

Hypermethylation of Apoptotic Genes in Oral Squamous Cell Carcinoma

ABSTRACT

Oral squamous cell carcinoma (OSCC) presents a substantial worldwide health challenge due to its aggressive behavior and resistance to standard treatments, resulting in high mortality rates. Epigenetic changes, notably DNA hypermethylation, are pivotal in OSCC development as they silence tumor-suppressor genes and drive tumor growth. This review highlights the hypermethylation status of apoptotic genes in the promoter region of CpG island which results in the formation of OSCC. This article utilizes different scientific databases such as Google Scholar, PubMed, NCBI, etc. to understand the interplay between external factors and epigenetic modifications which provides valuable insights for preventive strategies and personalized approaches to OSCC. The review discusses the epidemiology and causes of OSCC, emphasizing its resistance to therapy and poor prognosis. It also assesses current treatment strategies targeting DNA hypermethylation, such as inhibitors of demethylation and histone deacetylase, for their potential to improve patient outcomes. Future research directions center on investigating combination therapies targeting various epigenetic regulators and creating non-invasive methods for early cancer detection and prognosis evaluation. Overall, this review underscores the significance of DNA hypermethylation in OSCC progression and highlights the therapeutic promise of addressing epigenetic changes, especially those affecting apoptotic genes, to enhance patient survival.

Keywords: Hypermethylation, Apoptotic genes, Cancer, Epigenetics.

1. INTRODUCTION

Cancer, a diverse and intricate set of conditions, presents itself through the unregulated multiplication of cells, leading to the potential invasion of nearby tissues and the prospect of spreading to distant organs. The worldwide significance of cancer is considerable, marked by the diagnosis of millions of new cases annually. Among non-communicable diseases (NCDs), cancer ranks as the second principal contributor to mortality, following cardiovascular disease [1]. A tumor refers to an irregular cell proliferation that can be classified either as malignant or benign. Benign tumors like skin warts, are limited to a specific area and do not attack neighboring tissues or metastasize to distant organs. In contrast, malignant tumors have the potential to invade nearby tissue and spread through the bloodstream or lymphatic system. Only malignant tumors are considered cancers due to their invasive and metastatic nature, posing a significant threat to health. While surgical removal is often effective for benign tumors, the metastasis of malignant tumors renders them resistant to localized therapies. Tumors, whether benign or malignant, are categorized according to their cellular origin.

OSCC, a form of oral tumor, constitutes more than 90% of all malignant cases in the oral region and contributes significantly to the overall incidence of tumors in the head and neck region, accounting for 38%. Its onset is marked by a gradual accumulation of genetic alterations and tumor evasion of the host immune response [2]. The abnormal proliferation of malignant tissue is widespread across various regions of the oral cavity, including the labial mucosa, floor of the mouth, gingiva, palatal area, vestibule, buccal mucosa, alveolar ridge, and tongue. Notably, the distribution pattern of OSCC cases reveals a higher prevalence in certain regions, with approximately 32% occurring in the buccal mucosa, 11% in the lower lip, 22% in the tongue, 11% in the palate, 5% in the floor of the mouth, 3% in the gingiva, 8% in the vestibule, and 5% in the alveolus.

47 In 2020, a report by GLOBOCAN showed 377,713 of new oral cancer cases on a global level, which
48 resulted in approximately 177,757 of deaths in a year. In India, it was observed that there were 135,929 of
49 new cases, and 75,290 of deaths were reported each year based on the available data. Oral cancer is
50 notably prevalent, particularly in Asian countries, constituting approximately fifty to seventy percent of all
51 cancer-related fatalities in India [3]. The progression of oral cancer involves various stages influenced by
52 both internal and external factors. There has been a notable rise in the incidence of oral cancer in certain
53 regions, including India, Sri Lanka, Bangladesh, Taiwan, and Pakistan, collectively contributing to
54 approximately 25% of newly reported cases. Notably, OSCC demonstrates a male predominance among
55 affected populations [4]. Despite advancements in therapeutic modalities such as radiation, surgery, and
56 chemotherapy, treatment-resistant OSCC remains a significant challenge. The 5-year survival rate for
57 individuals with OSCC has persistently hovered around 50% for an extended period, indicating ongoing
58 obstacles in achieving substantial improvements [5]. Projections suggest a tripling of cancer incidence by
59 2030, despite extensive research efforts currently underway. OSCC is characterized as a highly
60 aggressive form of cancer, displaying considerable variability in etiological, clinical, and molecular
61 aspects [6].

