

Seed Morphology and Tetrazolium Quick Seed Viability Test of Malabar Neem (*Melia dubia* Cav.) under Laboratory Condition

ABSTRACT

Melia dubia Cav. commonly known as Malabar Neem, is an economically important fast-growing indigenous multipurpose tree species of India. It has a wide range of applications from medicinal utilities such as treating pain, fever, and infections, to agricultural and industrial uses viz., fodder, seed oil, resin, fuel wood, timber and its wood being utilized in the paper, matchbox, and plywood industries. Malabar Neem generally is distributed in tropical and sub-tropical regions of India and introduced in many countries of South Africa, the Middle East, North and South America, Brazil, Bermuda, Argentina, Southern Europe, Southeast Australia, and Asia Pacific regions. Nevertheless, it is a most promising tree species which is highly suitable for agroforestry or farm forestry with a life cycle of 8 to 12 years that gaining economic importance both in domestic and global markets and plantations in degraded lands. Consequently, the demand of Melia seedlings among Indian farmers has been increased, however the poor seed viability and very low seed germination hampered the good quality nursery production and large-scale plantations. Before seed sowing in nursery, there is need to determine the actual seed viability and to know the required seed quantity of Melia to be sown in forest nursery. Therefore, the present research study was conducted at Dr. Y. S. Parmar University of Horticulture and Forestry, Nauni, Solan, Himachal Pradesh of India to study the seed morphology and seed viability testing protocol using tetrazolium (TZ) test. The seeds of Melia species stained with 0.5, 1.0, 1.5 and 2.0 % solution of TZ and incubated at 30°C temperature for 24 hours. The tetrazolium test performed with 100 seeds in 4 replications in CRD design and analyzed with OPSTAT statistical software. The length, width and weight of fruit, drupe and seed were recorded in the range of 9.40-12.97 mm, 6.55-9.70 mm & 2.84-2.99 g; 5.84-14.32 mm, 2.76-9.72 mm & 1.672-1.798 g; and 1.2-1.98 mm, 1.02-2.98 mm & 3.12-3.74 g, respectively. The colour of fruit was yellow with oval shape having smooth surface. The result of TZ test showed that the seeds of Malabar Neem soaked in 0.5 % tetrazolium solution at 30 °C temperature for 24 hours found to be the highest seed viability percentage (40.25 %) which is recommended as a quick seed viability protocol in *M. dubia*. Overall, the results of the present research study fulfilled all the gaps to test quick seed viability of a seed lot before seed sowing in nursery for accurate knowledge of the seed requirement in quantity for the purpose of quality seedling production in nursery.

Keywords: *Melia dubia*, Agroforestry tree species, Seed Morphology, Seed Viability, Tetrazolium Test, Nursery Seedlings

INTRODUCTION

Melia dubia Cav. commonly known as Malabar Neem, is a member of the Meliaceae family. It is an industrially and economically important fast growing indigenous multipurpose tree species of India, which can be harvested on a short rotation period [1]. It is a deciduous tree which can grow upto 30 m height. The leaves are 2-3 pinnate, imparipinnate, estipulate, attenuate, rachis 10-30 cm long, slender, swollen at base, terete, scurfy tomentose when young; pinnae 3-7 pairs, 10-20 cm long; leaflets 2-11 on each pinna, opposite, estipulate; petiolule 3-10 mm long, slender; leaf lamina 4.5-9 x 2-4 cm in size, ovate-lanceolate, acute, base oblique, obtuse, round or attenuate, apex acuminate, margin crenate, glabrous at maturity. Flowers bisexual, 5-6 mm long in size, greenish-white. Fruit is a drupe, 2 x 1 cm in size, dorsally compressed with longitudinal ridges, yellow in ripen colour, fleshy; seeds 1 - 3 or 4, 1 cm in size [2]. Malabar Neem generally is distributed in tropical moist deciduous forests in the states of India such as Tamil Nadu, Kerala, Karnataka, Himalayas, Sikkim, North Bengal, Upper Assam, Khasi Hills, Gujarat, Hills of Orissa, Deccan plateau, Nallamalai Hills and Western Ghats at an altitude ranging between 400 and 1800m above mean sea level [2, 3]. This species is native to Southern Asia, and has been introduced in many countries in South Africa, the Middle East, North and South America, Brazil, Bermuda, Argentina, Southern Europe, Southeast Australia, and Asia Pacific regions [4].

