

ISOLATION AND CHARACTERIZATION OF GLUCOCORTICOID STERIOD FROM THE LEAF OF *RAUVOLFIA VOMITORIA*

Abstract

Isolation and characterization of glucocorticoid steroid was carried out on the leaf of *Rauvolfia vomitoria*. Initial phytochemical screening of the sample revealed the presence of Saponins, flavonoids, alkaloids, tannins, steroids, triterpenoids and cardiac glycoside and were quantitatively determined as follows; saponins (2.37 ± 1.09), tannins (0.09 ± 0.01), alkaloids (0.82 ± 1.05), and flavonoids (0.82 ± 0.05). Isolation of the compound was carried out with the aid of column chromatography and isolate subjected to thin layer chromatography. Sample was subjected to spectroscopic analysis with the aid of FT-IR, ^1H NMR, ^{13}C NMR and HSQC, the structure of compound was determined as Cis-3-(2,2-dimethyl-26-(((2S,3R,4S,5R,6R)-2,3,4trihydroxy-5-(hydroxymethyl)tetrahydro-2H-pyran-2-yl)oxy)butan-2-yl)-8-methoxy-15-(S)-3methoxy-4,4-dimethylpentyl)-14-(3-methoxy-4,4-dimethylpent-1-yn-1-yl)-

1,2,3,4,11,12,13,14,15, 15a-octahydrophenanthro[2,1-b]furan-16(10H)-one with a molecular formulae of $\text{C}_{44}\text{H}_{68}\text{O}_{11}$. A glucocorticoid steroid.

Key words; steroids, phytochemicals, column chromatography, glucocorticoids, spectroscopy

Introduction

The World Health Organization recognizes medicinal plant parts such as; leaves, roots, rhizomes, stems, barks, flowers, fruits, grains, or seeds as having therapeutic values. These parts are used in the management or treatment of disease conditions. The most useful chemical components are phytochemicals which are secondary plant metabolites. (1, 2, 3, 4). Medicinal plants has been used to treat a variety of conditions, such as fever, asthma, constipation, esophageal cancer, and hypertension (5). *Rauwolfia vomitoria* Afzel is mostly found in regenerating forests where extended periods of fallow are prevalent. It is a member of the Apocynaceae family. Within the Apocynaceae family, *R. vomitoria* Afzel is a small tree or big shrub that can reach a height of 15 meters. It features shiny, oval, or oblong leaves arranged in whorls, straight veins, and a cluster of small, white, or greenish flowers that yield red berries. While the Igbo people in Nigeria and the

Ashantes people in Ghana called *R. vomitoria* wada, its name is mmoneba (Efik), and utoenyin (Ibibio), respectively, the Yoruba people in Nigeria refer to it as "Asofeyeje," which means bearing fruits for the birds (6, 7, 8). The plant belongs to two distinct species: *R. vomitoria*, which is African variety, and *R. serpentina*, which is Indian variety (9). In natural medicine, this plant is also known for its several medicinal benefits, which include alleviating male infertility, hypertension, malaria, and mental disorders, among other conditions (10). The leaf of the plant is said to contain Alkaloids, saponins, and flavonoids (11). Though this plant has so much medicinal applications, its constituents are not fully characterized.

Materials and Method

Plant Collection and Preparation

The leaves of *R. vomitoria* were obtained in December 2021 from a thicket near Yaradua Drive in Owerri, Imo state, in eastern Nigeria. The plant sample, bearing authentication number UBHR421, was identified by Mr. Obiajunwa, Udoka of Forest Ecology and Conservation in the Department of Forestry

and Environmental Management at Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria. They were air-dried, coarsely ground with a hand blender, and then stored in an airtight container. 800 g of the sample was measured into a glass jars and 3.8 litres of analytical-grade chloroform solvent were added until the leaf samples were fully submerged. After that, the glass jars were securely sealed and stored for five days in a secure location. Using Whatman No. 1 filter paper, the extract was filtered out of the solution after the extraction procedure. The solvent and extract mixture were separated using a rotary evaporator.

PARTITIONING OF THE CRUDE CHLOROFORM EXTRACT

20 g of the crude chloroform extract of the sample was partitioned using 200 cm³ of chloroform and 200 cm³ of distilled water. The crude extract was dissolved first with the solvent which was continuously stirred to ensure complete dissolution and 200 cm³ of water was added. The mixture was then poured into a separation funnel, shaken vigorously until a homogenous mixture was obtained and left to stand for 24 hours to separate. The chloroform extract was allowed to evaporate completely. The dried chloroform fraction was mixed with 50 g silica gel (200 mesh) to form a homogenous mixture called slurry for column chromatography.

COLUMN CHROMATOGRAPHY

The glass column was washed and rinsed with petroleum ether. A plug of cotton wool was introduced into the glass column and was pushed to the neck of the column with a steel rod to prevent silica gel from entering into the collection vials and to provide an even baseline for the slurry. The silica gel (200 mesh) was mixed with petroleum ether and poured into the column. This was repeated until a lengthy column was obtained. The column was allowed to drop to ascertain the smooth flow of the solvent. Then the slurry was gently introduced into the column and 100 cm³ of petroleum ether was added. poured/ gradually to wash down the sides of the column. After which

Silica gel was introduced and to prevent disturbance to the plant sample when pouring the different solvents. the collection of fractions in 40 cm³ was carried out started with 100 cm³ beakers after the introduction of 100 cm³ petroleum ether. followed by introduction of solvent mixture of petroleum ether and chloroform (90:10). The column was gently tapped with a rubber hose to release trapped air bubbles in order to forestall column cracking. After which the process continued with petroleum ether: chloroform (80:20, 70:30, 60:40, 50:50, 40:60, 30:70, 20:80, 10:90) and 100 cm³ chloroform. Then chloroform and ethyl acetate (90:10, 80:20, 70:30, 60:40, 50:50, 40:60, 30:70,

20:80, 10:90) and 100 cm³ ethyl acetate. Followed by ethyl acetate and methanol (90:10, 80:20, 70:30, 60:40, 50:50, 40:60, 30:70, 20:80, 10:90) and lastly 100 cm³ methanol. 68 fractions were collected in total.