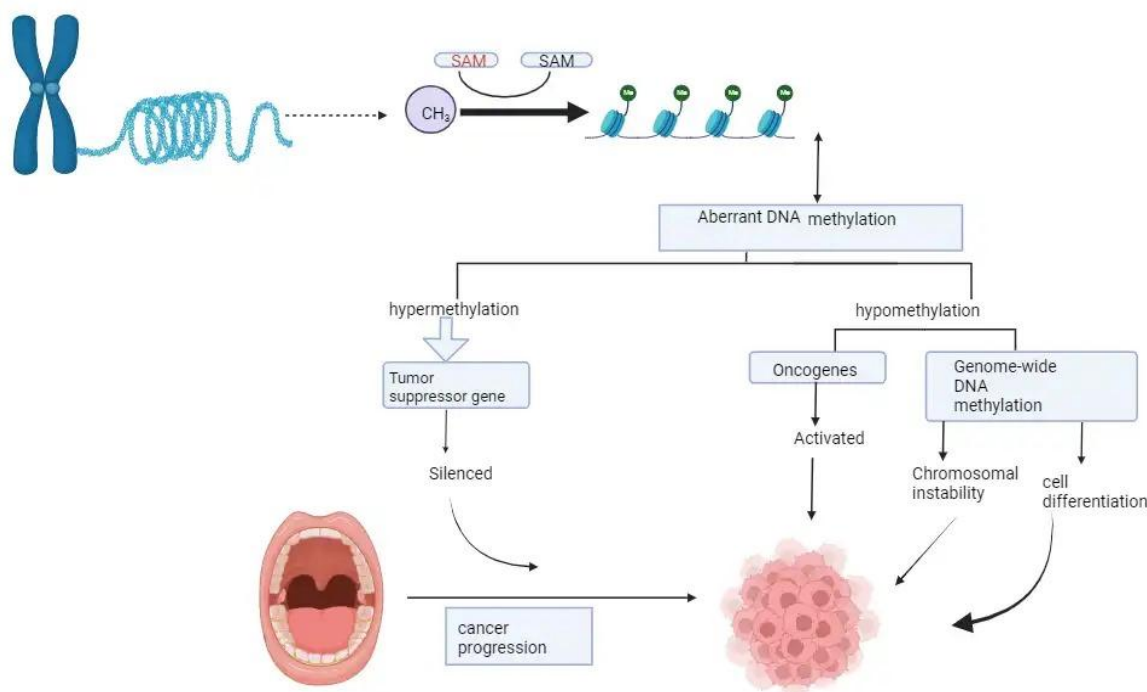
62 Significant contributors include consistent consumption of tobacco and alcohol, along with persistent
63 infection of human papillomavirus (HPV) [7]. These risk factors may trigger various genetic and epigenetic
64 pathway which enables the development and metastasis of tumors in addition to genomic instability. Oral
65 premalignancy conditions and OSCC arise from genetic alterations, encompassing permanent
66 modifications such as deletions, amplifications, and mutations in DNA sequence that initiate the activation
67 of oncogenes or the suppression of tumor suppressor genes (TSGs). Epigenetics, as defined by Cavalli
68 and Heard (2019), investigates the molecules and processes that maintain distinct patterns of gene
69 activity while preserving the same DNA sequence [8]. Environmental and lifestyle factors interact with
70 genetic information, significantly influencing genome activity. Epigenetic mechanisms produce various
71 adaptable structural configurations that influence gene expression independently of genomic alterations.
72 These mechanisms involve coordinated adjustments to DNA and chromatin which are classified into
73 various types such as histone alterations, DNA methylation, and regulatory small non-coding RNAs.
74 Epigenetic mechanisms could also contribute to the silencing of Tumor-Suppressor Genes
75 [9]. Additionally, alterations caused by epigenetic changes contribute to this process which involves many
76 gene expression variations that occur without any changes in DNA sequence. Changes in epigenetics
77 can have a significant impact on metastasis, chemotherapy response, and tumor progression [10].
78 At present, three primary forms of epigenetic mechanisms are identified: histone modification, ribonucleic
79 acid interference (RNAi), and DNA hypermethylation. Any disruption in these interdependent epigenetic
80 mechanisms results in aberrant gene expression, which can cause cancer and other “epigenetic
81 diseases” [11]. Methylation, a prevalent DNA modification in eukaryotic organisms, has garnered
82 considerable attention owing to its capacity that influence gene expression. Alterations in the pattern of
83 methylation may lead to either hypomethylation or hypermethylation. Hypomethylation of DNA is linked to
84 gene reactivation and chromosomal instabilities [12-13]. On the contrary, gene repression is associated
85 with hypermethylation in the promoter regions of the genes [14]. The hypermethylation of DNA, which
86 results in the silencing of genes, affects a multitude of genes related to various cellular pathways. This
87 includes functions like suppressing tumors, repairing DNA, responding to hormones, adhering to cells,
88 and metabolizing drugs, among other roles [15].

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90 2. EPIGENETIC REGULATION OF GENE EXPRESSION: FOCUS ON DNA METHYLATION

91 DNA methylation is an inheritable epigenetic modification characterized by the addition of a methyl group
92 to cytosine bases within DNA molecules, facilitated by enzymes known as DNA methyltransferases
93 (DNMTs), as illustrated in Figure 2. In mammals, DNA methylation primarily targets cytosine residues
94 within various genomic contexts and is one of the extensively studied forms of chromatin modification. In
95 somatic cells, the majority of DNA methylation occurs within CpG dinucleotide sequences, constituting
96 over 98% of total methylation. However, in embryonic stem cells, a significant proportion of methylation
97 occurs outside CpG contexts, accounting for up to a quarter of total methylation. DNA methylation is
98 regulated by a group of enzymes called DNA methyltransferases (DNMTs) [16-17].

99 DNA methylation is essential for normal developmental processes and significantly influences critical
 100 mechanisms such as X-chromosome inactivation, genomic imprinting, and the regulation of repetitive
 101 element expression and movement. Dysregulation of DNA methylation is implicated in pathological
 102 conditions, including cancer [18]. Hypomethylation induces instability in chromosomes and the
 103 reactivation of silenced genes, including proto-oncogenes thus enhancing cancer progression.
 104 Hypomethylation is also responsible for contributing to oral carcinogenesis by resulting in loss of
 105 imprinting due to which alteration in the expression of genes occurs. Similarly, hypermethylation in the
 106 promoter region leads to the inactivation of TSGs, disrupts DNA repair mechanisms, and facilitates
 107 evasion of the immune system. Furthermore, numerous hypermethylated genes have been identified that
 108 could potentially impede OSCC progression in certain instances. These epigenetic alterations serve as
 109 fundamental molecular events in OSCC tumorigenesis, offering promising avenues for diagnostic as well
 110 as therapeutic interventions [19] (Fig. 1).



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 112
 113 **Fig. 1.** Schematic diagram of aberrant DNA methylation pattern. Diagram showing the impact of both
 114 Hypermethylation and hypomethylation in OSCC progression. Hypermethylation of tumor-suppressor
 115 genes is responsible for their inactivation, restricting their ability to control cell growth and suppression of
 116 tumor development. Conversely, hypomethylation inhibits the transcription of cell division-inhibiting
 117 genes, such as oncogenes. Genome-wide DNA hypomethylation induces chromosomal instability and
 118 gene mutations while also contributing to normal cell function and differentiation. Understanding these
 119 patterns of DNA methylation is essential for comprehending the underlying mechanisms of cancer
 120 development.

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 122 CpG islands frequently display hypermethylation in tumor regions, leading to the transcriptional
 123 suppression of TSGs and promoting cancer progression. Conversely, hypomethylation or demethylation
 124 of CpG islands has been observed to reduce proto-oncogene transcription, resulting in chromosome
 125 instability, which is an early characteristic feature of tumorigenesis [20].

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127 **3. APOPTOSIS**

128 Apoptosis is a controlled cellular process designed to preserve cellular balance and eliminate impaired
129 cells. Cells undergoing apoptosis display distinct characteristics, such as condensed nuclei, fragmented
130 DNA, membrane blebbing, and heightened cell membrane permeability [21-22]. It encompasses three
131 primary pathways: intrinsic, extrinsic, and granzyme B pathways.

132 Regardless of the pathway involved, the outcome is the activation of caspase proteins, initiating a
133 cascade of proteolytic events responsible for breaking down and eliminating the dying cell [23]. This
134 process serves as one of the primary defenses against the development of cancer since cancer is
135 characterized by resistance to cell death. It is also an essential part of normal cell turnover and tissue
136 homeostasis [24].

