

Utility of Parthenocarpy in Vegetable Crops: A review

ABSTRACT

Parthenocarpy is a phenomenon observed in vegetable crops where fruits are produced without fertilization or pollination. This process leads to the development of seedless fruits, which are highly desirable in the market due to their convenience and improved quality. In parthenocarpic vegetables such as cucumbers, tomatoes and eggplants, fruit development occurs without the need for pollination by insects or wind. This can be advantageous in areas with limited insect activity or in greenhouses where pollinators may not be present. The development of parthenocarpy in vegetable crops is influenced by both genetic and environmental factors. Certain cultivars have been bred to exhibit this trait, while others may require specific environmental conditions such as temperature and light to induce parthenocarpy. One of the key benefits of parthenocarpy is the production of seedless fruits. This is particularly important in seedless cucumber varieties as it eliminates the need for seed removal, making them more convenient for consumption. Seedless tomatoes and eggplants also offer improved texture and taste as the absence of seeds reduces bitterness and enhances sweetness. Parthenocarpy can help improve crop yields and reduce crop losses. Since parthenocarpic fruits develop without pollination, they are less prone to damage caused by pests and diseases that target developing seeds. This can result in higher yields and better overall crop quality. However, it is important to note that parthenocarpy may have some limitations. In some cases, seedless fruits may be less flavorful compared to their seeded counterparts. Additionally, parthenocarpic varieties may require specific management practices and careful monitoring to ensure optimal fruit development.

KEY WORDS: Parthenocarpy, Vegetable crops, Seedless vegetable and PGRs

INTRODUCTION

The term parthenocarpy coined by Noll in 1902. The term parthenocarpy derived from two Greek word words Parthenos means virgin and carpy means fruit. Parthenocarpy defined as the development of the ovary into a seedless fruit without the need of pollination and fertilization (Benoit Gorguet, 2007). Absence of pollination is due to Male sterility, Self incompatibility and adverse environmental condition. It may occur naturally or can be induced artificially by exogenous application of hormones or their enhanced endogenous level. Several vegetable crops have been successfully bred to exhibit parthenocarpy including cucumber, tomato, pepper, and eggplant. The development of parthenocarpic varieties involves selecting and breeding plants with specific genetic traits that promote fruit development without fertilization. These traits can be naturally occurring or induced through genetic engineering techniques. For example parthenocarpy in cucumbers is controlled by a single recessive gene known as Parthenocarpy (Pat). By selecting and breeding plants with the Pat gene, breeders have been able to develop cucumber varieties that produce seedless fruits. Similarly in tomato, the application of plant growth regulators such as auxins, can induce parthenocarpy and result in the production of seedless fruits (Dhatt and Kaur, 2016).

IMPORTANCE OF PARTHENOCARPIC VEGETABLES

Parthenocarpic vegetables offer a solution to the problem of low fruit set in certain crops. In some cases, environmental conditions or lack of pollinators can hinder the natural fruiting process. However, with parthenocarpy farmers can still obtain a good yield of fruits even under unfavorable conditions. This is particularly significant in regions with unpredictable weather patterns or where pollinators are scarce. Parthenocarpic vegetables provide a consistent and high-quality harvest. Since they do not rely on pollination, the fruits are less likely to develop deformities or irregularities. This ensures that consumers

receive visually appealing and uniform produce which is important for both marketability and consumer satisfaction. The absence of seeds in parthenocarpic fruits makes them more convenient for culinary purposes which reducing the need for seed removal. Furthermore, parthenocarpy allows for extended growing seasons and increased productivity. As these vegetables do not require pollination, they can be grown in protected environments such as greenhouses or polytunnels. This enables farmers to cultivate crops year-round, regardless of external factors that may affect pollination. Additionally, parthenocarpic vegetables often have shorter maturation periods which allowing for multiple harvests in a single growing season. Parthenocarpic vegetables contribute to food security and sustainability. By providing a reliable source of produce, they help meet the demands of a growing global population. Additionally, their ability to thrive in diverse conditions reduces the reliance on specific climates or pollinators, making agriculture more resilient and adaptable to changing environments. This is particularly important in the face of climate change, where traditional crop production may be compromised (Dalal *et al.*, 2006).