THIN LAYER CHROMATOGRAPHY

The sample fractions were subjected to preliminary Thin Layer chromatography (TLC) which was used to determine the purity and number of compounds that could be present in each of the fractions collected from column chromatography. With the use of a capillary tube, each of the fractions was applied on pre-coated aluminum thin-layer chromatographic plates and at half inch apart from the lower edge of the plate. The plates were kept in different developing chambers of 250 cm³ beakers containing solvent mixtures for specific time until the developing solvent reaches top of the upper edge of the TLC plate. The plates were taken out from developing chamber, dried and solvent front was marked with lead pencil. The compound bands/spots on TLC chromatographic plates were visualized in Iodine chamber and marked. The retention factor (R_f) of each spot was calculated using the formula;

$$R_f = \frac{\text{Distance travelled by the sample}}{\text{Distance travelled by the solvent}}$$

Afterward, the sample fraction was obtained with R_f value of 0.75. The fraction was subjected to spectral analysis such as Infra-red (IR) and Nuclear Magnetic Resonance (NMR).

Screening for alkaloid

5g of the sample was extracted using 20% acetic acid in ethanol .5cm³ of the extract was treated with Wagner's reagent (iodine crystals and KI). A yellowish brown precipitate indicates the presence of alkaloid. Mayer's reagent (potassium mercuric iodide) was added to 10cm³ of the filtrate. Alkaloids were present when a yellow-colored precipitate formed (12).

Screening for saponins

Froth Test: 10 g of sample was dissolved in 100 cm³ of water and extracted after 4 hours. 10 cm³ of the extract was measured into a 50 cm³ beaker and shaken vigorously. The formation of a 1 cm layer of foam suggested the presence of saponins (12).

Screening for cardiac glycosides

Chloroform was used to treat the extract before it was filtered. The filtrate was then heated and cooled shortly after being treated with a few drops of acetic anhydride. Sulfuric acid concentrate was added. The presence of cardiac glycosides was revealed by the formation of a brown ring at the junction (12, 13).

Screening for Steroids

Salkowski's Test: Chloroform treatment and filtering were applied to the extract. To create a lower layer, a few drops of concentrated sulfuric acid were then added to the filtrate. The presence of steroids were indicated by a golden-yellow appearance.

Screening for tannins

Gelatin Test: A 1% sodium chloride-containing gelatin solution was added to the water extract. The presence of tannins was revealed by the formation of a white precipitate (13).

Screening for flavonoids

Shinoda's Test: The extract was dissolved in ethanol, and the filtrate was then mixed with a few drops of conc. HCl. After that, magnesium turnings were added. Flavonoids were detected by the formation of a pink-red coloring (12).

Screening for terpenes

After adding 3cm³ of chloroform to the extract, it was filtered. 3cm³ of concentrated sulfuric acid was added to the filtrate. Pink colouration of the interphase indicates the presence of terpenes. (12)

Alkaloids determination

A 250 cm³ beaker was filled with 15 g of the sample, 200 cm³ of 20% acetic acid in ethanol, and agitated. The beaker was then capped and allowed to stand for four hours. The Whatman filter paper was used to filter the mixture, and a water bath was used to concentrate the extract to a quarter of its initial volume. Concentrated ammonium hydroxide was applied drop by drop until the precipitation process was completed to remove the alkaloid. Using weighed Whatman filter paper,

the precipitate was collected, using the filtering process. After dried and the precipitate was weighed again.

Saponins determination

200 cm³ of a 20 % aqueous ethanol solution was added to a 250 cm³ beaker containing 15 g of the sample, which was then swirled with a glass rod. The mixture was heated over a water bath for four hours, stirring continuously to keep the temperature at 55 °C. The Whatman filter paper was used to filtered the mixture, and 200 cm³ of 20 % aqueous ethanol solution was used to extract the residue for one hour. The extracts were combined and boiled in a water bath at 90 °C until they weighed about 40 cm³. After transferring the concentrated extract into a 250 cm³ separation funnel, 20 cm³ of diethyl ether was added, and the mixture was vigorously shaken. The ether layer was laid out, and Partitions were formed and so that the aqueous layer was could then be recovered. Three times this technique was carried out, and n-butanol (60 cm³) was added. 10cm³ of 5 % sodium chloride were used to wash the mixture twice. In a weighed evaporation dish, the residual solution was heated to dryness over a water bath and then reweighed. The saponin content was determined.

Flavonoids Determination

15 g of the plant material was extracted several times using 100 cm³ of 80% aqueous methanol. Whatman filter paper No. 45 was used to filter the resultant solution. After being poured into a crucible, the filtrate was dried over a water bath and weighed (14). The flavonoid content was calculated.

Determination of tannins

A beaker was filled with 15 g of the sample. It was mixed with 150 cm³ of distilled water and allowed for four hours. After placing a stirring rod and allowing the liquid to sit for four hours, the Whatman

filter paper was used to filter. The mixture was filtered and After adding a few drops of concentrated hydrochloric acid was added to the filtrate to make it more acidic, then ethyl acetate was added. Using a separating funnel, this solution was thoroughly shaken and partitioned. The ethyl acetate layer was disposed of and an aqueous solution was produced. Following three repetitions of this, the aqueous solution was boiled to dryness, and the tannins that were produced were weighed. The quantity of tannins were calculated.

Results and discussion

Qualitative Screening of *R. vomitoria* leaf

The result of the initial assessment of the phytochemical composition of *R. vomitoria* leaves is contained in table 1 below.

Table 1. Result of phytochemical screening of *R. vomitoria* leaves

Phytochemical	Inference
Saponins	++
Flavonoids	++
Tannins	++
Alkaloids	++
Cardiac glycosides	+
Steroids	++
Terpenoids	++

Key; ++ present, + slightly present, – absent the results of the qualitative phytochemical screening of the *R. vomitoria* leaf extract reveals the presence of saponins, flavonoids, alkaloids, tannins, steroids, and triterpenoids, with a slight presence of cardiac glycoside (11). Most of these compounds were quantitatively determined in table 2.

