

Advancements in Enhancing Oil Quality in Rapeseed and Mustard: A Comprehensive
Review

ABSTRACT

Rapeseed and mustard are important oilseed crops cultivated worldwide for their high oil content and versatile applications in food, feed, and industrial sectors. The quality of oil derived from these crops is influenced by various genetic, environmental, and agronomic factors. Breeding efforts aimed at improving oil quality traits in rapeseed and mustard have garnered significant attention in recent years due to their impact on market value, nutritional attributes, and industrial utility. In this review, we provide a comprehensive overview of breeding strategies and methodologies employed to enhance oil quality traits in rapeseed and mustard. We explore the genetic basis of oil quality traits, including fatty acid composition, erucic acid content, glucosinolate content, and tocopherol content, and discuss the importance of these traits for different end-uses. Furthermore, we highlight the role of molecular markers, genomics-assisted breeding, and biotechnological approaches in accelerating the breeding process and achieving targeted improvements in oil quality. The review also addresses the challenges and constraints associated with breeding for oil quality in rapeseed and mustard, including genotype-environment interactions, trait stability, and regulatory considerations. Additionally, we discuss emerging trends and future prospects in oil quality breeding, such as genome editing, metabolic engineering, and precision breeding, which offer novel avenues for achieving desired oil quality profiles while addressing sustainability and consumer preferences. Overall, this review underscores the significance of breeding for oil quality in rapeseed and mustard and provides insights into the latest advancements, challenges, and opportunities in this field. By integrating multidisciplinary approaches and harnessing the power of modern breeding tools and technologies, rapeseed and mustard breeders can continue to drive innovation and deliver oilseed crops with enhanced nutritional value, functional properties, and market competitiveness.

Key word: Rapessed mustard, erucic acid, glucosinolate, fatty acid.

Introduction

Introduction:

Rapeseed and mustard, belonging to the Brassicaceae family, are among the most economically important oilseed crops globally. The oils extracted from these crops serve as valuable sources of edible oil, biofuel, and industrial feedstocks. Moreover, they contribute significantly to global agriculture and food security due to their adaptability to diverse agro-climatic conditions and high oil yields per unit area. The quality of oil derived from rapeseed and mustard is a critical determinant of their market value and utilization across various sectors. Oil quality traits, such as fatty acid composition, erucic acid content, glucosinolate content, and tocopherol content, not only influence the nutritional attributes and flavor of the oil but also impact its suitability for specific industrial applications. Consequently, there is a growing emphasis on breeding efforts aimed at improving oil quality traits to meet the evolving demands of consumers, industries, and regulatory standards. In this context, this review aims to provide a comprehensive overview of the breeding strategies, methodologies, challenges, and future prospects associated with enhancing oil quality in rapeseed and mustard. We will delve into the genetic basis of oil quality traits, elucidate the role of environmental factors and agronomic practices in modulating oil quality, and discuss the significance of molecular breeding tools and biotechnological interventions in accelerating the breeding process. Furthermore, we will examine the market dynamics, consumer preferences, and regulatory frameworks influencing oil quality breeding initiatives in rapeseed and mustard. By synthesizing insights from scientific literature, breeding programs, and industry perspectives, this review seeks to offer valuable insights into the complexities, opportunities, and future directions of breeding for oil quality in these economically important oilseed crops. Ultimately, the knowledge generated from this review can inform strategic decision-making, guide research priorities, and facilitate the development of improved rapeseed and mustard varieties with enhanced oil quality attributes.

Classification of Rapeseed & Mustard

Rapeseed and mustard are broad terms that encompass several species and varieties within the Brassicaceae family. The classification of these oilseed crops involves a hierarchical system that categorizes them based on various criteria, including botanical characteristics, genetic traits, and agronomic features. Here is a simplified classification of rapeseed and mustard:

Genus: Brassica

1. Brassica napus (Canola/Rapeseed):

- Subspecies:

- a. Spring Canola/Rapeseed (*Brassica napus* subsp. *oleifera*): Grown in regions with milder winters, it is characterized by spring planting.

- b. Winter Canola/Rapeseed (*Brassica napus* subsp. *biennis*): Planted in the fall and overwinters to resume growth in the spring.

2. Brassica juncea (Indian mustard/Brown mustard):

- Varieties:

- a. Brown Mustard (*Brassica juncea* var. *juncea*): Known for its pungent flavor, it is widely used in condiments and spice production.

