

## **Biopesticide effect of beneficial microorganisms' formulation against *Phytophthora infestans* (mildew) and *Aternariaspp.* (alternariosis) on *Capsicum annuum* L. in field**

### **Abstract**

The production of pepper, a staple food of millions of people in the world is facing a severe parasitic pressure of pests and diseases. The objective of this work aimed at developing a biopesticide formulation (T1) made up with combination of *Trichoderma harzianum* and *Metharizium anisopliae* and test its performance against the virulence of *Alternaria solani* and *Phytophthora infestans* isolated from *Capsicum annuum* L fruits and leaves. A significant difference ( $p = 0.05$ ) was revealed between the formulation (T1), chemical pesticide as positive control (T2) made up with Mandipropamid and the control (T0) in the field on disease incidence and severity. Mildew incidence ranged from 12.72% to 53% for the V1 variety (Safi) and 10.01% to 49% for the V2 variety (*Burkina yellow*) in agro-ecological area III. Similarly, 8.99% to 60.9% for variety V1 and 7.12% to 55% for variety V2 in agro-ecological area V between 4 and 12 weeks. The incidence of alternariosis varied from 7.12% to 77.12% for variety V1 (*Safi*) and 7.01% to 69.15% for variety V2 (*Burkina yellow*) in agro-ecological area III. Also 4.32% to 71.9% for variety V1 and 5.12% to 67.08% for variety V2 in agro-ecological area V between 4 and 12 weeks. Gas chromatography combined with mass spectrometry revealed an overproduction of bioactive molecules in the hydro-methanolic extracts of pepper leaves treated with the formulation, which are responsible for its phytoprotection in the two agroecological areas. In conclusion, the use of the *Trichoderma harzianum* and *Metharizium anisopliae* formulation significantly induced systemic resistance of *Capsicum annuum* L plants against alternariosis and mildew agents.

**Keywords:** Alternariosis, Mildew, *Trichoderma harzianum*, *Metharizium anisopliae*; *Capsicum annuum* L; bioprotective molecules.

### **1. Introduction**

Agriculture is an important economic sector in Cameroon. It contributes 15.6% of the national Gross Domestic Product [1]. However, Cameroon is committing substantial resources to ensuring the success of the agricultural sector, which has not yet made any

significant headway due to a major problem linked to the attack of cultivated fields by insect pests and phytopathogens, causing serious damage to economically important crops [2].

Food crops are threatened annually by a wide range of pests, including micro-organisms, animals (vertebrates and invertebrates) and plants (weeds). In order to reduce crop damage, predator control strategies must be put in place. The use of synthetic pesticides has proved effective in protecting crops, but it has also increased the environmental impact in terms of soil and water contamination and phytotoxicity. There have also been a number of cases where these products have lost their efficiency, due to pest resistance [3]. Pepper diseases cause quantitative and qualitative crop damage throughout the world [4]. In Cameroon, the spectrum of occurrence and development of pathologies is increasing over the years, and the literature provides few details of the disease. One reason why farmers find it difficult to implement an effective control strategy [4]. Phytopathogenic fungi are species of parasitic fungi that cause cryptogamic diseases in plants. These fungi belong to the various groups of the oomycete and/or eumycete kingdom ("true fungi"): ascomycetes, basidiomycetes, chytridiomycetes, zygomycetes and deuteromycetes (imperfect fungi) [5]. They are able to infect any tissue at any stage of plant growth, undergoing a complex life cycle that may involve sexual or asexual reproductive stages [6]. Despite its nutritional and economic importance, there is a growing shortage in capsicum production. This is mainly due to the negative impacts of phytopathogenic agents that cause diseases of various levels of severity on this plant. *Alternaria* spp. and mildew are common fungi in our environment. They have saprophytic and phytopathogenic lifestyles that can affect field crops or plant products during harvest and post-harvest [7]. *Alternaria solani*, one of the *Alternaria* pathovars, is a phytopathogenic fungus of the Pleosporaceae family. It has been reported for several decades as a pathogen of Solanaceae and has long been described as affecting pepper as well as tomato, eggplant, potato, and several species of this botanical family [4]. It causes a disease called "Alternariosis" or "Alternaria blight" by secreting toxins such as tenuazonic acid, alternariol, AOH-9-glucoside, altenariol monomethylether-3-glucoside [8]. Pepper mildew, on the other hand, is a disease that can destroy entire fields over long distances. It must be regarded as a disease of "collective scope", and it is vital that preventive and curative control measures are adopted for better biocontrol. Under certain conditions, *P. infestans*, the pathogen linked to this infection, can produce survival structures (oospores) that enable it to survive for several years in the soil, without a host plant. However, oospore production is only possible when two reproductive types (type A and type B) are present concurrently in the

same field. High relative humidity (>90%) and medium temperatures (between 10 and 15°C at night and 15 and 21°C during the day) are favorable conditions for mildew development. On the other hand, prolonged periods of drought and hot temperatures (> 30°C) are detrimental to its survival and dispersal [9].

Over the past few years, interest in biological protection has been renewed, due to a growing concern for better protection of the environment, and the desire for product quality imposed by consumers. The use of biological control agents has now become a reality in agriculture, particularly for the control of plant pathogens. Biological control using effective bacterial and fungal antagonists is also one of the strategies adopted to manage pests [10]. Indeed, these micro-organisms generally control their target pathogens through various mechanisms, including antibiosis, mycoparasitism, competition for space and nutrients, production of lytic enzymes and secondary metabolites, induction of plant resistance systems and inactivation of pathogen enzymes [11]. Thus, the inhibitory effects of microorganisms on plant pathogens and pests are increasingly being explored [12]. Commercial microbial inoculants (commonly used as biofertilizers or biostimulants) containing single species or strains of Rhizobia, *Pseudomonas* spp., *Azotobacter* spp., *Bacillus* spp., *Trichoderma* spp., *Aspergillus* spp., and *Glomus* spp. have been widely used in small agroecosystems for crop production [13]. Taking for example the case of certain *Bacillus subtilis* strains, which are the most extensively studied. Their inhibitory effect has been demonstrated against *Fusarium oxysporum*, *Sclerotinia sclerotiorum*, *Rhizoctonia solani* and *Colletotrichum gloeosporioides* [14]. In addition, *Bacillus subtilis* is a bacterium widely known for its insecticidal effects in agriculture. Hence their phytoprotective effect.

