

# Review Article Hydroponics in Spices and Aromatics: A Review

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## ABSTRACT

The global shift towards urbanization, with 55% of the population residing in urban areas and an anticipated rise to 68% by 2050, presents challenges to traditional farming. The increasing world population, further intensifies the strain on available agricultural land due to urban expansion. Climate change combines with these issues, make conventional cultivation more challenging. Alternative agricultural practices, such as hydroponics, gain significance in addressing these concerns. Aromatic and spices have been used for centuries for their medicinal and culinary properties. They contain various essential oils, flavonoids, and other phytochemicals responsible for their distinctive flavour and aroma, as well as their health benefits. The rising global demand for natural products from herbs and spices has led to its market growth. Hydroponics, a technology for cultivating plants in nutrient solutions with or without an artificial medium such as sand, gravel, vermiculite, rockwool, peat moss, coir or sawdust, offers a promising solution. This paper explores the potential of hydroponic cultivation for planting material production, enhancing the yield, nutritional composition, essential oil content and medicinal properties of various spice and aromatic crops, including black pepper, saffron, ginger, garlic, coriander, mint, celery, fenugreek, parsley, thyme, rosemary, green mustard, basil and others. Hence by reviewing recent literature, this paper aims to provide insights into the classification of hydroponics and its different techniques, various growing media and nutrient solutions used, types of hydroponics utilized and their purposes in the production of diverse herbs and spices. Embracing hydroponic techniques could contribute to sustainable agricultural practices, particularly in urbanized environments where traditional farming faces constraints.

*Keywords: Urbanization; Climate change; Hydroponics; Herbs; Spices; Sustainable agriculture.*

## 1. INTRODUCTION

As of today, 55% of the global population resides in urban areas, a figure projected to rise to 68% by 2050 [1]. Additionally, the world's population is anticipated to increase by nearly 2 billion persons in the next 30 years, from the current 8 billion (mid-November 2022) to 9.7 billion in 2050 and could peak at nearly 10.4 billion in the mid-2080s [2]. The land available for agriculture is shrinking because of urbanisation and the allocation of land for domestic as well as other needs. Additionally, climate change makes farming more challenging. To overcome this, alternative agricultural practices need to be strengthened, with hydroponic cultivation becoming increasingly important in this context.

Hydroponics offers several benefits, including the ability to reserve traditional farmland for major crops and save over 90% of irrigation water by using a recycled, fixed amount of water. It can be run in various places, such as balconies, rooftops, greenhouses, and lands unsuitable for conventional farming. Fertilizers are applied in precise quantities based on plant needs. In hydroponic cultivation, it is easier to manage plants and to conduct the required protection operations against various pests. Additionally, hydroponics allows for year-round cultivation, as climate conditions can be controlled in greenhouses. Since plant roots do not need to reach for nutrients, planting density can also be increased [3].

Aromatic plants and spices have long been valued for their medicinal and culinary uses, as they contain essential oils, flavonoids, and other phytochemicals that give them their unique flavour, aroma, and health benefits. The demand for spices and herbs is rising in both developed and developing nations, driven by increasing consumer interest in their potential health benefit [4]. The market for essential oils is also steadily increasing, because of their use in products like air fresheners, lotions, and other items. The most commonly traded essential oil compounds include menthol, menthone, carvone, eugenol, and linalool [5]. Aromatic plants are rich sources of antioxidants and natural antibacterial agents, making them valuable for the food industry, while spices are aromatic substances of plant origin used to season dishes [6]. In this context, promoting hydroponic cultivation can help boost the yield, nutritional value, and essential oil content, as well as the antioxidant and medicinal properties of certain spices and aromatic plants.

## 2. HYDROPONICS

Hydroponics is a method of growing plants in nutrient solutions (water and fertilizers) with or without the use of an artificial medium, such as sand, gravel, vermiculite, rockwool, peat moss, coir, or sawdust, to provide mechanical support [7]. The term hydroponics comes from the Greek words "HYDRO" (water) and "PONOS" (labour), meaning "water working." This system is scientifically feasible because soil is not required for photosynthesis. The fundamental requirements for any hydroponic system include maintaining optimal levels of electrical conductivity (EC) and pH, proper aeration and temperature, the buffering effect of water and nutrient solution on the growing medium, and providing all essential micro and macronutrients to the plants [8]. A typical hydroponic system consists of several key components, including a growing tray, reservoir, timer, controlled submersible pump, delivery system, air pump, air stone for oxygenating the nutrient solution, and grow lights (for indoor hydroponics) within the required range [9]. The basic plant requirements in any hydroponic system are as follows [3].

- 1. Growth demands:** Various factors, including nutrition, light, temperature, air, pH, and salinity, influence plant growth, regardless of whether they are cultivated in soil or soilless systems. While sunlight and air are accessible in outdoor settings, indoor systems require sufficient light sources, such as fluorescent lights, metal halide lamps, or sodium vapour lamps.
- 2. Nutrient solution:** The availability of well-balanced nutrient solutions suitable for all stages of plant growth.
- 3. Water quality and disinfection:** Water used in hydroponics should be of high quality and free from pathogens. A drawback of closed systems is the potential for rapid spread of soil-borne pathogens through the recirculating nutrient solution. To eliminate these pathogens, various disinfection methods can be employed, including ozone treatment, UV disinfection, heat treatment, slow sand filtration, electrolyzed water, hydrogen peroxide, membrane filtration, and chlorination.

### 2.1 Merits and Demerits of Hydroponic Cultivation

The key advantages of hydroponic cultivation include a lower occurrence of soil-borne pathogens and diseases, no need for soil disinfection or treatment, precise control over environmental factors, higher yields, improved product quality, increased water and nutrient efficiency, and year-round production [8]. Hydroponics is ideal for areas with limited or contaminated land, allowing for higher planting density and eliminating the risk of weed infestation [10]. It also requires less space compared to traditional farming and reduces the use of harmful plant protection chemicals [11].

The main limitations of hydroponics are the high initial investment, the need for advanced technical knowledge, and the requirement for precise monitoring [8]. Some diseases, such as fusarium and verticillium, can spread quickly through the system [11], and algal growth presents an additional challenge [9].

## 3. CLASSIFICATION OF HYDROPONICS

Hydroponic systems are categorized into passive and active types. In passive systems, plant roots remain in contact with the nutrient solution while being supported by the suspension method, making these systems simple to set up. An example of a passive system is the wick system. On the other hand, active systems use an electric pump to deliver nutrient solutions to the plant roots and rely on gravity to drain any excess solution, which can then be recycled and reused. Examples of active systems include the Nutrient Film Technique (NFT), Deep Flow Technique (DFT) and Ebb and flow system [12].

Hydroponic cultivation can be classified based on the type of substrate and container, the method of nutrient delivery to the plant, and drainage. The two main types are solution culture or liquid hydroponics, and solid media culture (also known as aggregate systems).

- 1. Solution culture or Liquid hydroponics:** Solution culture, or liquid hydroponics, involves growing plants in a fully liquid medium contained within pipes or appropriate containers [8]. This method can be further divided into circulating systems (closed systems) and non-circulating systems (open systems).

