

1

High Levels of EFNA2 in Lung

2

Adenocarcinoma Predicts Poor Prognosis

3

12

13 **ABSTRACT**

14 **Background:** The incidence of lung adenocarcinoma (LUAD) is increasing worldwide with different
15 prognosis. Ephrin-A2 (EFNA2), a member of the Eph/ephrin family, is associated with tumor
16 progression. However, the correlations of EFNA2 with prognosis in LUAD remain unclear.

17 **Methods:** In this study, we investigated the relationship between the expression and prognostic value
18 of the EFNA2 gene in LUAD patients in The Cancer Genome Atlas (TCGA) database. Sequential data
19 filtering (survival analysis, independent prognostic analysis, and clinical correlation analysis) was
20 performed. EFNA2 expression was analyzed by the Oncomine database and Tumor Immune
21 Estimation Resource (TIMER). We evaluated the influence of EFNA2 on clinical prognosis using
22 Kaplan-Meier plotter, the PrognoScan database and Gene Expression Profiling Interactive Analysis
23 (GEPIA). The correlation between EFNA2 and cancer immune infiltrates was investigated by TIMER.
24 In addition, correlations between EFNA2 expression and gene marker sets of immune infiltrates were
25 analyzed by TIMER and GEPIA. In addition, gene enrichment analysis was performed by Metascape.

26 Finally, a co-expression analysis was performed by the Oncomine database.

27 **Results:** A cohort of LUAD patients showed that high EFNA2 expression was associated with poorer
28 overall survival (OS), disease-free survival (DFS) by TCGA, and EFNA2 was significantly associated
29 with stage in LUAD. In addition, EFNA2 expression was positively correlated with infiltrating levels
30 of B cells and CD8⁺ T cells. Moreover, the differential expression of EFNA2 was significantly higher in
31 lung adenocarcinoma compared with that in normal controls. Specifically, EFNA2 was positively
32 associated with ADAMTSL5, REEP6, PCSK4, C19orf25, and ANAPC2.

33 **Conclusions:** Our data indicate that EFNA2 is a potential diagnostic and prognostic biomarker and a
34 promising molecular therapeutic target to attenuate LUAD progression.

35 Keywords: EFNA2, LUAD, prognosis, TCGA

36

37 INTRODUCTION

38 Lung cancer is the leading cause of cancer-related deaths with an increasing incidence of lung
39 adenocarcinoma (LUAD) subtype worldwide^[1]. Prognosis may vary in patients with the same stage tumor
40 because cancer is characterized by genetic, epigenetic, and phenotypic changes that result in a tremendous
41 variability in clinical behavior. Therefore, the development of additional molecular markers for survival
42 prediction of LUAD is required. Lung cancer is a malignant tumor caused by abnormal growth of bronchial
43 cells, and primary lung cancer is prone to metastasis. The most common pathological types include:
44 squamous cell carcinoma, adenocarcinoma, small cell lung cancer, and so on. Among them, LUAD accounts
45 for 40%-55% of the total number of lung cancers, most of which originate from the bronchial mucosal
46 epithelium, and more than 3/4 of the patients' lesions occur in the periphery. The disease progresses
47 slowly, and the initial symptoms are generally not obvious, but it is easy to metastasize.

48 In recent years, molecular progress has changed the treatment of LUAD, and genetic testing has become
49 a standard diagnosis and prognostic indicator and could determine the treatment target. The progress of
50 bioinformatics and high-throughput sequencing was whether we could identify many tumor biomarkers,
51 which could help improve the accuracy of predicting the prognosis of LUAD, and find increasingly effective
52 treatments. EFNA2 was a member of the Ephrins family. Ephrins, ligands for the Eph receptors, its
53 physiological role not only involved cell-to-cell communication, cell adhesion, cell migration, and invasion,
54 but also involved the regulation of blood vessel development and angiogenesis. They were promiscuous in
55 a very complex web of relationships (FIGURE 1). At present, the targeted therapy of LUAD has made
56 outstanding progress, but there were still about 10% of patients with negative genetic testing, so we
57 needed more genetic testing sites to improve the prognosis. using CGGA and TCGA data, Liu^[2] speculate on
58 genes that may affect the survival time of patients with low-grade gliomas. In this study, data filtering
59 (survival analysis, independent prognostic analysis, and clinical correlation analysis) from TCGA data was
60 used to screen the EFNA2. bioinformatics analysis of EFNA2 was performed. In addition, the relationship
61 between EFNA2 and clinical characteristics was also studied. Furthermore, gene enrichment analysis was
62 performed. Finally, a co-expression analysis was performed.

63

64 MATERIALS AND METHODS

65 Oncomine Database Analysis

66 We analyzed the EFNA2 mRNA levels in different tumors and normal tissues of multiple cancer types using
67 the Oncomine database. (<https://www.oncomine.org/resource/login.html>) . The threshold was determined
68 according to the following values: P-value of 0.001, fold change of 1.5, and gene ranking of all.

69

70 Data Download and Preprocessing

71 Gene expression data and corresponding clinical data from LUAD patients were downloaded from TCGA
72 (<http://www.cggg.org.cn/>). This dataset that contained 594 samples (DataSet ID: mRNAseq_594, Data Type:
73 RNA sequencing) were downloaded. The gene expression data from LUAD samples were corrected in
74 batches and integrated by loading them into the limma (14) and sva (15) packages in R software (R version
75 3.6.1:<https://www.rproject.org/>).

76

77 Survival Analysis Filtering

78 Survival and survminer packages were loaded in R software, and Kaplan–Meier (K-M) (16) and univariate
79 Cox analyses were used to filter gene expression data and survival data at a significance level of $P < 0.05$.

80

81 Independent Prognostic Analysis Filtering

82 The gene expression data obtained from the survival analysis and integrated clinical information were
83 analyzed using multivariate Cox analysis with R software, at a significance level of $P < 0.05$.

84

85 Analysis of the Correlation Between EFNA2 Expression and Clinical Characteristics

86 The correlation between EFNA2 expression and various clinical characteristics was plotted using
87 UALCAN(<http://ualcan.path.uab.edu/analysis.html>).

