

Original Research Article

An Evaluation of Anti-hyperlipidemic Activity of Ethanolic extract of *Moringa oleifera* on High Fat Induced Hyperlipidemic Rat Model

Abstract

The art or practice of herbal remedies refers to the use of herbs and herbal treatments for the purpose of maintaining health and preventing, treating, or curing sickness. In some areas, herbal treatments can also be referred to as herbal medicine. To assess the lipid profiles of *Moringa oleifera* extract, rats were studied. For both the SGPT and the SGOT, it was seen that groups 5 and 6 exhibited statistically significant ($p < 0.05$) outcomes in the case of the SGPT. However, in the case of the SGOT, there were no statistically significant findings. When conducting the renal function test, it was observed that the levels of creatinine and urea were statistically significant ($p < 0.05$) in the cases of groups 4, 5, and 6. In the case of high-density lipoprotein (HDL) and low-density lipoprotein (LDL), groups 4, 5, and 6 showed statistically significant findings ($p < 0.05$) in HDL levels, while groups 5 and 6 showed statistically significant LDL levels. The triglyceride levels in the group were found to be statistically significant ($p < 0.05$), while the findings obtained from groups 5 and 6 were also found to be statistically significant.

Keywords: Herbal medicine, *Moringa oleifera*, HDL, LDL, Phytochemicals.

Comment [A1]: Methodology like how many rats used, what is the dosage studied should be mentioned

Comment [A2]: In results section, Since its about lipidemia it is better to mention the lipid levels followed by Liver and Kidney function

Introduction

The liver, the largest glandular organ, is responsible for controlling the majority of a person's bodily functions. Numerous times during the day, the whole blood supply of an individual goes through the liver. The liver is very critical for human metabolism. [1]. Hepatotoxicity, the most frequent form of liver illness, is a leading cause of death and impairment in both animals and humans. Several medications can cause it. [2]. Addiction to alcohol or drugs, exposure to harmful chemicals, infections with viruses or parasites, and high levels of reactive oxygen species (ROS) are among the most damaging factors that may affect liver cells (OH, H₂O₂, O₂). [2]. The ability to neutralize free radicals is why ascorbic acid and the tripeptide L-glutathione (L-cysteine, glycine, and L-glutamate) are taken orally as dietary supplements. Because of their anti-inflammatory, antibacterial, and immune-enhancing qualities, they are highly prized by many. [3]. On the flip side, they might trigger an allergic response, such as dermatitis, or gastrointestinal issues, such as gas, diarrhea, indigestion, or even difficulties breathing due to airway narrowing. There are 1.5 billion individuals worldwide who suffer from chronic liver disease (CLD). Among Americans aged 45–64, the prevalence of CLD has risen by 31% in recent years. [4]. Some chemical components derived from medicinal plants may have therapeutic uses, say specialists in the subject. It follows that researchers are always on the lookout for new herbal cures and other plant-based therapies to treat a wide range of illnesses. [5]. While phytotherapy is based on scientific study, herbalism is more concerned with the practical applications of medicinal plants. Plants have played a significant role in human medicine for thousands of years due to the wide diversity of chemicals they contain, many of which have medicinal characteristics [6]. The vast variety of chemical components found in medicinal plants allows them to exert a broad spectrum of pharmacological and therapeutic effects. Tanning agents, glycosides, alkaloids, saponins, polysaccharides, essential oils, terpenoids, resins, and plant lipids are all examples of such components [7-9]. Modifying plants genetically allows for the precise regulation of chemical concentrations, allowing for the desired therapeutic effect. Reverse genetics has many potential applications, one of which is to enhance the production of secondary metabolites like alkaloids [10]. Recent scientific progress around the

world has led to more research into the healing properties of plants [11]. This is because plants are safe, have strong pharmacological activity, and are more cost-effective than man-made drugs.

Comment [A3]: Paper is about anti hyperlipidemic property but introduction discuss about liver & hepatoprotective. There is no mention about cholesterol, lipidemia etc.

