

# Inhibitory effect of aqueous extract of *Sacoglottisgabonensis* on weight gain in Swiss mice exposed to aspartame

## ABSTRACT

**Aim:** To evaluate the **Inhibitory effect of aqueous extracts of *Sacoglottisgabonensis* on weight gain in Swiss mice exposed to aspartame.**

**Study Design:** The study was a completely randomized design employing relevant statistical tools for analysis and interpretation.

**Place and Duration of Study:** The study was carried out in the Department of Animal and Environmental Biology, Rivers State University. The experiment lasted for 90 days between April and June 2022.

**Methodology:** A total of 30 adult male Swiss mice (mean weight  $23.07 \pm 0.12$ g) were divided into six groups of five animals each. Group A was the negative control which received neither *Sacoglottisgabonensis* nor Aspartame, group B was the positive control which received 50mg/kg/bw/day of Aspartame only, group C received 50mg/kg/bw/day of Aspartame with 250mg/kg/bw/day of aqueous bark extract of *S. gabonensis*, group D received 50mg/kg/bw/day of Aspartame with 250mg/kg/bw/day of aqueous leaf extract of *S. gabonensis*, while groups E and F received 50mg/kg/bw/day of Aspartame with a combination of 250mg/kg/bw/day and 500mg/kg/bw/day of extracts of *S. gabonensis* bark and leaf extracts respectively. The experimental animals were weighed twice weekly for 30, 60 and 90 days duration and recorded to the nearest 0.01g. Mean weight from each duration was taken and subjected to statistical analysis using Statistical Analyses System SAS 9.4

**Result.** There was significant increase ( $p < 0.05$ ) in weight in the 30, 60 and 90-days exposure periods in group B. The groups that received the combination of aspartame and *S. gabonensis* particularly at higher doses showed a significant decrease in weight across the exposure periods when compared to the group that received aspartame only and the control group.

**Conclusion:** This result suggests that *S. gabonensis* reduced aspartame-induced weight gain, indicating its potential benefits for weight management. Further research is needed to explore the underlying mechanisms and health implications.

**Keywords:** **Bodyweight, extract, *Sacoglottisgabonensis*, sweeteners, toxicant**

## Introduction

Majority of people all over the world consume foods containing one artificial sweetener or the other [1]. "Artificial sweeteners are used worldwide as sugar substitutes in dietary soda, chewing gum, ice cream, breakfast cereal and most sugar free foods. It is also used in food

products including dry beverage mixes, chewable multi-vitamins, breakfast, puddings and fillings, carbonated beverages, refrigerated and non-refrigerated ready to drink beverages, yogurt type products and pharmaceuticals” [2,3]. Aspartame is an artificial (non-nutritive) sweetener used to replace sugar in food and drinks [4]. It is an odourless, white crystalline powder with a clean, sweet taste without any aftertaste or cooling effect [5]. “Aspartame is one of the most widely used artificial sweeteners in the world. It is found in more than 6000 products” [6]. The accepted daily recommended dose by the FDA is 50mg/Kg body weight/day while the accepted or/and approved daily dose by the World Health Organization (WHO) is 40mg/kg/day [7]. “Although there is concern suggesting possible adverse neurological and behavioural effects due to aspartame’s metabolic components phenylalanine, aspartic acid (aspartate), diketopiperazine and methanol, which are produced during its breakdown [8]. Aspartame represents 62% value of the intense sweetener marketed and consumed worldwide in over 100 countries” [9,10]. “Majority of the populace in underdeveloped nations rely on common, affordable, and culturally acceptable plants (leaves, stem, bark, roots) to cure a variety of ailments and diseases” [9]. These plants such as *Acanthus montanus* [11], *Lepidium meyenii* [12], *Stelleria media* [13], *Sacoglottisgabonensis*[14], and many others are natural resources explored, accepted and thought to have fewer adverse effects. *Sacoglottisgabonensis* a tree found in the tropical rainforest region of Africa and America is commonly used as an additive to palm wine, a local alcoholic brew which is an exudate from the phloem of *Raphia* species and *Elaeisguinensis* [15,16]. *S. gabonensis* extract is used as a stimulant, pain killer, the sap is medicinal and prolongs the shelf-life of fresh palm wine, . The stem bark extract is taken to treat fever, diarrhoea, gonorrhoea, abdominal pain, and as a spice in food to induce heat in nursing and pregnant mothers in Sierra Leone [17,18]. The stem bark and the leaf extract reduced altered electrolytes concentrations in Swiss mice [19,20]. It is reported to have aphrodisiac properties. [21]. Several reports have linked aspartame to weight gain, obesity, increased appetite. [22] reported increased body weight and fat mass mainly due to an increase in energy efficiency in rats exposed aspartame and sucralose while [23] also suggested a long-term intakes of aspartame, saccharin or diet soda may increase adipose tissue deposition

