

## **Effect of different nitrogen levels and plant growth regulators on stomatal resistance, leaf temperature and transpiration rate and yield of banana cv Ney Poovan**

### **Abstract**

The present Study was carried out to understand the effects of certain plant growth regulators under different nitrogen levels on different gas exchanging parameters viz., stomatal resistance, leaf temperature, transpiration rate and yield of banana cv. neypoovan. Different levels of nitrogen viz., 150 g N plant<sup>-1</sup> (45g + 75g + 30g N plant<sup>-1</sup> at 3, 5 and 7 MAP, respectively), M<sub>1</sub> + Urea 2% foliar spray, 200 g N plant<sup>-1</sup> (60g + 100g + 40g N plant<sup>-1</sup> at 3, 5 and 7 MAP respectively), M<sub>3</sub> + Urea 2% foliar spray and foliar spray of salicylic acid 100 ppm, mepiquat chloride 500 ppm, chlormequat chloride 1000 ppm, nitrobenzene 50 ppm, benzyl adenine 20 ppm and 25 ppm of 2, 4-D at 3<sup>rd</sup>, 5<sup>th</sup> and 7<sup>th</sup> month after planting were given and compared with untreated control. Salicylic acid treated leaves showed high stomatal resistance with low transpiration.

The yield components were favourably influenced by higher level of N application. Soil application of 200 g N plant<sup>-1</sup> and giving two per cent urea spray on 3<sup>rd</sup>, 5<sup>th</sup>, and 7<sup>th</sup> month after planting distinctly enhanced all the yield components. Salicylic acid spray at 100 ppm improved the number of hands, fingers and finger size. Benzyl adenine at 20 ppm spray also was found beneficial in increasing yield components. More number of fingers per bunch were observed in benzyl adenine treatment. With maximum fruit weight, salicylic acid spray surpassed benzyl adenine by recording maximum bunch weight of 12.58 kg (13.9 % increase over control), while benzyl adenine recorded 12.52 kg (13.4 % increase).

### **Key words**

Banana, growth regulators, Stomatal diffusiveresistance, Leaf temperature, Transpiration rate and yield

## **Introduction**

Banana is one of the important fruit crops of the world, especially in the tropics. The world produces 40 million tonnes of banana each year, but most of them are consumed locally. In India, to meet the increasing food demand of the ever increasing population, stepping up of fruit production is part of the strategy to solve the food crisis. And, banana has a dominant role to play among the fruit crops. India produces about 30 million tonnes of bananas from an area of 0.83 million ha. Among horticultural crops, contribution of banana to Agricultural Gross Domestic Product (AGDP) is the highest (Turner, 1990).

There is a need to focus on standardization of improved production technologies suitable for different systems of cultivation to realize potential yields in many commercial cultivars for targeted banana production. Selection of high-yielding varieties, planting of healthy, disease-free planting material, choosing the right planting density, need-based and timely application of inputs, viz., irrigation water and nutrients, maintenance of weed-free conditions, etc., are important to bridge the gap between actual yield and potential yield per unit area. (David, 2002). Farmers that grow bananas use nonscientific management techniques that result in inefficient use of fertilizers and water which results in low output. (Nisarga et al., 2022). Foliar feeding of nutrients provides a considerable scope not only for the effective utilization of nutrients but also to safeguard the economy of the farmer by improving the yield potential and quality of the produce (Sreekanth et al., 2017).

With a view to manipulate vegetative growth, source size and its activity by varying nitrogen levels, and to convert the increased biomass to yield advantage by improving the

assimilate translocation to developing sink by using various growth regulating chemicals, the present study has been taken up.

## **Materials and methods**

A field experiment in banana cv. Ney Poovan was conducted with various levels of nitrogen and plant growth regulators. The main plots are, M<sub>1</sub> (Control-150 g N plant<sup>-1</sup> (45g + 75g +30g N plant<sup>-1</sup> at 3,5 and 7 MAP, respectively), M<sub>2</sub> (M<sub>1</sub> + Urea 2% foliar spray), M<sub>3</sub> (200 g N plant<sup>-1</sup> (60g + 100g + 40g N plant<sup>-1</sup> at 3,5 and 7 MAP respectively), M<sub>4</sub> (M<sub>3</sub> + Urea 2% foliar spray).The sub plot treatments are, S<sub>1</sub>(Control (water spray), S<sub>2</sub> (Mepiquat chloride (MC) 500 ppm), S<sub>3</sub>(Chlormequat chloride (CCC) 100 ppm), S<sub>4</sub> (Ethrel 500 ppm), S<sub>5</sub> (Salicylic acid (SA) 100 ppm), S<sub>6</sub> (Nitrobenzene 100 ppm), S<sub>7</sub> (Benzyl adenine (BA) 20 ppm), S<sub>8</sub>(2,4-Dichlorophenoxy acetic acid (2, 4-D) 25 ppm).The leaf temperature, transpiration rate and stomatal diffusivity resistance were estimated during 3<sup>rd</sup>, 5<sup>th</sup> and 7<sup>th</sup> month after planting and at harvest stages of the crop with measuring procedure was given below:

### **Leaf temperature**

Leaf temperature was recorded between 10.00 A.M to 12.00 noon at 3<sup>rd</sup>, 5<sup>th</sup> and 7<sup>th</sup> month after planting and at harvest stage using Steady State Porometer (LICOR 1600, Licor Inc, Nebraska, USA) and expressed as °C.

