

Gram Positive Cocci associated Urinary Tract Infections,their prevalence and Antibiotic susceptibility patterns.

ABSTRACT

Background

Urinary tract infections (UTIs) are the third most common types of infection in humans globally. Gram positive bacteria are said to be responsible for ten percent of urinary tract (UTI)infections. The study's goal was to profile gram-positive cocci associated UTIs and their antibiogram, as they were observed at LASUTH.

Methods

This was a retrospective assessment of the Medical Microbiology Laboratory records of the LASUTH to review the in vitro antibiotic susceptibility patterns of gram-positive urinary bacterial isolates between April 2020 and March 2021. The bacteria were isolated and identified from routine urine samples using standard bacteriological methods and the API.In vitro antibiotic susceptibility test (AST) was routinely performed by the modified Kirby-Bauer disk diffusion test and susceptibility break points determined using the Clinical and Laboratory Standards Institute (CLSI) guidelines.

Results

2,253 urine samples were processed in the medical microbiology laboratory over the one year and 662 (29.4%) samples yielded Positive cultures. Of the 662 isolates, 494 (74.6%) were gram-negative bacteria. 164(24.8%) were gram-positive cocci while 4 (0.6%) were gram positive rod.Among the gram-positive cocci's isolated *Enterococcus faecalis* had the highest frequency 58(35.4%).

Aminoglycosides (Gentamycin andAmikacin) and Linezolid antibiotics were found to be the most effective drugs against gram-positive cocci bacteria except *Enterococcus spp*.For empirical treatment of *Enterococcus spp* in our facility Fosfomycin and Tigecyclinearethe best options, while for *Streptococcusagalactiae* associated UTI, Amikacin, Cefuroxime,Linezolid, and levofloxacin can be used for empirical treatment.

Conclusion

The prevalence rate of gram-positive cocci associated UTI in this study was 7.3% (164/2253).The emergence of drug resistance in these pathogens to commonly used

antibiotics is a thing of concern. Therefore, efficient antimicrobial stewardship programmes must be in place.

Keywords

UTIs, Uropathogens, Gram-Positive Cocci, Enterococcus, MRSA.

INTRODUCTION

When compared to other uropathogens, bacterial urinary tract infections are the most common and dangerous infections in humans, and they frequently occur in both the community and in hospitals. (1-4)

Most Gram-negative bacteria that cause illness are classified as pathogenic bacteria; they include *Escherichia coli*, *Klebsiella species*, *Enterobacter species*, *Proteus species*, *Pseudomonas species*, *Acinetobacter species*, *Serratia species*, and *Citrobacter species*. 90% of UTI cases are caused by them. The remaining ten percent of UTI infections are caused by gram-positive bacteria, specifically *Enterococcus species*, group B streptococci, and *Staphylococcus species* (3, 4).

Although coagulase-negative staphylococci (CoNS) and *S. aureus* were previously thought to be uncommon etiological agents in ascending UTIs in outpatients, they may play a more significant role in hospitalised, immunocompromised patients. When *S. aureus* is isolated from urine, it can also be a sign of a more serious illness (like endocarditis or bacteraemia), in which the germs spread hematogenously and end up in the kidneys (5). The literature indicates that the isolation frequency of *S. aureus* from UTIs ranges from 0.5 to 13% (6,7). Conversely, *S. saprophyticus* is a well-studied pathogen in catheter associated UTIs as well as simple cystitis. After *S. saprophyticus*'s pathogenic role in UTIs—also known as "honeymoon cystitis"—was identified in the 1960s, an increasing amount of information about the pathophysiology of this illness has been discovered (8).

S. saprophyticus is thought to be the etiological agent in 5–20% of UTIs according to epidemiological research, but a Swedish study discovered that it was the cause of more than 40% of females' uncomplicated UTIs (9). *Enterococcus species* are among the few Gram-positive bacteria that are resistant to bile, and they are widely distributed in the gut microbiota of both humans and animals (10). Given their high prevalence in aquatic habitats,

enterococci should be considered as a sign of faecal contamination in urban areas (11). The most frequent species in bacteraemia, endocarditis, infections of the central nervous system, and urinary tract infections are *Enterococcus faecalis* and *E. faecium*; nevertheless, the appearance of non-faecalis enterococci should be considered (12,13). These organisms are important in nosocomial infections worldwide, much like *Staphylococcus aureus* (14).