Table 2. Result of the quantification of *R. vomitoria* leaves.

Phytochemical	Concentration mg/100g
Alkaloids	0.82 ± 1.05
Saponins	2.37 ± 1.09
Flavonoids	0.82 ± 0.05
Tannins	0.09 ± 0.01

Values are means ± standard deviation of triplicate determination

Table 2 shows that the extract from *R. vomitoria* leaves includes different levels of tannins, alkaloids, flavonoids, and saponins. When compared to alkaloids and flavonoids, tannins have the lowest concentration in *R. vomitoria* leaf extract, whereas saponins have a substantially larger concentration. The majority of phytochemicals are present in plant-based diets; they are absent from dairy and animal products. Most green leafy plants have been reported to contain alkaloids, which have a wide range of medicinal uses. These are secondary metabolites that have long since been discovered to be crucial for human health. They are found in foods and beverages consumed by humans daily, as well as in stimulant medications. According to (15), the majority of alkaloids have been used medicinally for their antimalarial, antioxidative, antibacterial, anti-inflammatory, anticancer, antiasthma, vasodilatory, analgesic, and neuropharmacological properties. Because of

the alkaloid concentration in the leaf extract, *R. vomitoria* leaves are used to treat fever, hypertension, jaundice, and rheumatism.

Groups of secondary metabolites of polyphenolic plants are called flavonoids. They add to the color of herbs, vegetables, and fruits. Among the bioactive flavonoids are quercetin, which has antibacterial, antiviral, anticarcinogenic, and anti-inflammatory properties, (17), and hesperidin, which has an antihypertensive and lipid-lowering impact (16). Therefore, these properties justify the application of *R. vomitoria* leaves in inflammatory and cancer therapies (18).

Among the phytochemicals found in plants are saponins. These are chemical substances found in many different kinds of plants, seeds, and herbs. It was discovered that *R. vomitoria* leaves have a high availability of saponins. They boast several health benefits and can foam when combined with water. To control immunological responses, saponins are used in vaccination formulations. By protecting the body from bacteria, viruses, and fungi, saponins have antibacterial, antiinflammatory, and immune-boosting qualities. They scavenge oxidative stress and function as antioxidants. Their hypoglycemic properties help to minimize insulin surges and maintain normal blood sugar levels. Saponins can reduce body fat and cholesterol, which helps to improve cardiovascular health. Both humans and animals have shown them to have a cholesterol-lowering effect (19). Many biological and therapeutic effects, including hemolysis, molluscicidal, anthelmintic, insecticidal, analgesic, antiviral, anticancer, sedative, and anti-tumor activity, have been found for saponins (20). It is widely believed that they are crucial to plants' defense against illnesses and insects. They are also utilized in the food supplement and cosmetic (soap, emulsions) industries, as well as in several pharmaceutical and medicinal applications (21). *R. vomitoria* leaves contain saponin, which makes them useful for treating conditions including headaches, inflammation, high blood pressure, and chest discomfort.

Plants such as *R. vomitoria* include flavonoids, alkaloids, and saponins that have enzymatic, antioxidant, anti-inflammatory, and immunomodulatory properties that can either delay or improve rheumatoid arthritis.

One type of phytochemical in the plant extract from *R. vomitoria* is steroids. They support numerous therapeutic uses, including those of cardiotoxic (digitoxin), vitamin D precursors (ergosterol), oral contraceptives (semisynthetic estrogens and progestins), anabolics (androgens), and anti-inflammatory (corticosteroids). Since inflammation is the primary cause of asthma, corticosteroids are mostly used to treat severe cases of the condition (22). This suggests that *R. vomitoria* plant leaves may be used similarly to treat inflammatory illnesses by acting as an antiinflammatory drug.

Tannins are a type of polyphenol that has several therapeutic and medical uses. They also function as antioxidants and have some pharmacological qualities, including the ability to heal wounds, prevent dysentery, fight bacteria, reduce inflammation, and be anti-toxic, anti-cancerous, antiallergic, and anti-inflammatory (23). In addition, tannins have several negative consequences, such as increased indigestibility, antinutritional effects, mutagenesis, and carcinogenic properties, ability to cause and co-promote several diseases, migraine intensity, hepatotoxic activity, and inhibitory actions when consumed in large quantities. Additionally, they have been used to treat throat infections, hemorrhoids, diarrhea, and dysentery (24). Since tannin was discovered to be somewhat prominent in *R. vomitoria* leaves, the plant's leaves may therefore be used as a treatment for the aforementioned illnesses.

Cardiac glycosides are steroid-based medications that lower heart rate and increase heart muscle contraction force without increasing oxygen consumption in the treatment of congestive heart failure and cardiac arrhythmia. Apart from their beneficial effects on heart health, cardiac

glycosides also exhibit anticancer capabilities (25). They do this by activating the heart muscle, which enhances cardiac output and efficiency. They also aid in the deceleration of irregular heartbeats. They are found in trace levels in the leaves, seeds, stems, roots, and bark of widely distributed plants. It was discovered that *R. vomitoria* leaves had a small amount of cardiac glycoside, which may be used to treat cardiovascular illnesses.

Triterpenes are found in several ethnomedicinal plants, including the leaves of *R. vomitoria*. Traditional healers regularly recommend herbal medicines containing triterpenes to treat or prevent a wide range of illnesses in several Asian nations (26, 27). In nutraceutical products, triterpenes can reduce pain and have anti-inflammatory, antiviral, antibacterial, anticancer, and immunomodulatory properties. As a result, *R. vomitoria* leaves can be used to treat a variety of illnesses, including malaria, rheumatism, diarrhea, and fever.

CHARACTERIZATION OF COMPOUND

The Compound was proposed from the complete elucidation of spectroscopic parameters which include; FT -IR, ^1H NMR, ^{13}C NMR, and HSQC.

FT-IR Spectral data of Compound

The spectrum of compound is presented in Figure 1.