**Circulating methods (closed system):** In this method, the nutrient solution is circulated through the plant root system, where an excess solution is collected, replenished, and reused [13]. Techniques within this system are the Nutrient Film Technique (NFT), Deep Flow Technique (DFT) and Ebb and flow system.

**Non-circulating method (open system):** In this system, the nutrient solution is utilized only once and is not recirculated. When its nutrient concentration reduces or when there are changes in pH or electrical conductivity (EC), it is replaced [13]. The different techniques, such as root dipping technique, floating technique and capillary action techniques, fall under this method. In the root dipping technique, plants are cultivated in small pots filled with a growing medium, with the bottom 2-3 cm of the pots submerged in a nutrient solution. The root dipping method is straightforward, cost-effective, and easy to set up. The floating technique involves using shallow containers about 10 cm deep where plants are placed in small pots and secured to a styrofoam sheet or a similar lightweight material, allowing them to float on the nutrient solution in the container [8]. The capillary action technique uses pots with holes at the bottom, filled with an inert medium where seeds or seedlings are planted. These pots are placed in shallow containers containing a nutrient solution, which is absorbed into the inert medium through capillary action. Proper aeration is crucial in this method [13].

## **2. Solid media culture (Aggregate systems)**

The chosen growing medium should be flexible, easily crumbled, capable of retaining both water and air and allow for efficient drainage. Additionally, it should be free from toxic substances, pests, disease-causing microorganisms, and nematodes. The medium must be thoroughly sterilized before use [13].

**Hanging bag technique:** In this verti-grow technique, 1-meter long, cylindrical UV-treated polythene bags (white on the outside, black inside) are filled with sterilized coconut fibre. These bags are sealed at the bottom and attached to small PVC pipes at the top, then suspended vertically from overhead support above a nutrient solution collection channel. Seedlings or planting materials, placed in net pots, are inserted into holes along the sides of the hanging bags. A micro sprinkler inside the top of each bag pumps nutrient solution, which drips down, moistening the coconut fibre and plant roots. Excess solution drains through holes at the bottom of the bags and is collected in the channel below, flowing back to the nutrient solution tank. This system is ideal for growing leafy vegetables, strawberries, and small flower plants [13].

**Grow bag technique:** This technique utilizes grow bags made from UV-stabilized polyethylene sheets, measuring 1 meter in length, 15-20 cm in width, and 8-10 cm in height, to cultivate plants. The bags can accommodate single or paired rows of plants. Fertigation is carried out using specialized stake drippers connected to poly tubes and lateral pipes [8].

**Trench or trough technique:** In this method, plants are grown in trenches or troughs constructed from UV-stabilized PVC/HDPE sheets, bricks, concrete, or other locally available materials [13]. These trenches or troughs are filled with inert organic, inorganic, or mixed materials, with a depth ranging from 30 to 60 cm based on the crop type. Fertigation is managed using specialized stake drippers connected to poly tubes and lateral pipes. Proper drainage is essential and can be ensured through holes or separate drainage pipes [8].

**Pot technique:** This method uses pre-made plastic pots for plant cultivation, which are filled with inert organic, inorganic, or mixed materials. Fertigation is managed using specialized single or multiple outlet stake drippers connected to poly tubes and lateral pipes [8].

## **3.1 Different Techniques of Hydroponic System**

Anyone can create their own design tailored to their specific needs and the type of plants they wish to grow based on the primary goal and the purpose of the hydroponic system [3]. The following are different systems of hydroponics which are mainly used.

**1. Nutrient film technique (NFT):** In this technique, plant roots are exposed to a thin film of nutrient solution, approximately 0.5 mm thick, flowing through the channel. This design allows the roots to access oxygen from the surrounding air, as they are not fully submerged in the solution [10]. Developed in the mid-1960s in England by Dr. Alen Cooper, this method is also referred to as gutter hydroponics [12] [14]. The system works by pumping nutrient solution from a holding tank through irrigators at the top of each sloping pipe. The excess solution is collected at the bottom and recycled back into the tank, creating a continuous flow [3]. Channels are typically made from flexible PVC or plastic sheets, with seedlings anchored in pots containing growing media. These channels, which are between five and ten meters in length, are set at a slope of 1 in 50 to 1 in 70, with a flow rate of two to three litres per minute [8]. While NFT systems do not require a timer for the submersible pump, they are highly vulnerable to power outages or pump failures, as the roots can dry out quickly without a continuous flow of nutrients [13]. This technique is particularly suitable for plants with small root systems, such as lettuce, strawberries, and herbs [3].

**2. Deep flow technique (DFT):** It is a hydroponic system where a nutrient solution, 2-3 cm deep, flows through PVC pipes. Plants are placed inside plastic pots, which are positioned at regular or desired intervals along the pipes. These main and sub-main pipes are fixed on raised platforms. The PVC pipes can be arranged either in a single horizontal plane or in a multi-level zig-zag vertical layout [8]. As the recycled nutrient solution flows back into the stock tank, it becomes aerated. The pipes are typically sloped at a gradient of 1 in 30 to 40 to ensure a smooth flow of nutrient solution. Painting the PVC pipes white can help to reduce the heating of the nutrient solution [13].

**3. Ebb and flow system:** This system which is also known as the flood and drain system, is an affordable type of hydroponic setup. It consists of two containers, the top one holds plants in pots with substrate, while the bottom one contains the nutrient solution. Nutrients are pumped in large volumes into the top container, flooding it to a level determined by an overflow pipe, with excess solution recirculating back to the bottom container. The pump operates intermittently, typically running for 30 minutes and switching off for 15 minutes, allowing the grow tray to flood periodically. When the pump is off, the nutrients drain out of the tray through the pump line, providing oxygen to the roots. For this reason, an air stone is not essential in ebb and flow systems. This setup is especially suitable for plants with large root systems [3]. However, issues like root rot, algae, and mould are common, making it necessary to incorporate a filtration unit in modified versions of the system [12].

**4. Deep water culture (DWC):** Plants are suspended on a floating platform above a bath of hydroponic nutrient solution, with oxygen supplied continuously by an air pump. This system can be easily set up in various containers, such as glass basins, fish ponds, plastic or ice boxes, concrete basins, or basins covered with polypropylene sheets. Since the plants are always in contact with the nutrient solution, they are not at risk of damage in the event of a power outage or air pump failure. DWC is particularly well-suited for growing plants like lettuce, strawberries, and herbs [3].

**5. Wick system:** This is the simplest type of hydroponic setup [3]. In this system, nutrient solution reaches the plant roots through capillary action, with the wick serving as the connection between the potted plant and the nutrient solution in the reservoir. It's especially useful in areas where electricity is unavailable or unreliable. The water supply to the plant can be adjusted by using a larger or wider wick, or multiple wicks. This system works well for small plants, herbs, and spices but is less effective for plants that require a large amount of water [15].

**6. Drip system:** The drip system consists of at least two containers, with plants placed in the top container and the nutrient solution in the bottom one. A pump delivers the nutrient solution to drips located at the base of each plant, and an aquarium air stone oxygenates the water. The nutrients filter down to the plant roots and then may or may not return to the bottom container. Both the water and air pumps typically run continuously in this system. It is particularly well-suited for plants with large root systems [3]. A key advantage of the drip system is its ability to withstand short-term power or equipment failures, while also conserving more water compared to other systems. However, it can be expensive and complex to set up, as the nutrient solution supply needs to be timed [12]. There are two types of drip systems in hydroponics: recovery and non-recovery. In a recovery system, the nutrient solution is collected and recycled continuously, while in a non-recovery system, the solution is not recycled but drained out after use [10].