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139 **3.1. Intrinsic pathway**

140 The mitochondrial pathway, also known as the intrinsic pathway of apoptosis, is a key mechanism by
141 which cells undergo programmed cell death in response to internal signals, such as cellular stress, DNA
142 damage, or developmental cues. This pathway is initiated by perturbations within the cell itself rather than
143 external stimuli. The process begins with the activation of pro-apoptotic proteins, particularly Bax and
144 Bak, which are members of the Bcl-2 protein family. These proteins undergo conformational changes and
145 oligomerization, leading to the formation of pores in the mitochondrial outer membrane. This results in the
146 release of pro-apoptotic factors from the mitochondrial intermembrane space into the cytoplasm, including
147 cytochrome c. Once released into the cytoplasm, cytochrome c interacts with the apoptotic protease
148 activating factor 1 (Apaf-1), leading to the formation of a large multimeric protein complex known as the
149 apoptosome. Within the apoptosome, procaspase-9 molecules are brought into proximity and undergo
150 autocatalytic cleavage, resulting in the activation of caspase-9. Activated caspase-9 serves as an initiator
151 caspase, triggering a cascade of caspase activation. Caspase-9 cleaves and activates downstream
152 effector caspases, such as caspase-3 and caspase-7, which are responsible for executing the final steps
153 of apoptosis. These effector caspases cleave specific cellular substrates, including structural proteins and
154 DNA repair enzymes, leading to cellular breakdown and programmed cell death [25].

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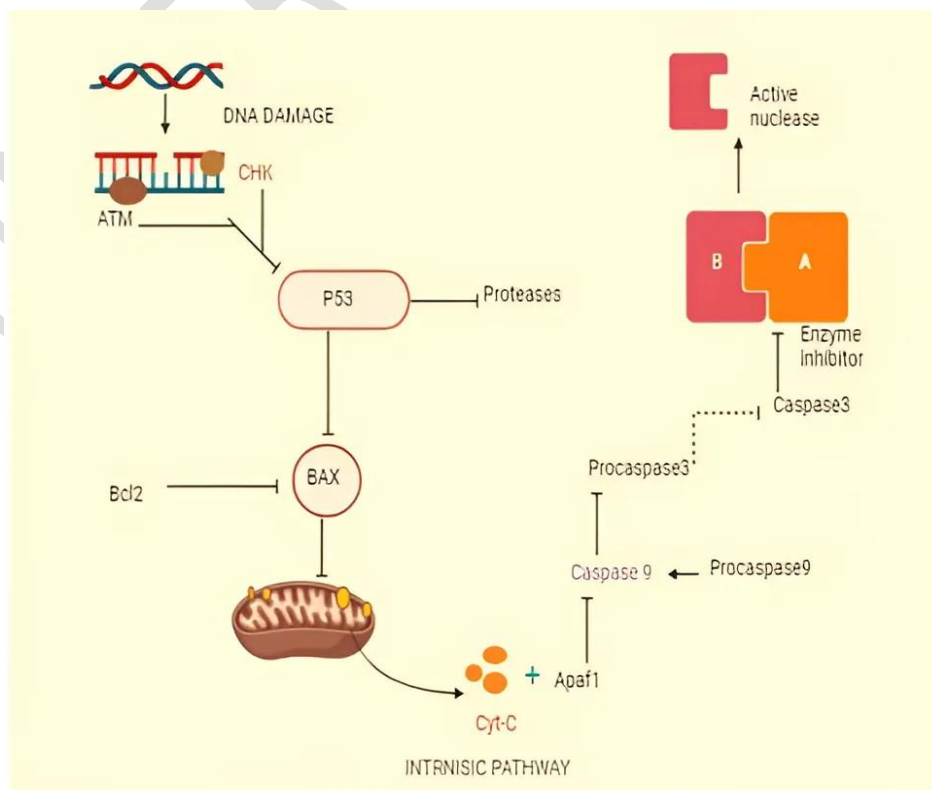
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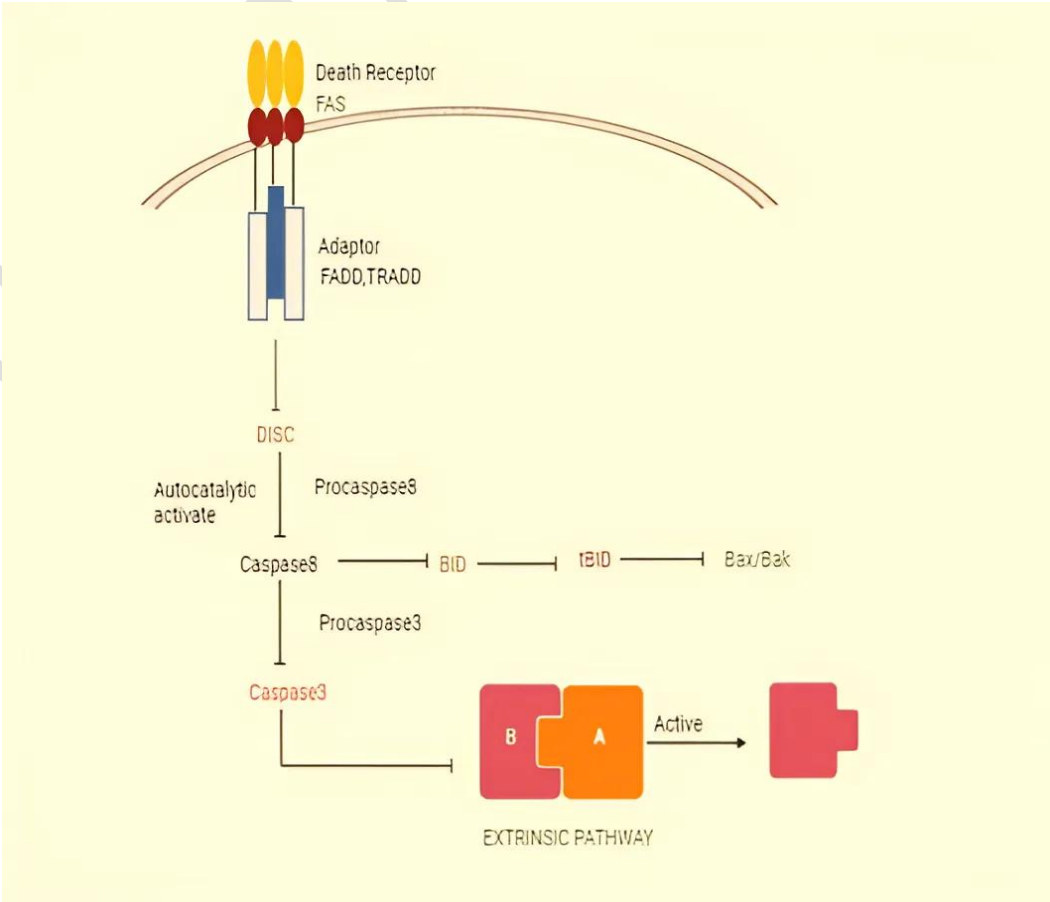