### **Stomatal resistance**

Stomatal resistance was measured at 3,5<sup>th</sup> and 7<sup>th</sup> month after planting and at harvest by using Steady State Porometer between 10.00 AM to 12.00 noon (LICOR 1600, Licor Inc, Nebraska, USA) and expressed as s cm<sup>-1</sup>.

### **Transpiration rate**

Transpiration rate was measured by using Steady State Porometer (LICOR-1600, Licor Inc, Nebraska, USA) at 3<sup>rd</sup>, 5<sup>th</sup>, 7<sup>th</sup> month after planting and at harvest and expressed as  $\mu\text{g H}_2\text{O cm}^{-2} \text{ s}^{-1}$ .

### **Yield and yield components**

Five plants in each treatment were tagged for recording yield and yield components and the mean values of each treatment were worked out and expressed.

### **Bunch weight**

Bunch weight was recorded including the peduncle measuring 20 cm above the first hand and expressed in kg.

### **Number of hands and fingers per bunch**

Total numbers of hands and fingers in each bunch were counted and recorded. The mean values were expressed in number.

### **Finger weight**

Two middle fingers each from top and bottom rows of the second hand of each bunch were removed with a sharp knife at the point of pedicel attachment with the fruit and the individual finger weight was recorded. The mean value was worked out and expressed in g.

### **Fruit length**

Length of the fruit was measured from the base of the pedicel to the tip of the fruit along the outer curvature and expressed as cm.

### **Statistical analysis**

The statistical analysis of the data and the correlation co-efficients were done by adopting the standard procedure of Gomez and Gomez (1984).

## Results

### Stomatal diffusive resistance

Stomatal resistance in banana clearly established showed an increasing trend upto shooting stage. There existed significant differences in main plot and subplot treatments at all the stages. The interaction effects were also found to be significant at all the growth stages. Among the doses of nitrogen, M<sub>4</sub> was found to be poorer in stomatal resistance than M<sub>3</sub>, M<sub>2</sub> or M<sub>1</sub> by recording 12.7, 29.1 and 5.7 per cent lower values than control at 5, 7 MAP and at harvest stages.

Among the plant growth regulators, salicylic acid spray (S<sub>5</sub>) recorded values of 1.46, 5.18, 6.55 and 1.24 at 3, 5, 7 MAP and at harvest which were 19.7, 12.4, 14.3 and 49.4 per cent increase over control respectively. Benzyl adenine (S<sub>7</sub>) and 2, 4-D performed poorly with low values in most of the stages. The results indicated that salicylic acid and growth retardant chemicals tended to increase stomatal diffusive resistance, while nitrobenzene, benzyl adenine and 2,4-D appeared to reduce the resistance. (Table 1)

M<sub>1</sub>S<sub>1</sub> and M<sub>2</sub>S<sub>5</sub> recorded maximum values at 7 MAP (7.98) and at harvest (1.37), while M<sub>4</sub>S<sub>5</sub> recording high value at 3 MAP (1.59) and M<sub>3</sub>S<sub>4</sub> at 5 MAP (5.49). Very low values were mostly seen in M<sub>4</sub>S<sub>7</sub> and M<sub>4</sub>S<sub>8</sub> which recorded the lowest mean values of 2.66 by pooling observations of all the stages

