

## Original Research Article

### N-Acetyl Cysteine and Zinc Sulfate Attenuates Acute Crude Oil-Induced Oxidative Stress and Testicular Structural Damage in Male Wistar Rats

#### ABSTRACT

**Introduction:** Crude oil toxicity is caused by the production of reactive oxygen species (ROS), leading to oxidative stress and tissue damage. Both natural and synthetic antioxidants have shown potential in reducing the impact of this stress. This study evaluated the protective effects of N-Acetyl Cysteine (NAC) and Zinc Sulfate ( $ZnSO_4$ ) in alleviating testicular cytoarchitecture damage and oxidative stress in male Wistar rats after short-term exposure to Bonny light crude oil (BLCO)

**Materials and Methods:** Fifty male Wistar rats (weighing 180–200g) were used in the study and divided into ten groups of five animals each. Group I served as the control, receiving distilled water, while Group II, the negative control, received BLCO (600mg/kg). Groups III and IV were given 100 and 200mg/kg NAC, respectively. Groups V and VI received BLCO (600mg/kg) along with 100 and 200mg/kg NAC, respectively. Similarly, Groups VII and VIII were administered 0.5mg/kg and 1mg/kg  $ZnSO_4$ , while Groups IX and X received BLCO (600mg/kg) combined with 0.5mg/kg and 1mg/kg  $ZnSO_4$ , respectively. All treatments were administered orally via an orogastric cannula for three weeks. Testicular tissue samples were stained with haematoxylin and eosin to obtain photomicrographs of the tissues. They were also homogenized for the assay of malondialdehyde (MDA), superoxide dismutase (SOD), glutathione peroxidase (GPx), glutathione (GSH) and total antioxidant capacity (TAC) by the method of ELISA.

**Results:** Exposure to BLCO resulted in a significant increase in MDA and GSH levels in BLO-exposed animals compared to control ( $P < 0.05$ ) which were mitigated by co-treatment with NAC and  $ZnSO_4$ . However, BLCO exposure caused a significant decrease in CAT and TAC levels compared to controls ( $p < 0.05$ ), but this decrease was significantly mitigated in rats co-treated with NAC and  $ZnSO_4$ . Histological analysis of the testes from rats exposed to BLCO revealed notable alterations in testicular morphology, including interstitial exudates, and atrophic and degenerated seminiferous tubules which indicate testicular degeneration. However, this was improved on supplementation with NAC and  $ZnSO_4$ .

**Conclusion:** This study demonstrates that BLCO-exposed rats experienced toxicity affecting both testicular antioxidants and testicular structure. Our findings suggest that NAC and  $ZnSO_4$  protected and improved testicular function in BLCO-exposed rats by thus enhancing testicular antioxidant levels and cytoarchitecture.

**Keywords:** Bonny Light Crude Oil, N-Acetyl Cysteine, Zinc Sulfate, Testicular dysfunction, Antioxidants

## **INTRODUCTION**

Environmental pollution from petroleum products is a significant issue in crude oil-producing regions, posing serious health risks to both aquatic and terrestrial animals, including humans [1, 2]. The Niger Delta region of Nigeria is particularly vulnerable to the harmful effects of crude oil spills. Although oil-producing regions are significantly affected by contamination, oil workers are at even greater risk. Furthermore, the practice of consuming crude oil for supposed medicinal benefits exacerbates this health threat [3-5]. The Bonny Light crude (BLCO) oil is a high-quality, light, sweet crude produced in Nigeria. Known for its low sulfur content and high American Petroleum Institute (API) gravity, it is highly sought after in global markets due to its ease of refining into high-value products like gasoline and diesel and remains one of Nigeria's key crude oil exports, playing a crucial role in the country's economy [6]. Bonny Light Crude Oil (BLCO) has the potential to cause both acute lethal and chronic sub-lethal toxicities, depending on the mode of exposure.