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Fig. 2. Schematic diagram of Intrinsic (mitochondrial) Pathway of Apoptosis

3.2. Extrinsic Pathway

The extrinsic pathway, also known as the death receptor pathway, is a critical mechanism by which cells initiate apoptosis in response to external signals. It begins when death ligands, such as Fas ligand (FasL), bind to death receptors located on the cell surface (Fig. 3). This binding event triggers a series of molecular events that ultimately lead to cell death. Upon binding of the death ligand to its receptor, conformational changes occur in the receptor, leading to the recruitment and activation of adaptor proteins such as FADD (Fas-associated death domain). FADD then facilitates the recruitment and activation of procaspase-8 or procaspase-10 molecules to form the death-inducing signaling complex (DISC). Within the DISC, procaspases undergo autocatalytic cleavage, resulting in the activation of caspase-8 or caspase-10. These activated initiator caspases then cleave and activate downstream effector caspases, such as caspase-3, caspase-6, and caspase-7. Effector caspases are responsible for executing the final steps of apoptosis by cleaving various cellular substrates, including structural proteins, DNA repair enzymes, and inhibitors of apoptosis (IAPs). This leads to cellular breakdown and ultimately programmed cell death[26].



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209 **Fig. 3.** Schematic diagram showing Extrinsic (death receptor) Pathway.

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211 **3.3. Granzyme B Pathway**

212 Granzyme B-mediated apoptosis, closely linked to natural killer (NK) cells and T cells' cytotoxic activity,
213 initiates the intrinsic apoptotic pathway in target cells. Granzyme B enters the target cell and cleaves Bid,
214 triggering mitochondrial outer membrane permeabilization (MOMP) and the release of cytochrome c.
215 Cytochrome c forms the apoptosome with Apaf-1 and procaspase-9, activating caspase-9 and initiating a
216 caspase cascade, leading to cellular breakdown and apoptosis [27].

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219 **4. ROLE OF APOPTOSIS IN CANCER BIOLOGY**

220 Apoptosis, termed programmed cell demise, serves as a critical guardian of cellular equilibrium,
221 meticulously orchestrating the removal of aberrant cells to uphold physiological harmony. Regarding
222 cancer development, cells obtain an enhanced potential to resist mechanisms that trigger apoptosis,
223 marking the evasion of this process as a crucial aspect of cancer cell evolution. Cancer cells utilize a
224 variety of mechanisms to resist programmed cell death, known as apoptosis. These processes
225 encompass the inactivation of TSGs through genetic variations, elevated expression of anti-apoptotic
226 proteins, activation of tumor-inducing genes, fortification of cell surviving signaling pathways, disruption of
227 apoptotic signaling molecules, malfunctioning of apoptosis execution pathways, and viral-mediated
228 inhibition of tumor suppressor proteins via interactions with viral proteins [28]. When apoptotic genes are
229 hypermethylated, it often results in the suppression of apoptosis, contributing to tumorigenesis and
230 cancer progression.

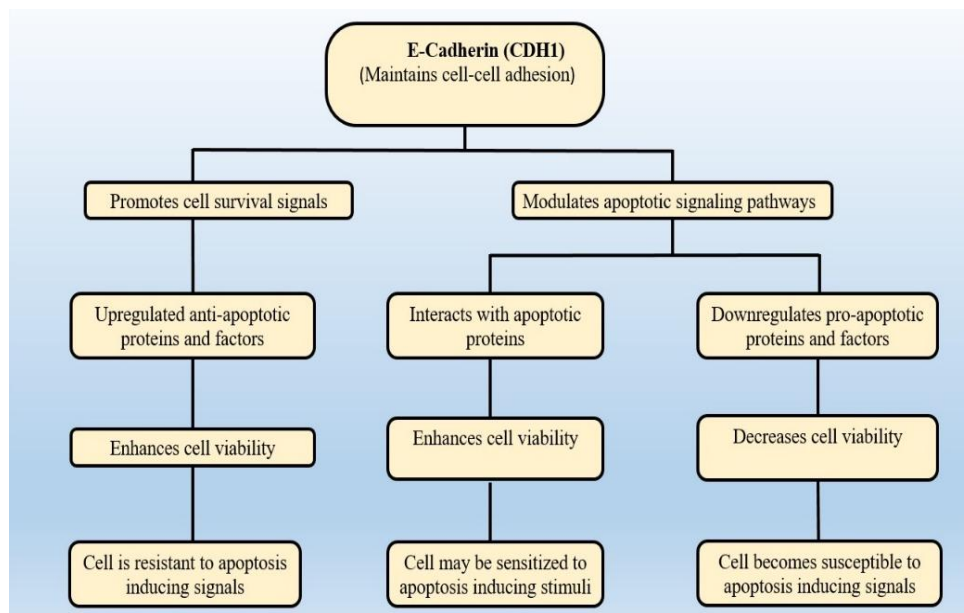
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232 **5. APOPTOTIC GENES HYPERMETHYLATED IN ORAL SQUAMOUS CELL CARCINOMA**

233 **5.1. Cadherin:**

234 Cadherin constitutes a sizable family of cell surface proteins pivotal in processes such as cell
235 differentiation, adhesion, and the establishment of cohesive tissue structures [29]. These proteins are
236 categorized into three distinct types: type I, type II, and type III cadherins. Type I cadherins,
237 encompassing epithelial (E), placental (P), neural (N), and retinal (R) cadherins, exhibit expression within
238 the mammary gland. Type II cadherin, represented by cadherin five, assumes a critical role in preserving
239 the structural integrity of blood vessels, alongside vascular endothelial cadherin. Cadherin 11, prevalent
240 in osteoblasts, is indispensable for upholding the integrity of bones and joints. Type III cadherin
241 comprises cadherin 13 and 15 [30]. Epithelial cadherin, a 120 kDa glycoprotein, encompasses three
242 functional domains: cytoplasmic, transmembrane, and extracellular. The CDH1 gene is located on
243 chromosome 16p22.1. This calcium-dependent adhesion molecule regulates essential physiological
244 activities such as polarity, differentiation, and cell movement. Decreased CDH1 expression is associated

245 with heightened invasiveness and progression of epithelial neoplasms, including oral carcinomas.
 246 Numerous reports emphasize CDH1 expression across various cancer types. The absence of CDH1
 247 strongly indicates potential changes in cell function and motility. In OSCC, reduced CDH1 expression is
 248 linked to an antagonistic relationship, indicating a connection between OSCC aggressiveness and
 249 decreased E-cadherin levels. The likelihood of metastasis, characterized as the “tumor avalanche” in
 250 cancerous cells, increases when mutations occur or when the gene expresses less [31]. Methylation of
 251 the CDH1 promoter is observed in around 17% to 85% of tumors found in the oral cavity.[32-34].