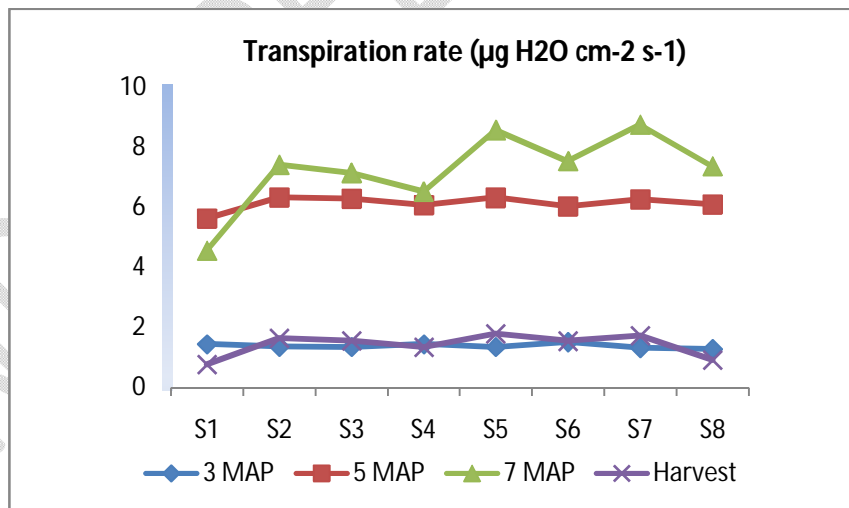
### Transpiration rate ( $\mu\text{g H}_2\text{O cm}^{-2} \text{s}^{-1}$ )

Data on transpiration rate revealed there were significant impacts of various plant growth regulators on transpiration rate at all growth stages. However, nitrogen levels failed to elicit any significant differences among themselves.

Among the subplots treatments, high transpiration rate was observed in control plants in all the chosen stages except at 3 MAP, when 2, 4-D and benzyl adenine treatments registered

enhanced rates over control. Almost all the growth regulator treatments distinctly recorded lower transpiration rate than control. Among them salicylic acid spray at 100 ppm recorded lowest rate at 3 and 7 MAP (21.8 and 21.4 per cent lower than control respectively), mepiquat chloride and CCC registered lower transpiration rate at 5 MAP and at harvest by 19.5 and 27.2 per cent respectively below the values of control. Among the growth regulators, nitrobenzene, benzyl adenine and 2,4-D showed high transpiration rate almost nearer to that of untreated control plants. (Fig 1 and table 2)

Significant interaction effects were observed in the stages of observation  $M_2S_8$  showed maximum transpiration rate at 3 MAP (8.44) which were comparable with that of  $M_3S_8$ . At 5 MAP and at harvest,  $M_1S_7$  revealed lower rates at 5 and 7 MAP; control recorded the highest rate (8.50), on the contrary,  $M_4S_5$  recorded lower rates at 5 and 7 MAP (6.91 and 6.14) respectively. Considering the mean values of all the stages,  $M_3S_8$  recorded the maximum value of 6.97, while  $M_1S_5$  recorded the lowest mean value of 5.10.



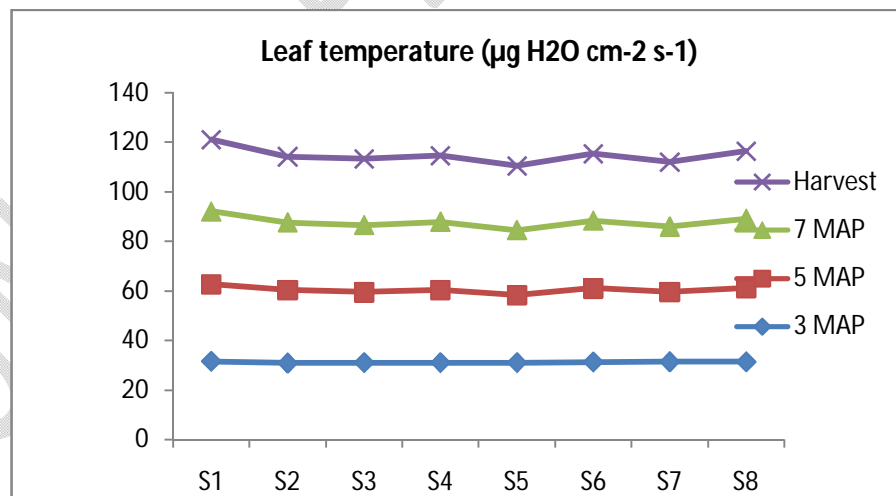
**Figure 1: Effect of different growth regulators and chemicals on transpiration at different growth stages**

## Leaf temperature (°C)

In banana, leaf temperature appeared to decrease consistently with advancement of age of the crop. Doses of nitrogen proved ineffective in producing significant variations during all the stages of crop growth. However, increasing dose of nitrogen decreased the leaf temperature marginally.

But, significant differences were observed among various plant growth regulators at all stages of crop growth. All the growth regulator treatments recorded the leaf temperature below that of control treatment and salicylic acid appeared to lower the leaf temperature consistently in all the stages. Mepiquat chloride and cycocel also reduced the temperature considerably. On the other hand, nitrobenzene (S<sub>6</sub>), benzyl adenine (S<sub>7</sub>) and 2, 4-D (S<sub>8</sub>) effected high leaf temperature among the chemicals tried. (Table 3 and Figure 2)

Distinctly, low leaf temperatures were observed in M<sub>4</sub>S<sub>2</sub>, M<sub>4</sub>S<sub>3</sub> and M<sub>4</sub>S<sub>5</sub> treatments among various interactions of main and subplot treatments. Main plot treatments in combination with control (S<sub>1</sub>) recorded uniformly higher leaf temperatures than other combinations.