Staphylococcus haemolyticus the second-most frequently isolated CoNS (*S. epidermidis* is the first) and it is considered a relevant hospital-acquired pathogen.[15]. It is associated with insertion of foreign devices into the human body like urinary catheters, prosthetic valves and cerebrospinal shunts.(16,17,18).

Streptococcus agalactiae or Lancefield group B Streptococcus (GBS), a gram-positive β -haemolytic chain-forming coccus, is a common bacterial cause of UTI. It is estimated to cause approximately 1%–2% of all single microorganism source UTIs (19,20). Pregnant, diabetic, immunocompromised, and persons with pre-existing urologic abnormalities are also susceptible to GBS-caused UTIs. In these cases, there is an increased risk of ascending pyelonephritis, which may develop to bacteraemia and/or urosepsis. (20,21,22,23)

Gram-positive cocci have numerous virulence factors, such as maintaining their high affinity for the epithelial cells of the urinary tract, allowing for their survival. These virulence factors include fibrillar proteins (Ssp) mediating cell–cell interactions, fibronectin-binding proteins, elastin-binding protein, adhesins, hemagglutinin, elastase, and lipase. Furthermore, urease is produced by the majority of *S. saprophyticus* and over 90% of *S. aureus* strains, which breaks down carbamide (urea) in the urine (5-14,24,25,26). Because *Enterococcus spp* are found in faecal matter and may colonise the rectum, their anatomical closeness to the urinary system may further increase their ability to cause UTIs (11, 27). Biofilm-production in these species is another important factor for the emergence and the persistence of UTIs, with some reports suggesting that some 80% of uropathogenic Gram-positive cocci are biofilm-producers (19). The presence of biofilm in urethral stents and catheters may lead to obstruction.

furthermore, microorganisms embedded in biofilm may survive 1000-times higher concentrations of antibiotics, compared to non-embedded cells (28,29,30).

MATERIALS AND METHODS.

Study setting:

The study was conducted in the department of medical microbiology of the Lagos State University Teaching Hospital, an 800-bedded tertiary centre located in Ikeja, Lagos southwest Nigeria. The hospital is dedicated to teaching, research and specialist services and serves Lagos State and neighbouring States in southwest Nigeria.

Study design:

This was a retrospective study that involved a review of the medical microbiology laboratory records to analyse the antimicrobial susceptibility profiles of Gram-positive cocci urinary tract infections isolates obtained between April 2020 to March 2021.

Isolation and antibiotic susceptibility pattern of bacterial isolates:

Normal processing of urinary samples in the laboratory during the period of the review involved macroscopic and microscopic examination. And then, urinary samples were inoculated into Cystine Lactose Electrolyte Deficient (CLED) and Blood agar plates and incubated aerobically at 35-37⁰C for 18-24 hours. Isolates were identified by conventional biochemical tests and using Analytical Processing Index (API) and antimicrobial susceptibility testing (AST) was performed using the modified Kirby-Bauer disk diffusion method. The break points for Susceptibility were determined using the Clinical and Laboratory Standards Institute (CLSI) guidelines (31).

Data Analysis

Data analysis was done using the Statistical Package for the Social Sciences (SPSS) version 22.

Ethical considerations

Ethical approval for the study was obtained from Lagos State University Teaching Hospital Research and Ethics Committee. As data were retrospectively obtained from the laboratory records and did not involve contact with patients nor was recruitment of patients, informed consent not deemed necessary. However, privacy and confidentiality of patients' data were protected.

RESULTS.

In the 1-year period under consideration, a total number of 2,253 urine samples were processed in the medical microbiology laboratory and 662 (29.4%) samples yielded Positive cultures (Table 1).

Of the 662 isolates, 494 (74.6%) were Gram-negative bacteria, while 164 (24.8%) were gram positive cocci and 4 (0.6%) were gram positive rod (Table 2).

The 164-gram positive cocci came from 74 (45.1%) inpatients urine samples and 90 (54.9%) outpatients urine samples (Table 3).