265 **Fig. 4.** Flowchart depicting the role of E-cadherin.

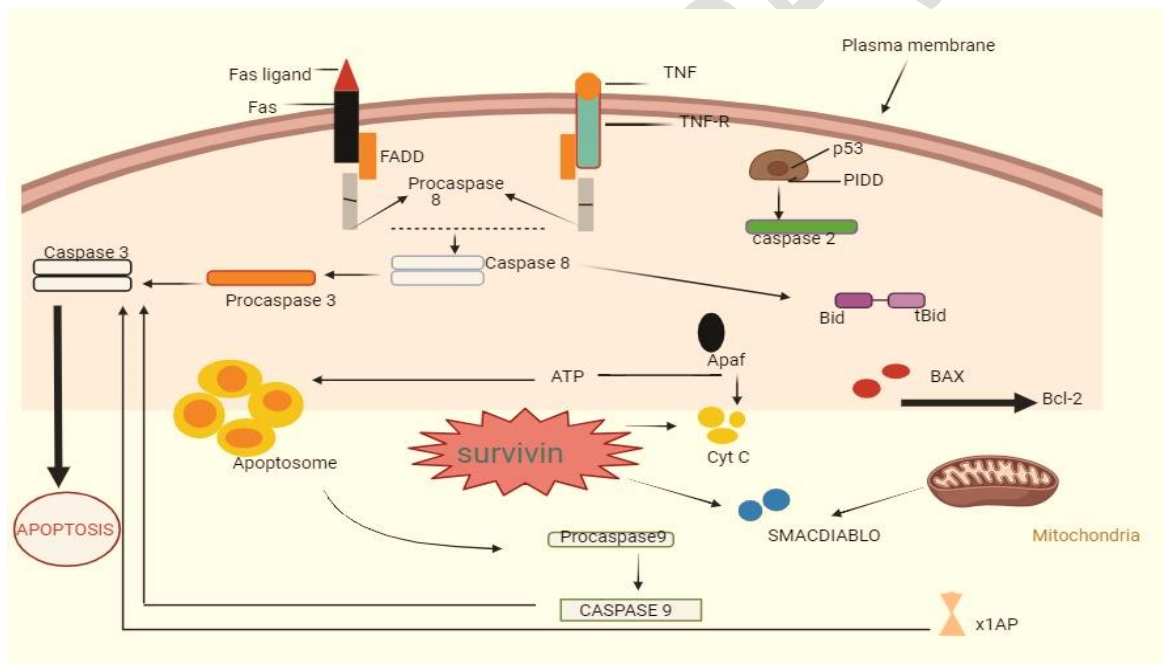
267 **5.2. CASP8:**

268 CASP8 encodes Caspase 8, also known as FLICE (FADD-like IL-1 β -converting enzyme), which is a vital
 269 enzyme involved in the initiation of apoptosis. It is situated on chromosome 2q33-34 and comprises 10
 270 exons spanning approximately 30 kilobases. There are findings indicating approximately 168 single
 271 nucleotide polymorphisms (SNPs) linked with CASP8, most of which are uncommon or nonoperative. In
 272 addition to its role in the FAS-FAS ligand-mediated extrinsic pathway, CASP8 interacts with the BH3
 273 interacting-domain death agonist protein to modulate the intrinsic pathway, thereby engaging caspases
 274 [35-37]. Caspase-8 has been identified as a contributor to various cellular processes, including the NF- κ B
 275 signaling activation, autophagy regulation, modification of endosomal trafficking, and facilitation of cell
 276 migration and adhesion. The multifaceted functions of CASP8 suggest that its impact on tumor
 277 malignancy depends on a particular cellular context, indicating that CASP8 can either enhance or
 278 suppress tumor development based on the prevailing conditions [38]. Caspase-8 becomes activated upon
 279 receiving signals from death receptors, triggered by the interaction between molecules like TRAIL or FasL
 280 and their corresponding death receptors. This activation takes place within the DISC, where Caspase-8 is
 281 induced and undergoes autocatalytic processing, leading to its conversion into its active form. Following
 282 this, Caspase-8, once activated, commences programmed cell death by cleaving and activating
 283 subsequent effector caspases such as Caspase-3 and Caspase-7. These events result in the impactful
 284 outcome execution of apoptosis, playing a crucial role in eliminating cells with damaged DNA or those
 285 experiencing cellular stress.

287 **5.3. Survivin:**

288 Survivin is the unique and the smallest member within the mammalian IAP family, characterized by its
 289 distinctive structure. It consists of a sole N-terminal BIR domain (Baculovirus IAP repeat), coupled with an
 290 extended C-terminal region featuring a coiled α -helix structure [39]. The survivin protein's BIR domain is
 291 crucial for its ability to prevent apoptosis, while its coiled domain enables interaction with tubulin
 292 structures and potentially regulates cell division. Human survivin, originating from the BRIC5 gene,
 293 possesses a molecular weight of 16.5 kDa and is situated on chromosome 17 at the telomeric end,
 294 spanning 14.7 kb of genomic DNA. Within cancer cells, survivin serves two primary roles: firstly, it
 295 regulates mitosis through the assembly of the CPC (Chromosomal passenger complex) in collaboration
 296 with other proteins, and secondly, it impedes the process of apoptosis [40]. Survivin, a key player in cell
 297 survival, exerts its anti-apoptotic effects primarily by disrupting the mitochondrial-mediated apoptotic
 298 pathway. The formation of the apoptosome complex occurs in the presence of dATP involving
 299 Cytochrome c, Apaf-1, and procaspase-9, leading to the activation of procaspase-9. Survivin intervenes
 300 in this process, likely hindering caspase-9 activation by inhibiting apoptosome formation. Moreover,
 301 survivin may directly inhibit both activator caspase-9 and effector caspases-3 (Fig. 5). Additionally,
 302 survivin opposes the proapoptotic effects of Smac/DIABLO, a protein that antagonizes IAPs. By
 303 counteracting Smac/DIABLO, survivin supports the role of other IAPs, including XIAP. XIAP acts as a
 304 potent suppressor of apoptosis by directly interacting with caspases and inhibiting their function.