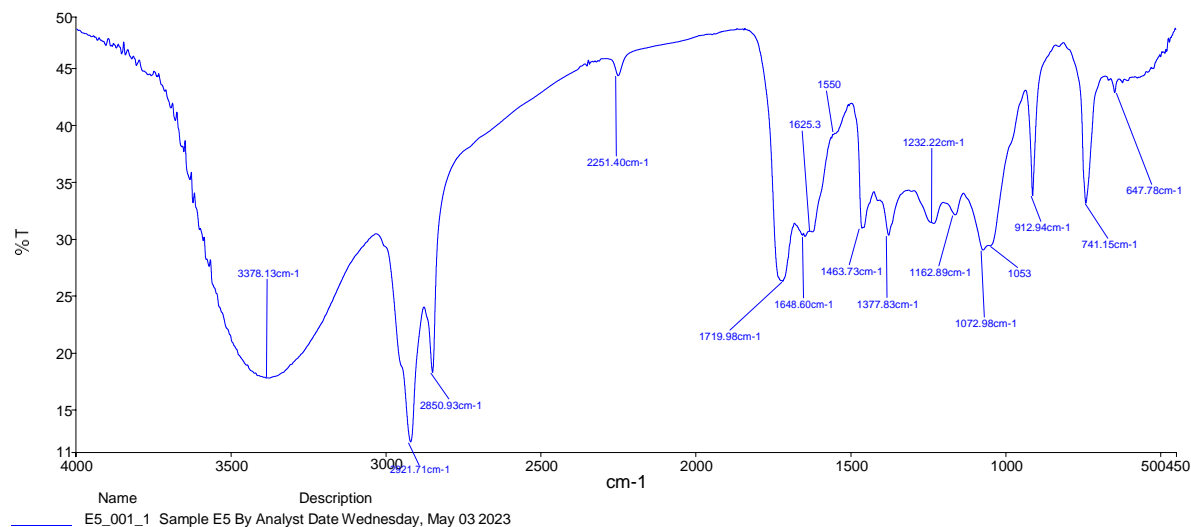


Figure 1: FTIR Spectrum of compound

The interpretation of the FT-IR spectra of compound is shown in Table 3.

Table 3: Interpretation of FT-IR spectra for compound

<u>Frequency of Absorption (cm⁻¹)</u>	<u>Characteristics/Intensity</u>	<u>Type of bond/Vibration</u>
3378.13		broad and intense -O-H stretching vibration of an alcohol
2921.71	sharp and intense	-C-H stretching vibration of an aliphatic
2850.93		
2251.40	medium and sharp	C≡C stretching vibration of an alkyne
1719.98		medium and sharp C=O stretching of an ester
1648.60	medium and sharp	C=C stretching of an aromatic
1625.30		
1550.00		
1463.73	medium and sharp	C-H deformation of CH ₂ and CH ₃
1377.83		
1162.89	medium and broad	C-O stretching vibration of an alcohol and an ether
1072.98		
1053.00		
912.94	medium and sharp	C-H deformation of an aromatic
741.15		
647.78		

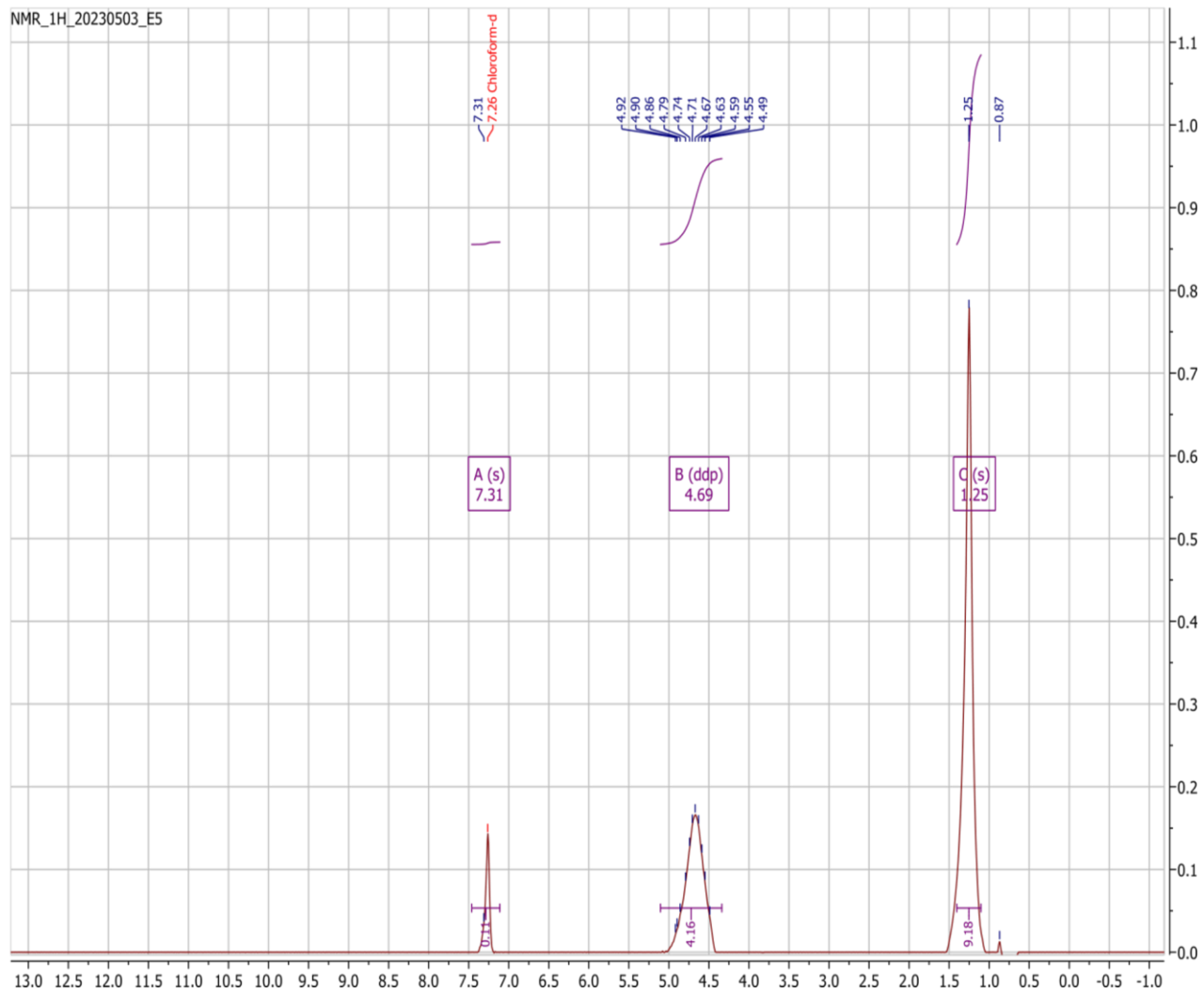
Table 3 shows data from the FT-IR spectrum of compound. The strong, broad, and intense absorption band at 3378.13 cm^{-1} depicted an O-H stretching of an alcohol. The broadness of the band is due to the intermolecular hydrogen bond that exists among the O-H groups of the sugar moiety (28). The sharp and intense absorption bands at 2921.71 cm^{-1} and 2850.93 cm^{-1} depicted the presence of C-H stretching vibration of an aliphatic group, which implies that hydrogen was attached to sp^3 carbons. A medium and sharp absorption band at 2251.40 cm^{-1} indicated the presence of a substituted $\text{C}\equiv\text{C}$ stretching vibration of an alkyne group. The strong and intense absorption at 1719.98 cm^{-1} indicated the presence of a C=O stretching vibration of an ester (29). Bands at 1648.60 cm^{-1} , 1625.30 cm^{-1} and 1550.00 cm^{-1} with medium and sharp intensity, depicted the presence of a C=C stretching of aromatic (30). Also, the medium and sharp bands at 1463.73 cm^{-1} and 1377.83 cm^{-1} , depicted the C-H deformation, that is, bending vibration of $-\text{CH}_2$ and $-\text{CH}_3$ respectively. Again, the medium and broad bands at 1162.89 cm^{-1} , 1072.98 cm^{-1} , and 1063.00 cm^{-1} indicated the presence of a C-O stretching of an alcohol and an ether. The medium and sharp bands at 912.94 cm^{-1} , 741.15 cm^{-1} , and 647.78 cm^{-1} , also, depicted a C-H bending vibration of aromatic (31).