Among the gram-positive cocci, isolated *Enterococcus faecalis* had the highest frequency 58 (35.4%), followed by *Staphylococcus epidermidis* 48 (29.3%), and *Enterococcus faecium* 24 (14.6%). MRSA and *Streptococcus agalactiae* were 10 (6.1%) respectively, *Staphylococcus aureus* 6 (3.7%), while *Staphylococcus saprophyticus* and *Streptococcus agalactiae* had a frequency of 4 (2.4%) each (Table 4).

Females had the greater incidence of gram-positive cocci associated UTI with a frequency of 120 (73.2%) while males had a frequency of 44 (26.8%). This is in a ratio of 3: 1 in favour of the females (Table 5).

In terms of age, among the males age group 51-60, 31-40 and 21-30 years were mostly affected in decreasing order, while for the females, age group 31-40 years were mostly affected, followed by age group 41-50 and 21-30 years (Table 5).

From table 6, Most species of *Enterococcus faecalis* and *Enterococcus faecium* were sensitive to Fosfomycin (70% -100%), Tigecycline (70 -79.3%), and Meropenem (86-93.1%). These species of *Enterococcus* had the greatest resistance to Streptomycin (51-58.3%), Cefotaxime (50-56.9%) and Amoxicillin-clavulanate (36-41.7%).

Furthermore, *Staphylococcus aureus* isolates were most sensitive to Amoxicillin, Amoxicillin-clavulanate, Linezolid and Gentamycin, with a sensitivity of 66.7% each. *Staphylococcus aureus* were most resistant to Clindamycin and Trimethoprim-sulfamethoxazole (66.7% each). While for Methicillin Resistant *Staphylococcus aureus* Linezolid was the most sensitive drug with a sensitivity of 80%.

For *Staphylococcus epidermidis* isolates, Nitrofurantoin and Levofloxacin had the most sensitivity with sensitivity rates of 83.3% and 54.2% respectively. Their greatest resistance was to Amoxicillin (83.3%) and erythromycin (62.5%).

Staphylococcus haemolyticus isolates were most sensitive to Gentamycin (75%) and Linezolid (50%). They were most resistant to Clindamycin (50%).

Again, *Staphylococcus saprophyticus* isolates were most sensitive to Nitrofurantoin (75%), levofloxacin (75%), Linezolid and gentamycin (50% each). *S. saprophyticus* isolates were most resistant to erythromycin (50%).

Streptococcus agalactiae isolates were most sensitive to Cefuroxime (90%), Amikacin and linezolid (80% each) and erythromycin and levofloxacin (60% each). They were most resistant to Nitrofurantoin (60%), Amoxicillin (40%) and Amoxicillin-clavulanate (30%).

DISCUSSION.

In this Study a total number of 2,253 urine samples were processed in the medical microbiology laboratory and 662 (29.4%) samples yielded Positive cultures while 1591 (70.6%) had negative cultures. The reason for the no bacterial growth among a good number of the urine samples may be because some of the patients have been on antibiotic therapy before reporting to the hospital or laboratory. These antibiotics may have inhibited bacterial growth [32].

The prevalence rate of gram-positive cocci Uropathogens causing UTI from this Study was 7.3% (164/2253). This was lower than that of a study done in Indian (33) that got a prevalence of 18.35%.

The 164-gram positive cocci came from 74 (45.1%) inpatients urine samples and 90 (54.9%) outpatients urine samples.

Females had the greater incidence of gram-positive cocci associated UTI with a frequency of 120 (73.2%) while males had a frequency of 44 (26.8%). This is in a ratio of 3: 1 in favour of the females. This may be due to the shortness of the Female urethra when compared with that of males. Furthermore, the moist vaginal introitus, which the urethral meatus opens into, is colonised by both pathogenic bacteria and normal flora, some of which could induce cystitis. UTI in females is also influenced by other significant factors, such as pregnancy, postmenopausal status, and sexual activity (34).

. Among the group of gram-positive cocci, *E. faecalis* was the predominant species (35.4%), which is not surprising, in view of worldwide epidemiological reports identifying the causing agents of UTI's. (35).

