

Sporicidal Treatments to Produce Germinated Finger Millet

ABSTRACT

Background

Aims: The current investigation was to ascertain the most efficient sporicidal treatment for germinated finger millet in order to inactivate the spores from interfering with the preparation of probiotic millet food.

Study design: By applying different dry and wet sporicidal treatments to the finger millet, the study attempted to inactivate the spores and create germinated finger millet. Following each treatment, the number of aerobic spores was enumerated.

Place and Duration of Study: Department of Dairy Microbiology, Dairy Science College, Hebbal, Bengaluru, Karnataka, India between June 2023 and April 2024.

Methodology: Both wet and dry sporicidal treatments were used. It comprised blanching, autoclaving for 15 min, hot air oven treatment, autoclaving for 30 min, combined treatment of hot air oven and autoclaving.

Results: Aerobic spores were reduced from 3.86 to 1.00 log₁₀ cfu/g after autoclaving finger millet for 30 min at 121°C. The autoclaving treatment did not promote germination. Therefore, finger millet was utilized for the germination process without any kind of treatment. Following a 24 h soaking and 48 h germination period, sporicidal and fungicidal treatments were followed. After autoclaving germinated finger millet flour, the initial counts of spores and fungus which were 4.98 and 3.56 log₁₀ cfu/g were completely eliminated. Statistically significant ($P=0.05$) difference were found between the treatment sample and the control.

Conclusion: Autoclaving of germinated finger at 121°C for 15 min resulted in the complete reduction of spores and fungus count from 4.98 and 3.56 log₁₀ cfu/g.

Keywords: Finger millet, spores, germination, fungi

1. INTRODUCTION

Finger millet (*Eleusine coracana*) is one of the major millet belonging to *Poaceae* family. With 11.33 lakh tonnes production, Karnataka was India's largest producer of finger millet [1]. Finger millet is known as ragi in kannada, kelvaragu in tamil, mandua in hindi and ragulu in telugu [2]. It is rich in calcium, phosphorus and essential amino acids such as lysine, isoleucine, leucine, phenylalanine, methionine, cysteine and tryptophan. Additionally, the important fatty acids palmitic and linolenic acid that are necessary for the growth of the brain and neural tissue are present in finger millet grains [3]. It offers several health advantages including antibacterial, anti-carcinogenic, antioxidant, lowering blood cholesterol, strengthening bones, regulating blood pressure, enhancing children's haemoglobin status and has the ability to cure wounds [4].

Finger millet has found numerous applications in the food industry. They are employed in the making of extruded products, fermented foods, weaning foods and baked products [5].

Formatted: Normal

Formatted: Font: (Default) +Body (Calibri), Not Bold

Formatted: Font: Italic

Spore-forming bacteria are ubiquitous in nature. Bacteria that generate spores are crucial for food deterioration and foodborne illnesses. Endospores that are dormant are resilient to a variety of environmental stressors such as radiation, heat, salt, acidity, oxygen and/or water deprivation and lack of nutrients. However, spores have the ability to detect changes in their immediate environment such as the availability of nutrients. Foods that are high in nutrients may induce the germination process of spores, after that spores can resume exponential cell division by reverting to its vegetative cell state. Food deterioration may result from spore germination in a finished food product, vegetative cell proliferation and may be even sporulation. When it comes to foodborne pathogens, foodborne disease can result from consuming foods containing pathogenic species spores that have the potential to germinate and grow in the gut or from consuming foods in which the spores have already started to grow and proliferate. In the second scenario, a foodborne illness could be caused by consuming toxins found in the food (food poisoning) or by consuming the pathogen's vegetative cells which would then create toxins in the gut (foodborne infection) and cause diarrhoea [6].

The aim of inactivation in many food processing methods is bacterial endospores, particularly those of the *Bacillus* species. Gram-positive, rod shaped bacteria that are naturally found in soil and vegetation are the *Bacillus* genus, which includes species like *B. subtilis*, *B. amyloliquefaciens*, *B. cereus*, *B. licheniformis*, *B. pumilus* and *B. thuringiensis*. These microorganisms are among the most significant human pathogens or those that cause quality damage. In particular, *B. subtilis*, *B. stearothermophilus* and *B. amyloliquefaciens* are the primary bacteria that cause processed food deterioration, while *B. cereus* and *B. anthrax* are representative harmful spore forming bacteria. Thus, it's critical to eliminate *Bacillus* spores linked to pathogenicity and degradation in order to sterilize processed foods [7].