Table 1: Quality parameters of some parthenocarpic vegetables.

Crop	Parthenocarpic	Reference
Watermelon	The shape, flavour and yield are as good as seed-producing cultivars and have a longer shelf life	Kihara, 1951
Gherkin	Seedless gherkins are more crunchy, firmer and fleshier than its seeded variety	Rudich <i>et al.</i> , 1977
Tomato	Seedless tomato fruits are tastier, more dry-matter (up to 1%), contain more sugars, less acidity and less cellulose	Lukyanenko, 1991
	More soluble solids	Gorguet <i>et al.</i> , 2005
Eggplant	High yield and fruit quality	Donzella <i>et al.</i> , 2000

NEGATIVE EFFECTS OF PARTHENO CARPY

Followings are negative effect of parthenocarpy on vegetable crops (Angelo and Giuseppe, 2001).

1. Fruits become misshapen, smaller and duller in appearance, softer in texture.
2. Decreases biodiversity, which reduces plant species resistance to disease.
3. Transfer of genes from seedless crops may cause non-modified plants to become sterile or not produce seeds.
4. Early application of auxins like phytohormones before anthesis damages the flowers. It results in the abortion of seed and fruit drop.

TYPES OF PARTHENO CARPY

1. Genetic/natural parthenocarpy

A. Obligatory: No external influence. It's commercially found in Ivy gourd.

B. Facultative: It occurs due to adverse conditions for pollination and fertilization. It's found in Tomato, brinjal and cucumber.

C. Stimulative parthenocarpy: pollination or other stimulation is required for parthenocarpy. It present in watermelon.

D. Vegetative parthenocarpy: pollination or other external stimulation is not required to produce parthenocarpic fruit *e.g.* cucumber.

E. Stenospermocarpy: Pollination and fertilization occur but the embryo gets aborted. Also produces seedless fruits, but here the seeds are aborted and remains traces of seeds.

2. Artificial induction of parthenocarpy

A. Use of plant growth regulators

Plant growth regulators (PGRs) are organic compounds, other than nutrients that modify plant physiological processes. Many studies have shown that phytohormones play an important role in fruit set regulation, growth and maturation in complex way. The parthenocarpic fruits are associated with the endogenous auxin and gibberellins in the ovary. The exogenous applications of plant growth hormones, like auxins, cytokinins and GAs, can influence parthenocarpic fruits in vegetable crops.

Table 2: Use of plant growth regulators for parthenocarpic fruit development.

Crop	Growth regulator	Stage of treatment	Types of parthenocarpy	Reference
Brinjal	GA ₃ @ 2700 ppm; 2-4-D@2.5 ppm	Foliar spray on freshly opened flower.	GA ₃ induced the completely seedless fruits during all seasons. 2,4-D, induced the development of degenerated seeds	Nothmann and Koller, 1975
Kokrol	2-4-D/2-4-5-T @100mg/L	Pre-anthesis sprays	Complete parthenocarpy	Vijay and Jalikop, 1980
Cucumber	GA ₃ @100mg/L	Pre-anthesis sprays	Complete parthenocarpy	Choudhury and Phatak, 1958
Bottle gourd	CPPU@ 10–100 mg/L	2 days before or after anthesis	Complete parthenocarpy	Jing, 1999

B. Distant hybridization

Interspecific hybridization have been utilized for producing a facultative parthenocarpic line suitable for a hot and dry climate (normal fruit at moderate temperature) was first introduced in tomato (Hawthorn, 1937). Altered ploidy through interspecific hybridization is a common approach to obtain parthenocarpic fruits in various crops such as banana, watermelon and citrus (Fortescue and Turner, 2005).

Table 3: Development of facultative parthenocarpy in tomato by distant hybridization.