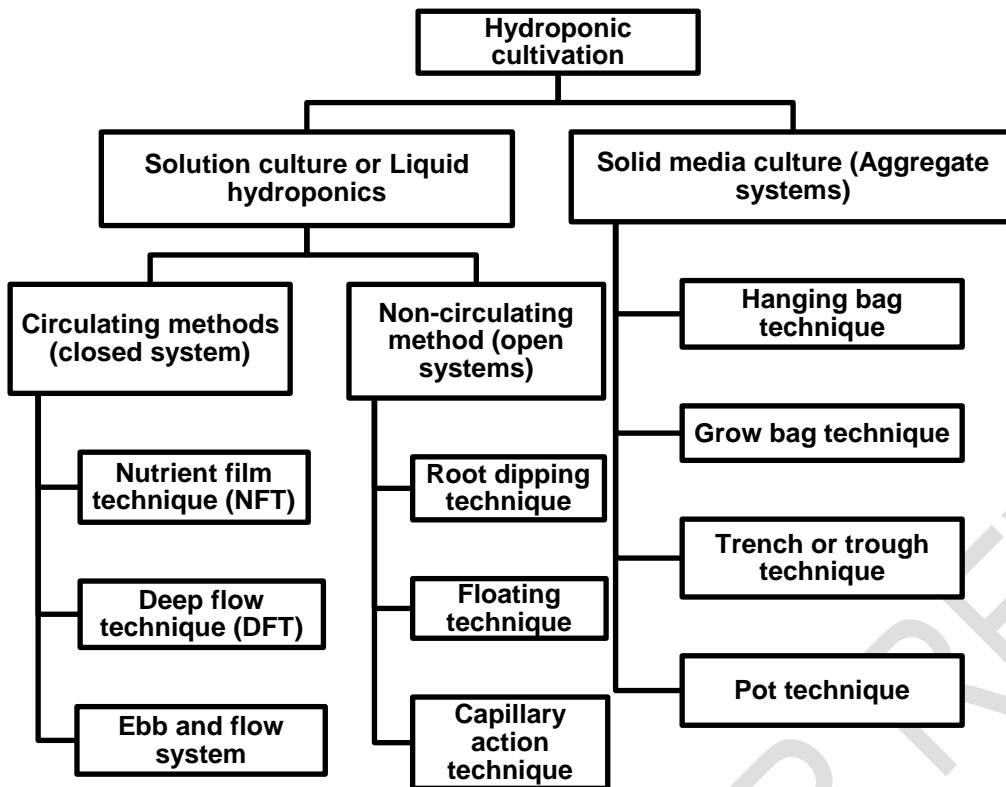


Fig. 1. Classification of hydroponics based on the type of substrate and container, the method of nutrient delivery to the plant, and drainage.

#### 4. NUTRIENT SOLUTIONS FOR HYDROPONICS

A nutrient solution must contain all the 17 essential elements for plant growth and development, as these nutrients are irreplaceable and vital for completing the plant's life cycle [13]. If the system experiences a water shortage, the concentration of nutrients can increase, potentially harming the roots and affecting their function. To prevent this, it is important to add fresh, filtered water to restore the nutrient solution to its original concentration. Additionally, maintaining the root zone temperature between 20 - 22°C is crucial. If the nutrient temperature is too high, it can lead to issues such as yellowing, falling flowers, damaged fruits, and stunted growth [3]. Therefore, the success of a soilless cultivation relies heavily on a strict nutrient management program. Properly controlling factors like pH level, temperature, and electrical conductivity, along with timely replacement of the nutrient solution, are essential for thriving soilless cultivation [13].

The optimal pH range for nutrient solutions in soilless cultivation is between 5.8 and 6.5. Deviating from this range can lead to nutrient deficiencies or toxicity symptoms, depending on the specific crop. Similarly, the ideal electrical conductivity (EC) for hydroponic nutrient solutions is between 1.5 and 2.5 dS m<sup>-1</sup>. If the EC is too high, osmotic pressure can inhibit nutrient absorption, while a lower EC can negatively affect plant health and yield. As plants absorb nutrients and water, the EC of the solution fluctuates. To maintain balance, fresh water should be added if the EC is too high, or nutrients should be added if it is too low [13].

There are several standard nutrient solutions used in hydroponics, such as the Hoagland and Arnon solution, Cooper solution, Steiner solution, and Albert medium. Commercial hydroponic fertilizers contain all the macro and micronutrients necessary for plant growth, with recommendations based on the electrical conductivity (EC) and pH levels of the nutrient solution [16]. The Hoagland solution, developed by Hoagland and Arnon (1950), is a well-known hydroponic nutrient solution that provides all essential nutrients for plant growth. Its composition per litre of solution includes KNO<sub>3</sub> (5 ml of 1 M), Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O (5 ml of 1 M), KH<sub>2</sub>PO<sub>4</sub> (1 ml of 1 M), MgSO<sub>4</sub>·7H<sub>2</sub>O (2 ml of 1 M), micronutrients (1 ml of stock solution), and Fe-EDTA (1–5 ml of 1000 mg L<sup>-1</sup>). The micronutrient stock solution contains H<sub>3</sub>BO<sub>3</sub> (2.86 g), MnCl<sub>2</sub>·4H<sub>2</sub>O (1.81 g), ZnSO<sub>4</sub>·7H<sub>2</sub>O (0.22 g), CuSO<sub>4</sub>·5H<sub>2</sub>O (0.08 g), and MoO<sub>3</sub>·H<sub>2</sub>O (0.02 g) per liter [17]. Alternatively, liquid organic nutrient solutions can be used in hydroponics as a chemical-free option derived from organic sources that are safe for health [18].

These organic solutions can be made from a variety of animal, plant, or algae-based substances, including manure extracts, fish products, vinasses, corn steep liquor, vermicompost tea, and algae extracts [19].

## 5. GROWING MEDIA FOR HYDROPONICS

In soilless cultivation systems, the growth medium serves as a substitute for soil. Its primary functions are to provide oxygen to the roots, ensure that water and nutrients reach the roots, and offer stability to plants to prevent them from toppling over [3]. Hydroponic substrates are classified into two types: fibrous and granular. Fibrous substrates can be either organic or inorganic, while granular substrates are typically inorganic [9].

**Table 1. Classification of materials used as substrate**

Substrate	Characteristics	Examples
Organic	This category consists of natural organic substrates, including organic residues, by-products, and waste materials of organic origin, typically derived from agriculture or industrial by-products from the wood industry.	Peat, coconut fibre
Inorganic	This category encompasses both natural materials and mineral products that are produced through industrial processes.	Sand, pumice, vermiculite, perlite, clay pellets, rockwool, zeolites
Synthetic	This category includes both ion-exchange synthetic resins and low-density plastic materials.	Expanded polystyrene, polyurethane foam

**Table 2. Different substrates used in hydroponic systems**

Substrate	Characteristics
Coco-Coir	Coco coir is a fully organic medium made from shredded coconut husks. It combines the water retention properties of vermiculite with the air retention qualities of perlite, providing protection against root diseases and fungi [13]. Additionally, coco coir is pH neutral and promotes good aeration for the roots [15]. Also referred to as coir pith, coir meal, coir dust, and coco peat, it consists of the dust and short fibers extracted from the mesocarp of the coconut [20].
Peat Moss	It retains moisture in growing mediums [13].