Moringa oleifera has nutritional and therapeutic features as a result of its tremendous medical potential; however, this is only the case if the economic worth of the plant's nutritional value, medicinal applications, and animal feed is significant. This tropical deciduous tree, which is endemic to the southern Himalayas in northern India, is a perennial and belongs to the Moringaceae family. Antioxidant, anti-inflammatory, neuroprotective, hypoglycemic, and blood lipid-lowering are just a few of the nutritional and medicinal benefits of *Moringa oleifera* extracts. *Moringa oleifera*'s rich phytochemical content—including flavonoids, glucosinolates, isothiocyanates, and phenolic acids—is strongly associated with its positive effects [12].

The purpose of our present study is to evaluate the hepatoprotective effects of *Moringa oleifera*.

Comment [A4]: The topic is antihyperlipidemic but its given hepatoprotective

MATERIALS AND METHODS

Plant Collection and Extract Preparation

Moringa oleifera were collected from local market of Dhaka. The material was authenticated by National herbarium, Bangladesh. *Moringa oleifera* was cleaned properly with water and it was then air-dried. Finally dried leaves were crushed in powder. The powder was soaked for 15 days in 70% ethanol. The solution was kept for 15 days. Vigorous shaking was also performed occasionally. Next, the solution was filtered. The collected filtrate was dried in a rotary evaporator at a low temperature and pressure. Finally, the crude residue was subjected to the required pharmacological testing.

Comment [A5]: herbarium

Comment [A6]: Abbreviation cant be used it should be in words First

Drugs and Chemicals

Atorvastatin drug was obtained from incepta pharmaceuticals as a gift sample. Ethanol was bought from Taj Scientific store.

Experimental Animal Procurement, Nursing, and Grouping

A total of 90 male rats weighing between 120 and 150 grams were obtained from Jahangirnagar University in Savar, Dhaka. Each of them was housed in a climate-controlled environment (temperature $25 \pm 3^\circ\text{C}$, relative humidity $55 \pm 5\%$, and a 12-h light/dark cycle) at the University of

Dhaka's Institute of Nutrition & Food Science (INFS). They were given a conventional food and were permitted to drink clean water. All of the animals were maintained in this habitat for at least one week prior to the research for **adaption**. All experimental methods followed the recommendations of the Institutional Animals Ethics **Committee (IEAC)**, 90 rats were randomly distributed into 9 groups were each groups contain 10 rats.

Comment [A7]: adaptation

Comment [A8]: IEAC approval no not mentioned

Experimental design

Rats were individually weighed and then divided into nine independent groups for research on anti-hyperlipidemic action. The distribution of rodents among the groups was based on their body weight, with each group consisting of five rats. The atorvastatin control group in Table 1 shows rats that were given atorvastatin with a high-fat diet since using simply atorvastatin would result in the animals dying. N/A indicates that rats in this group did not receive any therapeutic treatment.

Table 1: Antihyperlipidemic activity analysis

Group number	Group Status	Treatment specimen & Dose	Group Abbreviation
1	Negative Control	Physiological Saline	N
2	Positive Control	High Fat Diet	P
3	High Fat Diet + Atrovastatin	High Fat Diet + Atrovastatin	HFD + ATV
4	High Fat Diet + <i>M. oleifera</i>	High Fat Diet+ MO ₂₅₀	HFD + MO ₂₅₀
5	High Fat Diet + <i>M. oleifera</i>	High Fat Diet +MO ₅₀₀	HFD + MO ₅₀₀
6	High Fat Diet + <i>M. oleifera</i>	High Fat Diet + MO ₂₅₀	HFD +MO ₇₅₀
7	<i>M. oleifera</i>	MO ₂₅₀	MO ₂₅₀
8	<i>M. oleifera</i>	MO ₅₀₀	MO ₅₀₀
9	<i>M. oleifera</i>	MO ₇₅₀	MO ₇₅₀

High Fat Diet: The high-fat diet was modified based on the composition supplied by Levin and Dunn-Meynell. The high fat diet is composed of 50% lipid, 40% carbohydrate, and 10% protein. The diet's composition is shown in Table 2.