**Figure 2: Effect of different growth regulators and chemicals on transpiration at different growth stages**

## Yield and yield components (Table 4)

Two levels of nitrogen as soil application with or without foliar application of urea were studied in banana cv. Ney Poovan. In each of the above main plot treatments, foliar spray of seven growth regulators in addition to untreated control were tried as sub plots. The effects of these treatments on yield and yield parameters are presented in Table 4.

Data on **finger length** revealed significant differences among main, sub plot and interaction treatments. Finger length ranged from 13.5 cm in  $M_1$  to 14.6 cm in  $M_3$ . Both  $M_3$  and  $M_4$  recorded comparable values with 8.1 and 7.4 per cent increased finger length over control. Foliar spray was effective at low nitrogen fertilization of  $150 \text{ g N plant}^{-1}$  ( $M_2$ ) with 5.2 per cent increased value over  $M_1$  (soil application of  $150 \text{ g N plant}^{-1}$ ). Foliar application of benzyl adenine ( $S_7$ ) excelled among the growth regulator treatments (15.0 cm) with 11.9 per cent increased fruit length over control. Salicylic acid ( $S_5$ ) and 2,4-D ( $S_8$ ) also produced increased fruit lengths by 11.2 and 10.4 per cent respectively over control.

By increasing nitrogen dose from 150 to  $200 \text{ g of plant}^{-1}$ , **finger girth** was increased by 5.52 per cent (from 12.7 in  $M_1$  to 13.4 in  $M_3$ ). However, giving foliar spray of urea in addition to soil application of N at  $150 \text{ g plant}^{-1}$  showed 5.5 per cent increase, and over  $200 \text{ g plant}^{-1}$  the foliar spray facilitated 10.2 per cent increase over control. Benzyl adenine spray ( $S_7$ ) recorded the maximum finger girths of 14.8 cm (22.3 % increase over control), which was closely followed by  $S_5$  with 14.6 cm (20%) increase. Ehrel produced to the least effect (6.6% increase) among the growth regulators. Benzyl adenine and 2, 4-D showed moderate effects in enhancing fruit girth.

The values of **number of hands** ranged from 8.2 in  $M_1S_1$  to 10.8 both in  $M_3S_5$  and  $M_4S_7$ . The differences were significant among growth regulator treatments and interactions. N levels failed to reveal significant differences. But,  $M_4$  recorded 10.2 hands per bunch, while  $M_1$  recording 9.7. Among the sub plots, number of hands ranged from 9.1 in control to 10.6 in  $S_5$ . Other than  $S_5$ ,  $S_7$  also performed better with 10.5 hands per bunch.

**Number of fingers** ranged from 148.5 in S<sub>1</sub> to 161.8 in S<sub>7</sub>. Benzyl adenine spray (S<sub>7</sub>) increased the number of fingers by 9.0 per cent over that of control. This was followed by S<sub>5</sub> and S<sub>8</sub> with 7.6 and 7.5 per cent increases over control. Though all the growth regulators have performed better than untreated control in terms of finger number, CCC and ethrel treatments showed only negligible increases over control treatment.

Among the interactions, M<sub>4</sub>S<sub>7</sub> performed well with maximum finger number of 168.6 followed by M<sub>4</sub>S<sub>8</sub> (165.4) and M<sub>4</sub>S<sub>5</sub> (165.2). The results indicated that the growth promoters, namely benzyl adenine and 2, 4-D or salicylic acid in collaboration with higher N dose might increase the finger number favourably.

The maximum bunch weight of 12.58 kg was recorded by giving foliar spray of salicylic acid at 100 ppm level, which resulted in 13.9 per cent increase over untreated control. Benzyl adenine treatment also performed better with 12.52 kg bunch weight and 13.4 per cent increase over control. 2, 4-D also revealed its usefulness by enhancing bunch weight upto 8.4 per cent. The other growth regulators revealed very low influence in increasing bunch weight. However, ethrel spray adversely affected the bunch yield by reducing the weight by one per cent below that of control.

As higher nitrogen level showed profound influence on bunch weight in the present study, the growth regulator treatments in combination with higher nitrogen levels also performed better. Among the interactions, M<sub>4</sub>S<sub>5</sub> recorded the highest bunch weight of 13.65 kg followed by M<sub>4</sub>S<sub>7</sub> (13.28 kg) and M<sub>3</sub>S<sub>7</sub> (13.16 kg). The results clearly indicated that 200 g N plant<sup>-1</sup> as soil application in addition to two per cent urea spray, and foliar spray of either 100 ppm salicylic acid or 20 ppm benzyl adenine might result in increased fruit yield in banana cv. Ney Poovan.