and risk of incident obesity independent of diet quality or caloric intake. [24] reported that BMI is positively associated with the consumption of diet carbonated beverages and people who regularly consume artificial sweeteners are at increased risk of excessive weight gain, metabolic syndrome, type 2 diabetes and cardiovascular disease [25]. Therefore, there is need to evaluate and source for medicinal plant to help reduce the overall weight gain of individuals with incessant consumption of aspartame.

## 2. Materials and Methods

A total of 30 adult male swiss mice (mean weight  $23.07 \pm 0.12$ g) were used for the study. All experimental animals were allowed to acclimatize for two weeks during which animals were allowed access to clean water and rodent pellet *ad libitum* before the commencement of treatments. The experiment lasted for 90 days. Dried leaves and bark of *S. gabonensis* were homogenized using kitchen blender. A tincture of both extracts was administered to the experimental animals at the dosage of 250 and 500 mg/kg/bw/day. 50mg of aspartame was used according to the recommendation of the FDA. The animals were divided into six groups of five each. Group A was the negative control and so received water and feed only. Group B was the positive control which received 50mg /kg/bw/day of aspartame only, group C received 50mg/kg/bw/day of aspartame with 250mg/kg/bw/day of bark extract of *S. gabonensis*, group D received 50mg/kg/bw/day of aspartame with 250mg of leaf extract of *S. gabonensis*, while groups E and F received 50mg/kg/bw/day of aspartame each with 250mg/kg/bw/day and 500mg/kg/bw/day of combined extracts of *S. gabonensis* respectively. All treatments were administered by oral gavage and followed the institutional animal care and treatment protocols laid down as approved guidelines for the ethical treatment of laboratory animals and at the Rivers State University. The weight of the experimental animals were taken twice weekly for 30, 60 and 90 days using a digital balance (Model PCE-BTS 15) and recorded to the nearest 0.01g. The experimental data were subjected to descriptive statistics that include means (M), standard deviation (SD), and standard error (SE) of means. Data management and statistical analyses were conducted using Statistical Analyses System SAS 9.4.

## 3. Results

### 3.1 Effects of aspartame and *Sacoglottisgabonensis* on Body weight of Swiss Mice.

The effects of aspartame and *S. gabonensis* on the mean body weights of each group of the experimental animals exposed for 30 days is shown in table 1. There was a significant difference ( $p < 0.05$ ) in weekly weights of mice within treatment groups. A significant ( $p < 0.05$ ) increase was observed within the first 30 days of the experiment in group B that received aspartame only. However, there was no significant ( $p > 0.05$ ) difference in weekly weight of mice from the group D (that received aspartame and bark extract of *S. gabonensis*), group E (that received aspartame 50mg/kg/bw/day and 250mg/kg/bw/day of combined extract of *S. gabonensis*) when compared to the control group at ( $p < 0.01$ ) and group F (that received aspartame and 500mg/kg/bw/day of combined extract (bark and leaf) of *S. gabonensis*)

Table 1: Comparisons of Weekly Weights of Swiss Mice after 30 Days administration of Aspartame and *Sacoglottisgabonensis*