The wood of Malabar Neem tree is used majorly in plywood industry and screened out as an alternate species for pulpwood [5]. The wood of *Melia* is also used for packing cases, pencils, match box sticks, photo frames, mini furniture like stools, benches, wooden tables, interior decoration, window, doors, wooden racks, musical instruments, tea powder boxes, cigar boxes, wooden building, ceiling planks, agricultural implements etc. [6]. Malabar Neem is one of the fastest growing trees in the world and considered as a money-spinning tree of short rotation due to its high demand in pulpwood, plywood and timber industries [3]. It is a good fuel and fodder yielding tree species. Its uses range from medicinal applications, such as treating pain, fever, and infections, to agricultural and industrial uses, with its seeds as a source of neem oil and its wood being utilized in the paper, matchbox, and plywood industries [7]. Malabar neem is one such alternative species suitable for timber, plywood, pulpwood and fuel wood [8]. Thus, the species has a ready and assured market due to its multifarious utilities. The species is also highly adaptable in different environment conditions.

Nevertheless, Malabar Neem is a most promising tree species which is highly suitable for agroforestry or farm forestry with a life cycle of 8 to 12 years that gaining economic importance both in domestic and global markets [1]. Consequently, there is good demand of *Melia* seedlings among farmers in Indian subcontinent. But the poor seed viability and very low seed germination hampered the good quality nursery production and large-scale plantations of *Melia* in India [9]. The low seed viability and prolonged germination period resulted uneven germination and poor-quality seedlings. Therefore, there is a need of precise seed viability determination before to seed sowing in nursery which has a great importance for the reproduction of the species. The accuracy of required quantity of seeds of *Melia* to be sown in forest nursery is mainly dependent upon the viability of the seeds. One of the most accepted and reliable method of tree seed viability is the tetrazolium test [10]. Historically, a topographical method or tetrazolium test (TZ) of seed viability evaluation was developed first for agricultural cereal crops [10], later on conifer tree species [11] and many more crops [12]. Seed dormancy is very common in most of the forest tree species [13,14]. However, TZ test provides a quick information on seed viability of any crop species even it is in a state of seed dormancy [15]. In addition, there is very scanty information available on seed viability testing in *Melia dubia* using tetrazolium test. Seed morphology is also important to identify the good quality seeds by measuring seed dimensions, seed weight, colour, shape and surface of fruit and seeds. A comparative research study of seed morphology in response to desiccation tolerance and physiological parameters in 71 Eastern Australian rainforest species was conducted by Hamilton et al. (2013) for the purpose of long-term biodiversity conservation [27]. Keeping all the points in mind, the present experiment was conducted in Tree Seed Technology Laboratory with two primary objectives viz., first to study the drupe and seed morphological characteristics of Malabar Neem, and second to evaluate the seed viability of Malabar Neem using quick tetrazolium test.

EXPERIMENTAL DETAILS

Seed collection and extraction

Fruits of Malabar Neem tree was collected from the experimental farm of forestry, College of Horticulture and Forestry, Neri, Hamirpur, Himachal Pradesh, India during the year of 2024 at latitude (31.695790 N), longitude (76.4677291 E) and altitude (620 m from MSL) as geolocation presented in Figure 1. The recorded data of mother tree were recorded and presented in Table 1. After the collection of fruits, they were sun dried for 2-3 days so that the fruit pulp gets dried and removed. This process is known as depulping. The pulp removed and then seeds (kernels) were extracted from the drupes (Figure 2). Then, seeds exposed for surface sterilization and dried, this is known as seed drying. Further, kernels were mixed thoroughly to maintain homogeneousness in the seed lot as per ISTA rules [28].