## **Discussion**

By increasing nitrogen dose or by giving urea spray, stomatal resistance was found to decrease (Tab.1). Among the growth regulators, salicylic acid was found more effective in increasing stomatal resistance, while benzyl adenine performed poorly with low values. Apart

from benzyl adenine, nitrobenzene and 2,4-D also showed low stomatal resistance. As stomatal resistance is an useful parameter in identifying drought resistant character, salicylic acid appears to be an useful tool for inducing resistance to drought. . Zhu et al. (2011) mentioned that increases in CO<sub>2</sub> concentrations will decrease stomata conductance, but carbon assimilation in the intercellular cell could be maintained at levels seen. Eris (1983) observed significant increase in stomatal resistance in pepper with salicylic acid treatment. Anitha (2003) reported promotary effect of cycocel treatment on stomatal resistance in banana leaves, which is in conformity with the present findings as all growth retarding chemicals including CCC (cycocel) maintained positive influence on stomatal resistance.

The effect of nitrogen application in altering transpiration was not noticed significantly (Fig.2 and Tab.2). However, all the growth regulators chosen for the present study showed their distinct impacts in lowering transpiration rate. Salicylic acid and mepiquat chloride were very effective in lowering rate of transpiration. The known growth promoters namely nitrobenzene, benzyl adenine and 2,4-D, on the other hand, failed to reveal any tangible effects. The effect of salicylic acid in reducing the rate of transpiration was confirmed in previous works (Saavedra, 1978).

Nitrogen produced only negligible effect on leaf temperature changes in banana. Salicylic acid, mepiquat chloride and cycocel were effective in reducing leaf temperature below that of control. On the other hand, nitrobenzene, benzyl adenine and 2,4-D had higher leaf temperature values. The reduced leaf temperature values in certain growth regulator treatments might be attributed to the maintenance of better water status in leaf tissues.(Robinson and Bower, 1988).

The ultimate aim of any farmer will be to get maximum fruit yield with quality. If all the growth and physiological parameters are favourably settled, then the cumulative response could be manifested in terms of high fruit yield.

Increase in **finger length** was favoured by higher dose of nitrogen but giving urea spray over and above 200 g N plant<sup>-1</sup> as soil application showed only negative influence on finger length. On the other hand, **finger girth** was substantially increased by urea spray in addition to soil application of N. Satheeshkumar (2002) also stated that urea spray tended to influence fruit circumference as compared to fruit length. Salicylic acid and benzyl adenine, which influenced many of the parameters in the present study, also revealed greater influence on finger girth than on finger length. Except ethrel, the other growth regulators tended to increase both finger length and finger girth. But, ethrel failed to show any influence on finger length, but had a moderate influence in increasing finger girth only.

Nitrogen levels showed only negligible influence on **number of hands per bunch**. But, the number was found high in higher dose of nitrogen. Ashok kumar and Shanmugavelu (1978), and Ramaswamy and Muthukrishnan (1974) obtained significant increases in number of hands by increasing N levels either by foliar application of urea or by soil application. Salicylic acid and benzyl adenine revealed greater influences in increasing the number of hands per bunch. Ethrel spray appeared to have a better influence on number of hands, while the treatment failed to show any positive effect on other yield components.

**Number of fingers** per bunch and **finger weight** decide the bunch weight and hence, are very valuable yield components in banana. The influence of nitrogen levels was more on finger weight than on finger number. However, maximum values in both the parameters were with high nitrogen dose. Giving urea spray, in addition to soil application, showed beneficial effect on these characters explicitly. Ramaswamy and Muthukrishnan (1974) observed significant correlation between leaf nitrogen and number of fingers. Chattopadhyay *et al*, (1980) also reported that number of fingers were increased significantly with N application. Among the growth regulators, the effect of benzyl adenine was in promoting finger number, while salicylic acid tended to increase finger weight. Ethrel which had very negligible influence on these two parameters.

The regulators had varying degrees of positive influences on these traits. Anitha (2003) recorded

increased fruit weight in cycocel and mepiquat chloride treated trees. Chattopadhyay *et al.* (1980) attributed increased fruit weight due to high N dose for obtaining greater bunch weights.

There was concomitant increase in **bunch weight** with increase in nitrogen dose. The results revealed the benefit of giving urea spray in addition to soil application, as was seen in bunch weight. Srikul and Turner (1995) observed increased fruit growth rate in banana with the increase in nitrogen dose. Several workers have reported increased bunch yield with high N levels at 200 g N plant<sup>-1</sup> or nearer to this dose (Ramaswamy and Muthukrishnan, 1974; Chattopadhyay *et al.*, 1980; Nalina, 2002). As a culmination of favourable effects of major yield components, namely number of hands, number of fingers and finger weight, obtained by salicylic acid treatment, the maximum bunch weight of 12.58 kg was recorded in this treatment. However, Anitha (2003) reported higher bunch weight in CCC treated plants than in salicylic acid treatment.