305



306 **Fig. 5:** Role of Survivin in the mitochondrial-mediated apoptotic pathway.

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308 In summary, survivin orchestrates a multi-faceted strategy to inhibit crucial steps in the apoptotic pathway,
 309 thereby promoting the survival of cells and providing resistance to apoptosis mechanisms [41]. The
 310 regulation of survivin mechanisms is still not completely understood. Nonetheless, it has been noted that
 311 a variety of signaling cascades and constituents stimulate survivin in cancerous cells. Currently, it is
 312 believed that cancer cells generally exhibit more active intracellular pathways that activate survivin
 313 compared to normal cells. In contrast to cancer cell lines, variable reporter gene assays suggest a
 314 minimal survivin promoter role in normal cells, indicating differential regulation of survivin expression
 315 between these two cell types [42].

316

317 **5.4. Death Associated Protein Kinase 1 (DAPK1):**

318 DAPK1 is located at the gene locus 9q34.1. DAPK1 operates within the serine/threonine kinase pathway,
319 regulating apoptosis through calcium/calmodulin signaling and exerting proapoptotic effects [43-45]. Its
320 location on chromosome 9q34.1 underscores its pivotal role in maintaining cellular homeostasis by
321 governing responses to diverse stimuli. Several studies have reported the promoter hypermethylation of
322 DAPK1 in OSCC, ranging from 18% to 27% [46-48]. This epigenetic modification hampers the expression
323 of DAPK1, potentially impeding its role in initiating apoptosis. DAPK1's significance spans both intrinsic
324 and extrinsic apoptotic pathways, reflecting its multifaceted involvement in orchestrating cell death
325 processes. Its precise control over apoptosis is crucial for preventing abnormal cell survival and
326 uncontrolled proliferation. In various cancers, particularly OSCC, the strong correlation between DAPK1
327 hypermethylation and the disease has been consistently demonstrated. The observed reduction in
328 DAPK1 gene expression due to hypermethylation indicates a blockade in the apoptosis process.

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330 **5.5. p53:**

331 The p53 gene, commonly known as the "guardian of the genome," serves as a pivotal tumor suppressor
332 protein present on chromosome 17p13.1. It has a pivotal role in controlling multiple cellular functions such
333 as cellular differentiation, cell cycle advancement, repair of DNA, and apoptosis, which is crucial for
334 preserving cellular balance. In response to unfavorable changes, whether arising from external or internal
335 factors, the expression of p53 is elevated, triggering a series of events that temporarily arrest cellular
336 functions, allowing for DNA repair. Significantly, in the context of OSCC and diverse cancer types, p53
337 gene mutations are commonly identified. The methylation status of p53's promoter region, an epigenetic
338 modification, falls within the range of approximately 25% to 69% [49]. This epigenetic alteration
339 contributes to the reduced activity of p53, compromising its tumor-suppressive functions. Moreover, p53,
340 as a critical regulator of apoptosis, exerts influence over the intrinsic (mitochondrial) apoptotic pathway. In
341 instances of cellular stress, DNA damage, or other unfavorable conditions, p53 becomes activated.

342 Activated p53 has a central role in triggering the mitochondrial apoptotic pathway. This pathway entails
343 the modulation of Bcl-2 family proteins, mitochondrial permeabilization, and the release of pro-apoptotic
344 factors, ultimately leading to the activation of caspases and the execution of programmed cell death. A
345 comprehensive understanding of the molecular intricacies involving p53 and the intrinsic apoptotic
346 pathway provides insights into OSCC pathogenesis and cancer development, offering potential avenues
347 for targeted therapeutic strategies [50-52].

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5.6. RASSF1 and RASSF2:

349 The Ras Association domain family proteins are components of the Ras/PI3K/AKT pathways. RASSF1 is
350 present on chromosome 3 at position 21.3, while RASSF2 is situated on chromosome 20 at position 13.
351 Studies involving patients undergoing radiotherapy revealed that in 50% of cases, activation of the
352 Ras/PI3K/AKT pathways occurred due to promoter methylation, leading to the silencing of the RASSF1A
353 and RASSF2A genes [53]. This suggests that the inactivation of RASSF1A and RASSF2A through
354 methylation-mediated gene silencing may lead to the initiation of cancer through the Ras/PI3K/AKT
355 signaling cascade, particularly in response to radiotherapy treatment. Researchers have embarked on a
356 promising endeavor to uncover biomarkers by discerning methylation patterns associated with tumor
357 invasion, particularly metastasis. Methylation levels of approximately 12-38% have been detected in the
358 RASSF1 gene, while at least one RASSF2 gene exhibits a methylation status of around 39%. This
359 exploration into methylation patterns within the RASSF1 and RASSF2 genes represents a significant
360 avenue for identifying potential biomarkers that could aid in understanding and monitoring tumor
361 metastasis. Moreover, elucidating the role of methylation in these genes may offer valuable insights into
362 the mechanisms underlying cancer progression and metastatic spread, presenting new opportunities for
363 targeted therapies and improved patient outcomes [54].

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365 **5.7. MGMT:**

366 MGMT also referred to as O6-methylguanine-DNA methyltransferase, is located at the gene locus
 367 10q26.3. This enzyme plays a crucial role in DNA repair by eliminating O6-guanine-DNA adducts induced
 368 by alkylating agents. Essentially, MGMT acts as a detoxifying agent by repairing DNA damage caused by
 369 these adducts. Histone modification serves as an alternative mechanism to DNA methylation, impacting
 370 gene expression in a nuanced manner. Unlike DNA methylation, which typically leads to gene silencing,
 371 histone modification can both silence and activate gene expression. It exerts this dual influence by
 372 altering the structure of chromatin, either tightening it to suppress gene transcription or loosening it to
 373 promote transcriptional activity. This dynamic interplay between histone modification and gene expression
 374 regulation adds a layer of complexity to epigenetic regulation and contributes to the intricate control of
 375 cellular processes [55].