Figure 1. Geolocation of fruit collection of *Melia dubia* for the present research study

Table 1: Recorded data of Malabar Neem mother tree

Tree Height (m)	DBH (cm)	Girth (cm)	CBH (m)	Crown Length (m)	Crown Width (m)
9.5	20.70	65	5.5	4	3.5



Figure 2. Extracted drupes from fruits after removing pulp and extracted seeds or kernels from the dried drupes of *Melia dubia*

Recording of Fruit, Drupe and Seed (Kernel) data

The length and width of fruits, drupes and kernels were measured with the help of Vernier Calliper (Mitutoyo) in millimetre. The Weight of fruits, drupes and kernels were taken with the help of Analytical Balance (Fisher Scientific). The colour, shape and surface of fruits, drupes and kernels were recorded by visual observation with the help of standard Munsell Colour Chart.

TetrazoliumSeed Viability Testing

The tetrazolium test (TZ) was employed with four replications of 100 seeds from the seed lot of *M. dubia*. Seeds of the Malabar Neem were stained in 0.5, 1.0, 1.5 and 2.0% solution of tetrazolium test prepared in the double distilled water. The seeds of *Melia* were placed in the Petridishes in four replications to incubate with tetrazolium solution at 30°C temperature in the darkness for 24 hours (Figure 3).

The chemical 2,3,5-Triphenyl Tetrazolium Chloride (TTC) enters into the living embryo of the seed which produce a reddish or dark purple colour due to water insoluble compound called formazan by the activity of dehydrogenase enzyme [16]. After completion of incubation time, the Tetrazolium solution was drained out and seeds of *Melia* were washed in distilled water. Then, red- or purple-coloured seeds were counted as viable seeds and non-coloured seeds as non-viable seeds (Figure 4).



Figure 3. Dipping of excited seeds of *Melia dubia* in tetrazolium solution

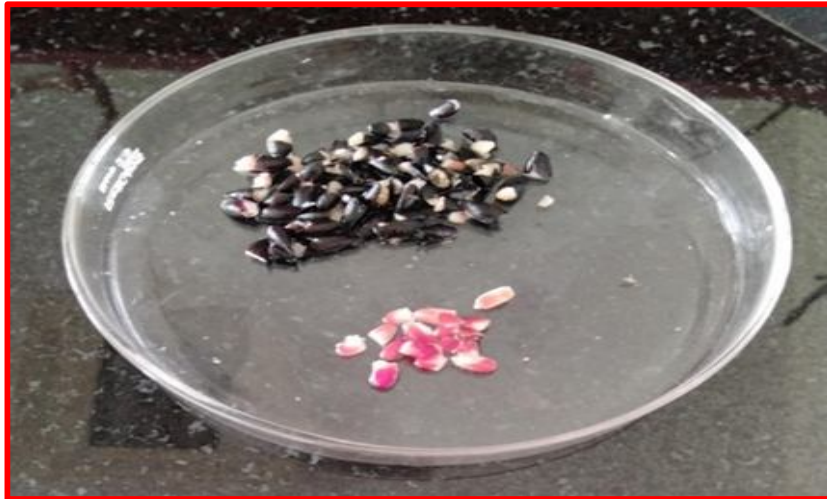


Figure 4: Data recording of purple-coloured seeds as viable and non-coloured seeds as non-viable seeds of *Melia dubia*

Statistical Analysis

The collected experimental data in the laboratory condition were analysed in CRD with four replications by using OPSTAT statistical software package [17] for appropriate interpretation of data and ANOVA was constructed. Then, treatment means were compared at 5 % level of significance to find out the best tetrazolium concentration for seed viability in *Melia* as per suggested per Gomez and Gomez [18].

RESULTS AND DISCUSSION

Fruit, Drupe and Seed Morphology

Fruit length of Malabar Neem was found in the range of 9.40 to 12.97 mm with the mean of 11.24 mm. Fruit width was recorded in the range (6.55-9.70 mm) with the mean of 8.35 mm. Fruit weight was recorded in the range between 2.84 to 2.99 g with the mean of 2.964 g. The colour of fruit was yellow with oval shape having smooth surface (Table 2).