Table 2: Composition of high fat diet

Food Ingredients	Composition
Lipid (50%)	Milk powder (10%) Ghee (30%) Mutton fat (40%) Coconut oil (10%) Butter (10%)
Carbohydrate (40%)	Boiled rice (40%) Smashed potato (40%) Boiled corn (20%)
Protein (10%)	Dry powdered prone (40%) Dry boiled mutton (20%) Cheese (20%) Egg (20%)

After mixing the ingredients thoroughly, the high fat diet was given to the rats to induce obesity for 10 weeks. [13]

Comment [A9]: This should come first prior to Experimental design with induction of high fat diet as title

Evaluation of anti-hyperlipidemic Activity

For this experiment, 100 rats were randomly picked and equally divided into fourteen groups

Comment [A10]: Only 90 rats and 9 groups

Table 3: Application of treatment efficacy

Group Number	Group Specification	Treatment species	Dose treatment species (mg/kg)	Abbreviation of Groups
1	Negative control	Physiological saline	10 ml/kg	N
2	High Fat	N/A	N/A	HF
3	HF+At ₁₀	Atrovastatin 10mg/kg	10	At ₁₀
4	HF+MO ₂₅₀	<i>Moringa oleifera</i>	250	MO ₂₅₀
5	HF+MO ₅₀₀	<i>Moringa oleifera</i>	500	MO ₅₀₀
6	HF+MO ₇₅₀	<i>Moringa oleifera</i>	750	MO ₇₅₀
7	MO ₂₅₀	<i>Moringa oleifera</i>	250	MO ₂₅₀
8	MO ₅₀₀	<i>Moringa oleifera</i>	500	MO ₅₀₀
9	MO ₇₅₀	<i>Moringa oleifera</i>	750	MO ₇₅₀

Biological Sample Collection.

We punctured a rat's tail to obtain blood samples for measuring blood glucose levels. On the other hand, blood was drawn from the animal as soon as its heart was punctured and transferred to a micro centrifuge tube after the killing. The samples were centrifuged at 5,000 rpm for 5 minutes to create the supernatant fluid. Biochemical testing subsequently required the transfer of this fluid to an additional micro centrifuge tube. We carefully took the kidney and liver from the animal after sacrifice and cleaned them in ice-cold saline to assess their function.

Estimation of Biochemical Parameters

The blood glucose level was measured using a glucometer. Aside from the Humaluzer 3000, lipid profile, kidney, and liver function tests were performed. In addition, the gluconeogenic and glycolytic enzyme activity of kidney and liver samples was examined

Statistical analysis:

All of our findings (raw data) in terms of numerical parameters were recorded and analyzed on a broadsheet using the MS Excel application. The gathered data were subjected to descriptive statistics, with the findings reported as mean SD. To evaluate statistical significance, we used the SPSS 16 software's "One-way Anova test" to interpret inter-group heterogeneity in terms of several biological factors. The occurrences are considered statistically significant since the 'p' value was less than 0.05 ($p < 0.05$).

Results and discussion:

For both the SGPT and the SGOT, it was seen that groups 5 and 6 exhibited statistically significant ($p < 0.05$) outcomes in the case of the SGPT. However, in the case of the SGOT, there were no statistically significant findings. There were two other investigations that came to the same conclusions [14, 15]. When conducting the renal function test, it was observed that the levels of creatinine and urea were statistically significant ($p < 0.05$) in the cases of groups 4, 5, and 6. Two separate investigations [16, 17] came to the same conclusions about the subject. In the case of high-density lipoprotein (HDL) and low-density lipoprotein (LDL), groups 4, 5, and 6 showed statistically significant findings ($p < 0.05$) in HDL levels, while groups 5 and 6 showed statistically significant LDL levels. The triglyceride levels in the group were found to be statistically significant ($p < 0.05$), while the findings obtained from groups 5 and 6 were also found to be statistically significant. There were two other investigations that came to the same conclusions [18, 19].