Research has demonstrated that crude oil can cause toxic effects in multiple organs of both humans and animals, including the kidneys, liver, lungs, neuroendocrine system, blood, and reproductive system [4, 7-9]. These toxicities are known to occur by way of the generation of reactive oxygen species (ROS) which causes oxidative stress and ultimately, tissue damage [10]. Oxidative stress arises when there is an imbalance between the production of reactive species and the body's antioxidant defence system. [11-13]. Cellular damage is mainly caused by the failure of antioxidants to neutralize oxygen radicals, ultimately resulting in infertility when reproductive organs and tissues are affected [4, 10]. Lipid peroxidation is a chain reaction that produces free radicals all the time, which start more peroxidation. There is a correlation between oxidative stress and enhanced lipid peroxidation, protein breakdown, and other macromolecule degradation [14, 15]. The presence of polyunsaturated fatty acids in crude oil may be the cause of the observed rise in lipid peroxidation levels as the fatty acids function as excellent substrates for lipid peroxidation due to their content of bis-allylic methylene groups [16]. Antioxidants are molecules that prevent oxidative damage by scavenging reactive oxygen species (ROS) or inhibiting their production, thereby reducing or preventing the oxidation of other molecules [17-20]. N-acetylcysteine (NAC) and Zinc Sulfate ( $ZnSO_4$ ) are widely used antioxidant compounds. N-acetylcysteine (NAC) is a powerful antioxidant that works by replenishing intracellular levels of glutathione, one of the body's most important antioxidants. NAC helps neutralize reactive oxygen species (ROS) and reduces oxidative stress by promoting the synthesis of glutathione, directly scavenging free radicals, and supporting the detoxification process in cells [21-23]. Zinc Sulfate on the other hand Zinc contributes to the function of various antioxidant enzymes, such as superoxide dismutase (SOD), which helps neutralize reactive oxygen species (ROS) [24, 25]. Research on the potential beneficial effects of these key antioxidant compounds on

reproductive health, particularly in relation to improving distorted testicular cytoarchitecture and oxidative stress caused by BLCO exposure, is limited. Our previous study emphasized their role in alleviating reproductive hormone and seminal fluid dysfunction resulting from BLCO exposure [4]. This study therefore, aims to evaluate the potential protective effects of NAC and ZnSO<sub>4</sub> in mitigating distorted testicular cytoarchitecture and oxidative stress in male Wistar rats following short-term exposure to Bonny light crude oil.

## **MATERIALS AND METHODS**

### **Experimental materials and research design**

Bonny Light crude oil was sourced from the Nigerian National Petroleum Corporation (NNPC) in Warri, Nigeria. N-acetyl cysteine (A7250-10G) was obtained from Sigma Aldrich, USA, while Zinc Sulfate was sourced locally (Bactolac Pharmaceutical Inc, USA). Fifty (50) male Wistar rats (weighing 180–200g) were acquired from the Animal House of the Department of Physiology at Delta State University, Abraka, for the study. The rats underwent a three-week acclimatization period under standard animal husbandry conditions. They were housed in clean, well-ventilated wooden cages with optimal conditions, including a 12-hour light/dark cycle, temperatures ranging from 28–31°C, and humidity levels of 45–50%. The rats had unrestricted access to standard rat pellets and tap water. They were divided into ten (10) groups of five (5) animals each. Groups I and II served as control and negative control and received distilled water and BLCO (600mg/kg)[3, 5], respectively. Groups III and IV received 100 and 200mg/kg NAC [26] only while groups V and VII received BLOC (600mg/kg) in addition to 100 and 200mg/kg NAC respectively. Similarly, groups VII and VIII received ZnSO<sub>4</sub> at 0.5 ZnSO<sub>4</sub> and 1mg/kg[27] respectively while Groups IX and X received BLOC (600mg/kg) in combination with 0.5 and 1mg/kg ZnSO<sub>4</sub> respectively. All supplementations were done orally via an orogastric cannula between 8 am and 10 AM daily for three weeks.

### **Tissue Sample Collection, Laboratory Assay and Histological Study**

The animals were subjected to overnight fasting and were euthanized through cervical dislocation and the testis was carefully dissected and used for tissue homogenate biochemical analysis and histological studies. Testicular tissue homogenate was done using 50mg of testis tissue in normal saline (3ml, 0.9% NaCl) in an ice water bath. The homogenate was then centrifuged at 3000 rpm for 10 min at 4°C[28, 29]. The clear supernatant was collected and used for formalondialdehyde (MDA), superoxide dismutase (SOD), glutathione peroxidase (GPx), glutathione (GSH) and total antioxidant capacity (TAC) assay via ELISA following instructions from standard laboratory kits (Shanghai Sunred Biological Technology Co., Ltd.). The tissues were dehydrated in ethanol, cleared in xylene, embedded in paraffin and stained using haematoxylin and eosin for the histological study[30]. Photomicrographs of the desired results were obtained using the Olympus research photographic microscope (BX51, Olympus, Japan).