## **2. Materials et methods**

### **2.1 Study location**

The study was carried out in agro-ecological areas III, IV due to the high trend of pepper cultivation. The study in area III took place in the locality of Njichom in the Noun Division. It is located at altitude 1112 m, with geographical coordinates 5°47'43" N and 10°56'29" E. Its climate is Sudano-Guinean, with an average temperature of about 21.2°C and average annual rainfall of 609 mm (Anonymous, 2019). Area IV is located in Mbele II, in the Obala Subdivision of the Lekie Division. This locality is at an altitude of 528 meters, with

geographic coordinates of 4°10'0" N and 11°31'60" E. Its climate is tropical humid, with a relative temperature of 24.7°C and average annual rainfall of 1,638 mm [15].

## 2.2 Material

The microbial strains used in the study were *Trichoderma harzianum* ( $2 \times 10^9$  CFU/g) and *Metharidium anisopliae* ( $10^5$  CFU/g), acquired from Dora Agritech laboratories in China. The plant material included two of Cameroon's most valued pepper varieties, *Safi* (Variety V1) and *Burkina yellow* (Variety V2). They have a high yield capacity of about 20 to 40 T/ha. The chemical pesticide Mandipropamid 25%, marketed under the name REVUSTOP or REVUS 250sc, was used as a positive control.

## 2.3 Morphological identification and compatibility tests between microorganisms

*Trichoderma harzianum* and *Metharidium anisopliae* isolates were morphologically identified. A 2 g sporal suspension of pure *Trichoderma* spp. and *Metharidium* spp. cultures was made in 98 ml sterile distilled water. After a series of dilutions ( $10^{10}$ ) and plating of 1 ml on PDA medium, followed by incubation for 7 days at 25°C in the dark, the germinated conidia were observed under a microscope with objective 40 and magnification X400. The conidia were transplanted with a needle onto PDA medium and incubated for one week at 25°C, with the appearance of green mycelium indicating the presence of *Trichoderma* spp. and *Metharidium* spp. [16].

To determine the additive and synergistic effects of the isolates, compatibility between the two microbial strains was determined by performing a diffusion test on modified agar (or Kirby-Bauer test). A 24-hour-old culture loop grown in LB agar (Luria-Bertani) was transferred to 4 glass test tubes containing 5 ml LB broth and incubated for 24 hours on an incubator shaker (New Brunswick Scientific CO., Inc, Edison, NJ 08817, USA) set at 200 rpm and 30°C. The concentration of *Trichoderma harzianum* suspension was then quantified and adjusted to  $\sim 10^8$  CFU/ml. Sterile 8 mm Whatman filter paper discs were aseptically immersed in the *Metharidium anisopliae* medium suspension and air-dried for 30 minutes. *Trichoderma harzianum* strain was dabbed evenly onto the plate using a sterile cotton-tipped applicator. Four discs impregnated with the suspension representing *Metharidium anisopliae* were gently pressed onto the *Trichoderma harzianum*-inoculated agar surfaces at four equidistant positions using sterile forceps. Plates were incubated at  $28 \pm 2^\circ\text{C}$  and observed over a 72 h period.

## 2.4 Formulation process

The "vehicle" used was rice bran according to the modified method of Olivera *et al.*[17]. This involved a mixture of 1kg of each microbial formulation with 1kg of brown sugar in 8l of non-chlorinated water. The mixture was fermented in a hermetically sealed bucket for 07days. The resulting product was then mixed with 50kg of rice bran and transferred to a drum for further fermentation of 7 days for fast growth of the microorganisms. The portion to be sprayed as a biopesticide was prepared using 5kg of the solid fraction mixed with 5L of molasses and 5L of mildew. This was mixed in a 100L drum, topped up with non-chlorinated water, then stirred and fermented for 7 days. The product was then ready for use in the field.

## 2.5 Field preparation and Bioinoculation

The two-factor split-plot design consisted of 06 basic plots per block for each pepper variety, separated from each other by an interval of 1 m and from each other by an interval of 2 m for each pepper variety. Plants were spaced 50 cm apart. All the plots in a block represented the same treatment. Each plot had 20 plants. Every two plots in the blocks consisted respectively of treatments T0 (control); T1 (*Trichoderma harzianum* and *Metharidium anisopliae*) and the positive treatment T2 (Mandipropamid). The biopesticide based on beneficial microorganisms and the chemical pesticide were applied to the crop soil a week before sowing in order to reduce the impact of phytopathogens. Other applications were made once every 7 days until harvest. For the treatment at T3, the application for the same frequencies was based on spraying by mixing 25 mL of Mandipropamid in 10 L of water.

## 2.6 Symptomatic identification and characterization of *Alternia* spp. and *Phytophthora infestans*

Observations on the leaves of seedlings in the field were made from 4 weeks after sowing to the first harvest corresponding to 10 weeks. Leaf symptoms were observed and described for each accession. Appearance, shape, size and coloration of symptoms were described by visual observation. Leaves and fruits from accessions showing different types of fungal attack symptoms were randomly sampled. Thus, five leaves with the same type of symptoms were collected from five plants per treatment for each variety in the different agro-ecological areas and placed in blotting paper, and then in a polyethylene plastic bag for laboratory analysis.