Table 4: Lipid profile of *Moringa oleifera*:

	SGPT	SGO T	Creati- nine	Urea	TC	HDL	LDL	TG
C	36.28±4.2 5	36.52 ±3.12	0.5±0.0 5	29.49± 2.32	94.82±3.19	66.25±3.62	35.22±3. 26	54.28±3.29
HF	95.32±6.8 0	88.73 ±8.18	2.24±0. 06	87.72± 6.91	177.84±9.5 3	38.54±3.84	69.32±5. 69	104.24±6.26
HF+At ₁₀	74.59±2.0 9	74.33 ±6.28	1.1±0.0 8	59.51± 6.39	119.26±7.5 6	57.36±4.58	45.40±4. 28	70.64±5.94
HF+MO ₂₅₀	93.28± 6.51	88.18 ±3.59	1.97±0. 08*	82.39± 4.81*	172.42±6.2 1	42.60±2.81 *	37.54±3. 57	97.29±6.82*
HF+MO ₅₀₀	90.27±6.5 8*	87.29 ±8.13	1.70±0. 07*	78.30± 3.59*	166.17±7.5 3*	47.84±5.33 *	41.66±4. 58*	93.10±5.28
HF+MO ₇₅₀	86.81±5.2 9*	85.52 ±3.61	1.29±0. 05*	74.53± 5.60*	162.23±6.1 8*	54.54±4.28 *	45.08±3. 28*	90.25±6.73
MO ₂₅₀	34.67±2.6 2	39.42 ±2.85	0.60±0. 08	28.08± 3.06	90.18±4.51	63.28±4.20	37.30±2. 21	56.29±4.10
MO ₅₀₀	36.37±4.1 2	34.73 ±2.80	0.70±0. 06	32.53± 2.84	92.30±2.81	66.53±3.29	35.18±3. 26	53.29±3.10
MO ₇₅₀	34.12±3.2 9	35.79 ±3.18	0.9±0.0 8	30.80± 2.35	94.95±3.22	65.19±4.24	38.42±4. 20	55.53± 4.23

Note: The results were expressed in Mean±SEM (standard mean error) *p< 0.05, **p< 0.01, and ***p< 0.001 were considered as statistically significant. The statistical analysis followed by one-way analysis of variance (Dunnett's test) compared to the control.

Conclusion

Within the scope of this investigation, the hepatoprotective properties of *Moringa oleifera* ethanolic extract were investigated. Based on the findings of this research, it seems that an ethanol extract derived from the plant *M. oleifera* may be able to provide protection against excessive cholesterol, damage to the liver, and impaired kidney function. As a result, more research is necessary in order to determine the active components in the entire extract that have

Comment [A11]: Anti hyperlipidemic property

the ability to reduce hyperlipidemia and diabetes. After the active chemicals have been discovered, it is possible to conduct a comprehensive investigation.

References:

1. Bhawna S, Kumar SU. Hepatoprotective activity of some indigenous plants. *Int J Pharm Tech Res.* 2009 Oct;4:1330-4.
2. Nadeem M.P.C, Dandiya P.C, K.V., Pasha M., Imran D., Balani K, Vohora S.B., Hepatoprotective activity of *Solanum nigrum* fruits. *Fitoterapia.*, 1997, 68, 245:51
3. FM SS, Juliana AB, Bornila M, Puja B, Nur-Neasha D, Rafat T. An Assessment of Hepatoprotective Activity of *Psidium guajava* Fruit Extract against Hepatic Injured Rodent Model. *Asian Journal of Medical Principles and Clinical Practice.* 2023 Oct 7;6(2):240-5.
4. Islam M, Rupak AH, Nasrin N, Chowdhury MM, Sen P, Foysal AU, Uddin MJ, Ferdous J, Tahsin MR, Aktar F, Kabir S. An Evaluation of Potential Hepato-Protective Properties of *Hylocereus Undatus* Fruit in Experimental Rat Model. *Biomedical Journal of Scientific & Technical Research.* 2022;43(2):34405-16.
5. Baroi JA, Hossian MR, Chowdhury MM, Dolon NN, Maliha F, Rupak MA, Lima NN, Ullah MR, Tahsin R. An assessment of anti-hyperlipidemic potentialities of ethanolic extract of *hemidesmus indicus* in high fat induced rat model. *Asian Journal of Food Research and Nutrition.* 2023 Jul 9;2(4):323-30.
6. Mim IJ, Peya FY, Chowdhury MM, Khan TR, Mandal SK, Maliha F, Alam M, Rahman T, Tashin R. An evaluation of anti-diabetic activity of ethanolic extract of *asparagus racemosus* in alloxan induced rat model. *International Journal of Advances in Nephrology Research.* 2023 Aug 2;6(1):60-8.
7. Lima NN, Dolon NN, Maliha F, Ullah MR, Humayra F, Chowdhury MM, Rupak MA, Baroi JA, Shohan FS, Tashin R. An Evaluation of Analgesic and Anti-Inflammatory Activity of *Ficus racemosa* in Rat Model.