88

89 Gepia Database Analysis

90 The examination of EFNA2 expression in homogeneous subsets of LUAD was performed in GEPIA. GEPIA, an
91 interactive web server containing RNA sequencing data based on 9,736 tumor samples and 8,587 normal

92 samples from the TCGA and GTEx databases, provides customizable functions such as tumor/normal
93 differential expression analysis, patient survival analysis, and correlation analysis.

94

95 Gene Enrichment Analysis

96 In this study, Metscape was used to generate an ordered list of all genes associated with the expression of
97 EFNA2. Then, Metscape was used to identify survival differences between the high and low EFNA2 groups.

98

99 Analysis of Immune Infiltration

100 Tumor Immune Estimation Resource (TIMER, <https://cistrome.shinyapps.io/timer/>) was used to
101 comprehensively study the molecular characteristics of tumor-immune interactions(18). The abundances of
102 six immune infiltrates, including B cells, CD4+ T cells, CD8+ T cells, macrophages, neutrophils, and dendritic
103 cells were evaluated. We analyzed the relationship between the expression level of EFNA2 and the level of
104 immune infiltration in LUAD using the TIMER “gene” module. The Kaplan-Meier method was used to plot
105 the effect of EFNA2 expression and immune cell infiltration on the prognosis of patients with LUAD, and
106 clinical factors were included to construct a multivariate Cox proportional risk model. Finally, the
107 relationship between copy number variations (CNVs) of EFNA2 in different somatic cells and the level of
108 infiltration in LUAD was analyzed using the “SCNA” module.

109

110 Co-expression Analysis

111 Oncomine (<https://www.oncomine.org>) was used to screen genes that were co-expressed with EFNA2. In
112 addition, the pheatmap (<https://github.com/taiyun/corrplot>) package was used to plot the first 20 genes
113 positively and negatively associated with EFNA2. The Corplot (<https://github.com/taiyun/corrplot>) and

114 Circlize packages in R were used to generate a circular plot of the top five genes positively and negatively
115 associated with EFNA2.

116

117 RESULTS

118

119 The Expression Levels of EFNA2 in Different Types of Human Cancers

120 The expression level of the EFNA2 gene in various types of cancers was identified in the Oncomine
121 database. This analysis revealed that the EFNA2 expression was higher in lung cancer compared to normal
122 tissues (**Figure 2A**). To further evaluate EFNA2 expression in human cancers, we examined EFNA2
123 expression using the RNA-seq data of multiple malignancies in TCGA determined by TIMER. The differential
124 expression between the tumor and adjacent normal tissues for EFNA2 across all TCGA tumors is shown in
125 **Figure 2B**. EFNA2 expression was significantly lower in LUAD (lung adenocarcinoma) compared with
126 adjacent normal tissues.

127

128 Kaplan-Meier survival analysis of the TCGA dataset showed that low EFNA2 expression was associated with
129 better prognosis in patients with lung adenocarcinoma (**Figure 3A**). Univariate Cox analysis showed that
130 EFNA2 (HR = 1.065; 95% CI = 1.022-1.110; $P < 0.05$), T stage, N stage, M stage and TNM stage were
131 high-risk factors, and pathology were low risk factors (**Figure 3B**). Multivariate Cox analysis showed that
132 EFNA2 (HR = 1.053; 95% CI = 1.007–1.102; $P < 0.05$) was independently associated with overall survival,
133 which suggested that EFNA2 could be an independent prognostic indicator for lung adenocarcinoma. In
134 addition, pathology and TNM stage may also be independent prognostic factors (**Figure 3C**).

135

136 Analysis of the Relationship Between EFNA2 Expression and Clinical Characteristics

137 Analysis of 573 samples from the TCGA database showed that the differential expression of EFNA2 was
138 significantly higher in lung adenocarcinoma compared with that in normal control by using UALCAN^[3]
139 (**Figure 4A**, $P < 0.001$). We also can see the significant different between normal tissues and 41-88years,
140 different stages, gender, N0-N3 nodal metastasis, different races, NOS, mixed and mucinous histological
141 subtypes, different smoking habits, different TP53 mutations (**Figure 4B-I**). In addition, analysis of 573
142 samples from the TCGA database showed that the differential expression of EFNA2 was significantly
143 associated with patients race and histological subtypes (**Figure 4F**). The EFNA2 expression of Caucasian was
144 significant different to African-american($P < 0.001$) or Asian($P < 0.05$). And the EFNA2 expression of mixed
145 subtype was significant different to lung bronchioloalveolar carcinoma, non-mucinous subtype($P < 0.05$) or
146 lung solid pattern predominant adenocarcinoma subtype($P < 0.05$). The EFNA2 expression of lung
147 adenocarcinoma not otherwise specified (NOS) subtype was significant different to the lung solid pattern
148 predominant adenocarcinoma subtype($P < 0.05$).

149

150

151 Relationship Between EFNA2 Expression and Prognosis of LUAD Patients

152 GEPIA is a public database established for expression profiling analysis of cancer and normal genes.^[4]
153 Prognostic analysis revealed that high expression of EFNA2 would lead to a short overall survival in patients
154 with LUAD based on GEPIA database (**Figure 5A**, $P < 0.05$). In addition, EFNA2 was significantly associated
155 with stage in LUAD analyzed by GEPIA database(**Figure 5B**, $P < 0.05$).

156

157 Analysis of the Correlation Between EFNA2 Expression and Clinical

158 Characteristics in TCGA Database

159 Analysis using TIMER showed that EFNA2 was negatively associated with B cells and CD8⁺ T cells (**Figure 6**).

160 Univariate cox survival analysis showed that EFNA2, B cell, and dendritic cells were associated with the

161 survival of patients with LUAD (**Figure 7**). Furthermore, only arm level decreases in copy number variations

162 (CNVs) of EFNA2 were associated with the extent of immune infiltration in LUAD immune cells (**Figure 8**).

163 These results showed that EFNA2 was associated with immune infiltration in LUAD using external data

164 analysis. These findings indicated that EFNA2 might be a prognostic biomarker of LUAD and may be a target

165 for immunotherapy.