presented in Figure 3.

Figure 2: ^1H NMR spectrum of compound

^1H NMR and ^{13}C NMR of Compound

The ^1H NMR spectrum of compound is shown in Figure 2 while the ^{13}C NMR spectrum is



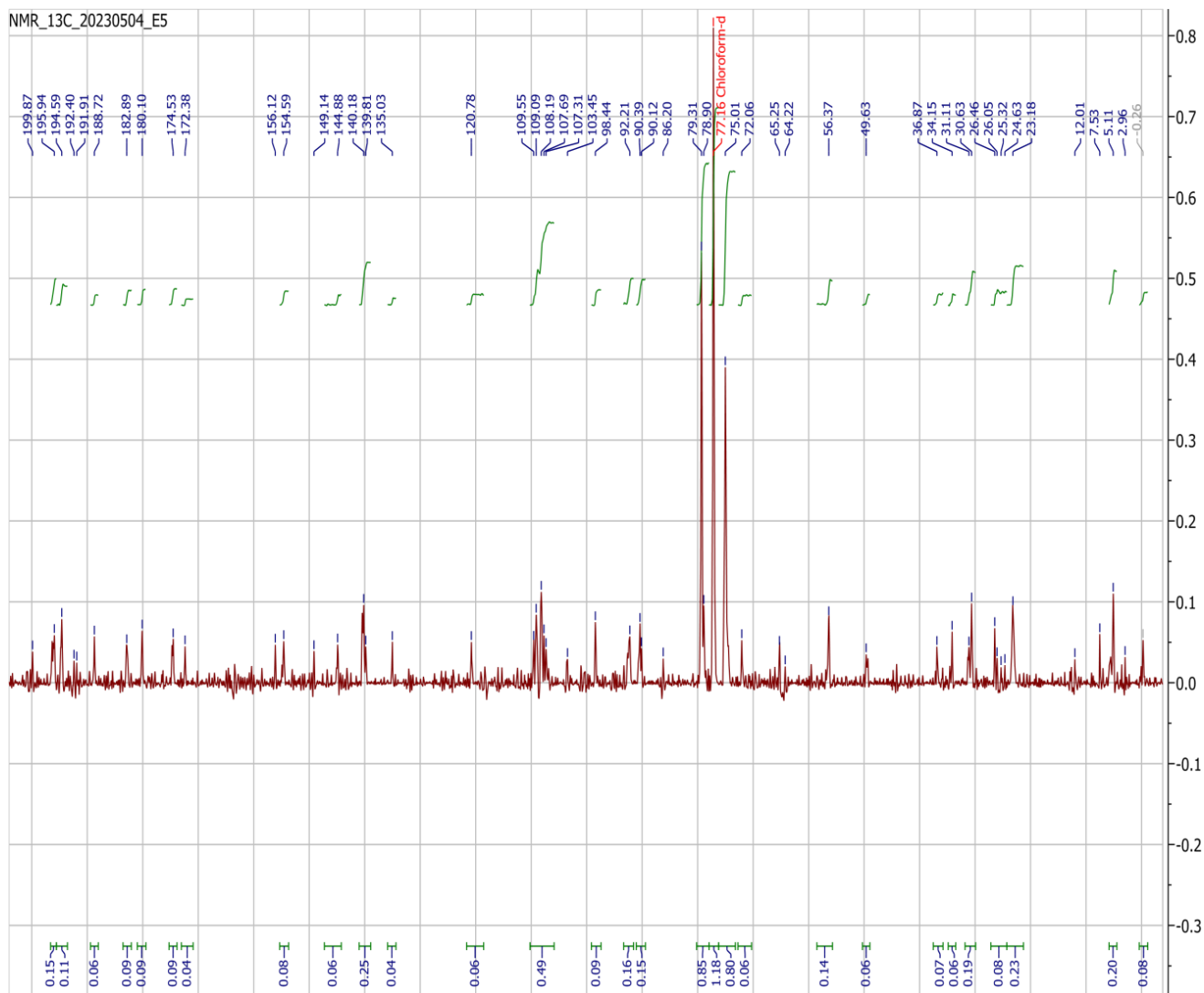


Figure 3: ¹³C NMR spectrum of compound The interpretation of the ¹H NMR and ¹³C NMR spectra of compound are shown in Table 4.

