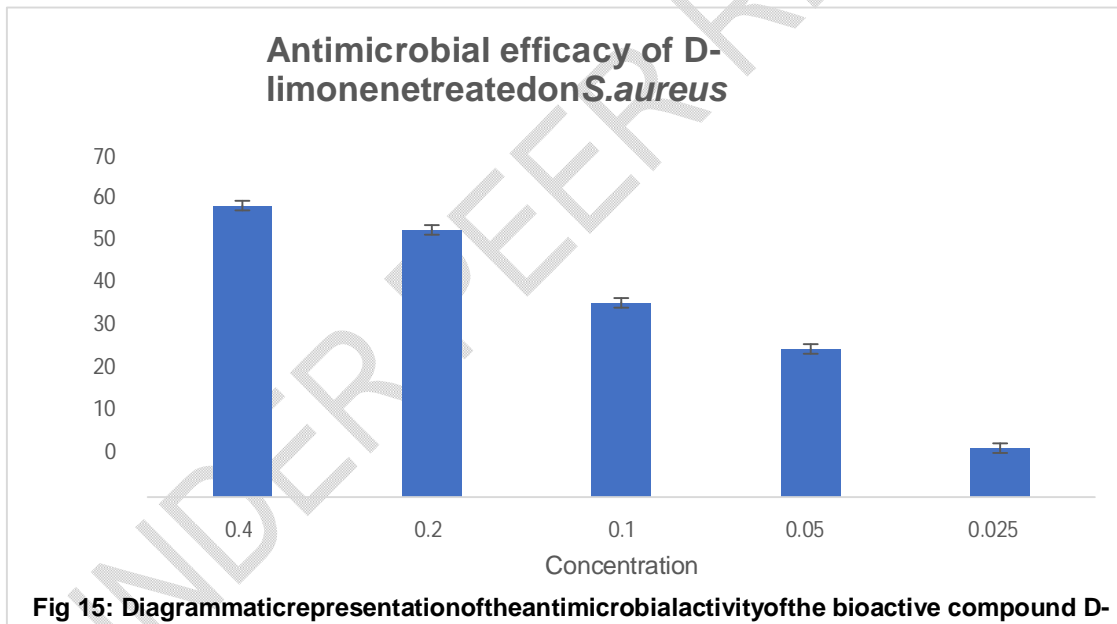


Fig 15: Diagrammatic representation of the antimicrobial activity of the bioactive compound D-Limonene on *S. aureus* bacteria



**Fig 16: A pictorial representation of the antimicrobial activity of the bioactive compound D-Limonene on *S. aureus* bacteria**

Treatment	Inhibitory zones (mm)
0.4	60±2.44
0.2	55±2.5
0.1	40±1.6
0.05	30.5±1.5
0.025	10±1.5

**Table 4. Range of concentration of d-limonene treatment along with their Inhibitory zones(mm)**

Different concentrations of **D limonene** have shown different inhibitory zones on *S.aureus*. Every time 40 µl of the sample has been used for the treatment 0.4 mg 60 ± 2.44 mm, 0.2 mg - 55± 2.5 mm, 0.1 - 40 ±1.6 mm, 0.05 mg - 30.5 ±1.5 mm, 0.025mg – 10 ±1.5 mm [ fig 15 16]

#### 4. CONCLUSION

In our research, we discovered that various parts of the citrus fruit exhibit a wide range of antimicrobial effects against both gram-positive and gram-negative bacteria. Our findings indicate that citrus reticulata peels demonstrated the most significant zone of inhibition at 65.33±6.16mm against *E. coli*, while the pulps of citrus aurantifolia displayed the highest zone of inhibition at 55±2.44mm against *S. aureus*. Furthermore, d-limonene, an essential bioactive compound present in different citrus species, exhibited the highest zone of inhibition.

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