	Body Weight (g)			
	Week 1	Week 2	Week 3	Week 4
	Mean ± SE	Mean ± SE	Mean ± SE	Mean ± SE
<b>Group A</b>	24.99±0.34	26.68±0.12	29.35±0.75	29.09±0.83 <sup>b</sup>
<b>Group B</b>	24.04±1.25	27.93±2.38	30.76±2.07	32.27±1.97 <sup>a</sup>
<b>Group C</b>	23.73±0.83	25.76±0.78	29.13±1.71	29.38±1.32 <sup>b</sup>
<b>Group D</b>	23.62±2.57	25.36±3.41	26.87±2.80	29.25±4.04 <sup>b</sup>
<b>Group E</b>	24.12±1.41	26.84±1.39	28.58±1.22	29.78±1.13 <sup>b</sup>
<b>Group F</b>	23.39±0.94	25.76±1.10	26.28±1.69	28.43±1.70 <sup>ab</sup>
<b>F- ratio</b>	2.15	0.54	2.43	6.79
<b>P-value</b>	0.1720 <sup>ns</sup>	0.6695 <sup>ns</sup>	0.1405 <sup>ns</sup>	0.0137 <sup>s</sup>

\*Values with the same superscripts are not significantly different @ $p>0.05$ . However, Values with different superscripts are significantly different @  $p<0.05$ .

The comparisons of weekly weights of Swiss mice after treatment with aspartame and *Sacoglottisgabonensis* (week 5-8) is presented in Table 2. There was increase in weight gain but no statistical significant difference in weekly mean weight values of all the experimental animals.

Table 2: Comparisons of weekly weights of Swiss mice after treatment with Aspartame and *Sacoglottisgabonensis* (Weeks 5-8)

Treatment	Weight (g)			
	Week 5	Week 6	Week 7	Week 8
	Mean ± SE	Mean ± SE	Mean ± SE	Mean ± SE
<b>Group A</b>	29.47±2.25 <sup>ab</sup>	30.76±2.47 <sup>c</sup>	30.46±1.50 <sup>c</sup>	30.76±0.95 <sup>c</sup>
<b>Group B</b>	34.07±0.85 <sup>a</sup>	34.96±0.08 <sup>a</sup>	35.65±0.44 <sup>a</sup>	39.93±0.18 <sup>a</sup>
<b>Group C</b>	29.24±0.60 <sup>ab</sup>	29.77±0.96 <sup>bc</sup>	29.76±1.40 <sup>bc</sup>	29.66±1.88 <sup>c</sup>
<b>Group D</b>	30.52±0.89 <sup>b</sup>	29.78±0.62 <sup>bc</sup>	29.86±1.28 <sup>bc</sup>	30.34±1.47 <sup>c</sup>
<b>Group E</b>	31.80±3.99 <sup>b</sup>	31.51±3.99 <sup>c</sup>	31.91±3.59 <sup>b</sup>	31.14±3.69 <sup>b</sup>
<b>Group F</b>	31.97±1.18 <sup>b</sup>	32.39±0.75 <sup>b</sup>	32.23±0.92 <sup>b</sup>	32.17±2.27 <sup>b</sup>
<b>F-Ratio</b>	<b>2.67</b>	<b>3.59</b>	<b>2.70</b>	<b>2.91</b>
<b>P-value</b>	<b>0.0431<sup>s</sup></b>	<b>0.059<sup>s</sup></b>	<b>0.02619<sup>s</sup></b>	<b>0.0302<sup>s</sup></b>

\*SE: Standard error, s=Not Significant ( $p>0.05$ ).

The comparisons of weekly weight of Swiss mice after treatment with aspartame and *Sacoglottisgabonensis* between week 9 to 12 showed significant ( $p<0.05$ ) difference in weight gain from week 10 to week 12, while there was non-significant( $p>0.05$ ) difference in weight gain in week 9 across treatment groups.

Table 3 Comparative analysis of Weekly Weights of Swiss Mice after Treatment with Aspartame and *Sacoglottisgabonensis* (Weeks 9-12).