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Composted organic waste	In the past 40 years, the use of composted organic waste in soilless growth media has grown significantly. Composts have been shown to inhibit pathogens, enhancing their economic value [20].
Rice Hulls	This material is a by-product of the rice industry. Although it is organic plant material, it decomposes very slowly, similar to coco coir, which makes it suitable for use as a growing medium in hydroponics [15].
Fibre from wood	The effectiveness of substantially processed wood fibre as a growing medium has been thoroughly investigated, revealing that it has enhanced total porosity and air-holding capacity [20].
Sand	Sand is a widely used growing medium in hydroponics. It is often blended with vermiculite, perlite, and/or coco coir, all of which contribute to moisture retention and improve aeration for the roots [15].
Perlite	Perlite is the most commonly used medium in containerized soilless culture systems. These lightweight granules are derived from a silicone mineral that forms in volcanoes [13]. Perlite can be used on its own or combined with other growing media [15]. It retains more air than water and has a neutral pH, exceptional wicking ability, and high porosity [12].
Vermiculite	This substance includes both potassium and magnesium. It retains a significant amount of water and helps with drainage and aeration, although it is less durable than other mediums like sand and perlite [13]. Vermiculite is a silicate mineral that expands when subjected to extreme heat. It is lightweight and often floats [15].
Rockwool	Rockwool is the most widely used growing medium in soilless cultivation. It is a sterile, porous, and non-degradable medium made mainly from granite and/or limestone. Rockwool absorbs water readily. Before use, it should be pH balanced by soaking it in pH-balanced water [15].
LECA (Light Expanded Clay Aggregate)	LECA baked clay pellets, are suitable for hydroponic systems and they are inert, pH neutral and do not contain any nutrient value [21].

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Oasis Cubes They are made from an open-cell material, allowing them to absorb both water and air. Unlike rockwool cubes, oasis cubes are less prone to becoming waterlogged [15].

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## 6. IMPORTANCE OF HYDROPONICS IN SPICES AND AROMATICS

Hydroponics plays a crucial role in the cultivation of spices and aromatics by enhancing both yield and product quality. This method allows for better control over pests and pathogens, ensuring the production of healthy planting materials such as corms and cuttings that are free from contamination. Additionally, hydroponics shortens the growing period and promotes the greater accumulation of medicinal components in spices and aromatics. It also improves the essential oil yield and quality by increasing the concentration of the major constituents present in the essential oil. Furthermore, hydroponics enables off-season production of various herbal and leafy spices, as well as aromatic herbs.

### 6.1 Hydroponics in spices and aromatics

Stem cuttings are the planting material for black pepper; however, soil propagation exposes the stems and developing roots to soil-borne pathogens, which can compromise plant quality. High-quality rootstock supports faster and more effective establishment of black pepper plants. Hydroponic farming provides an excellent platform for producing quality rootstock. A research study evaluated seven nutrient media compositions, finding that Hoagland solution supplemented with potassium silicate at 0.005 mM (T4) and salicylic acid (SA) at 2 mM (T6) promoted faster root growth and initiation. Silicon fertilization plays a regulatory role in nutrient uptake, while SA enhances cellular auxin synthesis at the root tip, aiding the development of adventitious roots. Stem cuttings without roots at planting showed greater increases in root numbers and extension due to their enhanced ability to absorb water and nutrients [22].

Saffron, one of the most expensive spices globally, is propagated vegetatively using corms, as the cultivated plants are sterile auto-triploids [23]. Its cultivation faces several challenges, including limited cultivable land in high-altitude, cold climates and the labour intensive process of harvesting and handling. Additionally, the low multiplication rate of cormlets and susceptibility to fungal infections further reduce saffron productivity. Hydroponics offers a promising alternative to traditional open-field cultivation, potentially increasing saffron yields and lowering production costs. This method also supports sustainable production by providing pathogen-free stock corms. However, for hydroponic saffron production to be effective, the system and growth conditions must be carefully optimized [24]. High germination rates for saffron was found in nutrient film technique (NFT), peat moss-perlite substrate cultivation, and soil cultivation. Despite lower flower productivity in NFT, it produced the highest quality corms, with 100% commercially viable and the heaviest corms. This was due to NFT's efficient water use and continuous nutrient supply [25].

In a study, saffron corms were sprouted by gradually lowering the temperature from 14°C to 8°C and cultivated hydroponically in perlite or volcanic rock for 24 weeks using ebb-and-flow or continuous immersion systems. They concluded that large corms grown in volcanic rock-based aerated continuous immersion hydroponics produced optimal yields of saffron stigmas and daughter corms [24]. The flowering potential of saffron corms across various weight groups in hydroponic conditions were evaluated and it was found that corms weighing less than 4 g generally lacked the ability to flower. However, corms weighing between 4-8 g appeared to produce more flowers in controlled environments compared to natural environments [26].

High-quality ginger is typically grown in highlands using shifting cultivation to avoid issues with infertile soil and soil-borne diseases that can harm rhizomes and reduce yields. However, this method is unsustainable, as it leads to land erosion in the highlands. Hydroponic systems offer a promising alternative, addressing land availability challenges while also increasing production yields, enhancing productivity, and shortening the growing cycle [27].

The effects of soilless substrates, such as coir dust and burnt paddy husks, on ginger growth using a drip irrigation system were studied. They found that substrates with 70-100% coir dust significantly improved growth and increased rhizome yield by up to 36%. The higher porosity of coir dust made it the most effective medium for ginger cultivation [28]. An experiment investigated the effects of hydroxyapatite nanoparticles (HANPs) on ginger growth in hydroponics, using a modified Hoagland solution with varying HANP concentrations (200, 500 and 1000 ppm) and the results showed that 200 ppm HANP significantly increased leaf number, chlorophyll content, and photosynthetic rate compared to the control. Regular use of the modified Hoagland solution provided adequate nutrients, promoting steady growth throughout the cultivation period [27].

Garlic, a member of the Liliaceae family, is widely used as both a spice and medicinal herb in many countries. One of its key bioactive compounds, ajoene, is formed through the enzymatic conversion of alliin into alliin by alliinase. Ajoene is known for its numerous health benefits, including anti-thrombosis, anti-tumor, anti-leukemia, and cholesterol-lowering effects, as well as its anti-fungal, antimicrobial, antioxidant, and antiviral properties [29]. There are two forms of ajoene: E-ajoene, the more stable cis isomer, and Z-ajoene, the trans isomer [30]. Hydroponic cultivation of garlic promotes faster growth and a higher accumulation of these medicinal compounds compared to traditional soil cultivation [29].