CCC treatment also increased the bunch weight by 3.8 per cent over that of control treatment, but the increase was not as high as that of salicylic acid or benzyl adenine, which are the better performing growth regulators as far as the present investigation is concerned.

## **Conclusion**

The findings of this research also showed that the leaf temperature, transpiration rate and stomatal diffusive resistance were greatly influenced by different nitrogen levels and plant growth regulators. Net photosynthesis and stomatal conductance revealed very high positive relationship with bunch yield, which were showing high rates at shooting stage in plants receiving nitrogen 200 g plant<sup>-1</sup> with urea two per cent foliar spray and salicylic acid 100 ppm as combined spray. The yield and yield components were enhanced by high nitrogen fertilization of 200 g N plant<sup>-1</sup> and foliar spray of urea. Salicylic acid and benzyl adenine spray produced further improvement in fruit yield. The enhancement in fruit yield by these treatments was by increasing finger number per bunch and finger weight. The results indicated that soil application of 200 g N

plant<sup>-1</sup> in three splits along with urea spray at two per cent concentration during third, fifth and seventh month after planting might be advocated for getting high fruit yield in banana cv. Ney Poovan. Additionally, by giving either salicylic acid spray at 100 ppm or benzyl adenine spray at 20ppm could further increase the yielding ability of the cultivar.

UNDER PEER REVIEW

**Table 1: Effect of different levels of nitrogen and plant growth regulators on stomatal resistance ( $s\ cm^{-1}$ ) at different growth stages of banana cv. Ney Poovan.**

Treatments	3 MAP		5 MAP		7 MAP		Harvest		Mean
<b>Main Plot</b>									
<b>M<sub>1</sub></b>	1.33	(0.0)	4.89	(0.0)	6.96	(0.0)	6.96	(0.0)	<b>5.04</b>
<b>M<sub>2</sub></b>	1.34	(0.8)	4.92	(0.6)	6.58	(-5.5)	6.58	(0.9)	<b>4.86</b>
<b>M<sub>3</sub></b>	1.28	(-3.8)	4.84	(-1.0)	5.90	(-15.5)	5.90	(-8.5)	<b>4.48</b>
<b>M<sub>4</sub></b>	1.43	(7.5)	4.27	(-12.7)	4.88	(-29.1)	4.88	(-5.7)	<b>3.87</b>
<b>Mean</b>	<b>1.34</b>		<b>4.73</b>		<b>6.08</b>		<b>6.08</b>		<b>4.56</b>
<b>CD (p=0.05)</b>	0.003		0.020		0.020		0.003		
<b>Sub Plot</b>									
<b>S<sub>1</sub></b>	1.22	(0.0)	4.61	(0.0)	5.73	(0.0)	1.12	(0.0)	<b>4.32</b>
<b>S<sub>2</sub></b>	1.46	(19.7)	4.96	(7.6)	6.22	(8.6)	1.68	(38.6)	<b>4.71</b>
<b>S<sub>3</sub></b>	1.41	(15.6)	4.90	(6.3)	6.48	(13.1)	1.78	(27.7)	<b>4.82</b>
<b>S<sub>4</sub></b>	1.33	(9.0)	4.76	(3.3)	6.72	(17.3)	1.38	(44.6)	<b>4.88</b>
<b>S<sub>5</sub></b>	1.46	(19.7)	5.18	(12.4)	6.55	(14.3)	1.24	(49.4)	<b>4.94</b>
<b>S<sub>6</sub></b>	1.29	(5.7)	4.44	(-3.7)	5.86	(2.3)	1.81	(24.1)	<b>4.36</b>
<b>S<sub>7</sub></b>	1.33	(9.0)	4.46	(-3.3)	5.71	(-0.3)	1.13	(0.0)	<b>4.30</b>
<b>S<sub>8</sub></b>	1.27	(4.1)	4.54	(-1.5)	5.37	(-6.3)	1.20	(4.8)	<b>4.13</b>
<b>Mean</b>	<b>1.34</b>		<b>4.73</b>		<b>6.08</b>		<b>6.08</b>		<b>4.56</b>
<b>CD (p=0.05)</b>	0.004		0.020		0.020		0.003		
<b>Interaction CD</b>									
<b>CD (p=0.05)</b>									
<b>M at S</b>	0.010		0.030		0.040		0.010		
<b>S at M</b>	0.010		0.030		0.040		0.010		