166

167 Gene Enrichment Analysis of EFNA2

168 Gene enrichment analysis is a computational method used to determine whether a group of genes is

169 differentially expressed in two biological states. Metascape gene enrichment analysis was used to identify

170 GO (Gene Ontology) and KEGG signaling pathways that were differentially expressed in LUAD between the

171 low and high EFNA2 expression groups. The results showed significant differences in enrichment using

172 Metascape(<http://metascape.org/>). The most significantly enriched GO and signaling pathways were

173 selected based on p value. As shown in **Figure 9**, A regulated exocytosis, blood vessel development, cell

174 cycle phase transition, gene ontology terms, extracellular matrix organization, cell cycle, mitotic,

175 hemostasis, collagen formation, PID integrin1 pathway, nervous system development signaling pathway

176 were enriched in the EFNA2 high expression phenotype.

177 Co-expression Analysis of EFNA2

178 A heatmap (**Figure 10A**) of the top 20 genes positively and negatively associated with EFNA2 was plotted.

179 In addition, a circular plot (**Figure 10B**) of the top five genes positively and negatively associated with

180 EFNA2 was generated. The results showed that EFNA2 was positively associated with ADAMTSL5, REEP6,
181 PCSK4, C19orf25 and ANAPC2, and was negatively associated with MSLNL, SLC35F2, RAB39B, BIRC3 and
182 KIAA1377.

183

184 Discussion

185 The treatment methods of LUAD included surgery, radiotherapy, chemotherapy, targeted therapy, and
186 immunotherapy. At present, surgery, radiotherapy, and chemotherapy have been used for many years, and
187 the therapeutic effect has been basically applied to the limit. There was still a lot of research space for
188 targeted therapy. Therefore, we studied the related genes of LUAD, hoping to find new treatment methods
189 or prognostic factors. We showed that EFNA2 was a high-risk factor and could be an independent
190 prognostic indicator in patients with LUAD using comprehensive univariate and multivariate Cox analyses.
191 Taken altogether, these results indicated that EFNA2 was upregulated in LUAD, and EFNA2 had a
192 prognostic value in LUAD, indicating that EFNA2 had important regulatory functions in LUAD. No previous
193 studies have reported a link between EFNA2 and lung cancer. However, EFNA2 is a member of the ephrin
194 family whose genes have been reported to be frequently overexpressed in a wide variety of cancer types
195 directly regulating critical steps of cellular adhesion, tumor growth, chemo-repulsion, invasion, metastasis,
196 angiogenesis, axon guidance, tissue border formation [5, 6] [7, 8] [9-12] [13-23]. The relative studies showed that
197 the expression levels of EFNA2 were negative and correlated with the prognosis of prostate cancer^[24],
198 CD133 high neuroblastoma^[25, 26], breast cancer^[11], hepatocellular carcinoma^[10], gastric cancer^[27] and
199 colorectal cancer^[28]. For example, Feng et al. found that EFNA2 is significantly upregulated in both
200 cancerous cell lines and clinical tissue samples of hepatocellular carcinoma (HCC) and compared with the
201 normal ones^[10]. In addition, Fox et al. reported that the expression of EFNA2 is significantly higher in CPTX

202 cells (human local prostate tumor) compared to NPTX cells (normal human prostate epithelium),
203 suggesting that EFNA2 may promote the transformation of the normal prostate epithelial cell into one with
204 a malignant phenotype^[12]. The transcript for EFNA2 is also highly and shows a relative increase in more
205 aggressive prostate cancer compared to its parent cell line, implying that EFNA2 may also play an important
206 role in promoting prostate cancer invasion. In addition, EFNA2 participates in the regulation of diverse
207 cellular processes and gene expression through chromatin remodeling, and the expression of EFNA2 at the
208 transcription level would lead to the activation of signaling pathways related to tumor progression. For
209 example, Liu et al. showed that blocking EFNA2 expression inhibits the metastasis ability of human liver
210 cancer cell line HepG2^[29]. Kuo et al. showed that EFNA2 might be a prognostic predictor of multiple DNA
211 methylation biomarkers for early-stage LUAD in Asian and Caucasian populations.^[30] Moreover, ephrins
212 family via Ephrin receptor (Eph)–ephrin interactions regulate critical steps of angiogenesis, blood vessel
213 formation malignant transformation, tumor metastasis, tumor differentiation, and outcome^[13-15, 23, 31, 32].
214 For example, upregulation of EphA2 has been observed in many malignant tumors and is associated with
215 accelerated cell proliferation, stimulating angiogenesis, and promoted cell migration and invasion,
216 increasing cancer cell survival ^[13-23]. In addition, research and clinical trials have confirmed the proteolytic
217 shedding of membrane-bound Ephrin-As, which releases soluble fragments at the cellular level^[33-35]. For
218 example, membrane-bound EFNA2 has been identified as the substrate of ADAM10 and released soluble
219 EFNA2 fragments into the cell medium ^[33, 34]. These data suggested that secreted EFNA2 may be useful
220 serum markers for the diagnosis and prognosis of many tumors ^[36].
221 Besides, gene enrichment analysis was performed to obtain further information about the role of EFNA2 in
222 tumor progression. The results of TIMER showed that a regulated exocytosis, blood vessel development,
223 cell cycle phase transition, Gene ontology terms, extracellular matrix organization, cell cycle, mitotic,

224 hemostasis, collagen formation, PID integrin1 pathway, nervous system development signaling pathways
225 were enriched in the EFNA2 high expression phenotype. The extracellular matrix regulates tissue
226 development and homeostasis, and its dysregulation contributes to neoplastic progression. The
227 extracellular matrix serves not only as the scaffold upon which tissues are organized but provides critical
228 biochemical and biomechanical cues that direct cell growth, survival, migration, and differentiation and
229 modulate vascular development and immune function.^[37, 38]

230 The gene ontology terms of EFNA2 were generally enriched in B cell related mediated immunity, humoral
231 immune response, and innate immune response. B lymphocyte was recognized to participate in regulating
232 the immune response to murine and human tumors^[39]. Regulatory B cell play an immunosuppressive role
233 in carcinogenesis and become a therapeutic target in solid tumors^[40].Recent studies indicated that the B
234 lymphocytes exists in all stages of cancer and plays important roles in shaping tumor development in lung
235 cancer and thus influences the prognosis of lung cancer patients ^[41, 42].

236 Finally, co-expression analysis showed that EFNA2 was positively associated with ADAMTSL5, REEP6, PCSK4,
237 C19orf25, and ANAPC2, and was negatively associated with MSLNL, SLC35F2, RAB39B, BIRC3 and KIAA1377.

