

# Predilection Site for the Meat Lover *Trichinella* spp Larvae and Its Pathogenesis and Potency in Human Host

---

## ABSTRACT

**Aims:** To revisited predilection site of *Trichinella* spp larvae including their pathogenesis and its potency

**Discussion:** The nematode *Trichinella* spp causes serious zoonosis called trichinellosis, a disease affecting muscles which consider as one of tropical disease. Even though its natural host varied, but infection among popular live stocks, such as pigs and other animals, which are raising community medicine concern. Human infection occurs after consumption of raw or undercooked meat or meat products contain muscle larvae of *T. spiralis*. The tropism of the parasite for individual muscles and/or muscle groups varies significantly. *Trichinella* spp. has a direct life cycle where all three life cycle stages (the infective muscle larvae, adult, and new born larvae) happen serially in one host only. Intestine-dwelling adults of *Trichinella* produce newborn larvae that bypass the enterocyte, enter the bloodstream and colonize skeletal muscle. The muscle larvae assemble excretory-secretory products which play crucial role in establishing and maintaining persistent parasitism and the host's immune modulation and evasion. It turns out that excretory-secretory products from muscle larvae and mature worm also have hidden medical potential that can be used to treat allergic problems, inflammation-based diseases, autoimmunity and even malignancy.

**Conclusion:** Trichinellosis is a serious and potentially fatal zoonosis which transmitted through consuming raw or uncooked contaminated meat or its comestibles. Its primary tropism is to the host's striated muscle and infection can persist for a long time facilitated by several reciprocities of its product (e.g., excretory-secretory) with the host's cell and immune system. Fortunately, there are several promising potency in the field of therapeutic and prevention medicine which should be explored intensely.

**Keywords:** tropical disease, nematode, tropism, excretory, secretory, muscle larvae, carcasses, trichinosis/trichinellosis

## 1. INTRODUCTION

The term tropical diseases encompass all diseases, communicable and no communicable, that occur principally in the tropical countries or tropics, areas that lie between, and alongside, the Tropic of Cancer and Tropic of Capricorn belts [1]. Among those communicable group of disease, neglected helminth infections including trichinellosis or trichinosis, which is still the major health problem [2]. There are eleven known species within the genus *Trichinella*. These eleven species subdivide into those that invade host muscle cells and encapsulate (surrounded by a collagen capsule) and those that do not encapsulate [3,4]. *Trichinella Spiralis*, the most common species in this genus, belongs to the

encapsulated group and causes most human infections and deaths from trichinosis [3]. This genus specifically causing a disease which affecting the host's muscles [1-5].

Even though its natural host varied, but infection among popular live stocks such as pigs [5-8] and other animals, e.g., horses [9], wild game meat (meat from an animal that is typically found in the wild and not raised domestically on a farm for mass consumption; usually free-roaming foragers and hunted for their meat) [10], rats [11], wild birds [12], wild and farmed reptiles [13] etc., which are raising public health concern [14], even though its global burden is much lower than that of other foodborne parasitic diseases (a mean estimated 76 healthy life years lost per billion people per year for human trichinellosis, globally [15]).

The aim of this study is to revisited predilection site of *Trichinella* spp larvae in its host including their pathogenesis and potency along with its comestible implication and effort conducted to prevent transmission.

## 2. LIFE CYCLE AND PATHOGENESIS

Among many member of helminths which affect human, *Trichinella* spp. are distinctive because it has a direct life cycle [16]; which means that all three life cycle stages of the parasite, namely infective muscle larvae, adult, and new born larvae; Intestine-dwelling adults of *Trichinella* produce newborn larvae that enter the bloodstream and colonize skeletal muscle [17]. Infection is acquired by consumption of infected raw or undercooked meat or meat based comestibles [3-17].

Under the biochemically pressure of low pH gastric juice, entrapped larvae which is basically anaerobic are released in the host's stomach, followed by the molting process (approximately four times in 30-40-time span) [19]. Proteases secreted by *Trichinella spiralis* intestinal infective larvae directly damage the surrounding junctions of the intestinal epithelial cell monolayer and also arbitrate larval invasion and develop into the adult stage inside the enterocytes of small intestine [20]. The results of study conducted by Song et al [20] stipulate that the parasite enzyme named serine proteases and cysteine proteases play crucial roles in larvae invasion, growth and survival inside the host and that they may be main candidate target molecules for vaccines against larval invasion and development.