A floating technique hydroponic system for garlic cultivation resulted in higher fresh weights and ajoene concentrations in the bulbs, roots, and leaves compared to soil-grown plants. The garlic plants in hydroponic culture were also more vigorous and healthier [31]. A research was done on how different levels of dissolved oxygen, achieved through various aeration methods, affected garlic growth and ajoene concentrations in floating technique hydroponics using a modified Hoagland nutrient solution. The treatments included no aeration (control), 12 hours of aeration in the dark, 12 hours in the light, and 24 hours of continuous aeration. The findings indicated that garlic plants with higher dissolved oxygen levels exhibited better growth rates and increased ajoene content [30].

Excessive use of nitrogenous fertilizers increases the nitrate concentrations in plant tissues while simultaneously reducing ascorbic acid levels. Therefore an experiment was done on the effects of three nitrogen concentrations (90, 180, and 270 ppm) on the growth, yield, and chemical characteristics of three celery cultivars (Utah, President, and Monterey) in a floating hydroponic system. The results showed that an intermediate nitrogen level of 180 mg L<sup>-1</sup> significantly improved plant height, number of leaves per plant, fresh weight, and total yield. Increasing nutrient concentration led to higher nitrate levels in leaf tissues, but these levels remained below the limits set by the European Community [32]. An investigation on the effects of vermiwash and vermicompost derived from neem (*Azadirachta indica*) and lime (*Citrus aurantifolia*) on celery growth in a hydroponic system, found that vermiwash and vermicompost from lime leaves significantly promoted plant growth, resulting in taller and wider celery plants. These amendments provide sufficient macro and micronutrients and release nutrients slowly for better accessibility [33]. A recent study compared fenugreek cultivation in NFT hydroponics and conventional systems. The results revealed that while the crops in the hydroponic system had a pale yellowish-green color, indicating micronutrient deficiencies, the NFT hydroponic system still produced slightly better crop growth and longer average root lengths than the conventional system [34].

Higher growth and production of coriander in DFT hydroponics were achieved with a recirculation interval of 0.25 hours compared to longer intervals of 12 and 24 hours. Adjusting the recirculation intervals is crucial for maintaining an optimal oxygen supply to the nutrient solution [35]. Two coriander cultivars, Verdao and Tabocas, were grown in both adapted DFT systems using pipes (with depths of 0.02 m and 0.03 m) and conventional DFT systems using wooden tanks (with depths of 0.013, 0.017, and 0.025 m). Results showed that Verdao was more productive, and the smallest nutrient solution depths performed similarly to larger ones, indicating that smaller depths are feasible and contribute to water and fertilizer savings [36]. An experiment demonstrated that varying concentrations of amino acid solutions (0, 100, and 200 ppm) influenced the growth, elemental content, and essential oil composition of hydroponically grown coriander under broad-range red and blue (BRB) light and white light. This amino acid solution rich in aspartic acid and other amino acids, at 100 ppm concentration, particularly under BRB light, led to significant improvements in plant morphology, chlorophyll, carotenoids, and the diversity of functional groups. It also promoted the synthesis of more aromatic monoterpenes, highlighting the potential of amino acids to modulate essential oil composition based on specific formulation needs [37].

The growth, essential oil quality, and content of thyme in a greenhouse hydroponic system (Drip system) were evaluated under varying planting densities (14, 28, and 71 plants m<sup>-2</sup>) and nutrient solution concentrations (Steiner- 100% and 50%). Optimal growth was achieved with a 100% Steiner solution and a density of 28 plants m<sup>-2</sup>. The hydroponic system produced 18.45 kg m<sup>-2</sup> of fresh thyme annually, a 386.8% increase compared to the 3.79 kg m<sup>-2</sup> yield from open-field cultivation [38]. An investigation was done to observe the relationship between plant density and total dry biomass production in hydroponically grown rosemary. Among the three densities tested, the highest biomass production was achieved at a population density of 8 plants m<sup>-2</sup>, outperforming the higher densities of 16 and 24 plants m<sup>-2</sup> [39]. Hydroponically grown parsley using Hoagland solution (T1) in a DFT system significantly improved plant morphology, physiological performance, and myristicin content in essential oil. Parsley treated with T1 also showed higher growth in leaf, stem, and root biomass and increased antioxidant potential, compared to those grown with the Thakur nutrient solution (T2) [40].

The green mustard plant (*Brassica juncea* L.), a vegetable that typically grows in sub-tropical climates, is also well-suited for tropical regions [41]. Its leaves are frequently used in traditional dishes from African, Indian, Asian, and African-American cuisines due to their distinct horseradish, peppery, and spicy flavor [42]. A study investigated the impact of using vegetable waste as a nutrient supplement in hydroponics on the growth of mustard greens. Six treatments were tested: P0 (control), P1 (20 ml Organic Liquid Fertilizer (OLF) + 40 ml AB mix nutrient solution), P2 (30 ml OLF + 30 ml AB mix), P3 (40 ml OLF + 20 ml AB mix), and P4 (60 ml OLF) in a wick hydroponic system. The findings indicated that P2 produced the best outcomes in terms of plant height, fresh weight, and dry weight, likely due to its role in promoting accelerated cell division

in the apical meristem (shoots) [41]. The growth performance of mustard greens was evaluated under wick hydroponic conditions using various nutrient sources, including Organic Liquid Fertilizer (OLF), OLF with bokashi, commercial liquid fertilizer, and pure water. Bokashi compost is one of the potential fertilizer sources for organic hydroponics. The study found that the organic liquid fertilizer from vegetable waste was as effective as commercial liquid fertilizer. Although plants in the OLF wick system slightly yellowed by the end of the season, no leaves dried out, unlike those in the nutrient solution, which had bright green leaves with few dried ones. Additionally, plants treated with OLF and bokashi wilted by the eighth day due to an unsuitable pH level for mustard greens [42].

An investigation was done on the impact of nitrogen levels on the growth, nutrient content, antioxidant activity, and essential oil quality and quantity of spearmint grown hydroponically. Five nitrogen concentrations (150, 175, 200, 225, and 250 mg L<sup>-1</sup>) were tested, while potassium and phosphorus levels were kept constant at of 325 mg L<sup>-1</sup> K and 50 mg L<sup>-1</sup> respectively, based on previous studies. The study found that a nitrogen level of 200 mg L<sup>-1</sup> resulted in high dry matter content in the upper plant biomass without affecting fresh weight. Higher nitrogen concentrations (>200 mg L<sup>-1</sup>) reduced the levels of several micronutrients in the plant tissue. For optimal carvone content and enzymatic activity, a nitrogen level of 200 mg L<sup>-1</sup> is recommended for spearmint cultivation aimed at essential oil production [43]. The feasibility of commercially growing peppermint and spearmint in a floating tray system under mild salinity conditions was explored, as freshwater availability is a challenge for hydroponic production. The study tested three NaCl concentrations (0 mM, 10 mM, and 20 mM) and found that plant growth was generally unaffected by salinity, although high salinity did reduce plant height. While the nutritional composition of peppermint remained stable, spearmint benefited from reduced nitrate levels and increased antioxidant capacity, total soluble phenols, total carotenoids, and essential oil content. Overall, both peppermint and spearmint can be successfully cultivated in floating systems with mild salinity without compromising yield or essential oil and phenolic composition [44].