238 As previously reported, ADAMTSL5 was an epigenetically activated gene underlying tumorigenesis and drug
239 resistance in hepatocellular carcinoma and pointed to a role for ADAMTSL5 in maintaining the function of
240 key oncogenic signalling pathways, suggesting that it may act as a master regulator of tumorigenicity and
241 drug resistance. ^[43]. Moreover, proliferation and metastasis of lung cancer cells lacking REEP5 and REEP6
242 were markedly decreased compared to the control group, and they could be novel regulators of
243 G-protein-coupled receptor signaling ^[44]. These reports suggested that ADAMTSL5 and REEP6 were
244 associated with the regulation of proliferation and metastasis of cancer. Our study showed EFNA2 to be
245 associated with ADAMTSL5 and REEP6, which indicated that EFNA2 might be associated with the regulation

246 of proliferation and metastasis of cancer.

247 Ephs and ephrins were regarded as promising candidates for drug development. However, Eph and ephrin

248 had been considered as undruggable target molecules because the interactions between Eph and ephrin,

249 shown in Figure 1, were not specific and promiscuous in a very complex web of relationships. Many

250 processes that involve fast changes in cellular motility and/or morphology depend on ephrin–Eph signaling

251 pathway. Therefore, there are no drugs against the Eph/ephrin family for medical use included in the

252 clinical guidelines so far. Dasatinib is the only drug for medical use that shows an inhibitory effect on EphA2

253 activity^[45]. However, dasatinib has not been used for any anti-EphA2 therapy so far. Recently, Richard

254 Huang et al. discovered EFNA2 targeted immunoliposomes incorporating pH-sensitive taxane prodrugs

255 were developed for sustained delivery of active drugs to solid tumors, and this drug had entered a Phase I

256 clinical trial^[46]. These results indicated that EFNA2 may be a useful molecular therapeutic target to

257 attenuate LUAD progression.

258 We speculated that EFNA2 may be used as a prognostic indicator for LUAD, and future studies will be

259 needed to explore the protein in a multidisciplinary way in the future, hoping to find a molecular predictor

260 with great clinical value.

261

262 CONCLUSION

263 In conclusion, this study investigated the relationship between EFNA2 and LUAD prognosis. First, sequential

264 data filtering was used to screen the key gene EFNA2. Then, EFNA2 was analyzed for correlation with

265 prognosis and clinical characteristics. The results show that high expression of EFNA2 was associated with

266 worse prognosis, and EFNA2 was a high-risk factor and could be used as an independent prognostic

267 indicator for patients with LUAD. Furthermore, Metascape gene enrichment analysis showed that EFNA2

268 could regulate the proliferation and metastasis of LUAD and that upregulation of EFNA2 in LUAD indicates
269 poor prognosis. EFNA2 mediates cell-to--to-cell interactions both in tumor cells and in the tumor
270 microenvironment, namely, the tumor stroma and vasculature. Thus, EFNA2 has been considered as
271 attractive targets for drug design, as targeting these molecules could simultaneously inhibit several aspects
272 of tumor progression. However, few studies have shown the association between EFNA2 and LUAD or
273 other cancers. Future studies will be needed to confirm the role of these genes as therapeutic targets and
274 biomarkers in LUAD. which suggests that this study may provide a scaffold for future development of
275 therapeutic strategies for LUAD.

276

277 DATA AVAILABILITY STATEMENT

278 The data that support the findings of this work are obtainable from the corresponding author based on
279 reasonable request.

291

292

293

References

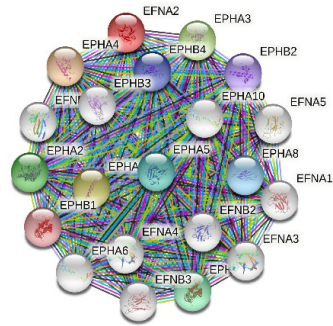
- 294 1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018:
295 GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries.
296 CA Cancer J Clin. 2018;68(6):394-424.
- 297 2. Liu W, Xu Z, Zhou J, Xing S, Li Z, Gao X, et al. High Levels of HIST1H2BK in Low-Grade Glioma
298 Predicts Poor Prognosis: A Study Using CGGA and TCGA Data. Front Oncol. 2020;10:627.
- 299 3. Chandrashekar DS, Bashel B, Balasubramanya SAH, Creighton CJ, Ponce-Rodriguez I,
300 Chakravarthi B, et al. UALCAN: A Portal for Facilitating Tumor Subgroup Gene Expression and
301 Survival Analyses. Neoplasia. 2017;19(8):649-58.
- 302 4. Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene
303 expression profiling and interactive analyses. Nucleic Acids Res. 2017;45(W1):W98-w102.
- 304 5. Zhang J, Hughes S. Role of the ephrin and Eph receptor tyrosine kinase families in
305 angiogenesis and development of the cardiovascular system. J Pathol. 2006;208(4):453-61.
- 306 6. Surawska H, Ma PC, Salgia R. The role of ephrins and Eph receptors in cancer. Cytokine
307 Growth Factor Rev. 2004;15(6):419-33.
- 308 7. Yates NJ, Martin-Iverson MT, Rodger J. The role of ephrin-A2 and ephrin-A5 in sensorimotor
309 control and gating. Behav Brain Res. 2014;275:225-33.
- 310 8. Diercke K, Sen S, Kohl A, Lux CJ, Erber R. Compression-dependent up-regulation of ephrin-A2
311 in PDL fibroblasts attenuates osteogenesis. J Dent Res. 2011;90(9):1108-15.
- 312 9. Hafner C, Schmitz G, Meyer S, Bataille F, Hau P, Langmann T, et al. Differential gene
313 expression of Eph receptors and ephrins in benign human tissues and cancers. Clin Chem.
314 2004;50(3):490-9.
- 315 10. Feng YX, Zhao JS, Li JJ, Wang T, Cheng SQ, Yuan Y, et al. Liver cancer: EphrinA2 promotes
316 tumorigenicity through Rac1/Akt/NF-kappaB signaling pathway. Hepatology.
317 2010;51(2):535-44.
- 318 11. Fox BP, Kandpal RP. Invasiveness of breast carcinoma cells and transcript profile: Eph
319 receptors and ephrin ligands as molecular markers of potential diagnostic and prognostic
320 application. Biochem Biophys Res Commun. 2004;318(4):882-92.
- 321 12. Fox BP, Tabone CJ, Kandpal RP. Potential clinical relevance of Eph receptors and ephrin
322 ligands expressed in prostate carcinoma cell lines. Biochem Biophys Res Commun.
323 2006;342(4):1263-72.
- 324 13. Zelinski DP, Zantek ND, Stewart JC, Irizarry AR, Kinch MS. EphA2 overexpression causes
325 tumorigenesis of mammary epithelial cells. Cancer Res. 2001;61(5):2301-6.
- 326 14. Ogawa K, Pasqualini R, Lindberg RA, Kain R, Freeman AL, Pasquale EB. The ephrin-A1 ligand
327 and its receptor, EphA2, are expressed during tumor neovascularization. Oncogene.
328 2000;19(52):6043-52.
- 329 15. Brantley-Sieders DM, Fang WB, Hwang Y, Hicks D, Chen J. Ephrin-A1 facilitates mammary