Table 4: ¹H NMR and ¹³C NMR of compound

	δC	δH	CHEMICAL	CHEMICAL	
	SHIFT	SHIFT	SHIFT	SHIFT	
	VALUE	VALUE	VALUE	VALUE	NO. OF
	POSITION (ppm)	CARBON (ppm)	TYPE OF	PROTON	MULTIPLICITY
1	30.63	CH ₂	1.60	2H	T
2	23.18	CH ₂	1.42	2H	Q

3	34.15	CH	1.71	1H	S
4	36.87	CH ₂	1.52	2H	D
5	120.78	Ar-C	-	-	-
6	120.78	Ar-C	- - - 7	109.55	Ar-H 7.31 1H S
8	135.03	AR-C-O	-	-	-
9	120.78	Ar-C	-	-	-
10	120.78	Ar-C	- - - 11	36.87	CH 1.81 1H T
12	65.25	CH-O	3.72	1H	Q
13	31.11	CH ₂	1.63	2H	T
14	49.63	CH	1.85	1H	T
15	86.20	CH	2.20	1H	Q
16	172.38	C=O	- - - 17	25.32	CH ₂ 1.95 2H Q
18	26.46	CH ₂	2.00	2H	T
19	79.11	CH-O	3.82	1H	M
20	64.22	C	-	-	-
21	90.12	O-CH ₃		3.51	3H S
22	12.01	CH ₃	1.25	3H	S
23	12.01	CH ₃	1.25	3H	S
24	12.01	CH ₃	1.25	3H	S
25	92.21	O-CH ₃		3.71	3H S
26	90.36	CH-O	3.82	1H	D
27	56.37	C	-	-	-
28	12.01	CH ₃	1.25	3H	S
29	12.01	CH ₃	1.25	3H	S
30	12.01	CH ₃	1.25	3H	S
31	75.01	C≡C	-	-	-
32	75.01	C≡C	- - - 33	98.44	CH-O 3.94 1H S
34	72.06	C	-	-	-
35	90.12	O-CH ₃		3.51	3H S
36	12.01	CH ₃	1.25	3H	S

37	12.01	CH ₃	1.25	3H	S
38	12.01	CH ₃	1.25	3H	S
1', 2', 3', 4', 5' & 6'	98.44	C ₆ H ₁₁ O ₆	4.69	11H	Ddp

s = singlet, d = doublet, t = triplet, q = quartet, ddp = doublet of doublet of pentet and m = multiplet

¹H NMR spectrum of compound revealed different kinds of proton present in compound E5. It also showed a cluster of peaks between 0.50 ppm to 2.50 ppm, which is an indication that compound is a steroid, and a cluster between 3.50 ppm to 6.00 ppm, which is also an indication that compound contains a sugar moiety (28). The protons revealed by the spectrum include; methyl proton (CH₃) presented in positions H-22, H-23, H-24, H-28, H-29, H-30, H-36, H-37, and H-38 with a resonance frequency signal at δ 1.25 ppm. The protons are chemically and magnetically equivalent which appeared as singlet (s), shielded by electron density, and also in the same environment. Methoxy protons (O-CH₃) in positions H-21 and H-35 are in the same environment and thereby resonated at δ 3.51 ppm, experienced de-shielding also, by inductive effect. H-25 (OCH₃) at δ 3.71 ppm is also de-shielded by inductive effect.

Another type of proton revealed by the spectrum is methylene proton (CH₂) presented in positions H-1 at δ 1.60 ppm with a triplet (t) multiplicity. It is in an electron-rich environment. H-2 which resonated at δ 1.42 ppm with a quartet (t) multiplicity, is also a methylene proton that is shielded by electron density. H-4 at δ 1.52 ppm, with a doublet (d), is shielded by electrons. H-13 at δ 1.63 ppm with the integration of 2H and splitting pattern of triplet (t), is also, shielded by electrons. H-

H-17 and H-18 at δ 1.95 ppm and δ 2.00 ppm with a quartet (q) and a triplet (t) respectively, are also methylene protons in an electron-dense environment.

Another type of proton detected in the spectrum is a methine proton (CH), which is presented in the positions H-3, H-11, and H-14 at δ 1.71 ppm, δ 1.81 ppm and δ 1.85 ppm with a singlet (s), a triplet (t) and a triplet (t) multiplicities respectively. They are also, in electron-rich environments. H-12 (CH-O), H-15 (CH), H-19 (CH-O), H-26 (CH-O) and H-33 (CH-O), with signals at δ 3.72 ppm, δ 2.20 ppm, δ 3.82 ppm, δ 3.82 ppm and δ 3.94 ppm and their multiplicities as quartet (q), quartet (q), triplet (t), doublet (d) and singlet (s) respectively. They are de-shielded inductively and anisotropically. Another proton detected in the spectrum is the aryl proton (Ar-H), presented in position H-7 with a resonance frequency signal at δ 7.31 ppm and a singlet (s) multiplicity. It is de-shielded by the anisotropic effect of the benzene ring. An overlap represented in positions H-

H-1, H-2, H-3, H-4, H-5, and H-6¹ at a signal of δ 4.69 ppm with a doublet of doublet of pentet (DDP) multiplicity indicated that a sugar moiety is attached to the aglycone unit.