The prevalence of MRSA isolates from urinary samples in our study was low (0.4%), and the levels of these isolates were in contrast to report from other literature (36,37).

Staphylococcus aureus in total (MRSA included) accounted for 16(9.8%) of the gram-positive cocci associated UTI's. This may be because *Staphylococcus aureus* is implicated in UTI in many sexually active females, as reported by some studies (38,39).

In our study, Aminoglycosides (Gentamycin and/ Amikacin) and Linezolid antibiotics were found to be the more effective drug against gram-positive cocci bacteria except *Enterococcus* spp. This was contrary to another Study (40) that found out that Teicoplanin, and Nitrofurantoin were most sensitive to gram positive cocci (GPC) urinary isolates.

Most *Streptococcus agalactiae* and all *Enterococcus* spp had the common resistance; Amoxicillin -clavulanate, while *Staphylococcus saprophyticus* and *Staphylococcus epidermidis* have come resistance to erythromycin. Furthermore, *Staphylococcus aureus* and *Staphylococcus haemolyticus* have similar resistance to Clindamycin. These findings were contrary to a study (40), where the most resistant drugs were Penicillin, Ampicillin, and Ciprofloxacin in all GPC (only Penicillin except *Enterococcus* spp.).

CONCLUSION.

The prevalence rate of gram-positive cocci associated UTI in this study was 7.3% (164/2253). Although urinary tract infections are mainly caused by gram-negative bacteria, gram-positive cocci have emanated as important agents of UTIs, particularly among elderly patients, mostly associated with co-morbidities, pregnant women, and catheterized patients, both in low- and high-income countries. In our study, *Enterococcus* spp. had the highest prevalence (3.6% (82/2253)) among other gram-positive cocci cause of UTI.

The emergence of drug resistance in these pathogens to commonly used antibiotics is a thing of concern.

The resistance rates for fluoroquinolones are worrisome and as such these agents are not recommended to be used empirically. In contrast, the use of nitrofurantoin for staphylococci may still be regarded as safe in our setting and the tested isolates are almost uniformly susceptible to the available last-resort antibiotics.

Generally, for empirical treatment of *Enterococcus* spp in our facility Fosfomycin and Tigecycline is our best option, while for *Streptococcus agalactiae* associated UTI Amikacin, Cefuroxime, Linezolid and levofloxacin can be used for empirical treatment.

To ensure the proper use of antibiotics for treating urinary tract infections, efficient antimicrobial stewardship programmes must be in place.

REFERENCES

1. G. Gebremariam, H. Legese, Y. Woldu, T. Araya, K. Hagos, and A. Gebreyesus Wasihun, "Bacteriological profile, risk factors and antimicrobial susceptibility patterns of symptomatic urinary tract infection among students of Mekelle University, northern Ethiopia," *BMC Infectious Diseases*, vol. 19, no. 950, pp. 1–11, 2019.
2. G. Beyene and W. Tsegaye, "Bacterial uropathogens in urinary tract infection and antibiotic susceptibility pattern in Jimma University Hospital, Ethiopia," *Ethiopian Journal of Health Science*, vol. 21, pp. 141–146, 2011.
3. G. Dalela, S. Gupta, D. K. Jain, and P. Mehta, "Antibiotic resistance pattern in uropathogens at a tertiary care hospital at Jhalawar with special reference to Esbl, Ampc β -Lactamase and MRSA production," *Journal of Clinical and Diagnostic Research*, vol. 6, pp. 645–651, 2012.
4. M. B. Ashagrie, "Bacterial profile and ESBL screening of urinary tract infection among asymptomatic and symptomatic pregnant women attending antenatal care of northeastern Ethiopia region," *Infection and Drug Resistance*, vol. 13, pp. 2579–2592, 2020.
5. Baraboutis, I. G. et al. Primary *Staphylococcus aureus* urinary tract infection: the role of undetected hematogenous seeding of the urinary tract. *Eur. J. Clin. Microbiol. Infect. Dis.* 29, 1095–1010 (2010).
6. Tong, S. Y. C., Davis, J. S., Eichenberger, E., Holland, T. L. & Fowler, V. G. *Staphylococcus aureus* infections: epidemiology, pathophysiology, clinical manifestations, and management. *Clin. Microbiol. Rev.* 28, 603–661 (2015).