Parthenocarpic line/cultivar	Cross involved	Reference
Line RP75/79	Multiple cross Atom × Bobjekosoko and Heinemanns Jubilaum × Piora (developed by R. Reimann-Philipp)	Philouze and Maisonneuve, 1978
Severianin	<i>L. esculentum</i> and <i>L. hirsutum</i> (bred by N. Soloviova)	Lin <i>et al.</i> , 1984
P-26, P-31, etc.	<i>L. esculentum</i> and <i>L. pennellii</i>	Stoeva <i>et al.</i> , 1985
Line RG	<i>L. esculentum</i> and <i>L. cheesmanii</i> var. <i>minor</i>	Mikhailov and Georgiev, 1987
IVT 1	<i>L. esculentum</i> and <i>L. hirsutum</i>	Zijlstra, 1985
IVT 2	<i>L. esculentum</i> and <i>L. peruvianum</i>	Zijlstra, 1985

C. Mutation

Spontaneous mutations occur naturally and are used in classical breeding programmes. Good example of this is the parthenocarpic *sha-pat* mutants in the tomato line Montfavet 191 (Pecaut and Philouze, 1978). Soft-X-ray used successfully to generate parthenocarpic mutants in watermelon

(Kawamura *et al.*, 2018). Gamma irradiation in *Citrullus lanatus* has been used successfully to generate parthenocarpic mutants (Sugiyama and Morishita, 2001). Alkylating agents (EMS and EES) has been used to generate parthenocarpic mutants of tomato (Vivian-Smith *et al.*, 2001).

D. Alteration in chromosome number

Unbalanced development of embryo and endosperm in triploid background has been utilized to yield parthenocarpic fruit. In watermelon seedless fruits with only residual integuments are obtained from F1 hybrid plants derived from cross between tetraploid and diploid parents (Kihara, 1951). Chromosome elimination in wide crosses may lead to the production of haploids, which are of enormous interest to the breeders. Haploid formation following interspecific hybridization is usually interpreted as parthenogenesis (Rowe, 1974).

Table.4 Parthenocarpic vegetables associated with various ploidy levels.

Vegetable	Species	Ploidy no.	reference
Tomato	<i>Solanaum esculentum</i> (2n = 2x = 24)	Triploid (2n = 3x = 36)	Habashy <i>et al.</i> , 2004
	<i>Solanaum esculentum</i> (2n = 2x = 24)	Aneuploid	Lesley and Lesley, 1941
Cucumber	<i>Cucumis sativus</i> (2n = 2x = 14) (Amphidiploid × Diploid)	Triploid (2n = 3x = 21)	Habashy <i>et al.</i> , 2004
Watermelon	<i>Citrullus lanatus</i> (2n = 22) (Autotetraploid × Diploid)	Triploid (2n = 3x = 33)	Kihara, 1951