A work was done to determine the effects of silicon (Si) amendments on hydroponically (using DFT) grown sweet basil (variety 'Genovese') at a constant temperature of 23°C. The study tested 0, 25, and 75 ppm Si levels, incorporated through K<sub>2</sub>SiO<sub>3</sub> and found that higher Si levels significantly improved plant growth and survival under cold stress. After an unexpected frost, basil treated with 75 ppm Si showed reduced cold injury, suggesting that Si amendments can enhance basil resilience in cooler conditions [45]. Basil grown in an open hydroponics was compared with that of soil to assess resilience to downy mildew, heat, and storage and shelf life performance. Hydroponically grown basil had higher antioxidant levels and better resistance to downy mildew, but it was less resilient to heat and more prone to browning during storage, leading to a shorter shelf life compared to soil-grown basil. The higher antioxidant content, while beneficial against pathogens and for health, contributed to reduced storability due to oxidation and browning [46].

Lemon balm (*Melissa officinalis* L.), a perennial herb from the Lamiaceae family with a lemon-like scent, is traditionally used in Europe to treat neurogenic disorders, insomnia, and stress due to its sedative and spasmolytic effects. Its essential oil also has antibacterial and antioxidant properties [47]. Studies have reported the use of hydroponic cultivation of lemon balm (ebb and flood system) to define the effect of wood fibre and sand substrates, drought stress on the herbal yield, amount and composition of essential oil, flavonoid content and antioxidative capacity [48] [49]. The antioxidant activity, total phenol, and flavonoid content of lemon balm grown in various media under hydroponic conditions were examined. The study found that different culture media, with distinct physical and chemical properties, influenced the plant's antioxidant levels. Peat moss substrate significantly enhanced the antioxidant properties of lemon balm compared to other media [50].

**Table 3. Types of hydroponic systems utilized for spice and aromatic crops cultivation as reported in previous studies**

Crop	Hydroponic system	Nutrient Solution	Growing media	Reference
Black pepper	Deep water culture	Hoagland solution and Modified Hoagland solution	Sponge cubes	[22]

Saffron	Nutrient Film Technique	Hoagland solution	Leca stone	[51]	
	Pot technique	Hoagland solution with different concentration of copper	Perlite	[52]	
	Ebb and flow system	Commercial liquid fertilizer	Perlite Rockwool	[24]	
	Continuous Immersion system	Commercial liquid fertilizer	Perlite Rockwool	[24]	
	Nutrient Film Technique	Hoagland solution	–	[25]	
	Drip system	Long-Ashton solution	Perlite	[53]	
	Ginger	Drip system	Formulated by MARDI (Malaysian Agricultural Research and Development Institute)	Burnt paddy husk and coco peat in various proportions	[28]
		Drip system	Modified Hoagland solution	Coco peat	[27]
Garlic	NFT	Cooper solution	–	[54]	
	Floating or hanging technique	Modified Hoagland solution	Styrofoam	[31]	
			particles	[30]	
				[55]	
				[29]	
Fenugreek	Non circulating culture	Albert's solution	Coco peat	[56]	
	Aggregate culture	Albert's solution	Coco peat	[56]	
	NFT	–	Coco peat	[34]	

Celery	Floating system	Cooper solution	Perlite	[32]
Coriander	DFT	Nutrient solution recommended by Furlani <i>et al.</i> (1999) [57] for leafy vegetables	Coconut fibre	[35]
	Adapted DFT system	Nutrient solution recommended by Furlani <i>et al.</i> (1999) [57] for leafy vegetables	Coconut fibre	[36]
	Conventional DFT system	Furlani <i>et al.</i> (1999) [57] for leafy vegetables		
	NFT	TV PLUS Mastermix	Coco-coir	[37]
Parsley	DFT	Hoagland solution Thakur nutrient solution	Pebbles	[40]
	DFT	12N-1.8P-13.4K water-soluble fertilizer	Stone-wool cubes	[58]
Thyme	NFT	–	Polyurethane foam	[59]
	Drip system	Steiner solution	Red volcanic sand	[38]
Rosemary	Closed hydroponic system	A standard hydroponic nutrient solution as per Rodríguez-Fuentes <i>et al.</i> (2011) [60].	Volcanic rock	[39]
Mustard greens	Wick system	Organic liquid fertilizer (OLF)	Rockwool	[41]
	Wick system	Organic liquid fertilizer, Organic liquid fertilizer with Bokashi, AB mix	Rockwool	[42]

Onion	Custom made hydroponics (Floating type)	–	Gravel	[61]
	Floating type	Growth nutrient solution	–	[62]
	Nutrient film technique,	Growth nutrient solution	–	
	Aggregate system	Growth nutrient solution	Perlite	
Dill	NFT	–	Polyurethane foam	[59]
	DFT	12N-1.8P-13.4K water-soluble fertilizer	Stone-wool cubes	[58]
Mint	Floating tray system	–	Peat	[44]
Sweet basil	DFT	Modified Sonneveld solution	Oasis cubes	[45]
Lemon balm	Ebb and flow system	70 g of Flory basis 1 along with 150 mg L <sup>-1</sup> N	Wood fibre and sand	[48]
	Ebb and flow system	70 g of Flory basis 1 along with 150 mg L <sup>-1</sup> N	Peat and wood fibre	[49]
Lavender	Drip system	–	Perlite	[63]
Geranium	Pot technique	–	Perlite	[64]

## 7. FUTURE PROSPECTS AND CONCLUSION

As the global population increases and arable land diminishes due to poor land management practices, there is a shift towards innovative technologies like hydroponics and aeroponics to expand crop production channels. The excessive use of pesticides and insecticides is contributing to soil infertility, further driving the movement towards soilless farming. Additionally, agriculture primarily relies on river water, but industrialization has led to pollution from toxic waste, contaminating these water sources with heavy metals and toxins. Consequently, the availability of water for conventional farming is diminishing, necessitating a transition to hydroponics, which uses 80-90% less water. Hydroponics is viewed as the future of agriculture, offering a sustainable and eco-friendly approach to enhance crop yield and quality, thereby supporting food security. Although the initial capital costs for setting up hydroponic systems pose a challenge, these costs are expected to decline over time, making hydroponics a more feasible option in the long run. Additionally, hydroponics could play a crucial role in the future of space exploration, as we haven't yet found any soil that can support life in space, and transporting soil via shuttles is impractical. Furthermore, hydroponics provides an excellent platform for biological research, facilitating the analysis of interactions among various factors that influence plant growth.