Table 2: Fruit characters of *Melia dubia*

Characters	Range	Mean
Fruit Length (cm)	9.40-12.97 mm	11.24 mm
Fruit Width (cm)	6.55-9.70 mm	8.35 mm
Fruit Weight (g)	2.84-2.99 g	2.964 g
Fruit Colour	Yellow	-
Fruit Shape	Oval	-
Fruit Surface	Smooth	-

Drupe length of Malabar Neem was found in the range of 5.84-14.32 mm with the mean of 9.53 mm. Drupe width was recorded in the range of 2.76-9.72 mm with the mean 5.16 mm. Drupe weight was recorded in the range between 1.672 to 1.798 g with the mean of 1.734 g. The colour of drupe was whitish yellow with ovoid-ellipsoid shape having **asperous** surface. The number of seeds per drupe was observed in the range of 1 to 4 having a mean value 2.2 (Table 3).

Table 3: Drupe characters of *Melia dubia*

Characters	Range	Mean
Drupe Length (cm)	5.84-14.32 mm	9.53 mm
Drupe Width (cm)	2.76-9.72 mm	5.16 mm
Drupe Weight (g)	1.672-1.798 g	1.734 g
Drupe Colour	Whitish Yellow	-
Drupe Shape	Ovoid-ellipsoid	-
Drupe Surface	Asperous	-
No. of seeds/ Drupe	1-4	2.2

Seed (Kernal) length of Malabar Neem was found in the range of 1.2-1.98 mm with the mean 1.42 mm. Seed width was recorded in the range (1.02-2.98 mm) with the mean of 1.84 mm. Seed weight of 100 seeds was recorded in the range between 3.12 to 3.74 g with a mean value of 3.45 g. The colour of seed was black with narrow fusiform shape having smooth surface (Table 4).

Table 4: Seed (Kernal) characters of *Melia dubia*

Characters	Range	Mean
Seed Length (mm)	1.2-1.98 mm	1.42 mm
Seed Width (mm)	1.02-2.98 mm	1.84 mm
100 Seed Weight (g)	3.12-3.74 g	3.45 g
Seed Colour	Black	-
Seed Shape	Narrow Fusiform	-
Seed Surface	Smooth	-

Suresh and Devakumar (2017) categorized fruits of Malabar Neem into three categories viz., small, medium and big size for the study of seed morphology [19]. They reported fruit length (small-27.50 mm, medium-31.75 mm and big-36.84 mm), fruit width (small-19.85 mm, medium-24.76 mm and big-27.89 mm), and fruit weight (small-9.68 g, medium-12.66 g and big-17.45 g) which was higher than the result of present study due to climatic conditions. They also reported seed length (small-14.25 mm, medium-14.36 mm and big-14.48 mm), seed width (small-3.64 mm, medium-3.74 mm and big-3.86 mm) and seed weight (small-0.16 g, medium-0.20 g and big-0.23 g) which was also higher than the result of present study due to climatic conditions. Malabar Neem was introduced in College of Horticulture and Forestry, Neri, Hamirpur to know the growth performance in the subtropical region of Himachal Pradesh at 680 m elevation height. It is also noticed that Malabar Neem tree is producing twice flowering and fruiting in a single year which divide the energy to produce flowerings due to climatic conditions. Therefore, the size of fruit is lower in the Northern India than the reported size by Suresh and Devakumar (2017) in Southern India.

Seed Viability

Analysis of variance showed significant differences ($p < 0.05$) among treatments of tetrazolium concentrations (Table 4). For imbibed seeds of Malabar Neem in tetrazolium solution, the lowest concentration (0.50 %) of tetrazolium solution was observed the highest (40.25 %) of viable and vigorous seeds, however the lowest seed viability percent (26.50 %) was recorded in 2.00 %

tetrazolium solution (Table 5& Figure 5). In case of 1.00 % TZ solution, the seed viability percent was recorded 30.25 % and 28.25 % seed viability in 1.50 % TZ solution. When the concentration of tetrazolium solution was increased then the seed viability percentage was decreased (Table 6).