### **Statistical Analysis**

The data from this study were analyzed using GraphPad Prism 8 Biostatistics Software (GraphPad Software, Inc., La Jolla, USA, version 8.0). Results are expressed as Mean  $\pm$  SEM. Further analysis was performed using one-way analysis of variance (ANOVA), followed by Tukey's post hoc test for multiple comparisons. A significance level of  $p < 0.05$  was applied to all tests.

### Ethical Considerations

The animals were handled in compliance with the highest ethical standards for animal experimentation. The research protocol and study design received approval from the University of Port Harcourt Research Ethics Committee.

### Results

**Table 1:** Effect of the effect of NAC and ZnSO<sub>4</sub> on sperm oxidative stress biomarkers, antioxidant enzymes of BLCO exposed-male Wistar rats

Groups/Doses(/kg)	MDA (unit/mg)	SOD (unit/mg)	Catalase (U/L)	Glutathione (mg/dL)	TAC (mmol/L)
Control	19.11 $\pm$ 0.26	49.60 $\pm$ 0.92	35.00 $\pm$ 1.00	3.41 $\pm$ 0.13	8.98 $\pm$ 0.06
BLCO (600mg/kg)	29.96 $\pm$ 0.66*	25.60 $\pm$ 0.50*	20.20 $\pm$ 0.58*	4.86 $\pm$ 0.07*	4.12 $\pm$ 0.03*
NAC (100mg/kg)	18.38 $\pm$ 0.40 <sup>#</sup>	35.60 $\pm$ 0.67 <sup>#</sup>	26.80 $\pm$ 0.48 <sup>#</sup>	2.65 $\pm$ 0.14 <sup>#</sup>	6.63 $\pm$ 0.18 <sup>#</sup>
NAC (200mg/kg)	20.56 $\pm$ 0.17 <sup>#</sup>	38.00 $\pm$ 0.70 <sup>#</sup>	27.00 $\pm$ 0.83 <sup>#</sup>	2.78 $\pm$ 0.10 <sup>#</sup>	6.91 $\pm$ 0.07 <sup>#</sup>
NAC(100mg/kg)+ BLCO(600mg/kg)	21.71 $\pm$ 0.26 <sup>#a</sup>	28.02 $\pm$ 0.53 <sup>#a<math>\beta</math></sup>	18.20 $\pm$ 0.66 <sup>#a<math>\beta</math></sup>	3.01 $\pm$ 0.06 <sup>#</sup>	4.51 $\pm$ 0.22 <sup>#a<math>\beta</math></sup>
NAC(200mg/kg) + BLCO(600mg/kg)	19.14 $\pm$ 0.49 <sup>#<math>\phi</math></sup>	29.56 $\pm$ 0.52 <sup>#a<math>\beta</math></sup>	21.80 $\pm$ 0.66 <sup>#<math>\phi</math>a<math>\beta</math></sup>	3.12 $\pm$ 0.07 <sup>#</sup>	4.85 $\pm$ 0.07 <sup>#a<math>\beta</math></sup>
ZnSO <sub>4</sub> (0.5mg/kg)	17.88 $\pm$ 0.47 <sup>#</sup>	27.64 $\pm$ 0.92*	25.40 $\pm$ 0.50 <sup>#</sup>	3.12 $\pm$ 0.07 <sup>#</sup>	6.26 $\pm$ 0.22 <sup>#</sup>
ZnSO <sub>4</sub> (1mg/kg)	17.59 $\pm$ 0.14 <sup>#</sup>	30.70 $\pm$ 0.68 <sup>#</sup>	23.40 $\pm$ 0.81*	2.61 $\pm$ 0.14 <sup>#</sup>	6.24 $\pm$ 0.21 <sup>#</sup>
ZnSO <sub>4</sub> (0.5mg/kg) + BLCO(600mg/kg)	19.92 $\pm$ 0.30 <sup>#</sup>	30.68 $\pm$ 0.76 <sup>#</sup>	20.20 $\pm$ 0.58 <sup>#a</sup>	3.06 $\pm$ 0.06 <sup>#</sup>	4.62 $\pm$ 0.14 <sup>#ab</sup>
ZnSO <sub>4</sub> (1mg/kg) + BLCO(600mg/kg)	17.65 $\pm$ 0.27 <sup>#</sup>	32.04 $\pm$ 0.82 <sup>#a</sup>	22.80 $\pm$ 0.80*	2.82 $\pm$ 0.06 <sup>#</sup>	5.19 $\pm$ 0.04 <sup>#ab</sup>

Values are expressed as Mean $\pm$ SEM (n = 5) (ANOVA followed by Tukey's test).