[Values in parentheses are per cent changes over respective control (M<sub>1</sub> and S<sub>1</sub>)]

**Table 2: Effect of different levels of nitrogen and plant growth regulators on leaf temperature (°C) different growth stages of banana cv. Ney Poovan (Main effect).**

<b>Treatments</b>	<b>3 MAP</b>		<b>5 MAP</b>		<b>7 MAP</b>		<b>Harvest</b>		<b>Mean</b>
<b>Main Plot</b>									
<b>M<sub>1</sub></b>	30.3	(0.0)	29.6	(0.0)	28.1	(0.0)	27.2	(0.0)	<b>28.8</b>
<b>M<sub>2</sub></b>	30.3	(0.0)	29.6	(0.0)	28.1	(0.0)	27.4	(0.7)	<b>28.9</b>
<b>M<sub>3</sub></b>	29.9	(-1.3)	29.6	(0.0)	27.8	(-1.1)	27.0	(-0.7)	<b>28.6</b>
<b>M<sub>4</sub></b>	29.3	(-3.3)	29.3	(-1.0)	27.9	(-0.7)	27.6	(1.5)	<b>28.5</b>
<b>Mean</b>	<b>30.0</b>		<b>29.5</b>		<b>28.0</b>		<b>27.3</b>		<b>28.7</b>
<b>CD (p=0.05)</b>	0.080		0.083		0.081		0.072		
<b>Sub Plot</b>									
<b>S<sub>1</sub></b>	30.7	(0.0)	30.7	(0.0)	29.4	(0.0)	29.0	(0.0)	<b>29.9</b>
<b>S<sub>2</sub></b>	29.0	(-5.5)	29.2	(-4.9)	27.2	(-7.5)	26.5	(-8.6)	<b>28.0</b>
<b>S<sub>3</sub></b>	29.5	(-3.9)	28.5	(-7.2)	27.2	(-7.5)	27.5	(-5.2)	<b>28.2</b>
<b>S<sub>4</sub></b>	29.6	(-3.6)	29.3	(-4.6)	27.4	(-6.8)	26.6	(-8.3)	<b>28.2</b>
<b>S<sub>5</sub></b>	29.1	(-5.2)	28.4	(-7.5)	26.9	(-8.5)	26.3	(-9.3)	<b>27.7</b>
<b>S<sub>6</sub></b>	30.6	(-0.3)	30.3	(-1.3)	28.7	(-2.4)	27.5	(-5.2)	<b>29.3</b>
<b>S<sub>7</sub></b>	30.4	(-1.0)	29.3	(-4.6)	28.1	(-4.4)	27.5	(-5.2)	<b>28.8</b>
<b>S<sub>8</sub></b>	30.6	(-0.3)	30.4	(-1.0)	29.0	(-1.4)	27.6	(-4.8)	<b>29.4</b>
<b>Mean</b>	<b>30.0</b>		<b>29.5</b>		<b>28.0</b>		<b>27.3</b>		<b>28.7</b>
<b>CD (p=0.05)</b>	0.080		0.089		0.087		0.079		
<b>Interaction CD</b>									
<b>CD (p=0.05)</b>									
<b>M at S</b>	0.184		0.184		0.171		0.165		
<b>S at M</b>	0.176		0.176		0.164		0.179		

[Values in parentheses are per cent changes over respective control (M<sub>1</sub> and S<sub>1</sub>)]

**Table 3: Effect of different levels of nitrogen and plant growth regulators on transpiration ( $\mu\text{g H}_2\text{O cm}^{-2}\text{s}^{-1}$ ) different growth stages of banana cv. Ney Poovan.**