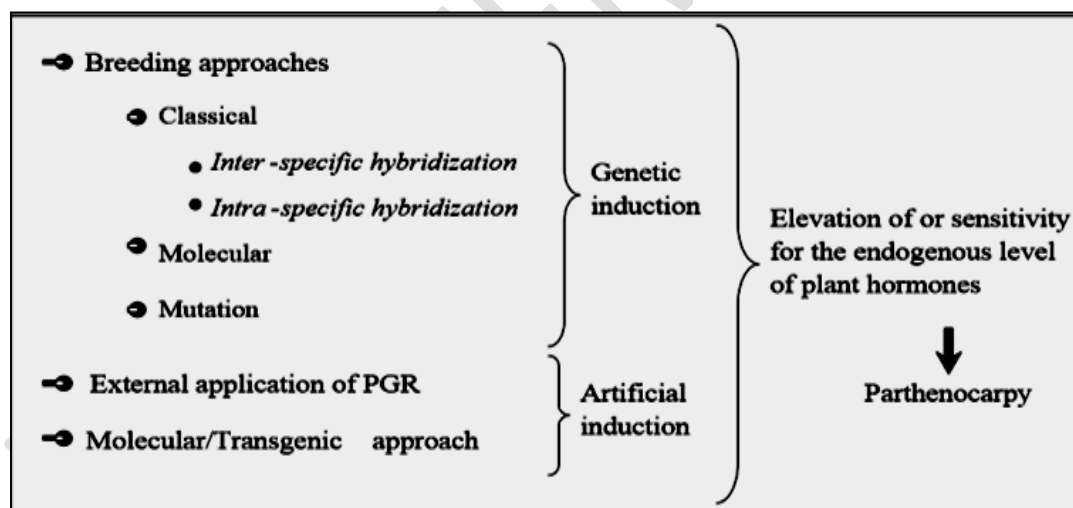


Figure 1: Development methods of parthenocarpic variety

MECHANISM OF PARTHENO-CARPY

The diagram illustrates the mechanism of parthenocarpy in a simplified manner. It shows the key steps involved in fruit development without fertilization. Firstly, the ovary is stimulated to grow by the activation of auxins and gibberellins. This leads to the enlargement of the ovary, which eventually develops into a mature fruit. Meanwhile, the absence of fertilization prevents the formation of seeds, allowing the fruit to divert its resources towards growth and development. Finally, the fruit undergoes ripening, aided by the suppression of ethylene production by gibberellins. The seed and fruit development was control by phytohormones (Pandolfini, 2009). GA₃, auxin and cytokinin involve signaling process

after fertilization for seed and fruit development (Fos *et al.*, 2001). Increase endogenous hormones during parthenocarpic fruit set not from source of seed (Tsao, 1980) Trigger the expression of auxin biosynthetic gene (Carmi *et al.*, 2003).

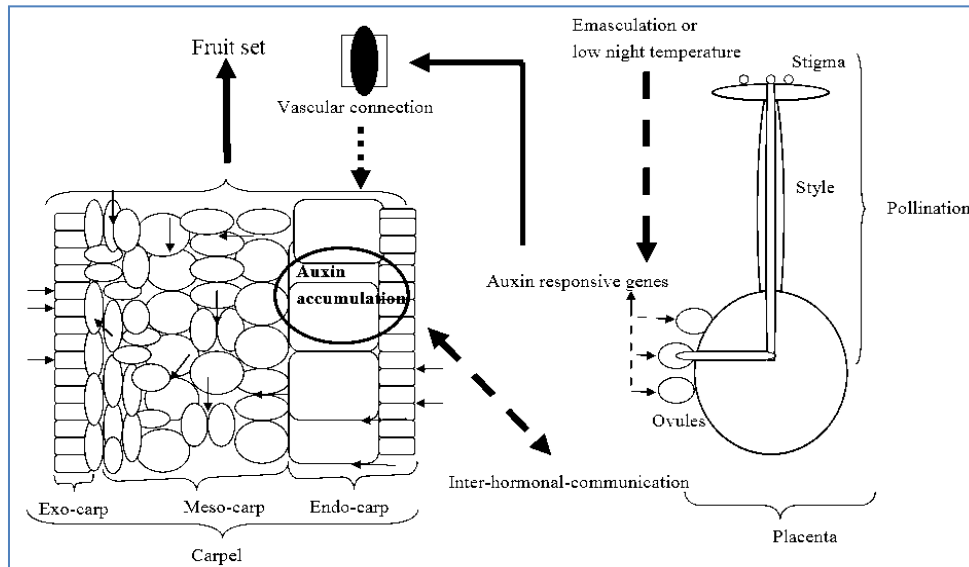


Figure 2: Mechanism of parthenocarpy

Table 5: Genetics of Parthenocarpy in Vegetable Crops

Crops	Genetics	Reference
Tomato	Several single gene recessive	Fos <i>et al.</i> , 2001
Brinjal	Single dominant gene	Kuno and Yabe, 2005
Capsicum	Single recessive gene	Tiwari <i>et al.</i> , 2011
Cucumber	Additive dominant epistatic gene and additive dominant polygenes	Yan <i>et al.</i> , 2010

FACTORS AFFECTING PARTHENO-CARPY

1. Genetic Factors

One of the primary mechanisms of parthenocarpy is genetic factors. Certain plant species possess genes that allow them to initiate fruit development without the need for pollination or fertilization. These genes are responsible for the production of hormones such as auxins and gibberellins, which play a crucial role in fruit development. Mutations or alterations in these genes can lead to the occurrence of parthenocarpy. The *SlAGL6* gene in tomato plants has been identified as a key regulator of parthenocarpy, as its mutation results in the production of seedless fruits (Fos *et al.*, 2001).