BLCO. (BLCO=Bony Light Crude Oil. NAC = N-acetylcysteine. ZnSO<sub>4</sub> = Zinc Sulfate).

\* $P < 0.05$ : significantly different when compared with the control group.

<sup>#</sup> $P < 0.05$ : Significantly different when compared with BLCO.

<sup>a</sup> $P < 0.05$ : significantly different when compared with NAC<sub>a</sub> (100mg/kg).

<sup>b</sup> $P < 0.05$ : significantly different when compared with NAC<sub>b</sub> (200mg/kg).

<sup>$\phi$</sup>  $P < 0.05$ : Dose-dependent significant difference.

<sup>a</sup> $P < 0.05$ : significantly different when compared with ZnSO<sub>4a</sub>(0.5mg/kg).

<sup>b</sup> $P < 0.05$ : significantly different when compared with ZnSO<sub>4a</sub>(1mg/kg).

<sup>c</sup> $P < 0.05$ : significantly different when compared with ZnSO<sub>4a</sub>(0.5mg/kg)

Table 1 presents the effects of NAC and ZnSO<sub>4</sub> on oxidative stress biomarkers in male Wistar rats exposed to BLCO for both short and long-term durations. Exposure to BLCO resulted in a significant increase in MDA levels compared to the control group ( $p < 0.05$ ). However, co-treatment with NAC and ZnSO<sub>4</sub> significantly reduced MDA levels in BLCO-exposed rats compared to those exposed to BLCO alone. Similarly, BLCO exposure caused a significant decrease in catalase levels compared to controls ( $p < 0.05$ ), but this decrease was significantly mitigated in rats co-treated with NAC and ZnSO<sub>4</sub>. Short-term BLCO exposure led to a significant increase in GSH levels compared to controls, which was notably reduced in rats co-

treated with NAC and ZnSO<sub>4</sub>. Additionally, the TAC of BLCO-exposed rats was significantly lower than that of the control group. Co-treatment with higher doses of NAC and ZnSO<sub>4</sub> significantly increased TAC levels in BLCO-exposed rats compared to those receiving only BLCO.

*MC – myoid cell, PS – primary spermatocyte, SG – spermatogonia, SC – Sertoli cells, ST - spermatids*

**Figure 1.** Photomicrograph of a testicular section of control (A) and BLCO-exposed rats (B) stained with Haematoxylin and Eosin (H&E) at x100 magnification.

Slide A of the testicular histology photomicrograph displays normal testicular architecture, with seminiferous tubules surrounded by the tunica albuginea. Visible structures include seminiferous tubules, Sertoli cells, and spermatocytes. The Sertoli cells exhibit a normal tubular shape with round nuclei located in the cytoplasm, while the spermatocytes are round with nuclei present in the cytoplasm. In contrast, Slide B reveals poor testicular architecture, characterized by atrophic seminiferous tubules lined by a stratified germinal epithelium consisting of a basal spermatogonia layer, indicative of testicular degeneration. Longitudinal Sertoli cells are also evident among the sparse spermatids, along with fibroconnective tissue (composed of myoid cells) encasing the seminiferous tubules.

*MC – myoid cell, PS – primary spermatocyte, SG – spermatogonia, SC – Sertoli cells, ST - spermatids*

**Figure 2:** Photomicrograph of a testicular section of NAC (C:100mg/kg and D: 200mg/kg) administered to rats stained with Haematoxylin and Eosin (H&E) x100 magnification.

The photomicrograph(C and D) shows normal testicular architecture in both doses with seminiferous tubules surrounded by tunica albuginea with normal Sertoli cells and spermatocytes. Myoid cells and spermatogonia are also interspersed at the periphery of the tubule.

*MC – myoid cell, PS – primary spermatocyte, SG – spermatogonia, SC – Sertoli cells, ST - spermatids*

**Figure 3:** Photomicrograph of a testicular section evaluating the effect of NAC (100mg/kg) + BLCO(E) and NAC (200mg/kg) + BLCO(F) co-administration in rats stained with Haematoxylin and Eosin (H&E) x100 magnification.