7. Gajdács, M. [Epidemiology and susceptibility patterns of *Staphylococcus aureus* isolates from STI samples of male patients (2008–2017)] (article in Hungarian). *Magyar Urol.* 31, 66–68 (2019).
8. Adegate, J., Juhász, E., Pongrácz, J., Rimanóczy, É. & Kristóf, K. Does *Staphylococcus saprophyticus* cause acute cystitis only in young females, or is there more to the story? A one-year comprehensive study done in Budapest, Hungary. *Acta Microbiol. Immunol. Hung.* 63, 57–67 (2016).
9. Eriksson, A., Giske, C. & Ternhag, A. The relative importance of *Staphylococcus saprophyticus* as a urinary tract pathogen: distribution of bacteria among urinary samples analysed during 1 year at a major Swedish laboratory. *APMIS* 121, 72–78 (2012).
10. Vu, J. & Carvalho, J. *Enterococcus*: review of its physiology, pathogenesis, diseases and the challenges it poses for clinical microbiology. *Front. Biol.* 6, 357–366 (2011).
11. García-Solache, M. & Rice, L. B. The *Enterococcus*: a model of adaptability to its environment. *Clin. Microbiol. Rev.* 32, e00058-18 (2019).
12. Ulricha, N., Vonberg, R. P. & Gastmeier, P. Outbreaks caused by vancomycin-resistant *Enterococcus faecium* in hematology and oncology departments: a systematic review. *Heliyon* 3, e00473 (2017).
13. Fisher, K. & Phillips, C. The ecology, epidemiology and virulence of *Enterococcus*. *Microbiology* 155, 1749–1757 (2009).
14. Guzman-Prieto, A. M. et al. Global emergence and dissemination of enterococci as nosocomial pathogens: attack of the clones? *Front. Microbiol.* 7, e788 (2016).
15. C. Vignaroli; F. Biavasco; P. E. Varaldo (2006). "Interactions between Glycopeptides and β -Lactams against Isogenic Pairs of Teicoplanin-Susceptible and -Resistant Strains of *Staphylococcus haemolyticus*". *Antimicrobial Agents and Chemotherapy.* 50 (7): 2577–2582. doi:10.1128/AAC.00260-06. PMC 1489795. PMID 16801450.
16. Falcone; et al. (2006). "Teicoplanin use and emergence of *Staphylococcus haemolyticus*: is there a link?". *Clin Microbiol Infect.* 12 (1): 96–97. doi :10.1111/j.1469-0691.2005.01307.x. PMID 16460556

17. Poyart; et al. (2001). "Rapid and Accurate Species-Level Identification of Coagulase-Negative Staphylococci by Using the *sodA* Gene as a Target". *Journal of Clinical Microbiology*. 39 (12): 4296–4301. doi:10.1128/JCM.39.12.4296-4301.2001. PMC 88539. PMID 11724835.
18. Viale, P.; Stefani, S. (2006). "Vascular catheter-associated infections: a microbiological and therapeutic update". *J Chemother*. 18 (3): 235–49. doi:10.1179/joc.2006.18.3.235. PMID 17129833. S2CID 25108301.
19. Magliano E, Grazioli V, Deflorio L, et al. Gender and age- dependent etiology of community- acquired urinary tract infections. *Scientific World J*. 2012;2012:349597. 10.1100/2012/349597 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
20. Kline KA, Lewis AL. Gram- positive uropathogens, polymicrobial urinary tract infection, and the emerging microbiota of the urinary tract. *Microbiology Spectrum*. 2016;4(2). 10.1128/microbiolspec.UTI-0012-2012 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
21. Edwards MS, Baker CJ. Group B streptococcal infections in elderly adults. *Clin Infect Dis*. 2005;41(6):839- 847. [PubMed] [Google Scholar]
22. Mathai E, Thomas RJ, Chandy S, Mathai M, Bergstrom S. Antimicrobials for the treatment of urinary tract infection in pregnancy: practices in southern India. *Pharmacoepidemiol Drug Saf*. 2004;13(9):645- 652. [PubMed] [Google Scholar]
23. Trivalle C, Martin E, Martel P, Jacque B, Menard JF, Lemeland JF. Group B streptococcal bacteraemia in the elderly. *J Med Microbiol*. 1998;47(7):649- 652. [PubMed] [Google Scholar]
24. Behzadi, P. et al. A survey on urinary tract infections associated with the three most common uropathogenic bacteria. *Maedica (Buchar)* 5, 111–115 (2010).
25. Murray, P. R., Baron, E. J., Jorgensen, J. H., Landry, M. L. & Pfaller, M. A. *Manual of Clinical Microbiology* 9th edn. (American Society for Microbiology, Washington, DC, 2007).
26. Kang, C. I., Song, J. H., Ko, K. S., Chung, D. R. & Peck, K. R. Clinical features and outcome of *Staphylococcus aureus* infection in elderly versus younger adult patients. *Int. J. Infect. Dis*. 15, e58–e62 (2011).