Weight (g)
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Treatment	Week 9 Mean ± SE	Week 10 Mean ± SE	Week 11 Mean ± SE	Week 12 Mean ± SE
Group A	31.26±0.20 <sup>ab</sup>	32.85±0.45 <sup>ab</sup>	32.12±0.40 <sup>a</sup>	33.28±0.69 <sup>a</sup>
Group B	40.22±1.19 <sup>a</sup>	40.54±0.63 <sup>bc</sup>	41.09±1.13 <sup>c</sup>	41.91±0.47 <sup>ab</sup>
Group C	30.07±0.63 <sup>b</sup>	31.00±0.63 <sup>bc</sup>	31.47±0.65 <sup>a</sup>	32.59±0.91 <sup>bc</sup>
Group D	31.76±1.45 <sup>ab</sup>	30.05±0.46 <sup>c</sup>	29.82±0.72 <sup>a</sup>	30.78±0.91 <sup>c</sup>
Group E	32.32±0.89 <sup>ab</sup>	32.96±0.69 <sup>ab</sup>	32.50±0.42 <sup>ab</sup>	32.86±0.10 <sup>bc</sup>
Group F	32.70±0.55 <sup>ab</sup>	35.23±0.35 <sup>a</sup>	34.06±0.63 <sup>a</sup>	34.84±0.61 <sup>b</sup>
<b>Test Statistics: F-Ratio</b>	2.254	9.330	10.223	8.141
<b>P-value</b>	0.0215 <sup>s</sup>	0.0008 <sup>***</sup>	0.0005 <sup>***</sup>	0.0015 <sup>**</sup>

SE: Standard error, Significance Level: \*\*= $p < 0.01$ ; \*\*\*= $p < 0.0001$ ; ns=Not Significant ( $p > 0.05$ ). Superscript with the same alphabet showed non significant difference while Superscripts with different alphabet showed significant difference.

#### 4. Discussion

Changes in body weight is major criterion used for the evaluation of potential systemic toxicity of harmful chemicals and toxicants in living system [26]. There was no significant ( $p > 0.05$ ) difference in weight across treatment groups from the initial weight to the final weight within the 30days exposure period in the groups that received the both leaf extract and 50mg/kg/bw/day of aspartame and 250mg/kg/bw of *S. gabonensis* bark extract compared with the control group. The groups that received aspartame along with 500mg/kg/bw combined bark and leaf extract of *S. gabonensis* and group B which received aspartame only showed no significant difference in weekly weight gain

Administration of aspartame and *Sacoglottisgabonensis* to experimental mice from weeks 5-8 is shown in table 2. There is no significant difference ( $p > 0.05$ ) in weight in group C compared to group A. However, a significant increase occurred in group B administered aspartame only compared with other groups coadministered *S. gabonensis* and the control. The body weights of Swiss mice after treatment with aspartame and *Sacoglottisgabonensis* in groups D, E and F showed no significant increase compared to the control group. This trend was also seen after 90days administration,

Group B, the positive control group had the highest weight gain which was statistically significant while the other groups had lower weights despite coadministration with either leaf, bark or combination of both extracts. This result implies that aspartame influenced weight gain by inducing hyperphagia while the other treatment groups and negative control group still maintained steady decrease compared with group B. The extracts of *S. gabonensis* reduced weight gain when compared with group B administered aspartame only.

Similar observation was made by [27] who reported increased weight gain and total feed intake in rats administered Monosodium glutamate alone. [28,29] also recognized various medicinal plants and herbal decoction of *Hibiscus rosa sinensis* flower and *Zingiber officinale* as a natural product source and therapeutically effective against overweight and obesity due to fewer side effects in comparison to synthetic drugs while [11, 13, 30] reported no significant effect on body weight of animals exposed to *Acanthus montanus*, dry leaf tincture of *Stelleria media* and sub-chronic concentrations of cypermetrin.

The impact of aspartame on body weight in Swiss mice suggests that, at least in short-term studies, aspartame does not consistently lead to weight gain but in a long term consumption. Moreso, this result indicates that the effects of aspartame might be influenced by multiple factors, including dosage and duration of exposure. Administration of the extracts of *S. gabonensis* to experimental mice has proved to be beneficial as it maintained a steady weight throughout the experimental period.

#### **4. Conclusion**

Comparison of weight gain of Swiss mice exposed to aspartame and *Sacoglottisgabonensis* for 30-, 60- and 90-days exposure periods strongly suggest that aspartame influenced feed intake that led to the observed increase in weight in the group that received aspartame alone, while the groups that received the extracts of *S.gabonensis* showed slow but steady growth, which significantly decreased compared to group B. This implies that *S. gabonensis* moderated weight gain in experimental mice and can serve as an additive for those who are at risk of developing increased body weight due to environmental factors and genetic predispositions.