Table 5: Analysis of variance (ANOVA) in CRD for seed viability of *Melia dubia*

Source of Variation	DF	Sum of Squares	Mean Squares	F-Calculated	Significance
Treatments	3	454.19	151.40	14.28	0.001
Error	12	127.25	10.60		
Total	15	581.44			

Table 6: Seed viability of *Melia dubia*

Treatments	Mean	S.E.
0.50 % TZ Solution	40.25	2.29
1.00 % TZ Solution	30.25	1.11
1.50 % TZ Solution	28.25	1.65
2.00 % TZ Solution	26.50	1.19
Mean	31.31	
SEm (±)	1.63	
C.D.	5.07	
C.V.	10.40	

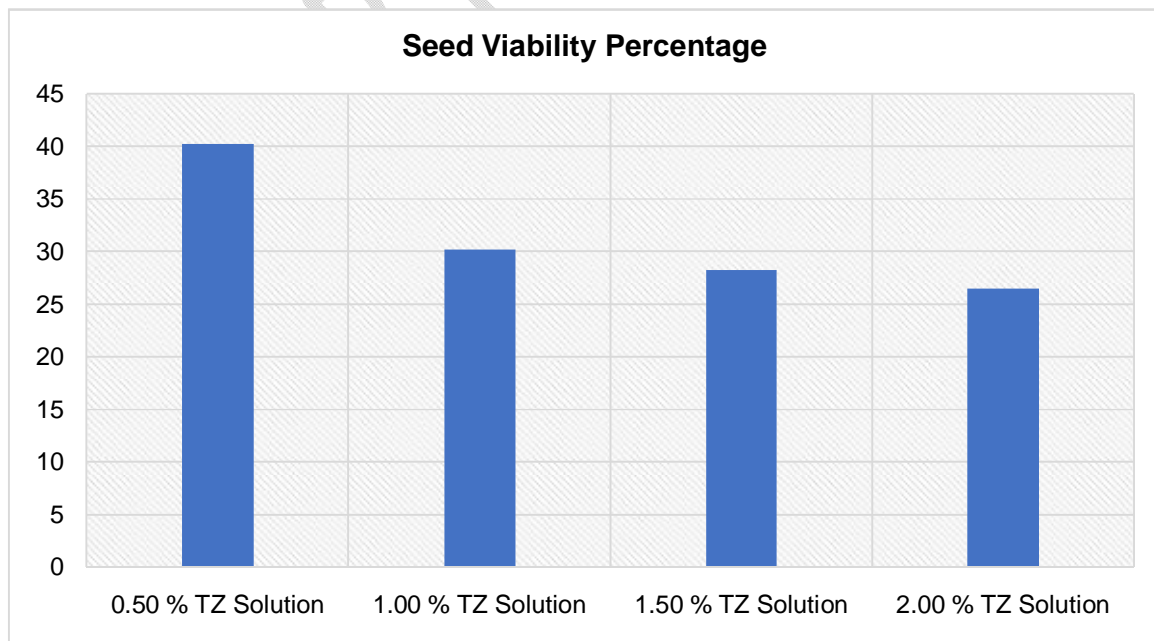


Figure 5: Seed viability of *Melia dubia*

Ajay *et al.* (2022) reported that GA₃ @ 100 ppm-soaked kernels of Malabar Neem for 24 hours significantly found to be the maximum seed viability percentage (41.68) with 0.1 % tetrazolium test with compare to control (20.87 %) [20]. This confirmed from the result of the present study that the seed viability ranged between 26.50 % to 40.25 % with a mean value of 31.31 % seed viability. Phartyal *et al.* (2003) reported that the seeds of *Ulmus wallichiana* took stain very lightly when put in low concentration of TZ (0.1%) for a short duration (4-8 hours) and the colour intensity increased with an increase in temperature, incubation time and concentration of TZ solution. They found that the seed soaked at 40°C for 24 hours in 1.0 and 0.5% solution of TZ showed 100% seed viability in *Ulmus wallichiana* [21]. Similarly, Aslam *et al.* (2010) studied the efficacy of tetrazolium test in determining the seed viability of *Pinus wallichiana* against the germination test. The percentage seed viability and laboratory germination recorded 79.82% and 82.69%, respectively which has further proved that tetrazolium test is a rapid, very effective and valuable research technique to know the seed viability of any crop [22]. The seeds of *Pinus roxburghii* soaked in 0.5 % tetrazolium solution at 25°C temperature for 24 hours was observed the highest seed viability percentage (64.75 %) which is recommended and standardized quick seed viability protocol in the chir pine species [23]. In various tree species, the tetrazolium test standardized by different scientists for quick seed viability testing such as 0.01 % TZ solution in *Swietenia macrophylla* [24]; 0.5 % TZ solution in *Moringa oleifera* [25]; 0.5 % TZ solution in *Platymiscium floribundum*; 0.1 % TZ solution in *Lonchocarpus muehlbergianus*; 0.5 % TZ solution in *Acacia polyphylla* [26]. A quick seed viability test was performed on four tropical forest tree species viz., *Albizia lebbek*, *Acacia nilotica*, *Azadirachta indica* and *Casuarina equisetifolia* of India and a positive correlation was found between seed viability percentage estimated Tetrazolium Test by with germination test [29]. Various seed viability tests viz., triphenyle tetrazolium chloride test, excised embryo test, electrical conductivity, and germination test were performed on seeds of Neem belongs to Meliaceae family obtained from mother tree of different age classes and the consistency was observed with the TTC (triphenyle tetrazolium chloride) and EC test in germination of seed in laboratory as well as nursery condition [30]. Thus, the results from the present research study validate to the previous research studies conducted by various scientists across the globe for different forest tree species and confirmed its suitability for prediction of seed viability in Malabar Neem.