8. Yang L, Stöckigt J. Trends for diverse production strategies of plant medicinal alkaloids. *Natural product reports* 2010;27(10):1469-1479.
9. Saxena M, Saxena J, Nema R, Singh D, Gupta A. Phytochemistry of medicinal plants. *Journal of Pharmacognosy and Phytochemistry*. 2013;1(6):168-182. Rupak MA, Chowdhury MM, Shurov
10. Rupak MA, Chowdhury MM, Shurovi FS, Ferdous J, Tahsin MR, Sarif S, Hasan MM, Chowdhury JA, Kabir S, Chowdhury AA, Aktar F. An Evaluation of Analgesic and Anti-Inflammatory Activity of Ethanolic Extract of *Cynodon Dactylon* on Stressed Rodent Model. *Biomedical Journal of Scientific & Technical Research*. 2022;42(3):33550-7
11. Pracheta SS, Sharma V, Paliwal R, Sharma S, Yadav S, Singh L et al (2011) Chemoprotective activity of hydro-ethanolic extract of *Euphorbia nerrifolia* Linn. Leaves against DENA-induced liver carcinogenesis in mice. *Biol Med*. 3(2):36–44.
12. El-Hadary AE, Ramadan MF (2019) Antioxidant traits and protective impact of *Moringa oleifera* leaf extract against diclofenac sodium-induced liver toxicity in rats. *J Food Biochem*. 43(2):e12704
13. Abdul, N. A., & Rahmat, A. (2015). jaafar Hz. Protective Effects of Tamarillo.
14. Puspitasari D, Lestari I, Rahayuningsih CK, Christyaningsih J. The Effectiveness of *Moringa Oleifera* Leaf Extract on Hepatotoxic Case Reviewing from Cadmium, SGOT and SGPT Blood Levels in White Rats (*Rattus Norvegicus*) Induced with Cadmium (Cd). *Jurnal Teknokes*. 2022 Sep 16;15(3):137-46.
15. Islam R, Alam MJ. Evaluation of liver protective activity of *Moringa oleifera* bark extract in paracetamol induced hepatotoxicity in rats. *BioRxiv*. 2019 Jan 7:513002.
16. Saleh SS, Sarhat ER. Effects of ethanolic *Moringa oleifera* extract on melatonin, liver and kidney function tests in alloxan-induced diabetic rats. *Indian Journal of Forensic Medicine & Toxicology*. 2019 Oct 1;13(4):1015-9.

17. Nafiu AO, Akomolafe RO, Alabi QK, Idowu CO, Odujoko OO. Effect of fatty acids from ethanol extract of *Moringa oleifera* seeds on kidney function impairment and oxidative stress induced by gentamicin in rats. *Biomedicine & Pharmacotherapy*. 2019 Sep 1;117:109154.
18. Madkhali HA, Alharthy KM, Asiri MA, Ganaie MA, Ansari MN, Rehman NU, Hamad AM. *Moringa oleifera* Lam.(family Moringaceae) leaf extract attenuates high-fat diet-induced dyslipidemia and vascular endothelium dysfunction in Wistar albino rats. *Tropical Journal of Pharmaceutical Research*. 2019;18(12):2597-604.
19. Helmy SA, Morsy NF, Elaby SM, Ghaly MA. Hypolipidemic effect of *Moringa oleifera* Lam leaf powder and its extract in diet-induced hypercholesterolemic rats. *Journal of medicinal food*. 2017 Aug 1;20(8):755-62.

UNDER PEER REVIEW