

Plant Growth Promoting Rhizobacteria Bio capsules: Enhancing Ginger Growth and Yield

Abstract

The study aimed to assess the impact of plant growth promoting rhizobacteria capsules on the growth and yield of ginger. PGPRs known for their role in nitrogen fixation, phosphate solubilization, and hormone secretion. These PGPR's tested at the Turmeric Research Station, Kammarpally across three consecutive years (2018-2021) in combination with three ginger varieties (Mahima, Maran, Local type) and five different treatments T₁-POP + *Trichoderma* (Talc formulations) + GRB 35 (Talc formulations), T₂- POP + *Trichoderma* capsules + GRB 35 capsule, T₃- POP + *Trichoderma* capsules, T₄- POP + GRB 35 capsule, T₅- POP (Recommended package of practices). The treatment T₄V₁ (POP + GRB 35 capsule with Mahima variety), produced the highest fresh rhizome yield with 6.28 kg plot⁻¹ and 21.53 t ha⁻¹ followed by T₄V₂ (POP + GRB 35 capsule with Maran variety) at 6.16 kg plot⁻¹ and 20.15 t ha⁻¹. These results suggest that GRB 35 capsules significantly enhance ginger yield, offering a sustainable approach to ginger cultivation.

Key words: Growth promoting rhizobacteria; bio capsules; ginger; yield enhancement

Introduction

Ginger (*Zingiber officinale* Rosc.) belongs to the Zingiberaceae family, is a perennial herbaceous monocot cultivated commercially for its rhizomes. It is traditionally grown as an annual crop in tropical and subtropical climates where, it thrives best under humid conditions.. Ginger has been grown for over 5000 years, with its origin in South East Asia. Although it is now cultivated in various regions, including Africa. Major ginger producing countries include Nigeria, which accounts for 56.23% of the world's production area, followed by Bangladesh, India, China, and Indonesia, which contribute approximately 23.6, 4.7, and 3.4 per cent of the total area of ginger cultivation, respectively. In India, ginger is widely used in pickles, food additives, confections, and traditional remedies. Its medicinal properties, such as antioxidant effects and the ability to alleviate nausea from various causes, including motion sickness, pregnancy, surgery, and chemotherapy make it a valuable ingredient in both traditional and modern medicine.

The importance of sustainable agriculture has grown in recent years due to its focus on long-term environmental and social benefits. Sustainable farming practices are crucial for reducing reliance on artificial pesticides and fertilizers, improving plant health, and maintaining soil quality (De Andrade *et al*, 2023). Preserving soil diversity and employing eco-friendly practices are critical for ensuring long-term environmental health and food security (Kumar, 2017). Plant growth promoting rhizobacteria (PGPR) represent a key component of sustainable agriculture. These beneficial microorganisms enhance plant growth through various mechanisms including nitrogen fixation, phosphorous solubilization, and production of phytohormones such as auxins, gibberellins, and cytokinins (Pantigoso *et al*, 2019). Researchers have thoroughly studied the positive impacts of PGPR, a naturally occurring soil bacterium, on plant vitality and output. In addition to protecting plants from

pathogens and harsh conditions, they can also boost nutrient availability, spur plant growth, fortify root development. Several different mechanisms are involved in how PGPR helps plants. Some PGPR form auxins, cytokinins, and gibberellins, which encourage root and shoot growth (VanPeer *et al.* 2018). PGPR also improve soil health by competing with harmful microbes, thereby reducing pathogen invasion and enhancing plant resistance to environmental factors like drought, salinity, and extreme temperatures.

The rhizosphere of a plant is home to numerous PGPR that play a crucial role in plant development. They contribute to increased tolerance to biotic and abiotic stress, reduced need for chemical inputs and enhanced soil fertility, and nutrient uptake. Additionally PGPR can aid in soil bioremediation and enhance crop yield, making them a valuable tool for promoting sustainable agricultural practices. This study aims to explore the role of PGPR bio-capsules in enhancing ginger growth and yield, highlighting their potential benefits and contributions to sustainable agriculture. PGPRs serve as the most major source of biofertilizer strains, enhancing soil quality and promoting sustainable agriculture with reduced reliance on fertilizers or pesticides. (Dandan Wang *et al.* 2023). Promoting soil health through sustainable agricultural practices is imperative for ensuring food security and environmental sustainability. Adoption of conservation tillage techniques, crop rotation and diversification play a pivotal role in enhancing nutrient cycling, suppressing pests, and mitigating diseases. Precision agriculture technologies aid in optimizing resource utilization, minimizing inputs, and mitigating environmental impacts. (Indira *et al.* 2023).