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- 2.
- 3.

#### **Ethical Approval**

Animal Ethic committee approval has been collected and preserved by the author(s)

## Reference

1. Whitehouse, C. R., Bisn, R.N., Boullata, J. & McCauley, L. A. (2008). The potential toxicity of Artificial Sweeteners. *AAOHN J.* 56 (6), 251-261.
2. Ashok, I., Sheeladevi, R. & Wankhar, D. (2013). Long term effect of aspartame (Artificial Sweetener) on membrane homeostatic imbalance and histopathology in the rat brain. *Free Radicals and Antioxidants*, 3, S42-S49.
3. Choudhary, A. K. & Lee, Y. Y. (2018). The debate over neurotransmitter interaction in aspartame usage. *Journal of Clinical Neuroscience*, 7(9), 77-100.
4. Choudhary, A. K. & Sheela Devi, R. (2014). Serum biochemical responses under oxidative stress of aspartame in wistar albino rats. *Asian Pacific Journal of Tropical Disease*, 4, 403-410.
5. Saleh, A. A. S. (2015). Anti-neuroinflammatory and antioxidant effects of N-acetyl cysteine in long-term consumption of artificial sweetener aspartame in the rat cerebral cortex. *The Journal of Basic & Applied Zoology*, 72, 73–80.
6. Bazzaz, A. A. & Al-Johani, N. S. (2018). Acute impact of Artificial Sweetener, Aspartame on Blood Parameter in Mice. *Advances in Bioscience and Biotechnology*, 9, 549-560.
7. Humphries, P., Pretorius, E. & Naude, H. (2008). Direct and indirect cellular effects of aspartame on the brain. *European Journal of Clinical Nutrition*, 62, 451–462.
8. Ashok, I., Wankhar, D., Sundareswaran, L., Sambantham, S., Ravindran, R. & Sheeladevi, R. (2017). Disruption of redox homeostasis in liver function and activation of apoptosis on consumption of aspartame in folate deficient rat model. *Journal of Nutrition and Intermediary Metabolism*, 8, 41-50.
9. Ashok, I., Wankhar, D., Wankhar, W. & Sheeladevi, R. (2015). Neurobehavioural changes and activation of neurodegenerative apoptosis on long-term consumption of aspartame in the rat brain. *Journal of Nutrition & Intermediary Metabolism*, 2, 76-85.
10. Maduka, H.C.C. & Okoye, Z.S.C. (2002). Elemental composition of *Sacoglottisgabonensis* A Nigerian Alcohol Beverage Additive. *Pakistan Journal of Biological Sciences*, 5 (1), 66-68.
11. Orlu, E.E. & Obulor, A.O. (2014). Investigation on the effect of aqueous leaf extract on *Acanthus montanus* on spermatogenesis in swiss mice. *Journal of Pharmacy and Biological Science*, 9(3), 44-49.
12. Ojo, O.A., Oyinloye, B.E., Ajiboye, B.O, Ojo, A.B., Akinlayo, C.O., Okozie, B. (2014). Dichlorvos induced Nephrotoxicity in rat kidney: protective effects of *Alstonia boonei* stem bark extract. *International Journal of Pharmacognosy*, 1(7), 429-437.
13. Iboroma, M., Orlu, E.E., Ebere, N. & Obulor, A.O. (2018). Androgenic and antioxidant activity of *Stellaria media* on rat following sub-chronic exposure to Dichlorvos. *Journal of Pharmacy and Biological Science*, 13(6), 38-46.
14. Wekhe-Emenike, A., Obulor, A. O. & Orlu, E. E. (2022). Influence of *Sacoglottisgabonensis* Ethanolic Extract on the Electrolytes of Swiss Mice Administered Aspartame. *Asian Journal of Biochemistry, Genetics and Molecular Biology*. 11(4), 35-42.
15. Maspalma, G .A., Fariku, S., Manu, J. M. & Ajide, M. A. (2013). Effect of *Sacoglottisgabonensis* (Urban humiraceae) stem bark extract, a palm wine additive on the rabbit jejunum. *Journal of Natural Product and Plant Resources*, 3 (2), 52-56
16. Dounias, E. (2015). *Sacoglottisgabonensis* (Baill) Urb. Protabase Record display [www.prota.org](http://www.prota.org). 01/03/2022.