¹³C NMR spectrum of compound revealed different kinds of carbon atoms in compound. These carbon atoms include; methyl (CH₃) carbon atoms, which are represented in positions C-22, C-23,

C-24, C-28, C-29, C-30, C-36, C-37, and C-38 with a chemical shift value of δ 12.01 ppm, which depicted that the carbons are equivalent and are in the same environment: this is analogous to tertbutyl group. C-21 and C-35 (CH₃O) at δ 90.12 ppm, are methoxy carbon atoms that are in the same environment and are de-shielded by inductive effects. C-25 at δ 92.21 ppm is another methoxy carbon that is de-shielded by the inductive effect of an oxygen atom.

Another type of carbon atom revealed by the spectrum is methylene (CH₂) carbon atoms which are represented in positions C-1, in an electron-dense environment with a resonance frequency at δ 30.63 ppm. C-2 at δ 23.18 ppm is another methylene carbon atom in an electron-dense environment,

including C-4 with a signal at δ 36.87 ppm, shielded by electron density, C-13 at δ 31.11 ppm, also shielded by electron density, C-17 at δ 25.32 ppm, also shielded by electron density and C-18, which resonates at δ 26.46 ppm, also in an electron-rich environment (28). Another type of carbon atom revealed by the spectrum is methine (CH) carbon atoms which are represented in positions C-3 at δ 34.15 ppm, shielded by electron density, C-11 at δ 36.87 ppm, shielded, C-12 (CH-O) at δ 65.25 ppm, de-shielded by inductive effect of an electronegative atom, oxygen, which tends to pull electrons close to itself, C-14 at δ 49.63 ppm is de-shielded by anisotropy, because, it is bonded to a carbon atom bearing a triple bond. C-15 at δ 86.20 ppm, is highly de-shielded by the anisotropic effect of C=O. Again, C-19 (CH-O) is an oxymethine that resonates at a high-frequency signal of δ 79.11 ppm, and it is de-shielded by the inductive effect of an oxygen atom. Also, C-26 at δ 90.36 ppm, is highly de-shielded by the inductive effect of an oxygen atom, and C-33 (CH-O) at δ 99.44 ppm, is also, highly de-shielded by anisotropy of the triple bond (28).

Another type of carbon atoms revealed were aryl carbons (Ar-C), which are the aromatic carbon atoms and are represented in positions C-5, C-6, C-9, and C-10 at a resonance frequency signal of δ 120.78 ppm, which indicated that they are in the same environment and are de-shielded by the anisotropic effect of the benzene ring. C-7 (Aromatic proton-H) at δ 109.55 ppm is de-shielded by anisotropy and C-8 (Ar-C-O) at δ 135.03 ppm is de-shielded by both anisotropic and inductive effects (31). Another type of carbon revealed is quaternary carbons which are represented in positions C-20 (C) at δ 64.22 and it is de-shielded. C-31 and C-32 (C \equiv C) are substituted alkyne carbon atoms that resonated at δ 75.01 ppm and are de-shielded by anisotropy. C-34 at 72.06 ppm, is also de-shielded by 2-J coupling to a methoxy carbon and 2-J coupling to a carbon atom bearing a triple bond, that is, it is de-shielded by both inductive and anisotropic effects. Another type of carbon revealed by the spectrum is a carbonyl group (C=O) which resonated at δ 172.38 ppm. It is highly de-shielded by anisotropic effect. A cluster of peaks at δ 98.44 ppm depicted the presence of a sugar moiety bonded to the aglycone unit (31).

HSQC of Compound

The Figure 4 shows the HSQC spectrum of compound displaying various correlations between different carbon atoms and the protons that are bonded to them.