Material and methods

The field experiment was conducted to study the effect of different PGPR biocapsules and talc formulations on growth and yield of ginger at Turmeric research station, Kammarpally, Nizamabad district, Telangana during 3 consecutive years from 2018-19 to 2020-21. The initial soil conditions included a slightly alkaline pH of (7.65), electric conductivity 0.15 dS m^{-1} , low organic carbon with medium available nitrogen (250 kg ha^{-1}), high available phosphorus (32.57 kg ha^{-1}) and high available potassium (332.7 kg ha^{-1}).

This experiment was laid in a randomized block design with three replications. This experiment is formulated with three varieties V₁-Mahima, V₂-Maran, V₃-Local type and five different PGPR combinations T₁-POP + *Trichoderma* (Talc formulations) + GRB 35 (Talc formulations), T₂- POP + *Trichoderma* capsules + GRB 35 capsule, T₃- POP + *Trichoderma* capsules, T₄- POP + GRB 35 capsule, T₅- POP (Recommended package of practices). Generally, the seed was sown in the month of June and harvested after completion of 8 months in the month of March during the experimental years.

Recommended cultural practices were adopted for all treatments. Growth parameters *viz.*, plant height, number of tillers, number of leaves, petiole length, leaf length, leaf width recorded in the second week of January month. Yield parameters data was recorded at harvesting time. In case of growth and yield parameters data was recorded from five plants from each replication and the means are used for statistical analysis (Panse and Sukhatme 1957)