Colonies of *Alternaria* spp. spores were identified through the symptoms observed on the plant and fruits using infected fruits, which were washed in 200ml of water. This solution was filtered through 0.45µm millispore membranes. The gelatin/glycerine cube was used to recover spores from the filter surface. These were then transferred to PDA culture medium in petri dishes. Culture was carried out in an incubator at 28°C for 7 days, with photoperiods of light and dark for 08 h/16 h. The spore-laden surface was scraped to remove spores using a metal spatula and distilled water. The resulting suspension was filtered through muslin to separate the spores from the mycelium [18]. The nature of mycelia growth, shape of conidia was noted. Numbers of septa were also recorded.

For plants infected with mildew, symptoms on leaves and fruits were sampled from 20 leaves and tomato fruits of each variety in both agroecological zones. Small fragments of infected plant organs (fruit and leaf) were surface-disinfected by dipping in 0.5 % (w/v) sodium hypochlorite for 2-5 min and rinsed with sterile water before they were placed onto potato dextrose containing streptomycin, in 90-mm Petri dishes. Height Petri dishes were used, for each plant organ, each with one pepper variety from each agroecological zone. Subsequently, plates were incubated at 25 °C until the mycelial growth was observed. After 3-5 days of growth on PDA, isolates were identified by morphological and cultural characteristics [19]. Disease severity on plants was evaluated in both areoecological areas on plants cultivars by counting infected plants above 20 plants in each of 6 plots during harvest.

## **2.7 Resistance of pepper plants to pathogens**

The incidence (I) or expansion rate of the disease is the frequency of occurrence of the disease on plants in a block was thus determined and expressed as a percentage according to the formula [20]:

$$I = \frac{n}{N} \times 100$$

I = Disease incidence

n = Number of infected plants in plot

N = Total number of plants in the plot

Once these two parameters had been calculated, mean values were compared between the treatment and the T0 control.

## **2.8 Pathogenicity Test**

The pathogenicity test was performed on healthy young pepper leaves placed on blotting paper soaked in distilled water in Petri dishes ( $\varnothing = 90\text{mm}$ ) and treated with the spore suspension (10  $\mu\text{L}$ ). Petri dishes containing 10-day-old pure isolates of both *Phytophthora infestans* and *Alternaria* spp. on PDA medium were selected for the preparation of spore suspensions. Healthy control leaves were treated with sterile distilled water (SDS). Inoculated dishes were sealed and incubated under laboratory conditions at 23°C. Observations were made from day 2 of treatment to day 15 [21].

## 2.9 Phytochemical screening

Extracts of leaves treated with combination of *Trichoderma harzianum* and *Metharidium anisopliae* in field and non-treated leaves (control) were analyzed using GC/MS. Leaf extraction was carried out on Soxhlet delipidated leaves (with methanol as solvent) according to the method of Oomahet *al.*[22] by reflux decoction of 50 g of powder in 500 ml of a hydro-methanolic mixture (20V/80V respectively) for 30 min, with the maximum aim of extracting polar compounds such as polyphenols. After filtration on Whatman No.1 paper, the filtrates obtained were evaporated using a rotary evaporator at 60°C. The filtrate residues were oven-dried for 48 h at 45°C to obtain the dry extracts. The methanolic extract was analyzed by capillary gas chromatography combined to mass spectrometry (GC/MS) using an Agilent 7890A GC coupled to an Agilent MSD 5975C inert mass spectrometer. The gas chromatograph was equipped with a VF-1MS capillary column (100% dimethylsiloxane, 20 m x 150  $\mu\text{m}$  (internal diameter) x 0.15  $\mu\text{m}$  (film thickness)) from Varian, liner internal diameter 4 mm. Carrier gas: helium (constant flow rate of 1 mL/min); oven programming: from 37°C (1 min) to 250°C at 5°C/min, then 11 min step-up to 250°C; source temperature: 230°C; transfer line temperature: 260°C; ionization energy: 70 eV. Electron ionization (EI) mass spectra were recorded between 40-400 u. In mass spectrometry, detection is based on the mass-to-charge ratio ( $m/z$ ) of an analyte. The analyte molecules in the vapor phase are bombarded with a high-energy electron beam. Positive ions are produced by bombardment in a magnetic field on the basis of their  $m/z$  ratio. The signal emitted by the ions from the spectrum is recorded and presented as a computer-generated graph. Constituent identification was based on comparison of mass spectra with commercial Wiley and in-house laboratory databases. Retention indices were calculated using the homologous alkane series (C7-C40) and compared with retention indices available in the NIST Webbook after Kondjoyan&Berdague,[23]. Each analysis was performed in triplicate after experimental optimization.

## 2.10 Statistical analysis

The results were analysed statistically by the one-way variance analysis method. The Tukey test was used to compare the different means in order to highlight any significant differences between them. Data were first registered on Excel version 2016 and then analyzed using the Rcmd package of R version 3.6.3 softwares.

## 3. RESULTS AND DISCUSSION

### 3.1 Test of compatibility

Test was carried out between combinations of the two microbial strains to assess their ability to cohabit in the same bioformulation. It has then been proved that *T. harzianum* and *Metharizium anisopliae* have developed a capacity to cohabit in the same environment. In the process of organic matter management technologies, more attention has to be paid to microbiology, and physical, chemical and technical approaches to ensure plant protection against phytopathogens while causing more environmental damage. All those factors are supposed to be taken into consideration when producing bioformulations.

Development of an easy applicable biopesticide most therefore take in account the delivery system for the application of the field. Thus, it is important to determine the size of the particle and flow properties which are parameters that can help to predict the distribution and to choose the ingredient that may favor good distribution of the active ingredient [24].

### 3.2 Symptomatic identification and characterization of *Alterniaspp.* And *Phytophthora infestans*

Direct and indirect techniques were used to obtain isolates from mycelium grown on pepper sections and leaves in the laboratory. Macroscopic (Figs. 1&2) and microscopic (Figs. 1B and 1C) characteristics of pure strains from the two agro-ecological areas of pepper production were observed. The pure strains showed short, whitish, aerial mycelial filaments. These mycelial filaments, observed under a light microscope at X100 magnification, are unpartitioned and display terminal sporocysts (Figs. 1B&2B) which differentiate after heat shock into zoospores (Fig. 1C) typical of *P. infestans*.