After successfully entering enterocyte and become mature, male and female are mating then produce new born larvae [21] are released into circulation and spread throughout the tissues and organs [22] and only those that enter striated muscles mature into muscle larvae [23]. During the muscular phase, the larvae invade the skeletal muscle fibers inducing a relevant inflammatory reaction aiming for the elimination of the parasite. However, the larvae eventually succeed to build their own home inside the infected myocytes [23]. Muscle invasion results in formation of a capsule surrounding muscle larvae in the region of infected muscles [24]. Once again, this eccentric meat lover *Trichinella* blessed with the capability to make itself "feel homey like being at home" by way of transforming the infected muscle cell for their own benefit and accomplishing a new type of cell inside the host affected musculature, the so-called nurse cell [25].

The lowest infectious dose of *Trichinella* larvae is remains unrevealed, but the clinical manifestations of trichinellosis starts to displayed as the number of parasite entering the host increases [26]. Asymptomatic infection could remain silent in human if it is only involving a minimum amount of larvae; gastrointestinal symptoms manifested as a specific syndrome consist of nausea, diarrhea, vomiting, fatigue, fever, and abdominal discomfort [27], starts very early to develop in case of unintentionally ingestion of hundreds of larvae, perhaps manifest itself clinically within the first 48 hours after consuming contaminated meat. The condition that followed by development of a series of condition which are serious, but scarcely fatal illness [5]. Clinical signs of the disease usually last 4–6 months, rarely longer (up to 2 years).

Chronic form of trichinellosis rarely reported, once in 1983 revealed by two German doctor in their case report regarding biopsies conducted on muscles of five patients with clinical diagnosis chronic neuromuscular disorder, mostly manifested as spinal muscle atrophy. All

of them had previous history of acute trichinellosis, the interval between acute parasitic infection and the appearance of the slowly progressive neuromuscular syndrome being of 13 to 26 years respectively. Analysis conducted on the biopsy specimens showed morphological and enzyme-histochemical alteration which indicative the presence of progressive neurogenic muscular atrophy. From the Parasite perspective, distinctive encapsulated but still living, enzyme-positive parasites were clearly identified with definite signs of focal myositis in the muscle portion surrounding the larva. The possibility pathogenesis correlations between the "chronic" trichinellosis and the "degenerative" neuromuscular disorder cannot yet be excluded and this still remains to be an uncharted sea of exploration.

### **3. THE POTENCY**

Infective larvae remain alive in striated muscles of the vulnerable host for years [20]; an evidence supported by the study of Sofronic-Milosavljevic et al which revealed the chronic existence of specific antibody responses that still could be recognized even 30 years post primary infection [29]. In case of invasion by *Trichinella* larvae against the host's immune system, it actually arouses a complex immune response; in human host is better designated by humoral immune response [30] rather than the cellular responses; and this emphasize future prospect of the human host's dynamic humoral response [30] for diagnostic [29] or even vaccine development purposes [31,32] such as reported by Bi et al [32] that revealed the newly identified rTs-ES-1 is potent immunodominant protein secreted by *Trichinella* stichocytes during natural infection and permits the arousal of fractional protective immunity in vaccinated mice inimical to intentional *Trichinella* infection. Therefore, findings of this rTs-ES-1 specific protein with the better understanding of its antigenic shift-dynamicity [31] is a potential candidate for vaccine development against trichinellosis. In contrast to what happened inside their vulnerable human host, in animals *T. spiralis* can outstretch a high worm burden without causing prominent clinical symptoms [33].

The initiation of infection depends on first by the annexation of prone intestinal epithelium by infective muscle larvae (ML) and followed secondly with the preservation of parasitism which is marked by the presence of ML in affected muscle cells. The parasite regulatory protein accountable for enzymatic process of these two steps are very important for future investigation.

Excretory-secretory products of invading larvae believed to be originate from stichocyte granules in the stichosome, the secretory organelle of the *Trichinella*'s mature muscle larvae [35]. These excretory-secretory products play a pivotal role in parasite's immune evasion and regulation inimical to the host's innate immune system by way of (1) suppressing NET (neutrophil extracellular traps which primary function as a trap for pathogens and facilitating phagocytosis and cytokine production) production and (2) negatively didacte cytokine secretion. The understanding of this excretory-secretory products function for the larvae or worm provides an encouraging area for manufacturing new intervention strategies in other areas of medicine, e.g., in tackling sepsis induced acute lung injury [36] or allergic plethora [29] or autoimmune condition/diseases such as colitis [37] and even malignancies [29].