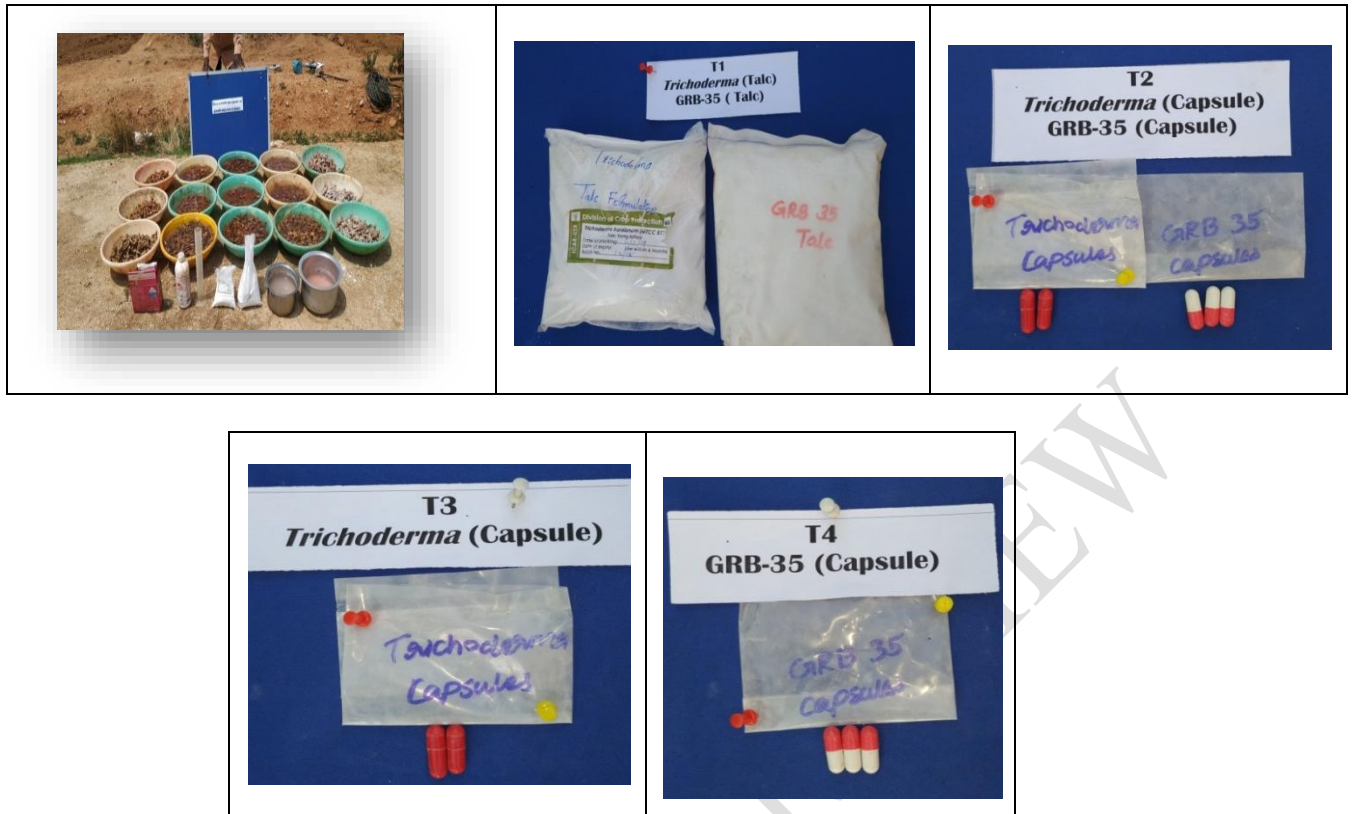


Fig 1: Treatments used for Randomized block design with three replications

Results and discussion

Growth characters:

“Significant differences were observed with the growth parameters plant height, number of shoots, height of shoot, leaf length, leaf width. The maximum mean plant height (56.0 cm), number of shoots (12.95), height of shoot (24.75), leaf length (20.72 cm), leaf width (1.79 cm) were recorded with T₄- POP + GRB 35 capsule with Mahima variety which may be due to the direct mechanisms observed in PGPR include N₂-fixation, mobilization of nutrients via production of phosphatases, siderophores, or organic acids, and production of phytohormones and enzymes which trigger the growth of the turmeric plants” (V. Jeyanthi *et al.*, 2018). “Many scientists reported that plant growth promoting rhizobacteria might enhance plant height and productivity by synthesizing phyto-hormones” (Prokryl *et al.*, 2000, Burd *et al.*, 2000, Gravel, 2007). “The beneficial effects of PGPR involve boosting key physiological processes, including water and nutrient uptake, photosynthesis, and source-sink relationships that promote growth and development” (Illangumaran and Smith, 2017). “One of the mechanisms by which bacteria are adsorbed onto soil particles is by ion exchange. A soil is said to be naturally fertile when the soil organisms are releasing inorganic nutrients from the organic reserves at a rate sufficient to sustain rapid plant growth” (Goswami *et al.*, 2016). Gray and Smith (2005) have shown that “the PGPR associations range in the degree of

bacterial proximity to the root and intimacy of association”. “The three distinct characteristics of PGPR are they must be able to colonize the root, they must survive and multiply in microhabitats associated with the root surface, in competition with other microbiota, at least for the time needed to express their plant promotion/ protection activities and they must promote plant growth” (Kloepper, 1994; Lucy et al., 2004). “Phytohormones are responsible for plant growth development and allow plants to tolerate different stress conditions” (Shaterian *et al.*, 2005). “Some rhizobacteria are able to produce phytohormones, including cytokinins, auxins, gibberellins, ethylene, and abscisic acid (ABA), which play a role in different growth processes in plants, including cell multiplication, which results in increased cell and root expansion” (Glick, 2014; Kaur et al., 2016). “However, the production of ABA by rhizobacteria is considered an indirect way of promoting plant growth. Several bacteria that have the ability to produce IAA and have positive effects on shoot and root weight and nutrient uptake on maize plants. Besides, activities like phosphorus solubilization, or even other non-evaluated PGPR traits that stimulate plant growth. PGPR may promote growth directly, e.g. through fixation of atmospheric nitrogen, solubilization of minerals (phosphorus and potassium), production of siderophores that solubilize and sequester iron, or production of plant growth regulating hormones” (Grover et al. 2009). “An understanding of ecological conditions effecting on bacterial inoculants is important when introducing microbes for increasing plant growth and productivity. The bacteria strains *Pseudomonas alcaligenes* PsA15, *Bacillus polymyxa* BcP26 and *Mycobacterium phlei* MbP18 had a much better stimulatory effect on plant growth and nitrogen (N), phosphorus (P) and potassium (K) uptake of maize in nutrient deficient calcisol soil. Their stimulatory efficiency reduced in relatively rich loamy sand soil where bacterial inoculants stimulated only root growth and N, K uptake of root”.(D. Egamberdieva, 2007).