Beninal[25]highlighted this difficulty when characterizing *Phytophthora infestans* populations in Algeria. This could be explained by the low competitiveness of *P. infestans* with saprophytic fungi. Morphological and morphometric characterization of *P. infestans* isolates showed significant variation in morphological characters. These morphological

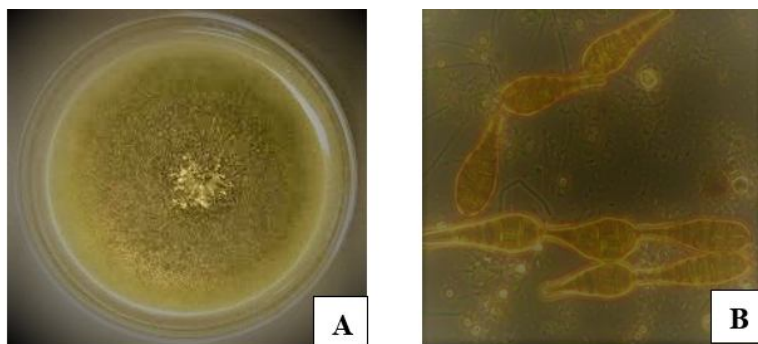
characteristics are similar to those described by Gallegly & Hong [26] in their work on the morphological identification of *Phytophthora* species. This can be explained by the fact that *P. infestans* populations are highly diverse in Cameroon. Indeed, Coulibaly *et al.* [27] had already pointed this out when working on the characterization of *Phytophthora* spp isolates from the cocoa orchard of Côte d'Ivoire. Indeed, the high genetic diversity observed in *P. infestans* populations in Northern Europe and Estonia has been attributed to the involvement of sexual reproduction [28]. The presence of these morphotypes in the two main pepper production basins is thought to result from the different origins of the pepper seeds used in the growing areas [29]. These seeds would be infected by strains of different sexual types.

Morphological characters of *Alternaria* spp. were quite close to those defined by Simmons [30]. Identification of *Alternaria* relied on morphological and cultural characters which cause identification of *Alternaria* spp. difficult. Purified fungal isolates were identified based on their morphological characters, including spore size, growth pattern and spore chain formation [31]. *Alternaria* spp. is an opportunistic fungus that assumes worldwide distribution and responsible for causing pepper leaf spot disease has not yet been reported in Cameroon. As the morphological characterization of *Alternaria* spp. is a challenging and often misleading task due to the striking similarities among the conidia of several species and the cultural features of some *Alternaria* spp. are unstable [32]. Qinet *al.* [8] reported that morphological characters used to delineate species in the *Alternaria* section *Alternaria* are phenotypically plastic and do not allow the reproducible differentiation of several morphospecies. These characteristics may therefore be misleading in the description and



comparison of small-spored catenulate *Alternaria* isolates in this study [29].

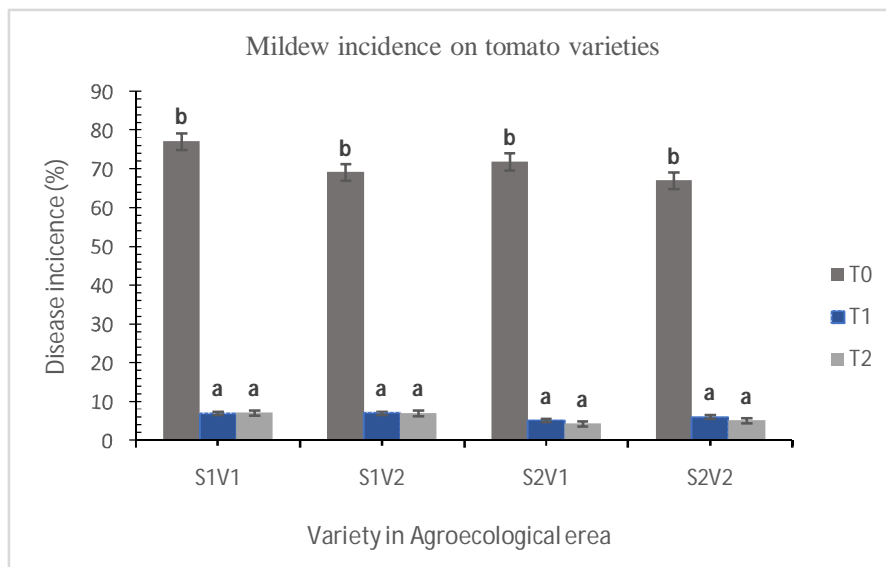
**Fig.1.** Macroscopic and microscopic characteristics of pure strains of *Phytophthora infestans* obtained in different agro-ecological areas. A- Pure strain. B- Sporocysts under light microscope; C- Zoospores under photonic microscope (magnification X 100).



**Fig.2.** Macroscopic and microscopic characteristics of pure strains of *Alternaria* spp. obtained in different agro-ecological areas. A- Pure strain. B- Sporocysts seen under a photonic microscope.