These important excretory-secretory products engage mainly in the reciprocity with various host cells: firstly, the immune cells, secondly the enterocytes and thirdly the muscle cells, and, through those interaction establishing their role in parasitism and immune response induction, modulation and even evasion [29-32,34-37]. Through these approaches, this nematode generates a perfect milieu for its own suitability and survival in two ways either by modulation of host immune response or affecting host cell gene expression. Extensive exploration of these molecules is important in order to build better understanding regarding (1) the establishment of triumphant parasitism, (2) the development of novel therapies and (3) preventive treatments for inflammatory based disorder.

#### 4. CONCLUSION

Trichinellosis with its related clinical syndrome must always be considered as serious and potentially fatal zoonosis. Transmission occurs through consuming raw or uncooked meat or its comestibles which contaminated with its muscle larvae. Its primary tropism is to the host's striated muscle and can affect the muscle strength and composition in long term. Infection can persist for a long time facilitated by several reciprocities of its product (e.g., excretory-secretory) with the host's organ specific cells (e.g., enterocytes, myocytes) and immune system. Fortunately, there are several promising potency in the field of therapeutic and prevention medicine which should be explored intensely.

#### CONSENT (WHERE EVER APPLICABLE)

Not needed

#### ETHICAL APPROVAL (WHERE EVER APPLICABLE)

Not needed

#### REFERENCES

1. Zumla A, Ustianowski A. Tropical diseases: definition, geographic distribution, transmission, and classification. *Infect Dis Clin North Am*. 2012 Jun;26(2):195-205. <https://doi.org/10.1016/j.idc.2012.02.007>.
2. Hotez PJ, Brindley PJ, Bethony JM, King CH, Pearce EJ, Jacobson J. Helminth infections: the great neglected tropical diseases. *J Clin Invest*. 2008 Apr;118(4):1311-21. <https://doi.org/10.1172/JCI34261>.
3. Pozio E, Zarlenga DS, La Rosa G. The detection of encapsulated and non-encapsulated species of *Trichinella* suggests the existence of two evolutive lines in the genus. *Parasite*. 2001 Jun;8(2 Suppl):S27-9. <https://doi.org/10.1051/parasite/200108s2027>.
4. Furhad S, Bokhari AA. Trichinosis. [Updated 2023 Jul 19]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK536945/>
5. Kalambhe D, Kaur H, Gill JPS. *Trichinella* spp. in Slaughtered Pigs of India: From Neglected Disease to an Emerging Food Safety Threat for Public Health. *Transboundary and Emerging Diseases*. 2024; 1-9. <https://doi.org/10.1155/2024/7550006>.
6. Gondek M, Knysz P, Pyz-Łukasik R, Łukomska A, Kuriga A, Pomorska-Mól M. Distribution of *Trichinella spiralis*, *Trichinella britovi*, and *Trichinella pseudospiralis* in the Diaphragms and *T. spiralis* and *T. britovi* in the Tongues of Experimentally Infected Pigs. *Front Vet Sci*. 2021 Jun 22;8:696284. <https://doi.org/10.3389/fvets.2021.696284>.
7. Eslahi AV, KarimiPourSaryazdi A, Olfatifar M, de Carvalho LMM, Foroutan M, Karim MR, Badri M, Ketzis JK. Global prevalence of *Trichinella* in pigs: A systematic review and meta-analysis. *Vet Med Sci*. 2022 Nov;8(6):2466-2481. <https://doi.org/10.1002/vms3.951>