Yield characters:

The maximum mean fresh rhizome yield per plot (6.28 kg/plot) and maximum mean fresh rhizome yield per hectare (21.53 t/ha) have been observed with T4V1- POP + GRB 35 capsule with Mahima variety followed by T4V2- POP + GRB 35 capsule with Maran variety which recorded fresh rhizome yield per plot (6.16 kg/plot) and mean fresh rhizome yield per hectare (20.15 t/ha). The results are in conformity with Kuan et al. (2016) who reported that plant growth-promoting bacteria may provide a biological alternative to fix atmospheric N₂ and delay N remobilization in maize plant to increase crop yield based on an understanding that plant-N remobilization is directly correlated to its plant senescence promoting high up to 30.9% with reduced fertilizer-N input. Di Salvo et al. (2018) reported that “PGPR used as inoculants of cereal crops including maize can improve their growth and grain yield. The crops responses to inoculation are complex because are defined by plant-microorganisms interactions, many of them still unknown”. “Thus, it is necessary to improve the knowledge about the microbial ecology of the rhizosphere of crops under different agricultural practices. Various processes, such as the mineralization of organic matter, nutrient immobilization, phosphate solubilization, nitrogen nitrification, and phytohormone production, combine to enhance soil fertility and crop productivity” (Van Peer *et al* 1989). “Plant growth promoting rhizobacteria in rhizosphere soil is highly dynamic, more versatile in transforming,

mobilizing and solubilising the nutrients. Therefore, the rhizobacteria are the dominant deriving forces in recycling the soil nutrients and consequently, they are crucial for soil fertility. They may be extensively used in plant growth promotion as it acts as a plant nourishment and enrichment source which would replenish the nutrient cycle between the soil and plant roots , exhibits detoxifying potential, controls phytopathogen thereby exerts a positive influence on crop productivity and ecosystem functioning, hence can be implemented in agriculture” (V. Jeyanthi, 2018)

Conclusion

The application of GRB 35 capsules, combined with recommended practices, significantly improves ginger growth and rhizome yield. Among the ginger varieties tested, ‘Mahima’ exhibited the highest growth and yield, followed by ‘Maran’. In order to increase yields and improve crop performance overall, it is suggested to use and incorporate GRB 35 capsules into ginger cultivation practices.

Future scope:

Further field research and testing must be conducted to completely discover the potential of PGPR and develop feasible, widespread applications. Eco-friendly PGPR approaches that significantly enhance plant growth and increase crop yields are now achievable due to these improvements.

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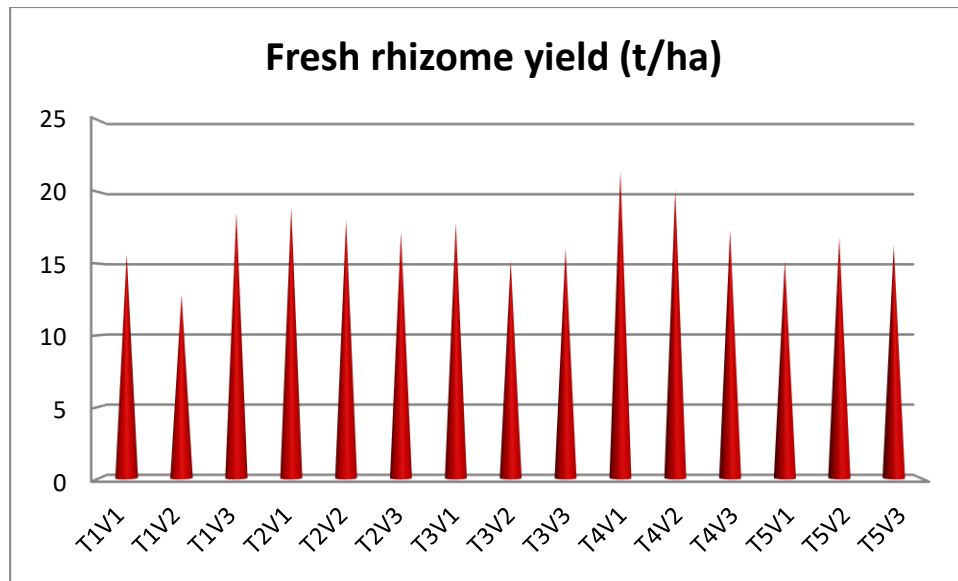