- 330 tumor metastasis through an angiogenesis-dependent mechanism mediated by EphA
331 receptor and vascular endothelial growth factor in mice. *Cancer Res.* 2006;66(21):10315-24.
- 332 16. Thaker PH, Deavers M, Celestino J, Thornton A, Fletcher MS, Landen CN, et al. EphA2
333 expression is associated with aggressive features in ovarian carcinoma. *Clin Cancer Res.*
334 2004;10(15):5145-50.
- 335 17. Kamat AA, Coffey D, Merritt WM, Nugent E, Urbauer D, Lin YG, et al. EphA2 overexpression is
336 associated with lack of hormone receptor expression and poor outcome in endometrial
337 cancer. *Cancer.* 2009;115(12):2684-92.
- 338 18. Taddei ML, Parri M, Angelucci A, Bianchini F, Marconi C, Giannoni E, et al. EphA2 induces
339 metastatic growth regulating amoeboid motility and clonogenic potential in prostate
340 carcinoma cells. *Mol Cancer Res.* 2011;9(2):149-60.
- 341 19. Miao H, Li DQ, Mukherjee A, Guo H, Petty A, Cutter J, et al. EphA2 mediates
342 ligand-dependent inhibition and ligand-independent promotion of cell migration and invasion
343 via a reciprocal regulatory loop with Akt. *Cancer Cell.* 2009;16(1):9-20.
- 344 20. Karidis NP, Giaginis C, Tsurouflis G, Alexandrou P, Delladetsima I, Theocharis S. Eph-A2 and
345 Eph-A4 expression in human benign and malignant thyroid lesions: an immunohistochemical
346 study. *Med Sci Monit.* 2011;17(9):Br257-65.
- 347 21. Strimpakos A, Pentheroudakis G, Kotoula V, De Roock W, Kouvatses G, Papakostas P, et al.
348 The prognostic role of ephrin A2 and endothelial growth factor receptor pathway mediators
349 in patients with advanced colorectal cancer treated with cetuximab. *Clin Colorectal Cancer.*
350 2013;12(4):267-74.e2.
- 351 22. Tan YC, Srivastava S, Won BM, Kanteti R, Arif Q, Husain AN, et al. EPHA2 mutations with
352 oncogenic characteristics in squamous cell lung cancer and malignant pleural mesothelioma.
353 *Oncogenesis.* 2019;8(9):49.
- 354 23. Yeddula N, Xia Y, Ke E, Beumer J, Verma IM. Screening for tumor suppressors: Loss of ephrin
355 receptor A2 cooperates with oncogenic KRas in promoting lung adenocarcinoma. *Proc Natl*
356 *Acad Sci U S A.* 2015;112(47):E6476-85.
- 357 24. Li S, Wu Z, Chen Y, Kang Z, Wang H, He P, et al. Diagnostic and prognostic value of tissue and
358 circulating levels of Ephrin-A2 in prostate cancer. *Tumour Biol.* 2016;37(4):5365-74.
- 359 25. Cournoyer S, Nyalendo C, Addioui A, Belounis A, Beaunoyer M, Aumont A, et al. Genotype
360 analysis of tumor-initiating cells expressing CD133 in neuroblastoma. *Genes Chromosomes*
361 *Cancer.* 2012;51(8):792-804.
- 362 26. Genander M, Frisén J. Ephrins and Eph receptors in stem cells and cancer. *Curr Opin Cell Biol.*
363 2010;22(5):611-6.
- 364 27. Kikuchi S, Kaibe N, Morimoto K, Fukui H, Niwa H, Maeyama Y, et al. Overexpression of Ephrin
365 A2 receptors in cancer stromal cells is a prognostic factor for the relapse of gastric cancer.
366 *Gastric Cancer.* 2015;18(3):485-94.
- 367 28. Herath NI, Boyd AW. The role of Eph receptors and ephrin ligands in colorectal cancer. *Int J*
368 *Cancer.* 2010;126(9):2003-11.
- 369 29. Liu JL, Yan-Juan LI, Liu JG, Yan LI, Dong XSJPIMB. Blocking EFNA2 Expression Inhibits
370 Metastasis Ability of Human Liver Cancer Cell Line HepG2. 2012.
- 371 30. Kuo IY, Jen J, Hsu LH, Hsu HS, Lai WW, Wang YC. A prognostic predictor panel with DNA
372 methylation biomarkers for early-stage lung adenocarcinoma in Asian and Caucasian
373 populations. *J Biomed Sci.* 2016;23(1):58.