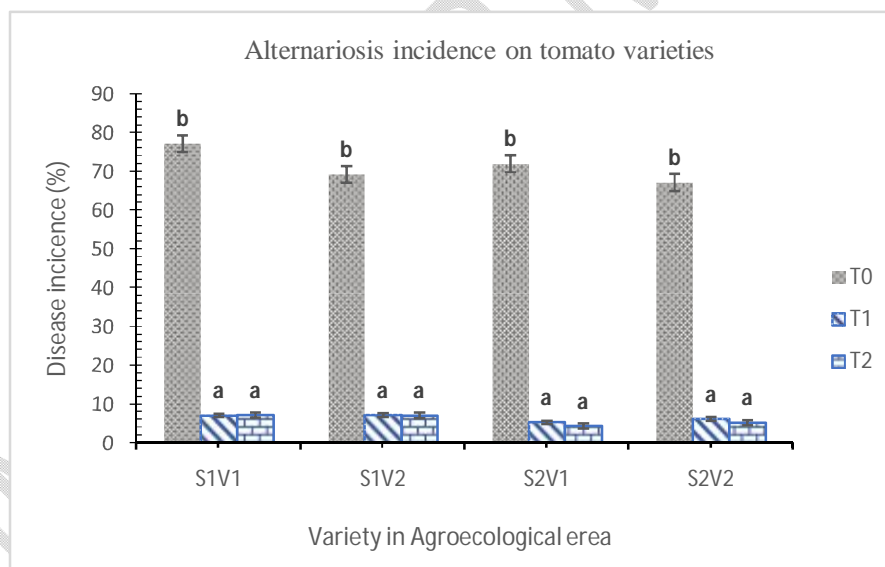
### 3.3 Disease average incidence and severity

Surveys carried out in agro-ecological areas III and V, in the localities of Njichom and Mbele II respectively, showed that the incidence and severity of mildew and alternariosis were fairly high in the two pepper varieties studied. All the plots visited in the different localities showed typical symptoms of both diseases. Figures 3 & 4 show respectively the evolution of the average incidence of mildew and alternariosis in the different localities. Incidences of mildew ranged from 12.72% to 53% for variety V1 (*Safi*) and 10.01% to 49% for variety V2 (*Burkinayellow*) in agro-ecological area III. Similarly, 8.99% to 60.9% for variety V1 and 7.12% to 55% for variety V2 in agro-ecological area V between 4 and 12 weeks. *Alternaria* incidence ranged from 7.12% to 77.12% for variety V1 (*Safi*) and 7.01% to 69.15% for variety V2 (*Burkina yellow*) in agro-ecological area III. Also 4.32% to 71.9% for variety V1 and 5.12% to 67.08% for variety V2 in agro-ecological area V between 4 and 12 weeks. Nevertheless, no significant difference ( $P > 0.05$ ) was observed between the incidences of the different localities for the different periods between the 2 pepper varieties for treatments T1 and T2 but present between these treatments and the T0 control in the above-mentioned agro-ecological areas.



\*Treatments with the same letters at the same zone are not significantly different according to Tukey's test  $P < 0.05$

**Figure 3:** Evolution of Mildew incidence on pepper varieties in agroecological areas



\*Treatments with the same letters at the same zone are not significantly different according to Tukey's test  $P < 0.05$

**Figure 4:** Evolution of alternariosis incidence on pepper varieties in agroecological areas

The field application of a biopesticide requires an adequate system to spread and deposit the microbial agents. The choice of correct equipment is therefore essential to ensure that the biopesticide is applied effectively at the correct rate and in the target surface. It

depends, among other things, on the size of the particles entering the compositions of the final products for example.

Temperature and humidity play a decisive role in the development of mildew, particularly during contamination phases: relative humidity above 90% and average temperatures above 16°C enhance the spread of the disease. In addition, the interaction between climate changes and thermal adaptation of *P. infestans* may have profound effects for the future of pepper. The appearance of the disease in untreated plots was therefore due to the maintenance of ambient temperature conditions favorable to the spread of phytopathogens during the growing season. Incidence and severity were very high in these plots, due to the absence of crop rotation, which accentuates the multiplication of pathogenic fungi [6]. Failure by some growers to comply with certain prophylactic measures, such as crop rotation, to limit primary inoculum [32]. The spatial importance of the crop could also be a factor influencing the importance of the disease in the plots, according to the work of Hammi [34] in the Sais region when he was working on the characterization of *P. infestans* populations. Disease incidence in the two agro-ecological areas ranged from around 49% to 71% for the two pepper varieties with greater susceptibility in zone V. Disease heterogeneity on treated plots is rather the subject of disparities in terms of field spraying frequencies. Based on the results reported here, it could be deduced that virulence of phytopathogens depends on the area where it's been collected. We found that the highly aggressive strains and the most complex races were detected in agroecological area V while weakly aggressive and less complex races were identified in area III.

The disease cycle of alternaria leaf blight is similar to that of other foliar alternaria diseases. Sources of inoculum include mycelium in overwintering pepper debris, diseased volunteer or wild peppers, and infected seed. In warm temperate areas, overlapping production seasons allow conidia produced in more mature fields to serve as inoculum for nearby more recently planted fields [7]. Conidiophores and conidia are produced at temperatures from 8 to 28°C and in the presence of 96 to 100% relative humidity or free water on the plant surface. Conidia are released and become airborne as the relative humidity drops soon after daylight and winds are above 2 to 3 m/s [5]. In a study of spore dispersal patterns in Ontario, the number of airborne spores detected displayed a circadian periodicity with peak dispersal from 1100 to 1400 h [8]. Spore dispersal increased with decreasing of relative humidity, transition from wet to dry leaves, and increases in temperature and wind speed. Spore dispersal patterns can then be interrupted by prolonged high winds, long periods

of leaf wetness, and cool temperatures [5]. Conidia deposited on leaf surfaces will germinate and infect under appropriate temperature and leaf wetness conditions. At 24°C, damage generally increases with increasing hours of leaf wetness from 8 to 56 h (for example). In addition, infection rate was markedly reduced at night temperatures below 12°C [8]. Pryor *et al.*[35] reported disease incidence of 63 to 99% on mature plants in a survey of eight fields in the Cuyama Valley of California. In some of the California cases, entire fields were abandoned before harvest due to excessive petiole decay. Estimates of disease incidence from storage houses and markets also reveal the prevalence of black rot.