CONCLUSION

From the present research, it can be concluded that the seeds of *Melia dubia* should be soaked in 0.5 % tetrazolium solution at 30°C temperature for 24 hours to test quick seed viability of a seed lot before seed sowing in nursery for accurate knowledge of the seed requirement in quantity for the purpose of quality seedling production.

Disclaimer (Artificial intelligence)

Author(s) hereby declares that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

REFERENCES

1. Dhaka RK, Gunaga RP, Sinha SK, Thakur NS, Dobriyal MJ. Influence of tree height and diameter on wood basic density, cellulose and fibre characteristics in *Melia dubia* Cav. families. *Journal of the Indian Academy of Wood Science*. 2020;17:138-44.
2. Brandis D. Indian Trees. International Book Distributors, Dehradun, India. 1906.
3. Warriar RR. *Melia dubia* Cav. Money Spinning Series - 2. Institute for Forest Genetics and Tree Breeding, Coimbatore. 2011: 1-16 p.
4. Mabberley DJ. A monograph of *Melia* in Asia and the Pacific. The history of white cedar and Persian lilac. *Gard. Bull. Sing.* 1984;37: 49-64.
5. Parthiban KT, Bharathi AK, Seenivasan R, Kamala K, Rao MG. Integrating *Melia dubia* in agroforestry farms as alternate pulpwood species. *Asia-Pacific Agroforestry Newsletter*. 2009;34:15-19.
6. Yadav D, Sahoo G, Wani AM. Growth performance and variability studies in different half sib families of *Melia dubia* under greenhouse condition. *Journal of Pharmacognosy and Phytochemistry*. 2019;8(2):1008-11.