27. Gajdács, M. The continuing threat of methicillin-resistant *Staphylococcus aureus*. *Antibiotics* 8, e52 (2019).
28. Shrestha, L. B., Baral, R. & Khanal, B. Comparative study of antimicrobial resistance and biofilm formation among Gram-positive uropathogens isolated from community acquired urinary tract infections and catheter-associated urinary tract infections. *Infect. Drug. Res* 12, 957–963 (2019).
29. Soto, S. M. Importance of biofilms in urinary tract infections: new therapeutic approaches. *Adv. Biol.* 2014, 13 (2014).
30. Gomes-Fernandes, M. et al. Accessory gene regulator (Agr) functionality in *Staphylococcus aureus* derived from lower respiratory tract infections. *PLoS ONE* 12, e0175552 (2017).
31. Clinical and Laboratory Standards Institute Performance standards for antimicrobial susceptibility testing. M100. Wayne, PA: Clinical and Laboratory Standards Institute, 2020.
32. Otajevwo, F.D. (2013) Urinary Tract Infection Among Symptomatic Outpatients Visiting a Tertiary Hospital Based in Midwestern Nigeria. *Global Journal of Health Science*, 5, 187-199. <http://dx.doi.org/10.5539/gjhs.v5n2p187>.
33. Gajdács, M., Ábrók, M., Lázár, A. et al. Increasing relevance of Gram-positive cocci in urinary tract infections: a 10-year analysis of their prevalence and resistance trends. *Sci Rep* 10, 17658 (2020). <https://doi.org/10.1038/s41598-020-74834-y>
34. Mandell, Douglas and Bennetts. Principles and practices of Infectious diseases. (2005) 6th edition: Vol. (I); pp – 242–317:875-905.
35. Sader, H. S., Farrell, D. J., Flamm, R. K. & Jones, R. N. Antimicrobial susceptibility of Gram-negative organisms isolated from patients hospitalised with pneumonia in US and European hospitals: results from the SENTRY Antimicrobial Surveillance Program, 2009–2012. *Int. J. Antimicrob. Agents* 43, 328–334 (2014).
36. European Antimicrobial Resistance Surveillance Network (EARS-Net). <https://ecdc.europa.eu/en/about-us/partnerships-and-networks/disease-and-laboratory-networks/ears-net>. Accessed on October 2023.

37. Hegstad, K., Mikalsen, T., Coque, T. M., Werner, G. & Sundsfor, A. Mobile genetic elements and their contribution to the emergence of antimicrobial resistant *Enterococcus faecalis* and *Enterococcus faecium*. *Clin. Microbiol. Infect.* 16, 541–554 (2010).

38. Alex, B., Siakwa, P.M., Boampong, J.N., Koffuor, G.A., Ephraim, R.K.D., Amoateng, P., Obodai, G. and Penu, D. (2012) Asymptomatic Urinary Tract Infections in Pregnant Women Attending Antenatal Clinic in Cape Coast, Ghana. *Journal of Medical Research*, 1, 74-83.

39. Murray, R.R., Rosenthal, K.S., Kobayashi, G.S. and Tenover, M.A. (1998) *Medical Microbiology*. 3rd Edition, Mosby Publishers, Maryland Heights, 186.