### **3.4 pathogenicity test**

The pathogenicity test of both phytopathogens showed the appearance of specific symptoms similar to those of mildew and alternariosis. Healthy young pepper leaves infected with inocula of both *P. infestans* and *Alternaria spp.* showed symptoms identical to those observed on infected organs in the field, characterized by mycelial growth; necrotic lesions of mildew, and brown spots; sunken lesions of alternariosis on the leaves, among others, similar to those confirmed by Qin *et al.*[8] and Rekadet *al.* [29]

### **3.5 Phytochemical screening**

Our present laboratory study revealed an overexpression of secondary metabolites in pepper leaves from the treated plot (Table 1), which are specific to treatment T1 compared with the control plot (Table 2). It also demonstrated that the treatments containing fungal conidia (*M. anisopliae* and *T. harzianum*) were effective for the control of *Phytophthora* and *Alternaria ssp.* In general, treatment with *T. harzianum* and *M. anisopliae* resulted in the reduction of plant diseases. Generally, a mixture of plant secondary metabolite with mycopathogen provides eco-friendly stable pest management system compared to environment-deteriorating chemical pesticides. However, the first step to develop a compatible mixture mainly depends on the interaction of these products that might lead to compatible or toxic interaction[36]. It is well-known that both the fungi and plant secondary metabolites demonstrated an entirely different mode of actions to overcome the target host defense mechanisms to cause mortality [37]. Fungal infection evades the host defense mechanism by triggering complex biochemical interactions in the form of series of events through cuticle adhesion, penetration, proliferation, and toxin production, which ultimately lead to the host mortality. On the other hand, plant secondary metabolites are known antagonists that act by interfering with the signaling of the nervous and cellular systems to

overcome the target host defense mechanism [38]. We speculate that secondary metabolites produced by T1 bioformulation like tetradecanoic and/or hexadecenoic acids and biological control effect on these two fungal infections. They also have an inhibitory effect on *Colletotrichum lagenarum*, agent of anthracnose of melon (*Cucumis melo*), and cucumber (*Cucumis sativus*) [39]. Benhamou & Picard [40] reported a positive correlation between overall plant response and changes in the biochemistry and physiology of plant cells pre-inoculated by *Trichoderma* spp., which were accompanied by structural modifications. Barakat *et al.* [41] reported that *T. harzianum* releases several volatile compounds such as benzene-ethanol, butanoic acid, propanoic acid, palmitoyl chloride, glycerol-1- palmitate and hexadecanoic acid. They have many functions in biological control (butanoic acid, propanoic acid, tetradecanoic acid, benzotriazepine) and as phytoalexins (benzoic acid; scoparone; scopoletin). The combination of both beneficial microorganisms revealed biopesticides compounds as Tetrachloroisophthalonitrile zinc ethylenebis(dithiocarbamate), Methyl-2, Benzimidazolecarbomate. Abid & Ahmed [37] reported that *M. anisopliae* can also release some mycotoxins as 1-chlorooctadecane, which is an acaricide. They have a significant inhibitory effect on the development of Chemical insecticide was less faster than the fungal treatments in reducing infection activity, but less lasting with no significant difference. These differences could be due to the mechanism of action of the different treatments evaluated. *M. anisopliae* could spread in the ant population, whereas *T. harzianum* causes a food shortage by destroying the symbiotic fungus. In contrast, the chemical insecticide killed large number of ants but did not affect the entire population nor did it affect the symbiotic fungus. Systemic resistance to pathogens is therefore induced by these metabolites, which develop allelopathic mechanisms in the plant [42]. It also happens that during the development of systemic resistance induced by the PGPMs of the T1 formulation while T2 (Mandipropamid) effectively exerts chemical control. The cyclohexylamine produced by the plant under the effect of T1 bioformulation is an analogue of mandipropamid.

Table 1. Specific molecules from the leaves of the two pepper varieties treated with T1 bioformulation.

Hypothetical specific molecules to T1	Retention time	Hypothetical formula	Molecular weight	Abundance (%)
<b>7-Hydroxy-3-(1,1-dimethylprop-2-enyl) coumarin</b>	5,51	$C_{14}H_{14}O_3$	230,26	100
<b>5-Chloro-2-pyridinol</b>	6,3	$C_5H_4ClNO$	129,54	79,08
<b>Tetrachloroisophthalonitrate zinc ethylenebis(dithiocarbamate)</b>	3,98	$C_4H_6N_2S_4Zn$	256,4	210,4
<b>Octadécanoïqueacid</b>	4,50	$C_{18}H_{36}O_2$	284,47	61,6
<b>Cyclohexylamine, N-(2-chlorocyclopentylidene)-, N-oxide</b>	7,14	$C_{12}H_{13}N_3O_2$	56,10	56,14
<b>Glycerol 1-palmitate</b>	5,64	$C_{19}H_{38}O_4$	330,50	100
<b>1-Chlorooctadecane</b>	6,99	$C_{18}H_{37}Cl$	279,29	98,74
<b>Methyl-2, Benzimidazolecarbomate</b>	6,55	$C_8H_8N_2$	298,9	191,19
<b>Palmitoyl chloride</b>	5,64	$C_{16}H_{31}ClO$	274,9	79,39
<b>1H-Imidazole-2-methanol</b>	4,97	$C_{19}H_{23}NO$	98,1	75,14
<b>Tetradecanoicacid</b>	3,99	$C_{14}H_{28}O_2$	228,37	95,76

from the leaves of the two pepper varieties with T0.