7. Sukhadiya ML, Thakur NS, Gunaga RP, Patel VR, Bhuvra DC, Singh S. *Melia dubia* Cav. drupe pulp: a new alternate livestock feed resource. *Range Management and Agroforestry*. 2019;40(2):299-305.
8. Valentina P, Ilango K, Kiruthiga B, Parimala MJ. Preliminary phytochemical analysis and biological screening of extracts of leaves of *Melia dubia* Cav. *International Journal of Ayurveda and Pharma Research*. 2013;4(3):417-419.
9. Tilakaratna D. Pretreatment for the seed of lunumidella (*Melia dubia* Cav.). *Sri Lanka Forester*. 1991;20(1/2): 27-28.
10. Lakon G. Topographical detection of viability of cereal seeds with tetrazolium salts. *Ber. Deut. Bot. Ges*. 1942;60:299-305.
11. Lakon G. The topographical tetrazolium method for determining the germinating capacity of seeds. *Plant physiology*. 1949;24(3):389-94.
12. Wood CB, Miles S, Rix C, Terry J, Daws MI. The effect of seed oil content on viability assessment using tetrazolium: a case study using 171 species. *Plant Genetic Resources Newsletter*. 2005;143:17.
13. Viswanath S, Singh RP, Thapliyal RC. Seed germination patterns in a Himalayan moist temperate forest. *Tropical Ecology*. 2002;43(2):265-73.
14. Dhaka RK, Jha SK. Evaluation of five teak (*Tectona grandis* LF) provenances for germination test to find out reasons of low germination. *International Journal of Pure and Applied Bioscience*. 2017;5(5):1420-6.
15. Gaspar S, Nagy J. A new method for tetrazolium analysis of wheat seeds without embryo excision. *Seed Science and Technology*. 1981;9:553-56.
16. Wortmann S, Kutta RJ, Nuernberger P. Monitoring the photochemistry of a formazan over 15 orders of magnitude in time. *Frontiers in Chemistry*. 2022;10:983342.
17. Sheoran OP, Tonk DS, Kaushik LS, Hasija RC, Pannu RS. Statistical software package for agricultural research workers. Recent advances in information theory, statistics & computer applications by DS Hooda & RC Hasija Department of Mathematics Statistics, CCS HAU, Hisar. 1998;8(12):139-43.
18. Gomez KA, Gomez AA. Statistical procedures for agricultural research. John Wiley & Sons; 1984.
19. Suresh T, Devakumar AS. Morphological characterization of *Melia dubia* seeds: implications to germination. *Mysore Journal of Agricultural Sciences*. 2017;51(3):721-725.
20. Ajay, Arya S, Nanda K, Redhu I, Bhupender. Effect of Pre-sowing Kernel Treatments on Viability, Germination and Seedling Performance of Malabar neem (*Melia dubia* Cav.). *Biological Forum- An International Journal*. 2022;14(3):1690-1693.
21. Phartyal SS, Thapliyal RC, Nayal JS, Joshi G. Assessment of viability of *Acer caesium* and *Ulmus wallichiana* seeds through the tetrazolium test. *Seed science and technology*. 2003;31(3):691-700.
22. Aslam M, Reshi ZA, Siddiqi TO. Standardization of seed viability protocol for *Pinus wallichiana* AB Jackson in Kashmir, India. *International Journal of Pharmaceutical Sciences Review and Research*. 2010; 4(3):93-98.
23. Dhaka RK, Thakur P, Kaler NS, Sharma SD, Negi C, Brahmi MK. Standardization of Quick Seed Viability Protocol for *Pinus roxburghii* Sarg. Using Tetrazolium Assay. *Journal of Advances in Biology and Biotechnology*. 2024;27(5):41-50.
24. Quintana YG, Naranjo RA, Crespo YA, Morán HR. Optimisation of tetrazolium concentration and immersion time in the viability test of *Swietenia macrophylla* seeds by using response surface methodology. *Mathematical and Computational Forestry and Natural Resource Sciences*. 2019;11(2):257.
25. Pallavi HM, Madalageri MB, Vishwanath K, Biradar IB, Thattimani M. Decortication to enhance seed value and standardization of quick viability test in drumstick (*Moringa oleifera* L.). *Environment and Ecology*. 2015;33(4):1558-61.
26. Masullo LS, Piña-Rodriguez FC, Figliolia MB, Américo C. Optimization of tetrazolium tests to assess the quality of *Platymiscium floribundum*, *Lonchocarpus muehlbergianus* and *Acacia polyphylla* DC. seeds. *Journal of Seed Science*. 2017;39:189-97.
27. Hamilton KN, Offord CA, Cuneo P, Deseo MA. A comparative study of seed morphology in relation to desiccation tolerance and other physiological responses in 71 Eastern Australian rainforest species. *Plant Species Biology*. 2013;28(1):51-62.

28. ISTA. International Seed Testing Association. International rules for seed testing. Rules-1999. 1999.
29. Vanangamudi K, Mani G, Umarani R. Tetrazolium Viability Testing–A Reliable Quick Viability Test for Some Tropical Tree Species. In IUFRO Joint Symposium on Tree Seed Technology, Physiology and Tropical Silviculture, College, Laguna (Philippines), 30 Apr-3 May 2001;179-185.
30. Kumar D. Assessment of seed viability and vigour in neem (*Azadirachta indica* A. Juss.). Journal of Forest and Environmental Science. 2013;29(4):282-91.

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