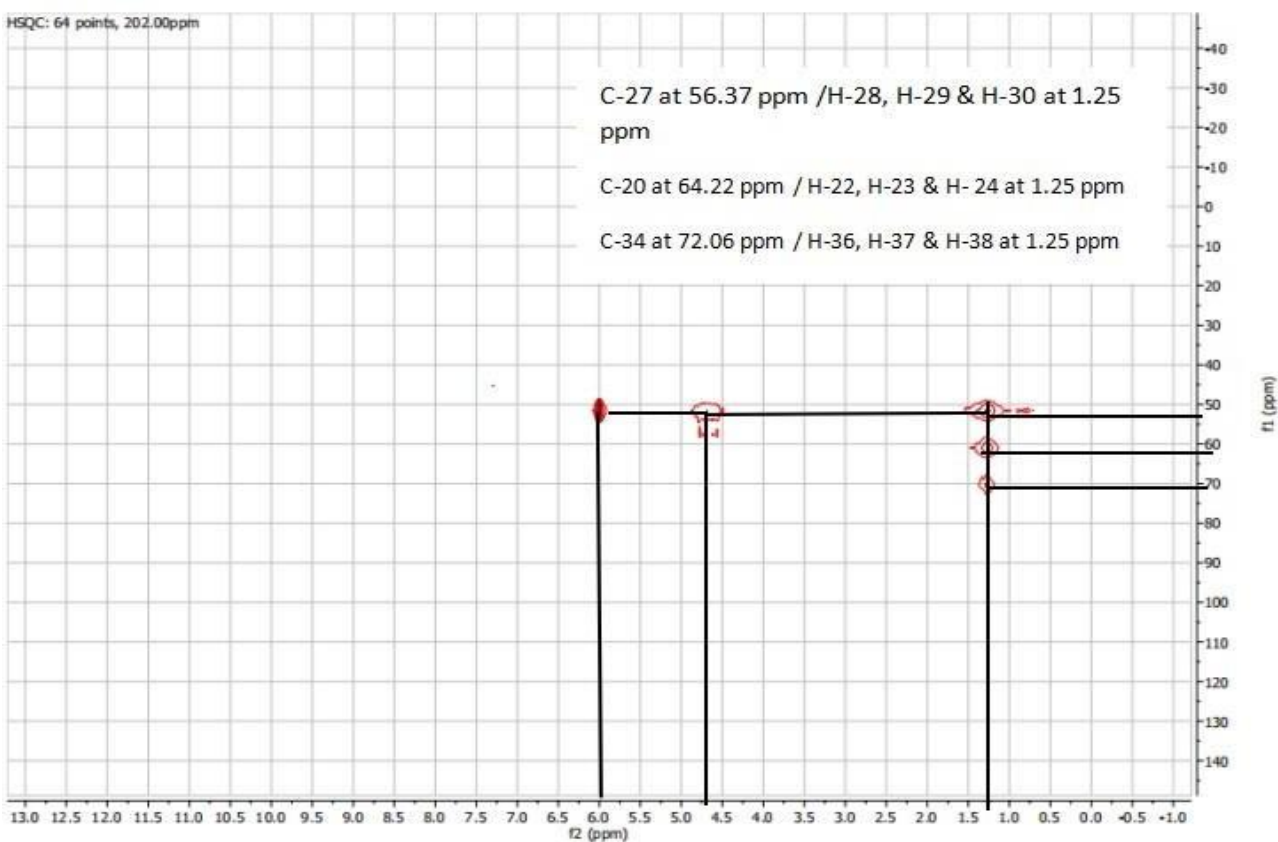


Fig .4 HSQC spectrum of compound

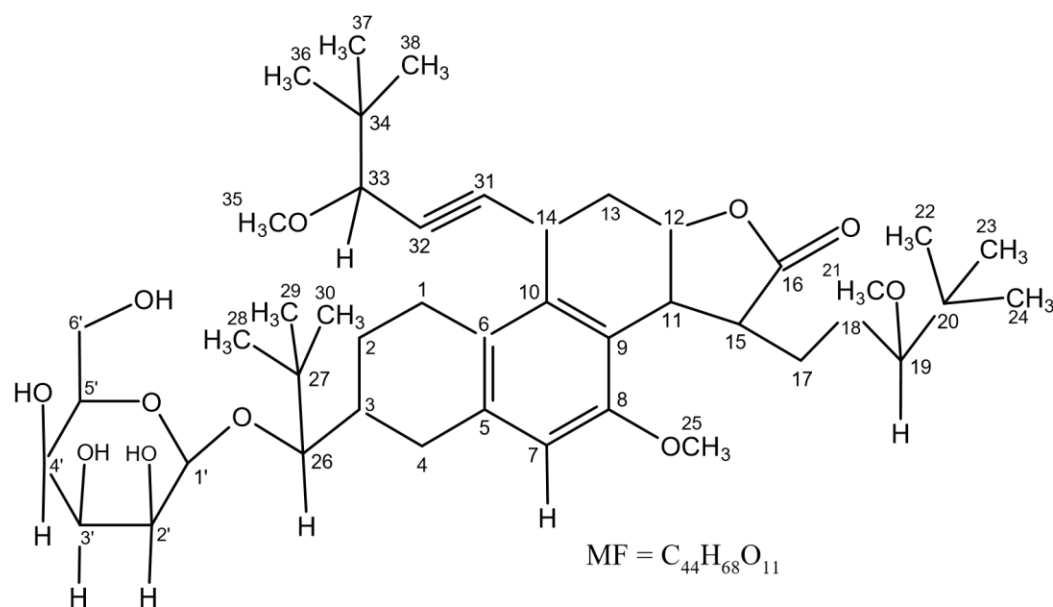
The HSQC spectrum of compound revealed a cross peak at δ 56.37 ppm which correlated with the proton at δ 1.25 ppm with a singlet multiplicity, and this corresponded to C-27 bonded to three methyl protons at H-28, H-29, and H-30. Another cross peak at δ 64.22 ppm which also correlated with the same proton at δ 1.25 ppm with a singlet (s) multiplicity, corresponded to C-20, bonded to

H-22, H-23, and H-24. Another cross peak was observed at δ 72.06 ppm which also correlated with the same proton at 1.25 ppm and this also, corresponded to C-34 and H-36, H-37 and H-38 that were bonded to it. This implies that the protons are chemically and magnetically equivalent and this depicted that three tert-butyl groups were attached to three different quaternary carbons.

The cross peak at δ 56.37 ppm also correlated with the proton at δ 4.69 ppm and δ 6.00 ppm which corresponded to the sugar moiety.

The compound is a glucocorticoid steroid with the IUPAC nomenclature, *Cis*-3-(2,2-dimethyl-26-(((2*S*,3*R*,4*S*,5*R*,6*R*)-2,3,4-trihydroxy-5-(hydroxymethyl)tetrahydro-2*H*-pyran-2*yl*)oxy)butan-2-yl)-8-methoxy-15-((*S*)-3-methoxy-4,4-dimethylpentyl)-14-(3-methoxy-4,4dimethylpent-1-yn-1-yl)-1,2,3,4,11,12,13,14,15, 15*a*-octahydrophenanthro[2,1-*b*]furan-16(10*H*)one, with a molecular formular of $C_{44}H_{68}O_{11}$. It is a glyco steroid because, there is an attachment of a sugar moiety to the aglycone unit, through a β - glycosidic linkage between the anomeric carbon, C-1¹ of the sugar, and the C-26 of the aglycone unit.

STRUCTURE OF COMPOUND 1 (E5)



Cis-3-(2,2-dimethyl-26-(((2*S*,3*R*,4*S*,5*R*,6*R*)-2,3,4-trihydroxy-5-(hydroxymethyl)tetrahydro-2*H*-pyran-2-yl)oxy)butan-2-yl)-8methoxy-15-((*S*)-3-methoxy-4,4-dimethylpentyl)-14-(3-methoxy-4,4-dimethylpent-1-yn-1-yl)-1,2,3,4,11,12,13,14,15, 15a-octahydrophenanthro[2,1-*b*]furan-16(10*H*)-one

Fig 5. Proposed structure of compound

Conclusion

The study of the leaf of *R. vomitoria* yielded a glucocorticoidal steroid, which can serve as immunomodulatory agent and immunosuppressants in acute chronic conditions Inflammation is the root cause of asthma, therefore the leaves of the plant is useful for the treatment of inflammatory diseases.

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