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

Steam sterilization is another name for autoclaving since microorganisms are killed using pressured steam. Compared to other sterilization techniques like dry heat, wet heat sterilization is much more effective. To reach the steam's temperature of 121.1°C and pressure of 15 psi, air must be evacuated during this operation. For effective sterilization, materials must be kept for 15 to 20 min. Any form of microbe, including spores will not survive the autoclaving process [8].

In this study, attempt was made to determine effective sporicidal treatments to produce germinated finger millet which is free from spore formers. Because spores are amylolytic in nature, they may interfere with the fermentation process and not provide the desired outcome. This endeavour to destroy spores in finger millet is intended to make it handy for the manufacturing of fermented probiotic millet food.

2. MATERIAL AND METHODS

2.1 Finger millet

Finger millet was obtained from reputed local market, Bengaluru, Karnataka, India.

2.2 Various sporicidal treatments given to the selected millet

To get rid of all the spores, the millet was subjected to many sporicidal treatments. Both wet and dry treatments were applied to the millet. Millet was treated with a dry sporicidal treatment by being exposed for 1 h inside a hot air oven set at 100°C. Moist sporicidal treatments included blanching the millet for 30 s at 98°C, autoclaving it for 15 min at 121°C; autoclaving it at 121 for 30 min; and giving it a combination treatment that involved heating the millet for 1 h at 100°C in a hot air oven and then autoclaving it for 15 min at 121°C. After each treatment, aerobic spores were enumerated.

2.2.1 Enumeration of aerobic spores

Using a sterile mortar and pestle, 11.0 g of millets were triturated using sterile phosphate buffer. To create a 1:10 dilution, it was then added to 99.0 mL of sterile phosphate buffer. After being heated to 80°C for 10 min in a water bath, the first dilution was cooled to less than 10°C. Then the needed dilutions were prepared serially by the first dilution. Serially diluted samples were then transferred to sterile petri plates that had been labelled. Molten sterile 2% nutrient agar maintained at 45°C was poured into marked petri dishes and allowed to solidify. By inverting the plates, all of the poured plates were incubated at 37°C for 24 to 48 h. The number of nutrient agar colonies was counted after the incubation time. The average count of the countable plates which ranged from 30 to 300 were reported as log₁₀ cfu/g [9].

Formatted: No underline

2.3 Germination of finger millet

The millet was submerged in clean potable water at a ratio of 1:3 (millet: water) for 24 h at 30°C, the surplus water was drained off. The soaked millets were wrapped in a sterile muslin cloth and placed in an incubator set at 30°C for 48 h to facilitate germination. After germination, millets were dried in an open incubator for 24 h at 45°C and were crushed and sieved to obtain germinated finger millet flour.

2.4 Different sporicidal and fungicidal treatments for germinated finger millet

Following germination, the spores and fungus count were enumerated. At various stages after germination, treatments of autoclaving at 121°C for 15 min were administered to ascertain the point at which the number of spores and fungi will decrease. Autoclaving was done after the milling and sieving of germinated millet in an open and sterile environment, as well as after germination. The spores and fungus count following each treatment were enumerated.

2.4.1 Enumeration of yeast and mold

About 99.0 mL of sterile phosphate buffer was mixed with 11.0 g of the weighed samples to prepare the first dilution. The first dilution was then used to prepare the subsequent dilutions in a sequential manner. Samples that had been serially diluted were thereafter transferred to labelled sterile petri dishes. Sterile malt extract agar kept at 45°C was poured into designated petri plates and were allowed to solidify. All of the poured plates were incubated at 30°C for 3-5 days by inverting the plates. Following the incubation period, the number of malt extract agar colonies was counted. The average of countable plate which varied from 30 to 300 was expressed as \log_{10} cfu/g [9].