2. Hormonal Factors

Hormones play a significant role in parthenocarpy. Auxins, particularly indole-3-acetic acid (IAA) are essential for the initiation and growth of fruits. In some cases high levels of auxins in the ovary can trigger fruit development without fertilization. Ethylene also has an impact on parthenocarpy. It can promote fruit growth and ripening and its application can induce parthenocarpy in certain plant species. Gibberellins, cytokinins and abscisic acid are other hormones that influence fruit development and can contribute to parthenocarpy (Nothmann and Koller, 1975).

3. Environmental Factors

Environmental conditions can also influence the occurrence of parthenocarpy. Temperature, light and water availability are among the key factors that can affect fruit development. High temperatures during the flowering period can stimulate the production of auxins which leading to parthenocarpy.

Similarly, exposure to specific light conditions such as prolonged periods of darkness also can trigger hormonal changes that promote fruit development without fertilization. Adequate water supply is crucial for fruit growth and water stress can inhibit the occurrence of parthenocarpy (Dalal *et al.*, 2006).

4. Pollination and Fertilization

While parthenocarpy is defined as fruit development without fertilization, the presence of pollination and fertilization can still influence its occurrence. In some cases, the presence of pollination signals triggers the production of hormones that initiate fruit development. Additionally, fertilization can enhance the growth and quality of parthenocarpic fruits. The interaction between pollination, fertilization and parthenocarpy is complex and varies among different plant species (Benoit Gorguet, 2007).

UTILITY OF PARTHENOCAOPY IN VEGETABLE CROPS

1. Tomato

Three source of facultative type parthenocarpy present in tomato *Viz.*, Soressi or Montfavet-191 (*Pat-1*), Severianin (*Pat-2*) and RP75/59 (*Pat-3/Pat-4*) (Gorguet *et al.*, 2005). Thien *et al.* (2021) Introducing *iaa9-3* mutation into tomato cultivar and they recorded that developed parthenocarpic cultivar retaining the normal stamen structure and fertile pollen, the parthenocarpic lines can produce sufficient seeds for self-pollinated propagation, average seedless fruit weight of experimental lines ranged from 25 g/fruit (small category) to over 50 g/fruit (intermediate category), together with good number of fruits/plant which can lead to a theoretical seedless fruit yield of over 3000 g/plant, which is acceptable for commercial varieties and increasing the quality of seedless fruits compared to seeded fruits which is attributable to an increase in the mass of placental area and a higher °Brix level than locule and pericarp areas. Optimized the CRISPR/Cas9 system to introduce somatic mutations effectively into *SlIAA9* a key gene controlling parthenocarpy with mutation rates of up to 100% in the T₀ generation which studied by Ueta *et al.* (2017). Federico *et al.* (2009) obtain parthenocarpic transgenic tomato plants (*cv MicroTom*) by the regulation of genes for auxin synthesis (*iaaM*) or responsiveness (*rolB*) driven by *DefH9* or the INNER NO OUTER (*INO*) promoter from *Arabidopsis thaliana*. Rotino *et al.* (2005) recorded that minimum percentage of fruit with seeds and number of seeds per fruit in transgenic parthenocarpic line Ri4. Andrea *et al.* (1998) revealed that 59% fruit with zero seeds in Pat genotype under 26 °C days and 18 °C night temperature.

2. Brinjal

Anupama *et al.* (2021) recorded that parthenocarpic line 93213-PC-2-1 and 93213-PC-2-3 were earlier for days to first harvest, at the top for the number of flowers per cluster, number of fruits per cluster and maximum number of marketable fruits per plant when grown under net house conditions. Saito *et al.* (2015) developed parthenocarpic cultivar Anominori 2 go which is an F₁ hybrid between two parthenocarpic inbred lines AE-P01 and AE-P24. Saito *et al.* (2009) developed parthenocarpic cultivar Anominori from AE PO8 and AE PO1 parthenocarpic line through pedigree methods. Acciarri *et al.* (2002) revealed that yield per plant and fruit weight recorded maximum in P₂ parthenocarpic hybrid.