8. Wang N, Bai X, Tang B, Yang Y, Wang X, Zhu H, Luo X, Yan H, Jia H, Liu M, Liu X. Primary characterization of the immune response in pigs infected with *Trichinella spiralis*. *Vet Res.* 2020 Feb 21;51(1):17. <https://doi.org/10.1186/s13567-020-0741-0>.
9. Różycki M, Korpysa-Dzirba W, Bełcik A, Bilska-Zajęc E, Gontarczyk A, Kochanowski M, Samorek-Pieróg M, Karamon J, Rubiola S, Chiesa F, Cencek T. Validation Parameters of the Magnetic Stirrer Method for Pooled Sample Digestion for *Trichinella* spp. in Horse Meat Based on Proficiency Tests Results. *Int J Environ Res Public Health.* 2022 Nov 2;19(21):14356. <https://doi.org/10.3390/ijerph192114356>.
10. McIntyre L, Pollock SL, Fyfe M, Gajadhar A, Isaac-Renton J, Fung J, Morshed M. Trichinellosis from consumption of wild game meat. *CMAJ.* 2007 Feb 13;176(4):449-51. <https://doi.org/10.1503/cmaj.061530>.
11. Bilska-Zajęc E, Różycki M, Antolak E, Bełcik A, Grądziel - Krukowska K, Karamon J, et al. Occurrence of *Trichinella* spp. in rats on pig farms. *Ann Agric Environ Med.* 2018; 25(4): 698-700. <https://doi.org/10.26444/aaem/99555>
12. Rugna G, Marucci G, Bassi P, Gelmini L, D'Annunzio G, Torreggiani C, et al. *Trichinella* surveillance program in wild birds, Emilia-Romagna (northern Italy), 2006–2021. First report of *Trichinella pseudospiralis* in western marsh harrier (*Circus aeruginosus*) in Italy. *International Journal for Parasitology: Parasites and Wildlife.* 2022; 19:191-195, <https://doi.org/10.1016/j.ijppaw.2022.09.006>.
13. Pozio E, Foggin CM, Gelanew T, Marucci G, Hailu A, Rossi P, Morales MA. *Trichinella zimbabwensis* in wild reptiles of Zimbabwe and Mozambique and farmed reptiles of Ethiopia. *Vet Parasitol.* 2007 Feb 28;143(3-4):305-10. <https://doi.org/10.1016/j.vetpar.2006.08.029>.
14. Yayeh M, Yadesa G, Erara M, Fantahun S, Gebru, Birhan M (2020). Epidemiology, diagnosis and public health importance of Trichinellosis. *Online J. Anim. Feed Res.*, 2020;10(3): 131-139. <https://dx.doi.org/10.36380/scil.2020.ojaf18>
15. Devleesschauwer B, Praet N, Speybroeck N, Torgerson PR, Haagsma JA, De Smet K, Murrell KD, Pozio E, Dorny P. The low global burden of trichinellosis: evidence and implications. *Int J Parasitol.* 2015 Feb;45(2-3):95-9. <https://doi.org/10.1016/j.ijpara.2014.05.006>.
16. Gamble HR. *Trichinella*. Ed(s): Batt CA, Tortorello ML. *Encyclopedia of Food Microbiology* (Second Edition), Academic Press. 2014. pp 638-643, ISBN 9780123847331. <https://doi.org/10.1016/B978-0-12-384730-0.00336-0>.
17. Lee JL, Rosenberg HF (eds) .Chapter 10 - Eosinophils: Mediators of Host-Parasite Interactions. In books *Eosinophils in Health and Disease*, Academic Press, 2013, pp 301-327, ISBN 9780123943859. <https://doi.org/10.1016/B978-0-12-394385-9.00010-9>.
18. Mostafa EM, Atwa HA. Intestinal mastocytosis in *Trichinella spiralis* infection: immunohistochemical study in murine model. *Parasitologists United Journal.* 2020;13(1): 52-59. <https://doi.org/10.21608/puj.2020.24540.1060>
19. Gagliardo LF, McVay CS, Appleton JA. Molting, ecdysis, and reproduction of *Trichinella spiralis* are supported in vitro by intestinal epithelial cells. *Infect Immun.* 2002 Apr;70(4):1853-9. <https://doi.org/10.1128/IAI.70.4.1853-1859.2002>.
20. Song YY, Lu QQ, Han LL, Yan SW, Zhang XZ, Liu RD, Long SR, Cui J, Wang ZQ. Proteases secreted by *Trichinella spiralis* intestinal infective larvae damage the junctions of the intestinal epithelial cell monolayer and mediate larval invasion. *Vet Res.* 2022 Mar 7;53(1):19. <https://doi.org/10.1186/s13567-022-01032-1>.
21. Gardiner CH. Habitat and reproductive behavior of *Trichinella spiralis*. *J Parasitol.* 1976 Dec;62(6):865-70.