- 374 31. Mosch B, Reissenweber B, Neuber C, Pietzsch J. Eph receptors and ephrin ligands: important
375 players in angiogenesis and tumor angiogenesis. *J Oncol.* 2010;2010:135285.
- 376 32. Hérault M, Schaffner F, Augustin HG. Eph receptor and ephrin ligand-mediated interactions
377 during angiogenesis and tumor progression. *Exp Cell Res.* 2006;312(5):642-50.
- 378 33. Hattori M, Osterfield M, Flanagan JG. Regulated cleavage of a contact-mediated axon
379 repellent. *Science.* 2000;289(5483):1360-5.
- 380 34. Janes PW, Saha N, Barton WA, Kolev MV, Wimmer-Kleikamp SH, Nievergall E, et al. Adam
381 meets Eph: an ADAM substrate recognition module acts as a molecular switch for ephrin
382 cleavage in trans. *Cell.* 2005;123(2):291-304.
- 383 35. Ieguchi K, Tomita T, Omori T, Komatsu A, Deguchi A, Masuda J, et al. ADAM12-cleaved
384 ephrin-A1 contributes to lung metastasis. *Oncogene.* 2014;33(17):2179-90.
- 385 36. Lisle JE, Mertens-Walker I, Rutkowski R, Herington AC, Stephenson SA. Eph receptors and
386 their ligands: promising molecular biomarkers and therapeutic targets in prostate cancer.
387 *Biochim Biophys Acta.* 2013;1835(2):243-57.
- 388 37. Pickup MW, Mouw JK, Weaver VM. The extracellular matrix modulates the hallmarks of
389 cancer. *EMBO Rep.* 2014;15(12):1243-53.
- 390 38. Walker C, Mojares E, Del Río Hernández A. Role of Extracellular Matrix in Development and
391 Cancer Progression. *Int J Mol Sci.* 2018;19(10).
- 392 39. Qiu GZ, Mao XY, Ma Y, Gao XC, Wang Z, Jin MZ, et al. Ubiquitin-specific protease 22 acts as an
393 oncoprotein to maintain glioma malignancy through deubiquitinating B cell-specific Moloney
394 murine leukemia virus integration site 1 for stabilization. *Cancer Sci.* 2018;109(7):2199-210.
- 395 40. Fremd C, Schuetz F, Sohn C, Beckhove P, Domschke C. B cell-regulated immune responses in
396 tumor models and cancer patients. *Oncoimmunology.* 2013;2(7):e25443.
- 397 41. Wang SS, Liu W, Ly D, Xu H, Qu L, Zhang L. Tumor-infiltrating B cells: their role and application
398 in anti-tumor immunity in lung cancer. *Cell Mol Immunol.* 2019;16(1):6-18.
- 399 42. Stankovic B, Bjørhovde HAK, Skarshaug R, Aamodt H, Frafjord A, Müller E, et al. Immune Cell
400 Composition in Human Non-small Cell Lung Cancer. *Front Immunol.* 2018;9:3101.
- 401 43. Arechederra M, Bazai SK, Abdouni A, Sequera C, Mead TJ, Richelme S, et al. ADAMTSL5 is an
402 epigenetically activated gene underlying tumorigenesis and drug resistance in hepatocellular
403 carcinoma. *J Hepatol.* 2021;74(4):893-906.
- 404 44. Park CR, You DJ, Park S, Mander S, Jang DE, Yeom SC, et al. The accessory proteins REEP5 and
405 REEP6 refine CXCR1-mediated cellular responses and lung cancer progression. *Sci Rep.*
406 2016;6:39041.
- 407 45. Chang Q, Jorgensen C, Pawson T, Hedley DW. Effects of dasatinib on EphA2 receptor tyrosine
408 kinase activity and downstream signalling in pancreatic cancer. *Br J Cancer.*
409 2008;99(7):1074-82.
- 410 46. Huang ZR, Tipparaju SK, Kirpotin DB, Pien C, Kornaga T, Noble CO, et al. Formulation
411 optimization of an ephrin A2 targeted immunoliposome encapsulating reversibly modified
412 taxane prodrugs. *J Control Release.* 2019;310:47-57.

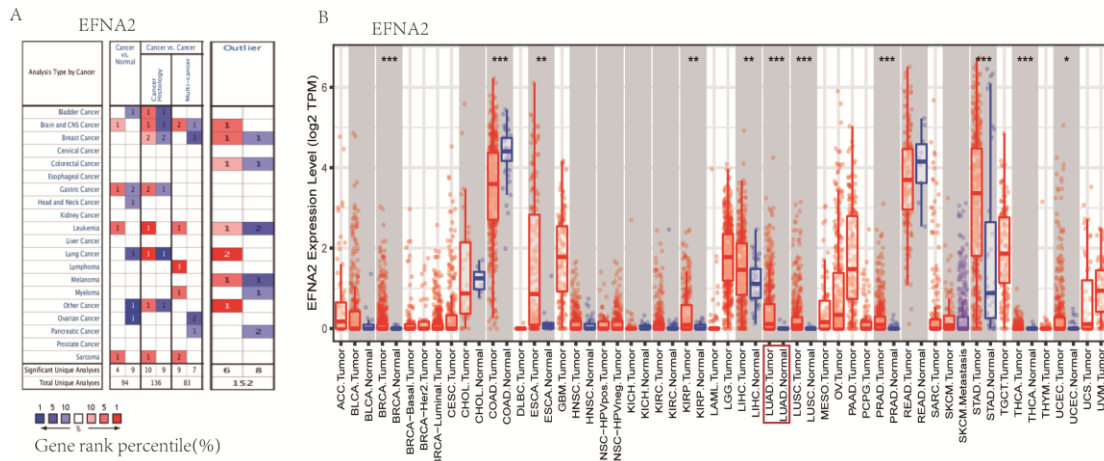
413

414 Figure legends



415

416 FIGURE 1 | Establishment of the PPI network.



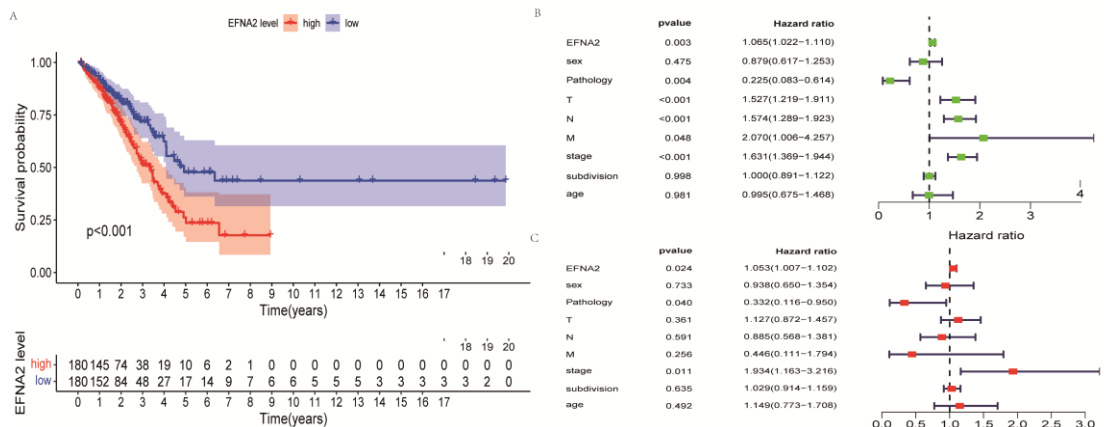
417

418 FIGURE 2 | EFNA2 expression levels in different types of human cancers. (A) Increased or decreased EFNA2 in data sets of different cancers