3. Cucurbits

Nagamani *et al.* (2019) noted that Alexious parthenocarpic hybrid of cucumber recorded maximum fruit length, fruit diameter, average fruit weight, fruit yield per vine and total yield under green house conditions. In cucumber maximum fruit firmness, fruit colour, fruit flavor and fruit texture revealed in JSCU 01 parthenocarpic cultivar under polyhouse which studied by Kumar *et al.* (2017). Kumar *et al.* (2015) also in cucumber, they recorded that parthenocarpic hybrid KPCH -1 perform superior related to yield parameters as compare to other parthenocarpic hybrid. Furthermore, Kumar *et al.* (2014) recorded utmost LAI, fruit yield and crude fiber with application of 150% RDF through fertigation and single stem straining system in parthenocarpic cucumber under NVPH. Sravani *et al.* (2018) observed minimum number of seeds per fruit with 2,4-D at 25 ppm concentration in watermelon. Rasul *et al.* (2008) revealed

that applications of 2,4-D at 100 ppm concentration observed parthenocarpic fruit development in spine gourd genotype Rangpuri (C13).

Conclusion

Parthenocarpy has proven to be a beneficial trait in vegetable crops. It allows for the production of seedless fruits, which are highly desirable in the market due to their improved taste, texture and convenience. Parthenocarpic varieties also have the advantage of being able to set fruit under unfavorable environmental conditions, such as low temperatures or high humidity. This trait has the potential to increase yields and improve crop productivity which contributing to food security and sustainability. However, further research is needed to fully understand the genetic and physiological mechanisms underlying parthenocarpy in vegetable crops and to develop new varieties that combine this trait with other desirable characteristics such as disease resistance and nutritional value. Overall, parthenocarpy holds great promise for the future of vegetable crop production and should be further explored and utilized to meet the increasing demands of a growing global population.

REFERENCES

- Angelo, S. and Giuseppe, L. R. (2001). Parthenocarpy: state of the art. *Curr. Trends in the Embryology of Angiosperms*, **1**: 435-450.
- Dalal, M.; Dani, R. G.; Kumar, P. A. (2006). Current trends in the genetic engineering of vegetable crops. *Scientia Hort.*, **107**(3):215-225.
- Dhatt, A. S. and Kaur, G. (2016). Parthenocarpy: a potential trait to exploit in vegetable crops a review. *Agril. Rev.*, **37**(4): 300-308.
- Fortescue, J. A. and Turner, D. W. (2005). Growth and development of ovules of banana, plantain and enset. *Sci. Hort.*, **104**(4): 463-478.
- Gorguet, B. (2008). Parthenocarpy and functional sterility in tomato. Ph.D. thesis, Wageningen University, Netherland, pp 139.
- Gorguet, B.; Heusden, A. W. and Lindhout, P. (2005). Parthenocarpic fruit development in tomato. *Pl. Biol.*, 131-139.
- Hawthorn, R. L. (1937). Seedlessness in tomatoes. *Sci.*, **85**(2199): 199-199.
- Jing, Q. Y. (1999). Parthenocarpy induced by N-(2- chloro 4- pyridyl)-N-phenylurea (CPPU) prevents flower abortion in Chinese white flower gourd. *Environ. and Exp. Botany*, **42**(12): 121-128.
- Kihara, H. (1951). Triploid water melons. *Proceedings of the Amer. Soc. for Horti. Sci.*, **58**:217-230.
- Lin, S.; George, W. L. and Walter, E. S. (1984). Expression and inheritance of parthenocarpy in severianin tomato. *J. Heredity*, **75**(1): 62-66.
- Lukyanenko, A. N. (1991). "In Monographs on theoretical and applied genetics: genetics improvement of tomato", pp 167-178.
- Mikhailov, L. and Georgiev, K. H. (1987). Line RG-a source of parthenocarpy in tomato. *Genet. Sel.*, **20**: 70-71.
- Nothmann, J. and Koller, D. (1975). Effects of growth regulators on fruit and seed development in eggplant. *J. Horti. Sci.*, **50**(1): 23-27.
- Philouze, J. and Maisonneuve, B. (1978). Heredity of the natural ability to set parthenocarpic fruits in a German line. *Report of the Tomato Genetics Cooperative*, **12**: 12-25.
- Rudich, J.; Baker, L. R. and Sell, H. M. (1977). Pathenocarpy in cucumber as affected by genetic parthenocarpy, thermo-photo period and femaleness. *J. American Society for Horti. Sci.*, **102**(2): 225-228.
- Stoeva, P. K.; Michailov, L and Georgiv, C. (1985). Parthenocarpy in hybrid *L. esculentlln* x *L. penellii*. *Rep. Tomato Genet. Croop.*, **35**: 19.
- Vijay, O. P. and Jalikop, S. H. (1980). Production of parthenocarpic fruit growth regulators in teasle gourd. *Ind. J. Horti.*, **37**: 167-169.
- Zijlstra, S. (1985). Parthenocarpy in tomato: two new line from interspecific crosses. *Aadbelangen*, 39: 92-94.
- Pecaut, P. and Philouze, J. (1978). A sha-pat line obtained by natural mutatuin. *Tomato Genetics Cooperative Reports*, **28**: 12.