Fig 2: Effect of plant growth promoting rhizobacteria bio capsules on growth and yield characteristics of ginger

UNDER PEER REVIEW

Table 1: Effect of plant growth promoting rhizobacteria bio capsules on growth and yield characteristics of ginger

	Plant height (cm)			Number of shoots			Height of shoot (cm)			Leaf length (cm)		
	2019-20	2020-21	Pooled mean	2019-20	2020-21	Pooled mean	2019-20	2020-21	Pooled mean	2019-20	2020-21	Pooled mean
T₁V₁	40.85	40.5	40.675	10.32	10.36	10.34	21.17	20.8	20.985	15.65	15.4	15.525
T₁V₂	53.65	50.8	52.225	8.65	8.69	8.67	20.2	19.2	19.7	15.95	16.3	16.125
T₁V₃	44.52	43.8	44.16	9.54	10.21	9.875	22.22	21.4	21.81	15.9	17.2	16.55
T₂V₁	44.05	43.0	43.525	10.36	9.65	10.005	15.42	16.4	15.91	17.52	18.4	17.96
T₂V₂	42.52	42.6	42.56	10.23	11.23	10.73	16.87	17.4	17.135	16.3	16.6	16.45
T₂V₃	37.17	38.5	37.835	7.69	8.54	8.115	20	20.1	20.05	14.21	16.34	15.275
T₃V₁	35.25	36.0	35.625	8.54	7.36	7.95	18.92	20.7	19.81	15.5	15.4	15.45
T₃V₂	46.3	45.4	45.85	9.63	10.25	9.94	18.45	20.7	19.575	14.27	15.1	14.685
T₃V₃	45.27	45.3	45.285	11.21	10.45	10.83	20.57	1.4	10.985	16.52	15.21	15.865
T₄V₁	55.23	56.78	56.005	13.21	12.69	12.95	25.36	24.15	24.75	20.12	21.33	20.725
T₄V₂	54.36	53.96	54.16	12.56	12.36	12.46	23.31	24.21	23.76	19.36	20.45	19.905
T₄V₃	45.65	49.0	47.325	10.36	9.21	9.785	22	21.6	21.8	16.75	18.1	17.425
T₅V₁	49.07	48.3	48.685	8.52	7.63	8.075	19.8	21.3	20.55	12.31	13.45	12.88
T₅V₂	44.45	35.2	39.825	7.65	8.12	7.885	21.25	20.1	20.675	16.3	16.4	16.35
T₅V₃	41.5	42.7	42.1	9.21	10.36	9.785	16.97	16.5	16.735	14.55	16.4	15.475
SE(m)	1.251	1.070	1.128	0.538	0.636	0.563	0.566	0.525	0.542	0.556	0.548	0.521
CD (5%)	3.584	3.065	3.345	1.540	1.823	1.634	1.622	1.505	1.534	1.593	1.569	1.563
CV %	5.682	4.927	5.178	6.175	7.282	7.165	5.851	5.434	5.634	6.634	6.388	6.513

T₁-POP + *Trichoderma* (Talc formulations) + GRB 35 (Talc formulations), **T₂**- POP + *Trichoderma* capsules + GRB 35 capsule, **T₃**- POP + *Trichoderma* capsules, **T₄**- POP + GRB 35 capsule, **T₅**- POP (Package of Parctice)

Table 2: Effect of plant growth promoting rhizobacteria bio capsules on growth and yield characteristics of ginger