## REFERENCES

1. UN [United Nations]. 2018 Revision of World Urbanization Prospects. United Nations. 2018; Accessed 4 October 2024. Available: <https://www.un.org/development/desa/en/news/population/2018-revision-of-world-urbanization-prospects.html>.
2. UN [United Nations]. Population | United Nations. United Nations. 2022; Accessed 4 October 2024. Available: <https://www.un.org/en/global-issues/population>
3. Kazzaz EKA, Kazzaz EAA. Soilless agriculture a new and advanced method for agriculture development: An introduction. *Agric Res Technol*. 2017;3(2):63-72.
4. Nguyen L, Duong LT, Mentreddy RS. The US import demand for spices and herbs by differentiated sources. *Journal of Applied Research on Medicinal and Aromatic Plants*. 2019;12:13-20.
5. Khaliq FA, Mushtaq A. The five most traded compounds worldwide: Importance, opportunities, and risks. *Int J Chem Biochem Sci*. 2023;23(1):185-94.
6. Vikou C, Semassa J, Diaconeasa Z, Roko G, Tohyouessou M, Dah-Nouvlessounon D et al. Diversity, chemical compositions and beneficial effects of some spices and aromatic leaves consumed in Benin and in the world: Critical review. *American Journal of Plant Sciences*. 2023;14(5):569-98.
7. Jensen MH. Hydroponics worldwide. In *International Symposium on Growing Media and Hydroponics*. 1997;481:719-30.
8. Hasan M, Sabir N, Singh AK, Singh MC, Patel N, Khanna M et al. Hydroponics technology for horticultural crops. *Tech. Bull*. 2018;188. I.A.R.I., New Delhi.
9. Sathyanarayana SR, Gangadhar WV, Badrinath MG, Ravindra RM, Shriramrao AU. Hydroponics: An intensified agriculture practice to improve food production. *Reviews in Agricultural Science*. 2018;10:01-114.
10. Dubey N, Nain V. Hydroponic—the future of farming. *International Journal of Environment, Agriculture and Biotechnology*. 2020 Jul 8;5(4):857-64.
11. Khan S, Purohit A, Vadsaria N. Hydroponics: Current and future state of the art in farming. *Journal of Plant Nutrition*. 2020;44(10):1515-38.
12. Macwan J, Pandya D, Pandya H, Mankad A. Review on soilless method of cultivation: hydroponics. *Int J Recent Sci Res*. 2020;11:37122-7.
13. Hussain A, Iqbal K, Aziem S, Mahato P, Negi AK. A review on the science of growing crops without soil (soilless culture)- a novel alternative for growing crops. *International Journal of Agriculture and Crop Sciences*. 2014;7(11):833-42.
14. Shinde CT, Marathe PS. Farming without soil in today's era. *Iconic Research and Engineering Journals*. 2021 Jan;4(7):24-7.
15. Gaikwad DJ, Maitra S. Hydroponics cultivation of crops. In: Maitra S, Gaikwad DJ, Shankar T, editors. *Protected Cultivation and Smart Agriculture*. New Delhi Publishers; 2020:279-87.
16. Brechner M, Both AJ, Staff CE. *Hydroponic lettuce handbook*. Cornell Controlled Environment Agriculture. 1996;834:504-9.
17. Hoagland DR, Arnon DI. 1950. The water culture method for growing plants without soil. *California Agricultural Experiment Station, Circular-347*.
18. Upendri HF, Karunarathna B. Organic nutrient solution for hydroponic system. *Academia Letters*. 2021;1893:1-0. Available: <https://doi.org/10.20935/AL1893>.

19. Park Y, Williams KA. Organic hydroponics: A review. *Scientia Horticulturae*. 2024;324:112604.
20. Joshi D, Nainabasti A, Awasthi RB, Banjade D, Malla S, Subedi B. A review on soilless cultivation: The hope of urban agriculture. *Archives of Agriculture and Environmental Science*. 2022;7(3):473-81.
21. George P, George N. Hydroponics-(soilless cultivation of plants) for biodiversity conservation. *Int. J. Mod. Trends Eng. Sci*. 2016;3(06):97-104.
22. Khadijah B, King JHP, Ong KH. Producing black pepper (*Piper nigrum* L. cv.'Kuching') rootstock in a deep-water culture hydroponic system. *Borneo Journal of Resource Science and Technology*. 2021;11(2):88-97.
23. Nardi L, Metelli G, Garegnani M, Villani ME, Massa S, Bennici E et al. Farming for pharming: Novel hydroponic process in contained environment for efficient pharma-grade production of saffron. *Molecules*. 2022;27(24):8972.
24. Dewir YH, Alsadon A, Ibrahim A, El-Mahrouk M. Effects of growing substrate, mode of nutrient supply, and saffron corm size on flowering, growth, photosynthetic competence, and cormlet formation in hydroponics. *HortTechnology*. 2022;32(2):234-40.
25. Razan K, Aziz R, Al-Maari K, Al-Biski F. Studying the growth and productivity of saffron under different agriculture techniques. *Iraqi Journal of Agricultural Sciences*. 2023;54(1):242-52.
26. Fallahi HR. Evaluation of flowering potential of saffron corms in different weight groups under hydroponic conditions. *Journal of Saffron Research*. 2022;10(2):331-44.
27. Chong WX, Kiew WY, Low YC, Lim TH, Tan HY. The effects of hydroxyapatite nanoparticles on the growth and yield of ginger (*Zingiber officinale* Roscoe) from seedling to tillering stage. In IOP Conference Series: Earth and Environmental Science 2023 Feb 1 IOP Publishing. 1139(1): 012005.
28. Suhaimi MY, Mohamad AM, Mahamud S, Khadzir D. Effects of substrates on growth and yield of ginger cultivated using soilless culture. *Journal of Tropical Agriculture and Food Science*. 2012;40(2):159-68.
29. Naznin MT, Kitaya Y, Hirai H, Alsanusi BW. Sulfur fertilization enhances ajoene accumulation in hydroponically grown garlic. In XXX International Horticultural Congress IHC2018: II International Symposium on Soilless Culture and VIII International 1273 2018;1273:217-22.
30. Naznin MT, Kitaya Y, Shibuya T, Hirai H. Growth and ajoene concentrations in garlic plants cultured hydroponically with different aeration regimes. *The Journal of Horticultural Science and Biotechnology*. 2010;85(2):161-5.
31. Naznin MT, Kitaya Y, Shibuya T, Hirai H. Development of hydroponic culture system for producing garlic and determination of ajoene concentration. *Eco-Engineering*. 2009;21(4):147-52.
32. Abd-Elkader DY, Alkharpotly AA. Effect of nitrogenous concentration solutions on vegetative growth, yield and chemical characters of celery (*Apium Graveolens* L.). *Journal of Plant Production*. 2016;7(11):1201-6.
33. Ansari AA, Raghubeer H, Jaikishun S. The effect of vermiwash and vermicompost produced from leaves of *Azadirachta indica* and *Citrus aurantifolia* on the growth parameters of celery through hydroponic system. *International Journal of Tropical Drylands*. 2019;3(1):11-6.
34. Begum AA, Rao BM, Gowd MHP, Ismail T. Design and evaluation of low cost rectangular framed hydroponic system in leafy vegetables. *The Pharma Innovation Journal*. 2022;11(12):2391-3.
35. Silva MD, Soares TM, Gheyi HR, Costa IP, Vasconcelos RS. Growth, production and water consumption of coriander grown under different recirculation intervals and nutrient solution depths in hydroponic channels. *Emirates Journal of Food and Agriculture*. 2020;32(4):281-94.
36. Silva DMG, Costa IP, Alves LS, Soares TM, Gheyi HR. Coriander cultivation under different nutrient solution depths in hydroponic systems: a comparison between conventional DFT and adapted DFT with PVC pipes. *Water Resources and Irrigation Management-WRIM*. 2023;12(1-3):29-43.