22. Stewart GL, Despommier DD, Burnham J, Raines KM. *Trichinella spiralis*: behavior, structure, and biochemistry of larvae following exposure to components of the host enteric environment. *Exp Parasitol*. 1987 Apr;63(2):195-204. [https://doi.org/10.1016/0014-4894\(87\)90162-7](https://doi.org/10.1016/0014-4894(87)90162-7).
23. Rayia DA, Othman A, Harras S, Helal D, Dawood L, Soliman S. Bevacizumab: A new take on therapy of muscle phase of *Trichinella spiralis* infection. *Acta Trop*. 2022 Jun;230:106409. <https://doi.org/10.1016/j.actatropica.2022.106409>.
24. Wu Z, Sofronic-Milosavljevic Lj, Nagano I, Takahashi Y. *Trichinella spiralis*: nurse cell formation with emphasis on analogy to muscle cell repair. *Parasit Vectors*. 2008 Aug 19;1(1):27. <https://doi.org/10.1186/1756-3305-1-27>.
25. Despommier DD. How does *Trichinella spiralis* make itself at home? *Parasitol Today*. 1998 Aug;14(8):318-23. [https://doi.org/10.1016/s0169-4758\(98\)01287-3](https://doi.org/10.1016/s0169-4758(98)01287-3).
26. Diaz JH, Warren RJ, Oster MJ. The Disease Ecology, Epidemiology, Clinical Manifestations, and Management of Trichinellosis Linked to Consumption of Wild Animal Meat. *Wilderness & Environmental Medicine*. 2020;31(2):235-244. <https://doi.org/10.1016/j.wem.2019.12.003>
27. Kociecka W. Trichinellosis: human disease, diagnosis and treatment. *Vet Parasitol*. 2000 Dec 1;93(3-4):365-83. [https://doi.org/10.1016/s0304-4017\(00\)00352-6](https://doi.org/10.1016/s0304-4017(00)00352-6).
28. Gullotta F, Fröscher W. Chronische Trichinose und neuromuskuläre Erkrankungen. Morphologische und pathogenetische Aspekte [Chronic trichinosis and neuromuscular diseases. Morphologic and pathogenesis aspects]. *Arch Psychiatr Nervenkr* (1970). 1983;232(6):479-87. German. <https://doi.org/10.1007/BF00344062>.
29. Sofronic-Milosavljevic L, Ilic N, Pinelli E, Gruden-Movsesijan A. Secretory Products of *Trichinella spiralis* Muscle Larvae and Immunomodulation: Implication for Autoimmune Diseases, Allergies, and Malignancies. *J Immunol Res*. 2015;2015:523875. <https://doi.org/10.1155/2015/523875>.
30. Alcántara P, Correa D. Human humoral immune responses against *Trichinella spiralis*. *Int J Parasitol*. 1993 Aug;23(5):657-60. [https://doi.org/10.1016/0020-7519\(93\)90173-v](https://doi.org/10.1016/0020-7519(93)90173-v).
31. Boireau P, Vallée I, Karajian G, Wang X, Liu M. Chapter 16 - Antigenic shift during *Trichinella* cycle, consequences for vaccine developments, Editor(s): Fabrizio Bruschi, *Trichinella* and Trichinellosis. Academic Press, 2021, pp 455-516, ISBN 9780128212097. <https://doi.org/10.1016/B978-0-12-821209-7.00014-7>.
32. Bi K, Yang J, Wang L, Gu Y, Zhan B, Zhu X. Partially Protective Immunity Induced by a 20 kDa Protein Secreted by *Trichinella spiralis* Stichocytes. *PLoS One*. 2015 Aug 19;10(8):e0136189. <https://doi.org/10.1371/journal.pone.0136189>.
33. Ribicich M, Pasqualetti MI, Fariña FA. Chapter 9 - Trichinellosis in animals. Ed(s): Bruschi F. *Trichinella* and Trichinellosis. Academic Press, 2021, pp 315-331, ISBN 9780128212097. <https://doi.org/10.1016/B978-0-12-821209-7.00013-5>.
34. Hao HN, Song YY, Ma KN, Wang BN, Long SR, Liu RD, Zhang X, Wang ZQ, Cui J. A novel C-type lectin from *Trichinella spiralis* mediates larval invasion of host intestinal epithelial cells. *Vet Res*. 2022 Oct 18;53(1):85. <https://doi.org/10.1186/s13567-022-01104-2>.
35. Despommier DD, Müller M. The stichosome and its secretion granules in the mature muscle larva of *Trichinella spiralis*. *J Parasitol*. 1976 Oct;62(5):775-85.
36. Li H, Qiu D, Yang H, Yuan Y, Wu L, Chu L, Zhan B, Wang X, Sun Y, Xu W, Yang X. Therapeutic Efficacy of Excretory-Secretory Products of *Trichinella spiralis* Adult

- Worms on Sepsis-Induced Acute Lung Injury in a Mouse Model. *Front Cell Infect Microbiol.* 2021 Mar 24;11:653843. <https://doi.org/10.3389/fcimb.2021.653843>.
37. Yang X, Yang Y, Wang Y, Zhan B, Gu Y, Cheng Y, Zhu X. Excretory/secretory products from *Trichinella spiralis* adult worms ameliorate DSS-induced colitis in mice. *PLoS One.* 2014 May 2;9(5):e96454. <https://doi.org/10.1371/journal.pone.0096454>.

UNDER PEER REVIEW