419 compared with normal tissues and different pathology of cancer in the Oncomine database. (B) Human EFNA2 expression levels in different tumor

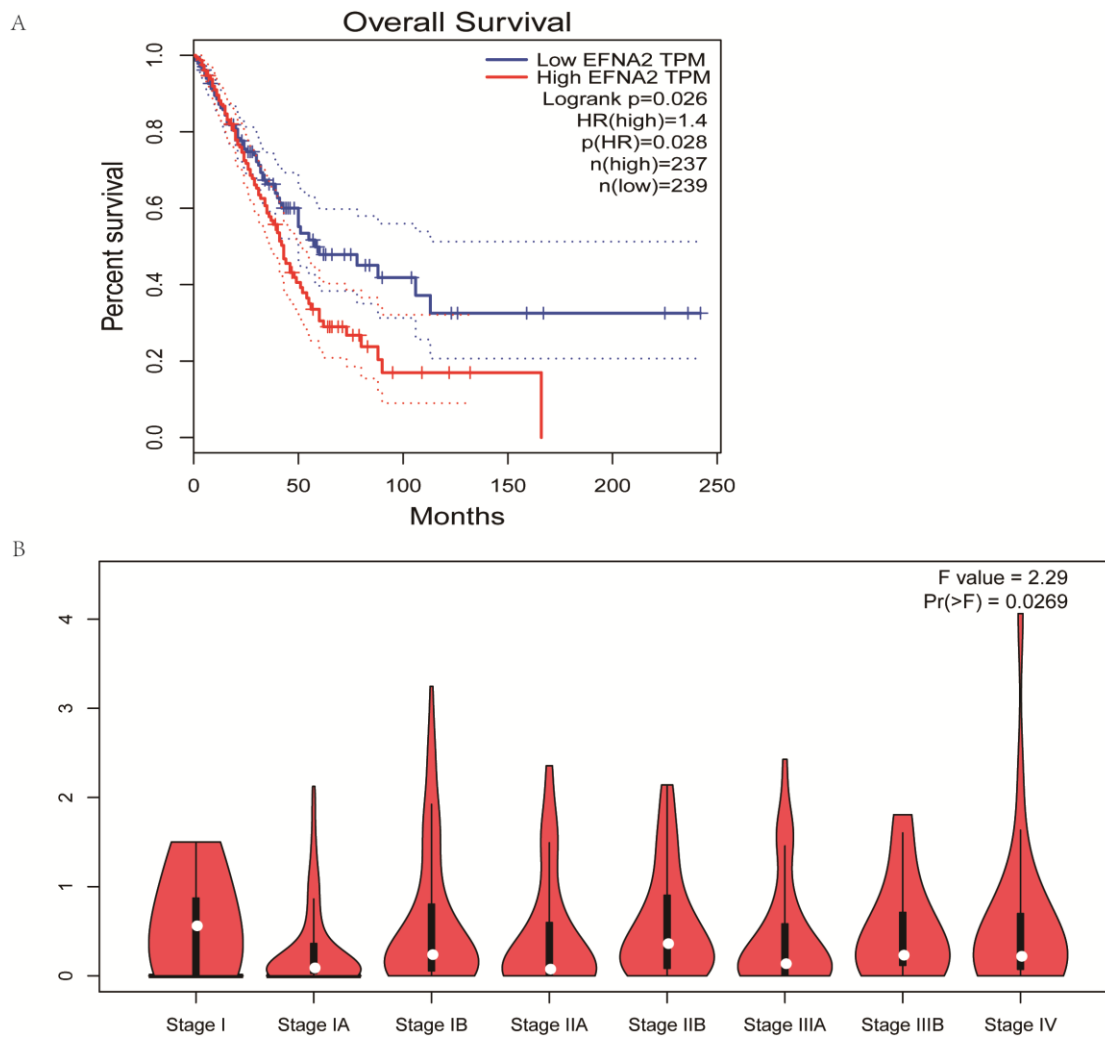
420 types from TCGA database were determined by TIMER. (*P<0.05,**P<0.01,***P<0.001).

421



422

423 FIGURE 3 | Bioinformatics analysis of EFNA2 using the TCGA database (LUAD, n = 367). (A) Survival analysis of patients with lung



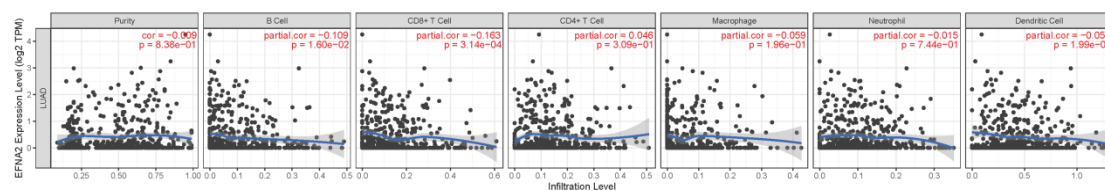
434

435 FIGURE 5 | Relationship between EFNA2 expression and prognosis of LUAD patients based on GEPIA database. (A) The relationship between

436 EFNA2 expression levels and overall survival in LUAD was analyzed by GEPIA database. $P < 0.05$. (B) EFNA2 was significantly associated with

437 stage in LUAD analyzed by GEPIA database. $P < 0.05$.