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379 **Table 1. List of different types of apoptotic genes hypermethylated in OSCC along with**
 380 **their location, function and the pathway they follow.**

Gene	Location	Function	Pathway	References
Cadherin	Chromosome 16p22.1	Mediate cell-cell adhesion in epithelial tissues	Intrinsic apoptotic pathway	[56]
CASP8	Chromosome 2q33.34	Function as a tumor suppressor	Extrinsic apoptotic pathway	[57]
Survivin	Chromosome 17q25	Promote cell survival and inhibit apoptosis	Both intrinsic and extrinsic apoptotic pathways	[41]
DAPK1	Chromosome 9q34.1	Cellular processes, including apoptosis, autophagy, and cell migration	Intrinsic apoptotic pathway	[58]
p53	Chromosome 17p13.1	Regulate cell cycle progression, DNA repair, apoptosis, and senescence.	Intrinsic apoptotic pathway	[50-52]
RASSF1 and RASSF2	Chromosome 3p21.3 and Chromosome 20p13	Cellular processes, including cell cycle regulation, apoptosis, and tumor suppression.	Intrinsic and extrinsic pathways	[59]
MGMT	Chromosome 10q26.3	Repair damaged DNA caused by alkylating agents.	follows the DNA repair pathway rather than directly participating in apoptosis pathways	[60]

Apaf-1	Chromosome 12q23.1	regulate programmed cell death by facilitating the formation of the apoptosome, a large multiprotein complex.	Intrinsic pathway	apoptotic	[58]
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382 **6. FUNCTIONAL CONSEQUENCES OF HYPERMETHYLATION ON APOPTOTIC GENE**
383 **EXPRESSION**

384 Hypermethylation-induced alterations in apoptotic gene expression have significant functional
385 consequences in OSCC, impacting various aspects of tumor development, progression, and response to
386 therapy.

387 **6.1. Suppression of Pro-Apoptotic Genes:** Hypermethylation-mediated silencing of pro-apoptotic
388 genes such as Adenomatous Polyposis Coli (APC), Death-associated Protein Kinase 1 (DAPK1), and
389 Phosphatase and Tensin Homolog (PTEN) in OSCC results in the suppression of apoptosis, facilitating
390 the survival and growth of tumor cells [61].

391 **6.2. Disruption of Apoptotic Signaling Pathways:** Epigenetic alterations, including
392 hypermethylation, can disrupt apoptotic signaling pathways by downregulating key apoptotic regulators.
393 For instance, hypermethylation-mediated silencing of the Apoptotic Peptidase Activating Factor 1
394 (APAF1) gene inhibits the development of the apoptosome complex, impairing the initiation of
395 downstream effector caspases and compromising the apoptotic response [62].

396 **6.3. Resistance to Apoptosis Induction:** Hypermethylation-mediated downregulation of apoptotic
397 genes facilitates the development of resistance to apoptosis induction in OSCC cells. For example,
398 hypermethylation of the CASP8 (Caspase 8) gene attenuates caspase-8 expression, thereby impairing
399 extrinsic apoptosis signaling and rendering tumor cells less susceptible to death receptor-mediated
400 apoptosis [63].

401 **6.4. Enhanced Tumor Survival and Metastasis:** The dysregulation of apoptotic gene expression
402 due to hypermethylation confers a survival advantage to OSCC cells, facilitating tumor growth, invasion,
403 and metastasis. Hypermethylation-mediated silencing of genes like RASSF1A and p73 disrupts apoptotic
404 pathways, promoting tumor cell survival and facilitating metastatic spread.

405 **6.5. Impact on Therapeutic Response:** Epigenetic alterations in apoptotic genes also influence the
406 response of OSCC to therapeutic interventions. Hypermethylation-induced downregulation of apoptotic
407 genes can confer resistance to chemotherapy and radiotherapy, limiting the efficacy of these treatments
408 and contributing to disease recurrence and poor patient outcomes.

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410 **7. THERAPEUTIC IMPLICATIONS AND FUTURE PERSPECTIVES**

411 **7.1. Targeting Hypermethylation for OSCC Treatment:**

412 Targeting hypermethylation as a therapeutic strategy for oral squamous cell carcinoma (OSCC) holds
413 significant promise. One example of a demethylating agent is 5-azacytidine, which has been investigated
414 for its ability to inhibit DNA methyltransferases and promote DNA demethylation. Another agent,
415 decitabine, also functions similarly and has shown efficacy in other cancer types. In preclinical studies,
416 these agents have demonstrated the potential to reactivate silenced genes and enhance the
417 effectiveness of chemotherapy and immunotherapy in models of OSCC. Additionally, targeted therapies
418 focusing on specific hypermethylated genes such as APAF1 and PTEN are being explored. For instance,
419 researchers are investigating small molecule inhibitors that selectively target proteins involved in gene
420 silencing through DNA methylation or histone modification. These inhibitors offer a more targeted

421 approach to reversing hypermethylation and restoring gene function. Furthermore, combination therapies
422 that include demethylating agents along with conventional chemotherapy or novel targeted agents
423 represent a promising approach. By merging these substances, there is potential to attain synergistic
424 results, ultimately enhancing the treatment results for individuals diagnosed with OSCC.

425 **7.2. Epigenetic Therapy Approaches in OSCC:**

426 Epigenetic therapeutic approaches for OSCC target the modulation of gene expression through
427 modifications in DNA methylation, histone acetylation, and ncRNA expression. A particular approach
428 involves employing demethylating agents like 5-azacytidine and decitabine, which hinder DNA
429 methyltransferases and facilitate DNA demethylation. This process results in the restoration of tumor-
430 suppressor gene expression [64]. Histone deacetylase inhibitors are a potent category of epigenetic
431 therapy agents that have shown promise in OSCC. Drugs like vorinostat and romidepsin reverse histone
432 hypo acetylation, resulting in the re-expression of silenced genes and induction of apoptosis in OSCC
433 cells [65].

434 Emerging epigenetic targets for OSCC therapy include lysine-specific demethylase 1 (LSD1) and
435 bromodomain and extra-terminal domain (BET) proteins. BET inhibitors disrupt chromatin structure and
436 inhibit oncogenic transcriptional programs, showing efficacy in preclinical models of OSCC [66]. Similarly,
437 LSD1 inhibitors reverse histone methylation patterns and suppress tumor growth in OSCC xenografts.
438 Moreover, dysregulated microRNAs are being targeted for OSCC therapy using antagomirs or microRNA
439 mimics. These molecules play critical roles in OSCC pathogenesis and their modulation offers another
440 avenue for epigenetic-based therapeutic intervention [67]. These epigenetic therapy approaches hold
441 promise for OSCC treatment by targeting key molecular mechanisms involved in tumor development and
442 progression. Nevertheless, additional investigation and clinical trials are imperative to comprehensively
443 grasp their effectiveness and potential adverse reactions in individuals with OSCC.