Slides E and F show poor testicular architecture with some normal seminiferous tubules. Seminiferous tubules with maturation arrest exhibiting widened lumen and interstitial space with normal Leydig cells are also seen. This is suggestive of the regenerative histoarchitecture feature of the testis which is more pronounced at NAC (200mg/kg).

*MC – myoid cell, PS – primary spermatocyte, SG – spermatogonia, SC – Sertoli cells, ST - spermatids*

**Figure 4:** Photomicrograph of a testicular section evaluating the effect of ZnSO<sub>4</sub> (0.5mg/kg)(G) and ZnSO<sub>4</sub> (1mg/kg)(H) co-administration, stained with Haematoxylin and Eosin (H&E) x100 magnification.

Section of the testes shows normal architecture with normal seminiferous tubules interspersed with Sertoli cells and spermatocytes which appear normal. The testes section suggests normal histoarchitecture displaying regenerative features (G) and complete regenerative property of testicular features (H).

*MC – myoid cell, PS – primary spermatocyte, SG – spermatogonia, SC – Sertoli cells, ST - spermatids*

**Figure 5:** Photomicrograph of a testicular section evaluating the effect of ZnSO<sub>4</sub> (0.5mg/kg) + BLCO(I) and ZnSO<sub>4</sub> (1mg/kg) + BLCO(J) co-administration in rats stained with Haematoxylin and Eosin (H&E) x100 magnification.

The testicular section displays poor testicular architecture with atrophic and degenerated seminiferous tubules and sloughed germinal cells. The interstitial spaces show normal Leydig cells with centrally placed spermatids which are scanty displaying regenerative properties in dose dose-dependent manner.

## DISCUSSION

Crude oil toxicity is prevalent in the Nigeria's Niger Delta Region. These toxicities are known to arise through the generation of reactive oxygen species (ROS), leading to oxidative stress and,

ultimately, tissue damage due to exposure to polycyclic aromatic hydrocarbons (PAHs) and volatile organic compounds (VOCs). Natural and synthetic antioxidants used as prophylactic or therapeutic measures have shown promise in the mitigation of the effects of oxidative stress [31, 32]. This present study assessed the potential protective effects of N-acetyl cysteine and Zinc Sulfate in mitigating distorted testicular cytoarchitecture and oxidative stress in male Wistar rats following short-term exposure to Bonny light crude oil (BLCO).

Our findings indicate that BLCO significantly increased malondialdehyde (MDA) levels while decreasing antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), glutathione (GSH), and total antioxidant capacity (TAC) compared to the control group. This suggests that BLCO may have generated reactive oxygen species (ROS) in the testes of exposed rats, leading to lipid peroxidation and increased MDA levels, a by-product of this process. Consequently, this could result in Sertoli cell death in the testes, negatively affecting spermatogenesis and male fertility [33]. The findings of the current study are consistent with earlier research. The testicular disruptions caused by BLCO exposure correspond with previous studies showing that the antioxidant systems in the testes and epididymal sperm of rats were impacted, evidenced by decreased levels of superoxide dismutase (SOD) and catalase (CAT), along with significantly increased levels of glutathione (GSH) and malondialdehyde (MDA) [29]. We also observed that NAC co-administration with BLCO significantly increased SOD, CAT, and TAC and decreased GSH levels in experimental animals when compared with BLCO as a result of its antioxidant activity. Antioxidants act by scavenging free radicals in the human system [34]. GSH is synthesized in the cytoplasm of the liver and thereafter distributed to other organs [35]. It is a non-enzymatic antioxidant known for ROS detoxification [36, 37] with studies showing that in light of exposure to different stressors causes markedly compromised defence mechanisms of GSH-related antioxidants, thus compromising GSH homeostasis [30, 38, 39]. Also, the observed increase in GSH level suggests stimulation of the antioxidant system in defence against the effect of stress, whereas, depletion of GSH level is an indication of tissue oxidative stress [40] which corroborates the results of increased GSH level following NAC co-administration from the present study. NAC has also been shown to reduce oxidative damage in sodium arsenide and tetracycline-induced oxidative stress [41, 42].