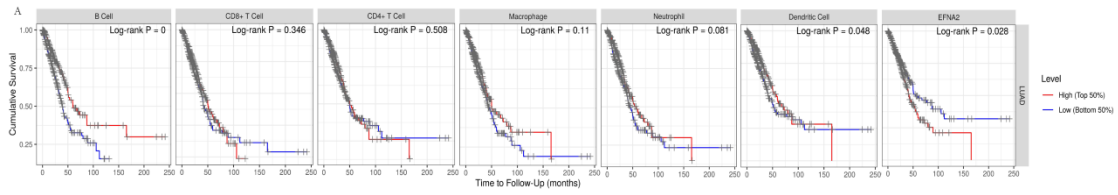
438



439

440 FIGURE 6 | Correlation between the expression of EFNA2 and immune infiltration of LUAD cells.

441



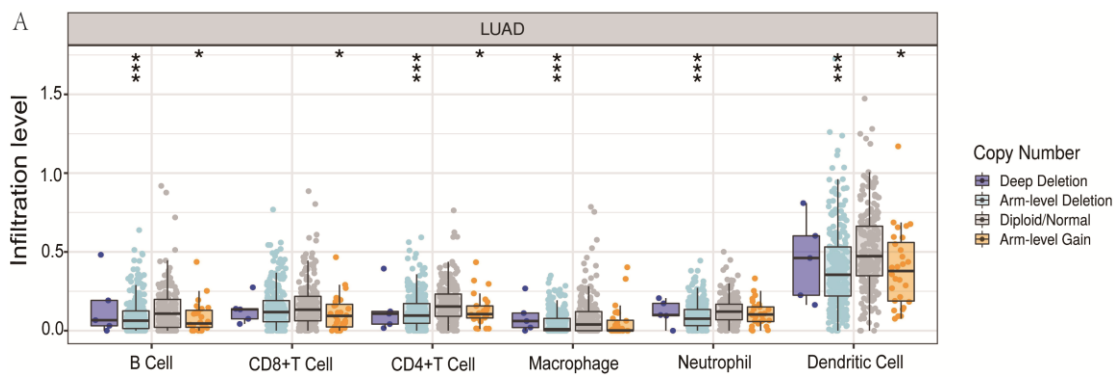
442

443 FIGURE 7 | Survival curve for immune cell infiltration. Kaplan–Meier survival curves based on top and bottom sample partitions with 50%

444 immune penetration. Red indicates a high degree of infiltration and blue indicates a low degree of infiltration. $P < 0.05$ was considered significant

445 and $P < 0.0001$ was reported as 0.

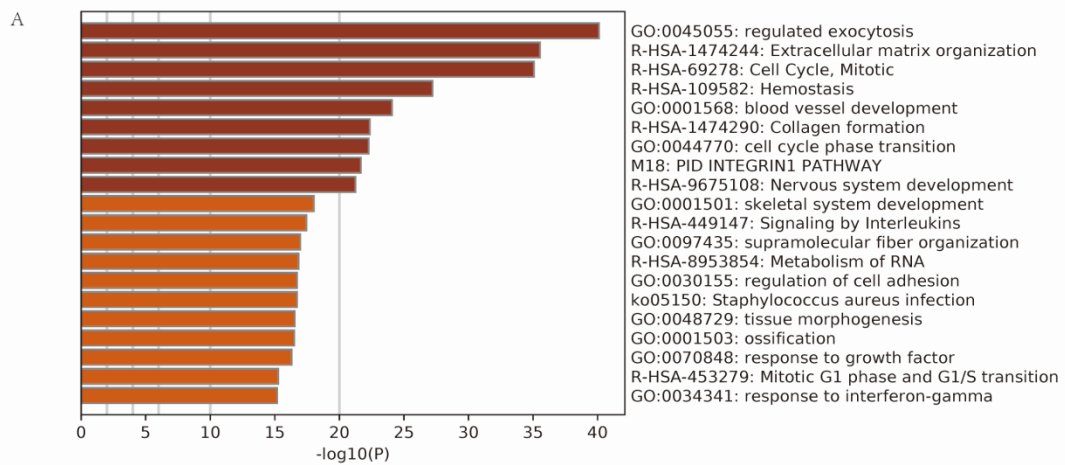
446



447

448 FIGURE 8 | Relationship between copy number variation of EFNA2 and immune infiltration level in LUAD. * $P < 0.05$; *** $P < 0.001$.

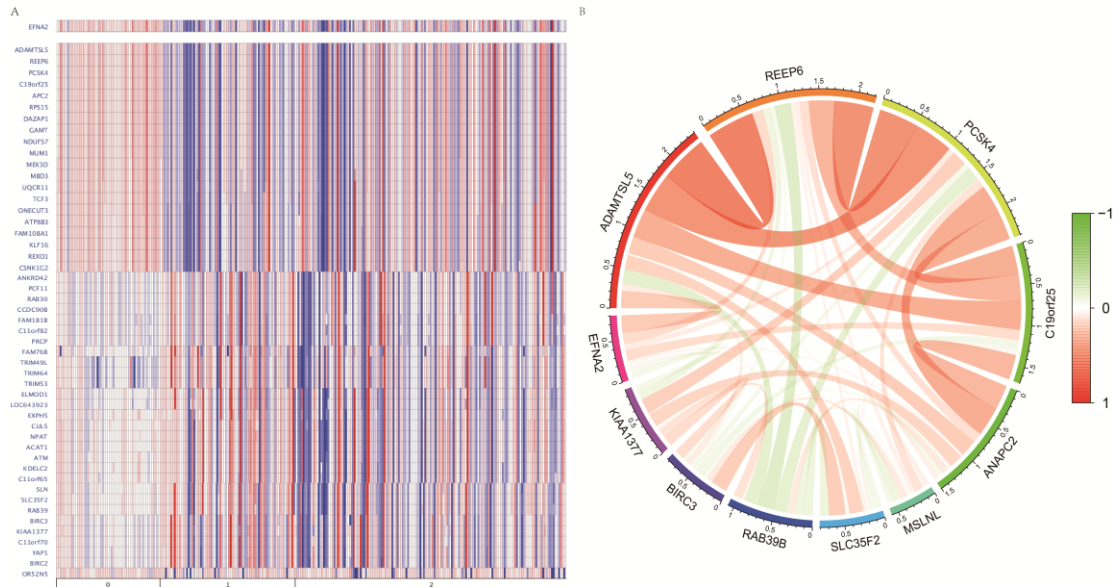
449



450

451 FIGURE 9 | Gene Set Enrichment Analysis of EFNA2. Heatmap of enriched terms across input gene lists, colored by p-values.

452



453

454 FIGURE 10 | Co-expression analysis of EFNA2 using the TCGA database. (A) Heatmap of the top 20 genes positively and negatively associated

455 with EFNA2. (B) Circular plot of the top five genes positively and negatively related to the EFNA2 gene. Green represents negative association, and

456 red represents positive association.

457

458