40. V.P. Sarasu, S. Ramalatha Rani Bacteriological profile and antibiogram of urinary tract infections at a tertiary care hospital *International Journal of Medical Microbiology and Tropical Diseases*, July-September, 2017;3(3):106-112

TABLE 1: TOTAL URINE SAMPLES, CULTURE OUTCOME AND FREQUENCY.

		Percentage
Total urine samples	2,253	100%
Positive culture	662	29.4%
Negative culture	1601	70.6%

TABLE 2: POSITIVE CULTURE BACTERIA AND FREQUENCY.

Positive culture	Frequency	Percentage (%)
Gram negative	494	74.6
Gram positive cocci	164	24.8
Gram positive bacilli	4	0.6
Total	662	100%

TABLE 3:FREQUENCY DISTRIBUTION OF GRAM-POSITIVE COCCI ISOLATES BETWEEN INPATIENTS AND OUTPATIENTS.

Gram positive cocci isolate	Frequency	Percentage (%)
Inpatient	74	45.1
Outpatient	90	54.9

TABLE 4: GRAM POSITIVE COCCI ISOLATES AND THEIR FREQUENCY

	Gram positive cocci	Frequency	Percentage (%)
1.	E. faecium	24	14.6
2.	E. faecalis	58	35.4
3.	S. aureus	6	3.7
4.	MRSA	10	6.1
5.	S. saprophyticus	4	2.4
6.	S. epidermidis	48	29.3
7.	S. agalactiae	10	6.10
8.	S. haemolyticus	4	2.40
Total		164	100

TABLE 5: GRAM POSITIVE COCCI BACTERIA OCCURRENCE BY AGE GROUP AND SEX.

Agerange(Year)	Male	Female	Total	
0-10	1	6	7	
11-20	2	13	15	
21-30	10	26	36	
31-40	12	40	52	
41-50	3	28	31	
51-60	14	3	17	
>60	2	4	6	
	44(26.8%)	120(73.2%)	164(100%)	

TABLE 6: ISOLATES AND ANTIBIOTICS SUBSCEPTILITY PATTERN.

ORGANISMS	ANTIBIOTICS	SENSITIVITY	INTERMEDIATE	RESISTANCE	TOTAL
<i>Enterococcus faecium</i>	Cefotaxime	10(41.7%)	2(8.3%)	12(50%)	24
	Amoxicillin - Clavulante	14(58.3%)	—	10(41.7%)	
	Levofloxacin	10(41.7%)	2(8.3%)	12(50%)	
	Tigercycline	17(70.8%)	2(8.3%)	5(20.8%)	
	Amoxicillin	20(83.3%)	3(12.5%)	1(4.3%)	
	Fosfomycin	17(70.8%)	5(20.8%)	2(8.3%)	
	Streptomycin	10(41.7%)	—	14(58.3)	
	Tetracycline	14(58.3%)	4(16.7%)	6(25%)	
	Meropenem	21(87.5%)	2(8.3%)	1(4.2%)	
<i>Staphylococcus epidermidis</i>	Amoxicillin	8(16.7%)	—	40(83.3%)	48
	Amoxicillin-Clavulanate	18(37.5%)	20(41.7%)	10(20.8%)	
	Cefoxitin	48(100%)	—	—	