Our data reveal that experimental animals treated with ZnSO<sub>4</sub> and BLCO had significantly higher levels of SOD, CAT, and TAC compared to those given only BLCO. Zinc Sulfate acts as an antioxidant through several mechanisms: it competes with iron and copper for binding to cell membranes, protects biomolecules from oxidation, enhances antioxidant enzyme activity, reduces oxidant-promoting enzyme activity, and promotes the expression of metallothionein, which scavenges hydroxyl radicals [43, 44]. Zinc has also been demonstrated to reduce oxidative stress due to cadmium treatment as manifested by a decrease in blood hydroperoxide and testicular MDA, a good indicator of oxidative stress [45]. Decreased total antioxidant capacity (TAC) as observed in the BLCO-exposed rats could have a significant role in the aetiology of

impaired sperm functions[46] as lower TAC in fructose-induced diabetic rats reduced sperm count, percentage motility and activity, suggesting that reduced antioxidant power was a major factor responsible for the impaired sperm quality, which could lead to infertility [47].

Histological analysis of the testes from rats exposed to BLCO revealed notable alterations in testicular morphology, including interstitial exudates, necrosis of spermatogenic interstitial (Leydig) cells, and degeneration. The current study similarly demonstrates that BLCO significantly disrupts testicular histology compared to the control group (Figure 1A & B). These findings suggest that BLCO exposure may contribute to infertility by altering testicular and sperm function, primarily through mechanisms such as oxidative stress and lipid peroxidation. These processes can lead to inflammation in the seminal fluid and trigger an immunological response, potentially resulting in the formation of antisperm antibodies (ASAs). Elevated levels of ASA can damage the sperm sheath, exhibiting cytotoxic effects on spermatozoa, causing seminal agglutination that immobilizes sperm cells, or even leading to sperm cell death[48, 49]. Our present finding is consistent with Orisakwe *et al.*, [50] where BLCO was found to have negatively impacted testicular histopathology in a dose-dependent manner. They observed that the severity of the damage was dependent on the amount of BLCO with the connective tissues, seminiferous tubules, spermatogonia, and spermatids were either completely absent, mildly, severely, or grossly injured, or the sperm cells were coagulated and necrotic. They also observed that the basement membrane was disorganized, with some tubules empty of cells in the 400 and 800mg/kg BLCO exposed group while spermatocyte necrosis was observed in the 800mg/kg BLCO exposed group.

The control and NAC-supplemented groups displayed densely packed Leydig cells within the seminiferous tubules, along with normal spermatozoa development at all stages, maintaining typical testicular architecture (Figure 2C & 2D). The current study also shows that NAC was given alongside BLCO significantly improved the testes architecture (Figure 3E&3F). Two potential mechanisms may explain NAC's protective effect on testicular tissue: first, NAC acts as a free radical scavenger in response to oxidative stress, and second, it prevents the depletion of GSH[51, 52]. Similar reports have demonstrated the ability of NAC to improve testicular architecture busulfan-induced testicular dysfunction[53]. Similar to NAC, the present study also observed that ZnSO<sub>4</sub> caused noticeable improvement in the pathohistological structure of the testes in BLCO-exposed Wistar rats (Figure 5I&5J). Zinc sulfate supplementation may have influenced testicular anatomy and function by preventing testicular damage caused by free radicals caused by BLCO exposure by binding directly to protein sulfhydryl groups, thereby inhibiting oxidation[54, 55]. The current findings align with previous reports showing that ZnSO<sub>4</sub> supplementation alleviated disruptions in spermatogenesis caused by cisplatin (CP)-induced testicular dysfunction. This included improvements in the degeneration of seminiferous tubules, severe perivascular fibrosis, disorganization and hyalinization of intertubular connective

tissue, reduced tunica albuginea thickness and tubular diameter, increased interstitial space, and sloughing of the germinal epithelium[56].

## CONCLUSION

This study demonstrates that BLCO-exposed rats experienced toxicity affecting both testicular antioxidants and testicular structure, which could contribute to male infertility by impairing testicular function. It also highlights potential risks to the reproductive health of animals and humans exposed to this environmental pollutant, particularly in regions prone to oil spills. Our findings suggest that NAC and ZnSO<sub>4</sub> improved testicular function in rats, including spermatogenesis and steroidogenesis, by reducing testicular reactive oxygen species, thus enhancing antioxidant levels in BLCO-induced gonadal toxicity. NAC and ZnSO<sub>4</sub> also protected the testicular cytoarchitecture from the damaging effects of BLCO exposure.

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