- Kawamura, S.; Ida, K.; Osawa, M. and Ikeda, T. (2018). No effect of seed presence or absence on sugar content and water status of seeded and seedless watermelon fruit. *Hortic. Sci.*, **53**(3): 304-312.
- Vivian-Smith, A.; Luo, M.; Chaudhary, A. and Koltunow, A. M. (2001). Fruit development is actively restricted in the absence of fertilization in Arabidopsis. *Development*, **128**: 2321-2331.
- Rowe, P. R. (1974). Parthenogenesis following interspecific hybridization. *Int. Symp., Univ. Guelph.*, pp 1-9.
- Habashy, A. A.; Testa, G.; Mosconi, P.; Caccia, R.; Mazzu, A.; Santange, E. and Soressi, G. P. (2004). Parthenocarp restores fruitfulness in sterile triploid tomatoes artificial obtained by crossing $4x \times 2x$ somaclones. *J. Hort. Sci. Biotechnol.*, **79**: 322-328.
- Lesley, M. M. and Lesley, J. W. (1941). Parthenocarp in a tomato deficient for a part of chromosome and its aneuploid progeny. *Genetics*, **26**: 374-386.
- Pandolfini, T. (2009). Seedless fruit production by hormonal regulation of fruit set. *Nutrients*, **1**: 168-177.
- Fos, M.; Proano, K.; Nuez, F. and Garcia, J. L. (2001). Role of gibberellins in parthenocarpic fruit development induced by the genetic system pat-3/pat-4 in tomato. *Physiol. Pl.*, **111**: 545-550.
- Carmi, N.; Salts, Y.; Dedicova, B.; Shabtai, S. and Barg, R. (2003). Induction of parthenocarp in tomato via specific expression of the rolIB gene in the ovary. *Planta.*, **217**: 726-735.
- Tsao, T. (1980). Growth substance: role in fertilization and sex expression. *Spring Verlag*, 345-348.
- Kuno, S. and Yabe, K. (2005). Genetic analysis of parthenocarp and spineless in the F2 segregating generation between parthenocarp F1 variety and spineless line in eggplant. *Res. Bulletin of the Aichi Ken Agril. Res. Center*, **1**: 21-28.
- Tiwari, A.; Smith, A. V.; Voorrips, R. E.; Habets, M. E.; Xue, L. B.; Remko, O. and Heuvelink, E. P. (2011). Parthenocarpic potential in capsicum is enhanced by carpelloid structures and controlled by single recessive gene. *BMC Pl. Biol.*, **11**: 143.
- Yan, L. Y.; Lou, L. N.; Li, X. L.; Feng, Z. H.; Lou, Q. F. and Chen, J. F. (2010). Inheritance of parthenocarp in monoecious cucumber. *Sci. Agric. Sin.*, **6**: 26.
- Ueta, R.; Abe, C.; Watanabe, T.; Shigeo, S. S.; Ishihara, R.; Ezura, H.; Osakabe, Y. and Osakabe, K. (2017). Rapid breeding of parthenocarpic tomato plants using CRISPR/Cas9. *Scientific Report*, **7**(1): 507.
- Federico, M.; Sandra, L. U.; Russell, L. R.; Chen, Y.; Tricoli, D.; Fiehn, O.; Rocke, D. M.; Gasser, C. S. and Dandekar, A. M. (2009). Gene regulation in parthenocarpic tomato fruit. *J. Exp. Botany*, **60**(13): 3873-3890.
- Andrea, M.; Taddei, A. R. and Soressi, G. P. (1998). The parthenocarpic fruit (pat) mutant of tomato (*Lycopersicon esculantum* M.) set seedless fruits and has aberrant anther and ovule development. *Development*, **125**(1): 107-114.
- Rotino, G. L.; Acciarri, N.; Sabatini, E.; Mennella, G.; Scalzo, R. L.; Maestrelli, A.; Molesini, B.; Pandolfini, T.; Scalzo, J.; Mezzetti, B. and Spena A. (2005). Open field trial of genetically modified parthenocarpic tomato: seedlessness and fruit quality. *BMC Biotechnology*, **5**(32): 105-118.
- Thien, L.; Nguyen, A. T.; Nguyen, M. H.; Nguyen, L. T.; Nguyen, M. T.; Trinh, L. T.; Tran, D. T.; Ta, S. V.; Hoshikawa, K.; Sugimoto, K. and Ezura, H. (2021). Developing new parthenocarpic tomato breeding lines carrying iaa9-3 mutation. *Euphytica*, **217**(7): 139.
- Gorguet, B.; Van, A. W. and Lindhout, P. (2005). Parthenocarpic fruit development in tomato. *Pl. Biology*, 131-139.
- Acciarri, N.; Restaino, F.; Vitelli, G.; Perrone, D.; Zottini, M.; Pandolfini, T.; Angelo, S. and Giuseppe, L. R. (2002). Genetically modified parthenocarpic eggplants: improved fruit productivity under both greenhouse and open field cultivation. *BMC Biotechnology*, **2**(4): 1472-1478.
- Saito, T.; Matsunaga, H.; Saito, A.; Hamato, N.; Takeshi, K.; Toshiyuki, S. and Yoshida, T. (2009). A novel source of cytoplasmic male sterility and a fertility restoration gene in eggplant (*Solanum melongena* L.) lines. *J. Japan. Soc. Hort. Sci.*, **78**(4): 425-430.