2.5 Statistical analysis

R software (version 4.1.2) was used to analyse the data and perform statistical computations. For every treatments, three replications of the corresponding variable data were gathered. The data were analysed using ANOVA tables and in cases where the F value is significant, the critical difference ($P=0.05$) was assessed to determine the presence of significant differences, which were then displayed in the tables with superscripts. The formula for critical difference (CD) is

$$CD = \frac{\sqrt{2 \times MSS(E)}}{R} t_{\alpha} @ 0.05$$

Where, MSS (E) = Mean Sum of Squares of the error
R = number of replications
 t_{α} = table t value of the α level of significance

3. RESULTS AND DISCUSSION

3.1 Various sporicidal treatments given to the finger millet

After being treated at 100°C for 1 h in a hot air oven, the aerobic spore count decreased from 3.86 to 2.54 \log_{10} cfu/g. The aerobic spore count of finger millet lowered from 3.86 to 2.18 \log_{10} cfu/g after blanching it for 30 s at 98°C and decreased from 3.86 to 1.70 \log_{10} cfu/g autoclaving of finger millet for 15 min at 121°C. After subjecting in hot air oven at 100°C for 1 h and autoclaving them for 15 min at 121°C, the aerobic spore count decreased from 3.86 to 1.78 \log_{10} cfu/g. In the same way, autoclaving for 30 min at 121°C decreased the aerobic spore count from 3.86 to 1.00 \log_{10} cfu/g (Table 1 and Plate 1). When compared to various dry and wet treatments, autoclaving finger millet at 121°C for 30 min was more effective in killing spores where the maximum reduction of 2 log was observed. The aerobic spore formers could have been killed by the moist heat used in autoclave sterilization and due to more time contact. All of the treatments showed statistically significant ($P=0.05$) differences, with the exception of autoclaving at 121°C for 15 min and autoclaving at 121°C for 30 min following hot air treatment at 100°C, which did not exhibit any significant differences.

On par with the above study, the impact of several dry and moist sporicidal treatments on black gram dhal was investigated. The aerobic spore with initial count of 3.47 \log_{10} cfu/g was completely destroyed when dhal was exposed to a hot air oven at 100°C for 1h, followed by sterilization at 121°C

for 30 min. It's probable that hot air sensitized the spores and autoclaving later assisted in destroying the remaining spores or vegetative cells [10].

Table 1. Various sporicidal treatments given to the finger millet

Treatments	Aerobic spore count (log ₁₀ cfu/g)
Finger millet (Control)	3.86 ^a
Dry treatment	
Hot air oven (100°C/1 h)	2.54 ^b
Wet treatments	
Blanching (98°C/30 s)	2.18 ^b
Autoclaving (121°C/15 min)	1.70 ^{bc}
Hot air oven at 100°C and Autoclaving (121°C/15 min)	1.78 ^{bc}
Autoclaving (121°C/30 min)	1.00 ^c
CD (P=.05)	0.72

- CD= Critical Difference, all the values are average of three trials; same superscripts indicate non-significance while different superscript indicates significant difference at P=.05

Plate 1. Various sporicidal treatments given to the finger millet



Finger millet (Control)



Hot air oven (100°C/1 h)



Blanching (98°C/30 s)



Autoclaving
(121°C/15 min)

Hot air oven at 100°C and
autoclaving (121°C/15 min)

Autoclaving
(121°C/30 min)

3.2 Different sporicidal and fungicidal treatments for optimized germinated finger millet

Sporicidal treatments before germination prevented the millets from sprouting. Hence, sporicidal treatments was followed after germination.

The initial aerobic spore count of 4.98 and the fungal count of 3.56 log₁₀ cfu/g were observed in germinated finger millet. After 15 min of autoclave sterilization at 121°C, the number of spores and fungus in the germinated finger millet decreased from 4.98 to 1.60 and from 3.56 to 0 log₁₀ cfu/g. The spore and fungal counts of the germinated finger millet flour which was made by grinding and sieving autoclaved germinated finger millet were 1.00 and 2.26 log₁₀ cfu/g. After being ground and sieved in an open environment, germinated finger millet was autoclaved for 15 min at 121°C. The results demonstrated reduction in the number of spores and fungus from 4.98 to 1.03 and 3.56 to 1.00 log₁₀ cfu/g. Likewise, autoclaving germinated finger millet that had been ground and sieved in a sterile atmosphere for 15 min at 121°C resulted in reduction of spore and fungal counts from 4.98 to 0.00 and 3.56 to 0.00 log₁₀ cfu/g respectively (Table 2). Moist heat sterilization might have killed spores and fungus.