17. Morah, F. N. I. & Robinson, I. G. (2015). SacoglottisGabonensis as a Potential Preservative for Palm-Wine. American Scientific Research Journal for Engineering, Technology, and Sciences, 13 (1), 97-101.
18. Titus, L.K., Orlu, E.E. & Obulor, A.O. (2019). Evaluation of the therapeutic role of *Citrullus lanatus* and *Annona muricata* fruit extracts on Cyhalothrin-induced toxicity. Journal of Advances in Biology & Biotechnology, 22(4), 1-10.
19. Wekhe-Emenike,A., Orlu, E.E & Obulor, A. O. (2022).Duration Dependent Impact of Aspartame and Sacoglottisgabonensis on the Liver of Swiss Mice. Journal of Advances in Biology & Biotechnology. 25(4), 39-49.
20. Ragi, M-E. E. El-Haber, R.El-Masri F. & Obeid, O. A. (2022). The effect of aspartame and sucralose intake on body weight measures and blood metabolites: role of their form (solid and/or liquid) of ingestion. British Journal of Nutrition (2022), 128, 352–360.
21. Pang MD, Goossens GH and Blaak, E. E. (2021). The Impact of Artificial Sweeteners on Body Weight Control and Glucose Homeostasis. Frontiers in Nutrition. 7, 598340.
22. Ramos-García, M.Ble-Castillo, J.L. García-Vázquez, C.Tovilla-Zárate, C.A. Juárez-Rojop,I.E. Olvera-Hernández, V.Genis-Mendoza, A.D. CórdovaUscanga, R. Álvarez-González, C.A.Díaz-Zagoya, J.C. (2021). Effects of Non-Nutritive Sweeteners on Energy Intake, Body Weight and Postprandial Glycemia in Healthy and with Altered Glycemic Response Rats. Foods, 10, 958.
23. Steffen,B.T., Jacobs,D. R. Yi,S-Y Lees,S. J. Shikany,J. M. Terry,J. G. Lewis,C. E. Carr,J.J.Zhou, X. and Steffen, L. M. (2023). Long-term aspartame and saccharin intakes are related to greater volumes of visceral, intermuscular, and subcutaneous adipose tissue: the CARDIA study. International Journal of Obesity. 47, 939 – 947.
24. Swithers, S.W. (2013). Artificial sweeteners produce the counter intuitive effect of inducing metabolic derangements. Trends Endocrinology metabolism. 24(9)
25. Forshee, R.A & Storey, M.I.(2003). Total beverage consumption of beverage choices among children and adolescent. Int. J. of food Science and Nutrition, 54(4),297-307.
26. Obulor, A.O & Orlu, E.E (2019). Protective role of lycopene on hormonal profile and & post testicular functions of male rats exposed to sublethal doses of cypermethrin. J.ofAdevnces on Biology and Biotechnology, 21(4), 1-9.
27. He, K., Zhao, L., Daviglius, M.L., Dyer, A.R., Horn, L., Garside, D. & Stamler, J. (2008). Association of Monosodium Glutamate Intake with Overweight in Chinese Adults: The INTERMAP Study. Obesity, 16(8), 1875–1880.
28. Iftikhar, N., Hussain, A. I., Chatha, S. A. S., Sultana, N., & Rathore, H. A. (2022). Effects of polyphenol-rich traditional herbal teas on obesity and oxidative stress in rats fed a high-fat–sugar diet. Food Science& Nutrition, 10, 698–711.
29. Gomathi, N., Malarvili, T., Mahesh, R., & Begum, V. H. (2008). Lipids low-ering effect of Hibiscus rosa-sinensis flower petals on monosodiumglutamate (MSG) induced obese rats. Pharmacology, 1, 400–409
30. Orlu, E.E. & Obulor, A.O. (2021). Impact of Coenzyme Q10 on hormonal profile in male sprague-dawley rats exposed to sub-chronic concentrations of cypermethrin. Asian Journal of Biology, 12(3), 1-9.