Hypothetical specific molecules to T1	Retention time	Hypothetical formula	Molecularweight	Abundance (%)
<b>1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester</b>	3,90	$C_{16}H_{24}O_4$	278,3	76,22
<b>Acide Phthalique, 8-chlorooctyl isobutyl ester</b>	3,78	$C_{20}H_{29}ClO_4$	230,3	14,14
<b>AcidePhthalique, hexylpropyl ester</b>	5,84	$C_{22}H_{34}O_4$	362,5	31,12

T

able  
au  
2.  
Spe  
cific  
mol  
ecul  
es

<b>Diisopropyladipate</b>	6,30	$C_{12}H_{22}O_4$	230,3	65,27
<b>N-(2-chlorocyclopentylidene) -, N-oxide</b>	7,14	$C_{18}H_{32}O_2$	280,4	53,40

---

UNDER PEER REVIEW

#### 4. CONCLUSION

This study aimed at assessing the phytoprotection effect of bioformulation with *Trichoderma harzianum* and *Metharizium anisopliae* on diseases caused by *Alternaria* spp and *Phytophthora* spp. The results showed that this formulation effectively reduced diseases on the two pepper varieties in both agroecological zones. This bioformulation significantly influences metabolite production in the pepper plants which had biological control effects. These metabolites stimulate the plant's defense mechanisms against agents causing pepper diseases such as Mildew and Alternaria leaf blight. This formulation could thus be useful for plants phytoprotection.

#### DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that generative AI technologies such as Large Language Models, etc have been used during writing or editing of manuscripts. This explanation will include the name, version, model, and source of the generative AI technology and as well as all input prompts provided to the generative AI technology.

#### REFERENCES

1. Ministry of Agriculture and Rural Development. Agricultural policies around the world: some examples. *Service Europe et International*. 2023; N° 03, 1p.
2. Shahid M., Zeyad M.T., Syed A., Singh U.B., Mohamed A., Bahkali A.H., Elgorban A.M., Pichtel J. Stress-Tolerant Endophytic Isolate *Priestiaaryabhattai* BPR-9 Modulates Physio-Biochemical Mechanisms in Wheat (*Triticum aestivum* L.) for Enhanced Salt Tolerance. *International Journal of Environmental Research and Public Health*. 2022; 19, 10883.
3. Martins P. M. M., Merfa M. V., Marco A., Takita M.A., & Alessandra A. Persistence in Phytopathogenic Bacteria: Do We Know Enough. Weihui Wu, Nankai University, *China*. 2018; p 14.
4. Blancard D., LATERROT H., MARCHOUX G. & CANDRESSE, T. A colour Handbook - Tomato Diseases: identification, biology and control. *Manson Publishing Ltd*. 2012; 688p.
5. Fritz M., Jakobsen I., Lyngkjær M.F., Thordal-Christensen H., Pons-Kühnemann J. Arbuscular mycorrhiza reduces susceptibility of tomato to *Alternaria solani*. *Mycorrhiza*. 2006; 16, 413–419.
6. Garrido C., Fernandez - Acero Francisco J., Carbu M., Gonzalez -Rodriguez Victoria E., Lineiro Eva et Cantoral Jesus M. "Molecular Microbiology Applied to the Study of

Phytopathogenic Fungi”, dans Sameh Maghdeldin, Gel Electrophoresis- *Advanced Techniques*.2012; pp 139-156.

7. Logrieco A., Moretti A. & Solfrizzo M. *Alternaria* toxins and plant diseases: an overview of origins, occurrence and risks. *World Mycotoxin Journal*. 2009; 2: 129-140
8. Qin Q.; Fan Y.; Jia Q.; Duan S.; Liu F.; Jia B.; Wang G.; Guo W.; Wang C. The Potential of *Alternaria* Toxins Production by *A. alternata* in Processing Tomatoes. *Toxins*. 2022; 14, 827.
9. Shi X., Tian Z., Liu J., van der Vossen EAG., Xie C. A potato pathogenesis-related protein gene, StPRp27, contributes to race-nonspecific resistance against *Phytophthora infestans*. *Mol Biol Rep*. 2012; 39: 1909–1916
10. Samada LH, Tambunan USF. Biopesticides as promising alternatives to chemical pesticides: A review of their current and future status. *OnLine Journal of Biological Sciences*. 2020;20: 66–76.
11. Mukesh, M.; Andleeb, Z.; Dubey, M.K.; Mohd, A.; Gupta, V.K.; Upadhyay, R.S. Comparative evaluation of biochemical changes in tomato (*Lycopersicon esculentum* mill.) infected by *Alternaria alternata* and its toxic metabolites (TeA, AOH, and AME). *Front. Plant Sci*. 2016; 7, 1408.
12. Deguine JP. 2023. Lutte biologique et biocontrôle : un besoin de clarification. *Cahiers Agricultures* 32: 11.
13. Adeyemi N.O., Atayese M.O., Olubode A.A. & Akan M.E. Effect of commercial arbuscular mycorrhizal fungi inoculant on growth and yield of soybean under controlled and natural field conditions. *J. Plant Nutr*. 2019; 16: 85-101.
14. Guillemette T, Bastide F. Trichoderma mushrooms, benefactors for our society. The Conversation. <https://thecon.com/les-champignons-trichoderma-des-bienfaiteurs-pour-notre-societe-184993>. N. Soulé Adam : 2022. *Cah. Agric*. 2024, 33, 5
15. Endamana D. & Sonwa D. Augmenter la productivité en milieu rural à forte pression foncière de la zone de forêt du Sud Cameroun. *Rapport des activités de recherche de Nkometou III*. 1998 ; 34p.
16. Samuels G.J., 2006. *Trichoderma*: systematics, the sexual state and ecology. *Phytopathology*, 96 : 195-206.
17. Olivera D., Ayala J., Calero A., Santana M., Hernandez A. Pratiques agroécologiques dans la province de Santi Spiritus, Cuba. *Les microorganismes efficaces, une technologie appropriée sur les bases agroécologiques*. 2014; 17p.
18. Manika S., Swati D., Dinesh SB., Chowdappa P., Selvamani R. and Pratibha S. Morphological, cultural, pathogenic and molecular studies of *Alternaria brassicae* infecting cauliflower and mustard in India. *African J. of Microb. Res*. 2013; 7: 3351-3363.
19. Brasier CM, Cooke DEL, Duncan JM, Hansen EM. Multiple new phenotypic taxa from trees and riparian ecosystems in *Phytophthora ganapodyides*-*P. megasperma* ITS Clade