With the exception of germinated finger millet flour made from autoclaved germinated finger millet and autoclaved germinated finger millet flour that was ground and sieved in an open environment, statistically significant ($P=0.05$) differences were seen for the spores count across all treatments. The literature regarding sporicidal and fungicidal treatments of germinated millets were very scanty.

Table 2. Different sporicidal and fungicidal treatments for optimized germinated finger millet

Treatments	Spores count	Yeast and mold count
	log ₁₀ cfu/g	
Control	4.98 ^a	3.56 ^a
Autoclaved millet	1.60 ^b	0.00 ^d
Flour from autoclaved millet	1.00 ^c	2.26 ^b
Autoclaved millet flour (open environment)	1.03 ^c	1.00 ^c
Autoclaved millet flour (sterile environment)	0.00 ^d	0.00 ^d
CD ($P=0.05$)	0.33	0.29

- CD= Critical Difference, all the values are average of three trials; same superscripts indicate non-significance while different superscript indicates significant difference at $P=0.05$

4. CONCLUSION

Among various sporicidal treatments given to finger millet, autoclaving at 121°C for 30 min resulted in maximum reduction of spores from 3.86 to 1.00 log₁₀ cfu/g. Finger millet subjected to various sporicidal treatments did not undergo sprouting process. Hence, sporicidal treatment was followed after 24 h soaking and 48 h germination. Autoclaving germinated finger millet flour that had been ground and sieved in a sterile atmosphere for 15 min at 121°C resulted in complete reduction of spores and yeast and mold.

REFERENCES

1. India Millets Production. Agricultural and Processed Food Products Export Development Authority (APEDA), Ministry of Commerce and Industry, Government of India, New Delhi. 2023. Available: <https://apeda.gov.in/milletportal/Production.html>.
2. Chandra D, Chandra S, Sharma AK. Review of Finger millet (*Eleusine coracana* (L.) Gaertn): A power house of health benefiting nutrients. *Food Sci. Hum. Wellness*. 2016;5(3):149-155. <https://doi.org/10.1016/j.fshw.2016.05.004>
3. Ramashia SE, Anyasi TA, Gwata ET, Meddows TS, Jideani, AIO. Processing, nutritional composition and health benefits of finger millet in sub-saharan Africa. *Food Sci. Technol*. 2019;39(2):253-266. <https://doi.org/10.1590/fst.25017>
4. Dhanushkodi V, Hemavathy AT, Shenbagavalli S, Sangeetha S, Anitha R, UmaMaheshwari T. A review on nutritional properties and health benefits of finger millet. *Int. J. Plant Sci*. 2023;35(18):753-761. <https://doi.org/10.9734/ijpss/2023/v35i183342>
5. Rathore T, Singh R, Kamble DB, Upadhyay A, Thangalakshmi S. (2019). Review on finger millet: Processing and value addition. *Pharma Innov. J*. 2019;8(4):283-291.
6. Bennik MH, Eijlander RT, Besten HM, Berendsen EM, Warda AK, Krawczyk AO *et al*. Bacterial spores in food: survival, emergence, and outgrowth. *Annu. Rev. Food Sci. Technol*. 2016;7(2):457-482. <https://doi.org/10.1146/annurev-food-041715-033144>
7. Cho WI, Chung MS. Bacillus spores: A review of their properties and inactivation processing technologies. *Food Sci. Biotechnol*. 2020;29(11):1447-1461. <https://doi.org/10.1007/s10068-020-00809-4>
8. Chauhan A, Jindal T, editors. *Methods of Sterilization and Disinfection*. In: *Microbiological Methods for Environment, Food and Pharmaceutical Analysis*. Springer: Switzerland; 2020. https://doi.org/10.1007/978-3-030-52024-3_2
9. Harrigan. *Laboratory methods in food and dairy microbiology*, Department of Food Science, Reading Univ., Reading Academy Press Inc. (London) Ltd. UK; 1998.
10. Akshaykumar, Manjunatha H, Ramachandra B. Production of Biomass of Lactic Acid Bacteria Using Optimized Solid Substrate in Self-Technique. *Int. J. Curr. Microbiol. App. Sci*. 2017;6(12):2890-2898. <https://doi.org/10.20546/ijcmas.2017.612.336>