37. Sowmya RS, Warke VG, Mahajan GB, Annapure US. Effect of amino acids on growth, elemental content, functional groups, and essential oils composition on hydroponically cultivated coriander under different conditions. *Industrial crops and products*. 2023;197:116577.
38. Guerrero-Lagunes LA, Ruiz-Posadas LD, Rodríguez-Mendoza MD, Soto-Hernández M, Castillo-Morales A. Thyme hydroponics crops effect (*Thymus vulgaris* L.) on essential oil quality and yield. *Revista Chapingo. Serie horticultura*. 2011;17(2):141-9.
39. Luna-Maldonado AI, Rodríguez-Fuentes H, Rodríguez-Ortiz JC, Vidales-Contreras JA, Márquez-Reyes JM, Flores-Breceda H. Cultivation of *Rosmarinus officinalis* in hydroponic system. *Journal of Experimental Biology and Agricultural Sciences*. 2016;4(6):637-43.
40. Rattan S, Partap M, Kumar S, Warghat AR. Nutrient feeding approach enhances the vegetative growth biomass, volatile oil composition, and myristicin content in hydroponically cultivated *Petroselinum crispum* (Mill.) Nyman. *Journal of Applied Research on Medicinal and Aromatic Plants*. 2022;26:100359.
41. Faruq H, Novelia E, Setyaningsih M, Nisa RA. The utilization of vegetable waste as a nutrient addition in hydroponic media for the growth of green mustard (*Brassica juncea* L.). In *IOP Conference Series: Earth and Environmental Science*. IOP Publishing. 2021;755(1):012078.
42. Muthu HD, Izhar TN, Zakarya IA, Saad FN, Ngaa MH. Comparative study between organic liquid fertilizer and commercial liquid fertilizer and their growth performances on mustard greens. In *IOP Conference Series: Earth and Environmental Science*. IOP Publishing. 2023;1135(1):012002.
43. Chrysargyris A, Nikolaidou E, Stamatakis A, Tzortzakis N. Vegetative, physiological, nutritional and antioxidant behavior of spearmint (*Mentha spicata* L.) in response to different nitrogen supply in hydroponics. *Journal of applied research on medicinal and aromatic plants*. 2017;6:52-61.
44. Danai-Christina A, Eirini S, Pavlos T, Paschalina C, Stefan M. Improvement of the quality in hydroponically grown fresh aromatic herbs by inducing mild salinity stress is species-specific. *Folia Horticulturae*. 2012;33(2):265-74.
45. Li Y, Heckman J, Wyenandt A, Mattson N, Durner E, Both AJ. Potential benefits of silicon nutrition to hydroponically grown sweet basil. *HortScience*. 2020;55(11):1799-803.
46. Maurer D, Sadeh A, Chalupowicz D, Barel S, Shimshoni JA, Kenigsbuch D. Hydroponic versus soil-based cultivation of sweet basil: impact on plants' susceptibility to downy mildew and heat stress, storability and total antioxidant capacity. *Journal of the Science of Food and Agriculture*. 2023;103(15):7809-15.
47. Son YJ, Park JE, Kim J, Yoo G, Nho CW. The changes in growth parameters, qualities, and chemical constituents of lemon balm (*Melissa officinalis* L.) cultivated in three different hydroponic systems. *Industrial Crops and Products*. 2021;163:113313.
48. Manukyan AE, Heuberger HT, Schnitzler WH. Yield and quality of some herbs of the Lamiaceae family under soilless greenhouse production. *Journal of applied botany and food quality*. 2004;78(3):193-9.
49. Manukyan A. Effect of growing factors on productivity and quality of lemon catmint, lemon balm and sage under soilless greenhouse production: I. Drought stress. *Medicinal and aromatic plant science and biotechnology*. 2011;5(2):119-25.
50. Farrokhi E, Samadi A, Rahimi A. Investigation of antioxidant activity, total phenol and flavonoid content of lemon balm (*Melissa officinalis* L.) in different media under hydroponic condition. *Eco-phytochemical Journal of Medicinal Plants*. 2021;8(4):19-33.
51. Souret FF, Weathers PJ. The growth of saffron (*Crocus sativus* L.) in aeroponics and hydroponics. *Journal of herbs, spices and medicinal plants*. 2000;7(3):25-35.
52. Sorooshzadeh A, Tabibzadeh Z. Effect of copper in hydroponic solution on leaves and roots of saffron (*Crocus sativus* L.). In *International Conference and Exhibition on Soilless Culture 1062* 2010;1062:83-88.

53. Stelluti S, Caser M, Demasi S, Herrero ER, García-González I, Lumini E et al. Beneficial microorganisms: A sustainable horticultural solution to improve the quality of saffron in hydroponics. *Scientia Horticulturae*. 2023;319:112155.
54. Abou-Hadid AF, El-Shinawy MZ, Omer EA. Cultivation of garlic in nutrient film technique (NFT). *Egyptian Journal of Horticulture*. 1998;25(3):271-280.
55. Naznin MT, Kitaya Y, Shibuya T, Endo R, Hirai H, Lefsrud MG. Ground based study on culturing garlic as a source of vegetable food and medicine in space-growth and ajoene accumulation in garlic plants cultured with different CO<sub>2</sub> regimes. *Biological Sciences in Space*. 2015;29:1-7.
56. Tuwar DA, Tuwar AR. Comparison in growth of hydroponic cultivated crop plants with normal soil cultivated crop plants by using non circulating culture and aggregate culture methods of hydroponics. *International Journal of Researches in Bioscience*. 2018;4(2):266-9.
57. Furlani PR, Silveira LC, Bolonhezi D, Faquin V. *Cultivo hidropônico de plantas*. Campinas: IAC; 1999.
58. Walters KJ, Lopez RG. Modeling growth and development of hydroponically grown dill, parsley, and watercress in response to photosynthetic daily light integral and mean daily temperature. *Plos one*. 2021;16(3):e0248662. Available: <https://doi.org/10.1371/journal.pone.0248662>.
59. Udagawa Y. Some responses of dill (*Anethum graveolens*) and thyme (*Thymus vulgaris*), grown in hydroponic, to the concentration of nutrient solution. *Hydroponics and Transplant Production* 396. 1994;396(24): 203-210.
60. Rodríguez-Fuentes H, Muñoz S, Alcorta-García ES. *El Tomate Rojo Sistema Hidropónico*, Editorial Trillas, México; 2011.
61. Khatun I, Karim S, Das SK, Hossen R. Onion cultivation approach by custom-made outdoor hydroponics: A very first attempt in Bangladesh. *Journal of Aridland Agriculture*. 2021;7:48-51.
62. Mouroutoglou C, Kotsiras A, Ntatsi G, Savvas D. Impact of the hydroponic cropping system on growth, yield, and nutrition of a Greek sweet onion (*Allium cepa* L.) landrace. *Horticulturae*. 2021;7(11):432.
63. Chrysargyris A, Panayiotou C, Tzortzakis N. Nitrogen and phosphorus levels affected plant growth, essential oil composition and antioxidant status of lavender plant (*Lavandula angustifolia* Mill.). *Industrial Crops and Products*. 2016;83:577-86.
64. Chrysargyris A, Petropoulos SA, Prvulovic D, Tzortzakis N. Performance of hydroponically cultivated geranium and common verbena under salinity and high electrical conductivity levels. *Agronomy*. 2021;11(6):1237.

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