	Nitrofurantion	40(83.3%)	—	8(16.7%)	
	TMP	10(20.8%)	8(16.7%)	30(62.5%)	
	Clindamycin	20(41.7%)	4(8.3%)	24(50%)	
	Erythromycin	16(33.3%)	2(4.2%)	30(62.5%)	
	Gentamicin	22(45.8%)	2(4.2%)	24(50%)	
	Levofloxacin	26(54.2%)	2(4.2%)	20(41.7%)	
	Linezolid	22(45.8%)	8(16.7%)	18(37.5%)	
<i>Enterococcus faecalis</i>	Amoxicillin-Clavulanate	20(34.5%)	16(33.3%)	22(37.9%)	58
	Amoxicillin	28(48.3%)	—	30(51.7%)	
	Cefotaxine	15(25.9%)	10(20.8%)	33(56.9%)	
	Tigercycline	46(79.3%)	12(20.7%)	—	
	Fosfomycin	58(100%)	—	—	
	Meropenem	54(93.1%)	2(3.4%)	2(3.4%)	
	Erythromycin	30(51.7%)	6(10.3%)	22(37.9%)	
	Levofloxacin	38(65.5%)	2(3.5%)	18(31.0%)	
	Tetracycline	40(83.3%)	18(16.7%)	—	
	Streptomycin	20(34.5%)	8(13.8%)	30(51.7%)	
<i>Staphylococcus aureus</i>	Amoxicillin	4(66.70%)	1(16.65%)	1(16.65%)	6
	Amoxicillin-Clavulanate	4(66.7)	2(33.3%)		
	Cefoxitin	6(100%)	—	—	
	Nitrofurantion	3(50%)	2(33.3%)	1(16.7%)	
	Clindamycin	1(16.6%)	1(16.7%)	4(66.7%)	
	Linezolid	2(66.7%)	2(33.3%)	—	
	TMP	1(16.7%)	1(16.7%)	4(66.6%)	
	Gentamycin	4(66.7%)	2(33.3%)	—	
	Rifampicin	3(50%)	2(33.3%)	1(16.7%)	
	Levofloxacin	2(33.3%)	2(33.3%)	(33.3)	
<i>Staphylococcus haemolyticus</i>	Levofloxacin	2(50%)	2(50%)	0(0%)	4
	Amoxicillin	2(50%)	1(25%)	1(25%)	

	Cefoxitin	4(100%)	—	—	
	Nitrofurantion	2(50%)	1(25%)	1(25%)	
	Clindamycin	1(25%)	1(25%)	2(50%)	
	Linezolid	2(50%)	2(50%)	—	
	Gentamycin	3(75%)	1(25%)	0	
	TMP	2(50%)	1(25%)	1(25%)	
	Erythromycin	2(50%)	1(25%)	1(25%)	
	Rifampicin	2(50%)	1(25%)	1(25%)	
<i>Staphylococcus saprophyticus</i>	Cefoxitin	4(100%)	—	—	4
	Nitrofurantion	3(75%)	1(25%)	0	
	Amoxicillin	2(50%)	1(25%)	1(25%)	
	TMP	2(50%)	1(25%)	1(25%)	
	Levofloxacin	3(75%)	0	1(25%)	
	Linezolid	2(50%)	2(50%)	—	
	Gentamycin	2(50%)	2(50%)	0	
	Erythromycin	1(25%)	1(25%)	2(50%)	
	Clindamycin	1(25%)	2(50%)	1(25%)	
	Rifampicin	2(50%)	2(50%)	—	
<i>Streptococcus agalactiae</i>	Levofloxacin	6(60%)	2(20%)	2(20%)	10
	Amoxicillin	4(40%)	2(20%)	4(40%)	
	Linezolid	8(80%)	1(10%)	1(10%)	
	Amikacin	8(80%)	2(20%)	—	
	Amoxicillin	4(40%)	2(20%)	4(40%)	
	Erythromycin	6(60%)	1(10%)	3(30%)	
	TMP	4(40%)	3(30%)	3(30%)	
	Nitrofurantoin	2(20%)	2(20%)	6(60%)	
	Cefuroxime	9(90%)	—	1(10%)	
	AMC	5(50%)	2(20%)	3(30%)	
<u>MRSA</u>	Amoxicillin	2(20%)	2(20%)	6(60%)	10
	Cefoxitin	--	--	10(100%)	
	Nitrofurantoin	4(40%)	2(20%)	4(40%)	
	Clindamycin	6(60%)	2(20%)	2(20%)	

	Linezolid	8(80%)	2(20%)	--	
	Gentamycin	6(60%)	3(30%)	1(10%)	
	Rifampicin	8(80%)	1(10%)	1(10%)	
	Amoxicillin- Clavulanate	6(60%)	2(20%)	2(20%)	
	Erythromycin	2(20%)	4(40%)	4(40%)	
		3(30%)	4(40%)	3(30%)	

UNDER PEER REVIEW