	Leaf width (cm)			Leaf petiole length (cm)			Fresh rhizome yield kg plot ⁻¹			Fresh rhizome yield (t ha ⁻¹)		
	2019-20	2020-21	Pooled mean	2019-20	2020-21	Pooled mean	2019-20	2020-21	Pooled mean	2019-20	2020-21	Pooled mean
T₁V₁	1.72	1.55	1.63	1.7	1.6	1.65	4.8	4.6	4.7	16.0	15.33	15.66
T₁V₂	1.35	1.45	1.4	1.65	1.7	1.675	3.8	3.91	3.855	12.66	13.03	12.85
T₁V₃	1.65	1.65	1.65	1.77	1.7	1.735	5.6	5.56	5.58	18.66	18.53	18.6
T₂V₁	1.5	1.65	1.57	1.55	1.7	1.625	5.7	5.67	5.685	19.0	18.9	18.95
T₂V₂	1.6	1.55	1.57	1.8	1.9	1.85	5.5	5.34	5.42	18.33	17.8	18.06
T₂V₃	1.5	1.6	1.55	1.6	1.8	1.7	5.2	5.12	5.16	17.33	17.06	17.2
T₃V₁	1.65	1.55	1.6	1.8	1.8	1.8	5.4	5.32	5.36	18.0	17.73	17.86
T₃V₂	1.52	1.35	1.43	1.5	1.7	1.6	4.5	4.63	4.565	15.0	15.43	15.21
T₃V₃	1.65	1.65	1.65	1.77	1.6	1.685	4.9	4.75	4.825	16.33	15.83	16.08
T₄V₁	1.82	1.77	1.79	1.77	1.8	1.785	6.2	6.32	6.28	20.66	22.41	21.53
T₄V₂	1.4	1.35	1.37	2.1	2.1	2.1	5.9	6.42	6.16	19.66	20.65	20.15
T₄V₃	1.57	1.58	1.57	1.82	1.8	1.81	5.3	5.12	5.21	17.66	17.06	17.36
T₅V₁	1.67	1.62	1.64	1.75	1.8	1.775	4.6	4.53	4.565	15.33	15.1	15.21
T₅V₂	1.65	1.7	1.67	1.67	1.7	1.685	5.1	5.01	5.055	17.0	16.7	16.85
T₅V₃	1.65	1.65	1.65	1.5	1.6	1.55	4.9	4.87	4.885	16.33	16.23	16.28
SE(m)	0.122	0.126	0.116	0.132	0.090	0.108	0.437	0.340	0.397	1.16	0.906	0.964
CD (5%)	N.S.	N.S.	N.S	N.S.	N.S.	N.S	1.251	0.974	1.145	3.33	2.596	3.174
CV %	15.29	15.991	15.234	15.36	10.094	12.654	13.12	10.016	11.964	13.1	10.016	11.234

T₁-POP + *Trichoderma* (Talc formulations) + GRB 35 (Talc formulations), **T₂**- POP + *Trichoderma* capsules + GRB 35 capsule, **T₃**- POP + *Trichoderma* capsules, **T₄**- POP + GRB 35 capsule, **T₅**- POP (Package of parctice)

References

- Aidin N, Sahiri, I. Madauna. The Effect of Rhizome Types and the Composition of Planting Media on The Growth of Red Ginger (*Zingiber officinale* Rosc.) (Pengaruh Jenis Rimpang dan Komposisi Media Tanam Terhadap Pertumbuhan Bibit Jahe Merah). *J. Agrotekbis*. 2016; 4(4): 394–402.
- Burd GI, Dixon DG and BR Glick. Plant growth promoting rhizobacteria that decrease heavy metal toxicity in plants. *Can.J.Microbiol.* 2000; 33:237-245.
- Dandan W, Chao W, Yinglong C and Zhihong X. Developing plant-growth-promoting rhizobacteria: A Crucial approach for achieving sustainable agriculture. *Agronomy*; 2023, 13(7), 1835; <https://doi.org/10.3390/agronomy13071835>
- De Andrade LA, Santos CHB, Frezarin ET, Sales LR, Rigobelo EC. Plant growth-promoting rhizobacteria for sustainable agricultural production. *Microorganisms* 2023, 11, 1088.
- Egamberdiyeva D. The effect of plant growth promoting bacteria on growth and nutrient uptake of maize in two different soils. *Applied soil ecology*, 2007; Volume: 36, Issue: 2, Pages: 184-189.
- Gohain N, Atibudhi HN, Kumar A. A study on economics of ginger cultivation in Assam—a case study of Tinsukia district. *Plant Archives*. 2020;20(2):2522 2526.
- Glick B R.. The enhancement of plant growth by free living bacteria. *Canadian Journal of Microbiology*, 1995. 41: 109–14.
- Goswami, D, Thakker JN, Dhandhukia PC. Portraying mechanics of plant growth promoting rhizobacteria (PGPR): A review. *Cogent Food Agric.*, 2016; 2:1-19.
- Gravel V, Antoun H and Tweddell RJ. Growth stimulation and fruit yield improvement of greenhouse tomato plants by inoculation with *Pseudomonas putida* or *Trichoderma atroviride*: Possible role of indole acetic acid (IAA). *Soil. Biol. Biochem.* 2007; 39:1968–1977.
- Gray EJ and Smith DL. Intracellular and extracellular PGPR commonalities and distinctions in the plant-bacterium signalling processes. *Soil Biol. Biochem.*, 2005; 37: 395–412.
- Indira G, Chandrakanth A, Anjali, Kanna T and Archana V. Sustainable agriculture practices for promoting soil health: A Crucial paradigm for environmental resilience. *Vigyan varta an international e-magazine for science enthusiasts*. E-ISSN: 2582-9467
- Lucy M, Reed E, Glick BR. Applications of free living plant growth promoting rhizobacteria. *Antonie Van Leeuwenhoek.*, 2004; 86 (1): 1–25.
- Grover M, Pandey AK, Mishra BK, Lata and Roy RC. Sugarcane Crops: Plant Growth Promoting Bacteria in Growth, Yield and Productivity. (In) *Sugar Beet Crops: Growth, fertilization and Yield*, pp 135–51. Hertsburg Claus T (Eds). 2009. Nova Science Publishers Inc., New York.