444 **7.3. Challenges and Opportunities in Translating Research Findings into Clinical
445 Practice:** Translating epigenetic therapies into clinical practice for OSCC faces significant challenges. A
446 major obstacle is the absence of biomarkers predictive of treatment response, making it difficult to select
447 patients who will benefit from these therapies and monitor their efficacy. For example, in a study by
448 Kurokawa et al., the authors emphasized the need for biomarkers to predict the response to epigenetic
449 therapy in OSCC patients [68]. Additionally, off-target effects and toxicity associated with epigenetic
450 drugs, such as vorinostat and romidepsin, may limit their clinical utility [69]. Moreover, the heterogeneity
451 of OSCC and the intricate interplay between genetic and epigenetic alterations present challenges for
452 developing personalized treatment strategies. For example, in a review by Vered et al., the authors
453 discussed the complexities of addressing the genetic and epigenetic heterogeneity of OSCC in clinical
454 practice [70].

455 However, advancements in high-throughput sequencing technologies and bioinformatics tools offer
456 promise for identifying novel epigenetic biomarkers and optimizing treatment regimens in OSCC patients.
457 These tools enable researchers to analyze large datasets and identify potential biomarkers associated
458 with treatment response and disease prognosis [71]. Furthermore, collaborative efforts between basic
459 scientists, clinicians, and pharmaceutical companies are crucial for conducting well-designed clinical trials
460 to validate the efficiency and safety of epigenetic therapies in OSCC. By pooling expertise and resources,
461 these stakeholders can overcome challenges and accelerate the translation of epigenetic therapies from
462 the bench to the bedside [72].

463 **7.4. Future Directions and Emerging Strategies:**

464 Future directions in epigenetic therapy for oral squamous cell carcinoma (OSCC) entail investigating
465 combination treatments that target multiple epigenetic regulators and pathways. **By merging
466 demethylating agents, histone deacetylase inhibitors (HDACis), and targeted drugs, researchers aim to
467 surmount resistance mechanisms and enhance treatment efficacy in OSCC patients.
468 Several DNA methylation inhibitors have been developed, including 5-Aza-2'-deoxycytidine (5-Aza-dc).
469 This nucleoside DNMT inhibitor (DNMTi) works by being incorporated into DNA during replication. Once
470 integrated, it irreversibly binds to DNMT1, inhibiting its activity and leading to demethylation of DNA [73].**

471 For example, Cheng et al. (2009) demonstrated the effectiveness of a demethylating agent combined with
472 HDACi in inducing apoptosis in OSCC cells. Their study concluded that demethylation of the APAF-1
473 promoter induces apoptosis in OSCC both in vitro and in vivo. Specifically targeting the APAF-1 promoter
474 region for demethylation restored APAF-1 expression, promoting apoptotic cell death in OSCCs. These
475 findings suggest that APAF-1 promoter demethylation could serve as a potential therapeutic strategy for
476 inducing apoptosis and inhibiting tumor growth in oral squamous cell carcinoma. Furthermore, there is
477 growing interest in uncovering new epigenetic vulnerabilities and actionable targets through
478 comprehensive omics profiling and functional genomics studies.
479 This methodology will pave the way for the formulation of bespoke medical approaches precisely
480 customized to the unique needs of each OSCC patient. For instance, Singh et al. highlighted the
481 therapeutic potential of targeting lysine-specific demethylase 1 (LSD1) in OSCC.

482 Additionally, integrating epigenetic therapies with immunotherapy and radiotherapy holds promise for
483 achieving lasting responses and enhancing survival rates in OSCC patients. Preclinical investigations
484 have demonstrated that epigenetic modulation can amplify the effectiveness of immunotherapy agents,
485 such as immune checkpoint inhibitors, in OSCC. Moreover, ongoing research focuses on developing non-
486 invasive epigenetic biomarkers for early detection, prognosis, and monitoring treatment response in
487 OSCC. For example, Kurokawa et al. showed the potential of demethylating the APAF-1 promoter as a
488 biomarker for inducing apoptosis in OSCC cells. These advancements have the potential to transform
489 clinical management strategies for OSCC, ultimately leading to improved patient outcomes.

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494 **8. CONCLUSION:**

495 Essentially, the hypermethylation of apoptotic genes in OSCC signifies a fundamental molecular
496 alteration that significantly impacts the progression and development of this cancer type. The atypical
497 DNA methylation seen in OSCC interferes with the typical operation of apoptotic genes, facilitating the
498 avoidance of programmed cell death and fostering unrestricted cellular proliferation. This disruption,
499 especially impacting tumor suppressor genes linked to apoptosis, plays an important role in the initiation
500 and progression of OSCC. The intricate interplay between epigenetic modifications, specifically DNA
501 hypermethylation, and the apoptotic pathway highlights the intricate nature of OSCC. This disruption in
502 the apoptotic process is a characteristic feature of cancer, enabling malignant cells to persist and
503 proliferate. The identified hypermethylation patterns may extend beyond individual genes, indicating a
504 broader epigenetic landscape that supports the survival and expansion of OSCC cells. From a clinical
505 perspective, comprehending the significance of hypermethylation in apoptotic genes holds potential as a
506 biomarker for diagnosing, prognosis, and responding to treatment in OSCC. Targeted therapies aimed at
507 reinstating normal apoptotic functions could improve the efficacy of conventional treatments, opening new
508 avenues for managing OSCC. Furthermore, environmental factors and lifestyle choices, such as tobacco
509 and alcohol use, have been recognized as influential in shaping hypermethylation patterns in apoptotic
510 genes. This understanding of the interplay between external factors and epigenetic modifications provides
511 valuable insights for preventive strategies and personalized approaches to OSCC. To advance our
512 knowledge and therapeutic approaches, future research should concentrate on identifying the specific
513 apoptotic genes affected by hypermethylation in OSCC and deciphering the intricate regulatory networks
514 at play. Integrating multi-omics approaches and conducting extensive clinical studies will enrich our
515 understanding of the role of hypermethylation in OSCC.

516

517 **ABBREVIATIONS:**

- 518 1. OSCC- Oral squamous cell carcinoma
 519 2. NCDs- Non-communicable diseases
 520 3. HPV- Human papillomavirus
 521 4. RNAi- Ribonucleic acid interference
 522 5. DNMTs - DNA methyltransferases
 523 6. TSG - Tumor suppressor genes
 524 7. HDACs- Histone deacetylases
 525 8. ncRNAs- Non-coding RNAs

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527 12. REFERENCES

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