Treatments	3 MAP		5 MAP		7 MAP		Harvest		Mean
<b>Main Plot</b>									
<b>M<sub>1</sub></b>	6.33	(0.0)	8.04	(0.0)	7.68	(0.0)	2.18	(0.0)	<b>6.06</b>
<b>M<sub>2</sub></b>	7.03	(11.1)	8.02	(0.2)	7.68	(0.0)	1.89	(-13.3)	<b>6.15</b>
<b>M<sub>3</sub></b>	6.86	(8.4)	8.10	(0.7)	7.35	(-4.3)	2.10	(-3.7)	<b>6.10</b>
<b>M<sub>4</sub></b>	6.72	(6.2)	8.07	(0.4)	7.37	(-4.0)	2.01	(-7.8)	<b>6.04</b>
<b>Mean</b>	<b>6.73</b>		<b>8.06</b>		<b>7.52</b>		<b>2.05</b>		<b>6.09</b>
<b>CD (p=0.05)</b>	0.002		0.001		0.001		0.005		
<b>Sub Plot</b>									
<b>S<sub>1</sub></b>	7.35	(0.0)	8.87	(0.0)	8.31	(0.0)	2.24	(0.0)	<b>6.69</b>
<b>S<sub>2</sub></b>	6.00	(-18.4)	7.17	(-19.5)	7.02	(-15.5)	1.65	(-26.3)	<b>5.46</b>
<b>S<sub>3</sub></b>	6.32	(-14.0)	7.31	(-17.6)	6.91	(-16.8)	1.63	(-27.2)	<b>5.54</b>
<b>S<sub>4</sub></b>	6.08	(-17.3)	7.43	(-16.2)	7.37	(-11.3)	2.09	(-6.7)	<b>5.75</b>
<b>S<sub>5</sub></b>	5.75	(-21.8)	7.50	(-15.4)	6.53	(-21.4)	1.71	(-23.7)	<b>5.37</b>
<b>S<sub>6</sub></b>	7.08	(-3.7)	8.54	(-3.7)	7.71	(-7.2)	2.18	(-2.7)	<b>6.38</b>
<b>S<sub>7</sub></b>	7.63	(3.8)	8.86	(-0.1)	8.22	(-1.1)	2.53	(12.9)	<b>6.81</b>
<b>S<sub>8</sub></b>	7.66	(4.2)	8.78	(-1.0)	8.09	(-2.6)	2.33	(4.0)	<b>6.71</b>
<b>Mean</b>	<b>6.73</b>		<b>8.06</b>		<b>7.52</b>		<b>2.05</b>		<b>6.09</b>
<b>CD (p=0.05)</b>	0.004		0.002		0.003		0.013		
<b>Interaction CD</b>									
<b>CD (p=0.05)</b>									
<b>M at S</b>	0.002		0.001		0.001		0.011		
<b>S at M</b>	0.003		0.001		0.003		0.011		

[Values in parentheses are per cent changes over respective control (M<sub>1</sub> and S<sub>1</sub>)]

**Table 4. Effect of different levels of nitrogen and plant growth regulators on yield and yield components of banana cv. Ney Poovan**

Treatments	Finger length (cm)	Finger girth (cm)	Number of hands/bunch	Number of fingers/bunch	Finger weight (g)	Bunch weight (kg)	[Values in parentheses are percentage changes over control (M <sub>1</sub> and S <sub>1</sub> )]
Main plot							
M <sub>1</sub>	13.5 (0.0)	12.7 (0.0)	9.7 (0.0)	152.3 (0.0)	50.1 (0.0)	10.70 (0.0)	
M <sub>2</sub>	14.2 (5.2)	13.4 (5.5)	10.0 (3.1)	155.7 (2.2)	53.6 (7.0)	11.51 (7.6)	
M <sub>3</sub>	14.6 (8.1)	13.4 (5.5)	10.0 (3.1)	154.2 (1.2)	56.6 (13.0)	12.20 (14.0)	
M <sub>4</sub>	14.5 (7.4)	14.0 (10.2)	10.2 (5.2)	158.9 (4.3)	55.4 (10.6)	12.36 (15.5)	
Mean	14.2	13.4	10.0	155.3	53.9	11.69	
CD (p=0.05)	0.10	0.09	NS	1.10	0.41	0.10	
Subplot							
S <sub>1</sub>	13.4 (0.0)	12.1 (0.0)	9.1 (0.0)	148.5 (0.0)	52.1 (0.0)	11.0 (0.0)	
S <sub>2</sub>	14.1 (5.2)	13.0 (7.4)	9.8 (7.7)	157.0 (5.7)	52.4 (0.6)	11.56 (3.8)	
S <sub>3</sub>	14.3 (6.7)	13.1 (8.3)	10.0 (9.9)	150.7 (1.5)	54.5 (4.6)	11.54 (4.5)	
S <sub>4</sub>	13.4 (0.0)	12.9 (6.6)	10.1 (11.1)	151.2 (1.8)	51.2 (-1.7)	10.93 (-1.0)	
S <sub>5</sub>	14.9 (11.2)	14.6 (20.7)	10.6 (11.6)	159.8 (7.6)	57.2 (9.8)	12.58 (13.9)	
S <sub>6</sub>	14.1 (5.2)	13.1 (8.3)	9.9 (8.8)	153.4 (3.3)	53.1 (1.9)	11.51 (4.3)	
S <sub>7</sub>	15.0 (11.9)	14.8 (22.3)	10.5 (11.5)	161.8 (9.0)	55.6 (6.7)	12.52 (13.4)	
S <sub>8</sub>	14.8 (10.4)	13.4 (10.7)	9.8 (7.7)	159.7 (7.5)	55.1 (5.8)	11.97 (8.4)	
Mean	14.2	13.4	10.0	155.3	53.9	11.69	
CD (p=0.05)	0.10	0.09	0.06	0.95	0.34	0.09	

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