He Y, Pantigoso HA, Wu Z, Vivanco JM. Co-inoculation of *Bacillus* sp. and *Pseudomonas putida* at different development stages acts as a biostimulant to promote growth, yield and nutrient uptake of tomato. *J. Appl. Microbiol.* 2019, 127, 196–207. [CrossRef] [PubMed]

Jeyanthi V and Kanimozhi S. Plant Growth Promoting Rhizobacteria (PGPR) - Prospective and Mechanisms: A Review-*Journal of Pure and Applied Microbiology*, June 2018. Vol. 12(2), p. 733-749.

Kloepper JW and Schroth MN. Relationship of in vitro antibiosis of plant growth promoting rhizobacteria to plant growth and the displacement of root microflora. *Phytopathology*. 1981; 71:1020–1024.

Kuan KB, Othman R, Rahim KA, Shamsuddin ZH. Plant growth-promoting rhizobacteria inoculation to enhance vegetative growth, nitrogen fixation and nitrogen remobilisation of maize under greenhouse conditions. *PLoS ONE* 2016, 11, 0152478.

Kumar A, Maurya BR, Raghuwanshi R, Meena VS, Islam MT. Co-inoculation with *Enterobacter* and Rhizobacteria on Yield and Nutrient Uptake by Wheat (*Triticum aestivum* L.) in the Alluvial Soil Under Indo-Gangetic Plain of India. *J. Plant Growth Regul.* 2017, 36, 608–617.

Nair KP, Nair KP. The agronomy and economy of ginger. In: Turmeric (*Curcuma longa* L.) and Ginger (*Zingiber officinale* Rosc.)-World's Invaluable Medicinal Spices: *The Agronomy and Economy of Turmeric and Ginger*. 2019. pp. 245-315.

Prasad S, Tyagi AK. Ginger and its constituents: Role in prevention and treatment of gastrointestinal cancer. *Gastroenterology Research and Practice*; 2015. Available:<https://doi.org/10.1155/2015/142979>

Prokryl Z, Vancura V, Wurst M. Auxin formation by rhizosphere bacteria as a factor of root growth. *Biol. Plant.* 1985; 27:159- 163

Salvo LPD, Cellucci GC, Carlino ME, de Salam IEG. Plant growth-promoting rhizobacteria inoculation and nitrogen fertilization increase maize (*Zea mays* L.) grain yield and modified rhizosphere microbial communities. *Appl Soil Ecol* (2018) 126: 113–120.

Soeparjono S. Pengaruh Komposisi Media Organik Terhadap Pertumbuhan, Hasil dan Kualitas Rimpang Tiga Varietas Jahe (*Zingiber officinale* Rosc.). *Prosiding Seminar Ilmiah Perhorti*. 2013;2013:613 618.

VanPeer R, Schippers B. Plant growth responses to bacterization with selected *Pseudomonas* spp. strains and rhizosphere microbial development in hydroponic cultures. *Can. J. Microbiol.* 1989, 35, 456–463.