- Anupama, S.; Kaur, S. M. and Singh, D. A. (2021). Performance of parthenocarpic lines of brinjal (*Solanum Melongena* L.) in net house and open field during the rainy season. *Vegetable Sci.*, 48(1): 79-85.
- Saito, T.; Matsunaga, H.; Saito, A.; Yoshida, T. and Monma, S. (2015). Development of the parthenocarpic eggplant cultivar anominori 2 go. *Bulletin of the national institute of Vegetable and Tea Sci.*, 14: 1-14.
- Kumar, P.; Khapte, P. S.; Saxena, A. and Kumar P. (2015). Performance of different Parthenocarpic cucumber F1 hybrids under poly house. *Ind. J. Agri. Sci.*, 89(3): 545-50.
- Kumar, S.; Patel, N. B. and Saravaiya, S. N. (2014). Response of Parthenocarpic cucumber to fertilizers and training systems under NVPH in subtropical conditions. *Int. J of Cur. Res.*, 6(8): 8051-8057.
- Sravani, V.; Ashok, P.; Sasikala, K. and Ramesh, B. B. (2018) Induction of parthenocarpy through growth regulators in watermelon (*Citrullus lanatus* Thunb.). *Int. J. Chemi. Studi.*, 6(6): 182-184.
- Rasul, G. M.; Abdul, K. M.; Yasuhiro, C.; Yukio, O. and Hiroshi, O. (2008). Application of Plant growth regulators on Parthenocarpic Fruit Development in Teasle Gourd (*Momordica dioica* Roxb.). *J. Fac. Agri. Kyushu University*, 53(1): 39-42.
- Kumar, S.; Chaudhari, V. I.; Saravaiya, S. N. and Dev, R. (2017). Potentiality of greenhouse cucumber cultivars for economics and nutritional realization. *Int. J. Farm Sci.*, 7(1): 1-7.
- Nagmani, G. V.; Kumar, J. S.; Reddy, T. B.; Rajesh, A. M.; Amarananjundeswara, H.; reddy, R. L. and Doddabasappa, B. (2019). Performance of different parthenocarpic cucumber (*Cucumis Sativus* L.) hybrids for yield and yield attributing trait under shade net house. *Int. J. Curr. Microbiol. And App. Sci.*, 8: 978-982.