- 6, which tend to be high-temperature tolerant and either inbreeding or sterile. *Mycol Res.* 2003;107 :277–290.
20. Tchumakov A. E. & Zaharova I. I. Influence and statistics of disease development. *In: Agronome*(eds). Disease damage to agricultural crops. *Moscou.* 1990. P 5-60.
21. Abdessemed N. Champignons A Potentiel Mycoherbicide (Doctoral Dissertation) ; 2021.
22. Oomah BD, Caspar F, Malcolmson LJ, Bellido AS. Phenolics and antioxidant activity of lentil and peahulls. *Food Research International.* 2010; 44:436-441.
23. Kondjoyan N, Berdagué JL. A Compilation of relative retention indices for the analysis of aromatic compounds, Edition du Laboratoire Flaveur, Theix. NIST *Webbook Website.* 1996;18/06/2013. Available: <http://webbook.nist.gov/chemistry/les>, R. B., and Wicks, T. J. 2003. The incidence of *Alternaria radicina* on carrot seeds, seedlings and roots in South Australia. *Australas. Plant Pathol.* 32:99-104.
24. Singh A., Srivastava S. & Singh H.B. Effect of substrates on growth and shelf life of *Trichoderma harzianum* and its use in biocontrol of diseases. *Bioresource Technology.* 2018 ;98 : 470-473.
25. Beninal L. Diversité génétique de *Phytophthora infestans* agent du mildiou de la pomme de terre en Algérie. Thèse de Magister. E.N.S.A. El Harrach. 2011; 92p.
26. Gallegly M.C., Hong C. *Phytophthora: Identifying Species by Morphology and DNA Fingerprints.* APS Press –The *American Phytopathological Society*, St. Paul, Minnesota. 2008; 158 p.
27. Coulibaly K., Kebe B. I, Koffi K. N., Mpika J., Kone D. Caractérisation des isolats de *Phytophthora* spp du verger cacaoyers de Cote d’Ivoire. *Journal of Applied Biosciences.* 2013; 70: 5567– 5579.
28. Runno-Paurson E., Kiiker R., Joutsjoki T., Hannukkala A. High genotypic diversity found among population of *Phytophthora infestans* collected in Estonia. *Fungal Biology.* 2016;120: 385–392. Doi 10.1016/j.funbio.2015.11.008.
29. Rekad Z. F. Phenotypic and genotypic characterization of *P.infestans*(Mont) de Barry causal agent of potato and tomato mildew in the northwest region of Algeria. Thesis, Algeria, University of *Abdelhamid IBN Badis.* 2018; 136.
30. Simmons EG. *Alternaria* themes and variations (244–286) species on Solanaceae. *Mycotaxon.* 2000;75 :1–115.
31. Rotem J. The genus *Alternaria* biology, epidemiology, and pathogenicity. APS Press, St. Paul, Minnesota, USA.1994; 48 :211–215.
32. Montarry J. Adaptive response of populations of *Phytophthora infestans*, agent of potato mildew, to crop deployment of its host *Solanum tuberosum.* *PhD thesis.* 2007; 192 p.

33. Park, M.S., Romanoski, C.E. & Pryor, B.M. A re-examination of the phylogenetic relationship between the causal agents of carrot black rot, *Alternaria radicina* and *A. carotiinculatae*. *Mycologia*. 2008; 100 (3), 511-527.
34. Hammi A. Characterization of *phytophthora infestans* (mont.) De bary populations in the saïs region. National doctoral thesis in biology. Specialization: *phytopathologie*. 2003; 272 p.
35. Pryor, B. M., Davis, R. M., and Gilbertson, R. L. Detection of soilborne *Alternaria radicina* and its occurrence in California carrot fields. *Plant Dis*. 1998; 82 :891-895.
36. Gonçalves Diniz A.; Barbosa L.F.S.; da Silva Santos A.C.; De Oliveira N.T.; da Costa A.F.; Carneiro-Leão M.P.; Tiago P.V. Bio-insecticide effect of isolates of *Fusarium caatingaense* (Sordariomycetes: Hypocreales) combined to botanical extracts against *Dactylopiusopuntiae* (Hemiptera: Dactylopiidae). *Biocontrol Sci. Technol*. 2020; 30, 1–12.
37. Abid H. & Ahmed M.A. Potential Synergy between Spores of *Metarhizium anisopliae* and Plant Secondary Metabolite, 1-Chlorooctadecane for Effective Natural Acaricide Development. *Molecules*. 2020; 25: 1900; doi :10.3390/molecules2508190.
38. Wink M. Plant secondary metabolites modulate insect behavior-steps toward addiction? *Front. Physiol*. 2018 ; 9, 364.
39. Benzohra I.E., Bendahmane B.S. & Youcef B.M. Essais de lutte biologique *in vitro* par l'utilisation de *Trichoderma harzianum* et de *Trichoderma viride* contre *Ascochyta rabiei*, agent de l'antracnose du pois chiche en Algérie. *Journal Algérien des Régions Arides*. 2016; 13: 51-60.
40. Benhamou N., & Picard K. Induced resistance: a new plant defense strategy against pathogens. *Phytoprotection*. 2000;80: 137-168.
41. Barakat I., Chtaina N. EL, Guilli M. & Ezzahiri B. Efficacy of *Bacillus amyloliquefaciens* and *Trichoderma harzianum* in the biocontrol of wheat septoria by seed coating. *Revue Marocaine de Protection des Plantes*. 2019; 13: 9-18.
42. Dantas T.L., Alonso Buriti F.C., Florentino E.R. Okra (*Abelmoschus esculentus* L.) as a Potential Functional Food Source of Mucilage and Bioactive Compounds with Technological Applications and Health Benefits. *Plants*. 2021; 10 :1683.