- b. Yellow Mustard (*Brassica juncea* var. *integrifolia*): Primarily used for producing the bright yellow mustard commonly found in condiment bottles.

3. Brassica rapa (Field mustard):

- Subspecies:

- a. Turnip rape (*Brassica rapa* subsp. *rapa*): Cultivated for its edible roots, leaves, and oil content in seeds.

- b. Chinese cabbage (*Brassica rapa* subsp. *pekinensis*): Primarily grown for its edible leaves, used in various culinary dishes.

4. Brassica carinata (Ethiopian mustard/Abyssinian mustard):

- Known for its high oil content and resistance to environmental stressors. It is cultivated in certain regions for biofuel production.

Additional Considerations:

- Geographical Variations: Different regions may have specific varieties or subspecies adapted to local climate and soil conditions.

- Hybrid Varieties: With advancements in breeding technologies, hybrid varieties with improved traits, including disease resistance and higher yield, are also part of the classification.

This classification provides a broad overview, and within each category, there are numerous cultivars and hybrids developed to meet specific agricultural, industrial, and culinary needs. It's essential to consider regional variations, breeding advancements, and specific traits when discussing the classification of rapeseed and mustard.

Rapeseed & Mustard scenario

Cultivation and Production:

Rapeseed and mustard are cultivated in diverse agro-climatic regions worldwide, including Europe, Canada, China, India, and Australia. Cultivation practices vary based on factors such as climate, soil type, and local agricultural practices. These crops are adaptable and can thrive in different environments, making them valuable for crop rotation and diversification.

Oilseed and Oil Production:

Rapeseed and mustard seeds are rich sources of oil, commonly used for culinary, industrial, and biofuel purposes. Canola, a variety of rapeseed, is known for its low erucic acid and glucosinolate content, making it suitable for human consumption and industrial applications. Mustard oil is popular in South Asian cuisines and is also used in traditional medicine and cosmetics.

Nutritional and Health Benefits:

Rapeseed and mustard oils are rich in monounsaturated and polyunsaturated fatty acids, including omega-3 and omega-6 fatty acids, which are beneficial for heart health. Mustard seeds contain phytochemicals with potential antioxidant and anti-inflammatory properties, contributing to their health benefits.

Industrial and Commercial Uses:

Rapeseed and mustard oils are utilized in food processing, including cooking oils, salad dressings, and margarine production. In addition to culinary uses, these oils are used in industrial applications such as lubricants, biofuels, and cosmetics.

Trade and Market Dynamics:

Global trade in rapeseed and mustard oils is significant, with major exporting and importing countries involved in trade agreements and partnerships. Price fluctuations, weather conditions, and geopolitical factors influence market dynamics and trade patterns.

Research and Development:

Ongoing research focuses on breeding high-yielding varieties with improved disease resistance, stress tolerance, and oil quality traits. Biotechnological advancements, including genetic engineering and genome editing, are being explored to enhance crop productivity and sustainability.

Environmental and Sustainability Considerations:

Sustainable cultivation practices, such as conservation tillage and integrated pest management, are promoted to minimize environmental impacts and preserve soil health. Crop rotation with rapeseed and mustard contributes to biodiversity, reduces pest pressure, and improves soil fertility.

Overall, the rapeseed and mustard scenario reflects the importance of these oilseed crops in global agriculture, food security, and economic development. Continued research, innovation, and sustainable practices are essential for addressing challenges and maximizing the potential of rapeseed and mustard cultivation in the future.

Chemical Composition

Chemical Composition:

Rapeseed and mustard seeds contain a diverse array of chemical compounds that contribute to their nutritional, industrial, and culinary properties. The chemical composition of these seeds varies depending on factors such as species, variety, growing conditions, and processing methods. However, some common components include:

1. **Fatty Acids:** Both rapeseed and mustard seeds are rich sources of fatty acids, including monounsaturated, polyunsaturated, and saturated fats. The predominant fatty acids in rapeseed oil are oleic acid (monounsaturated) and linoleic acid (polyunsaturated), while mustard oil contains a higher proportion of erucic acid and linolenic acid.

2. **Proteins:** Rapeseed and mustard seeds are also rich in proteins, which are essential for human nutrition. These proteins contain a variety of amino acids, including lysine, methionine, and cysteine, which contribute to their nutritional quality.

3. **Glucosinolates:** Mustard seeds contain glucosinolates, sulfur-containing compounds responsible for their pungent taste and aroma. Glucosinolates have potential health benefits, including antioxidant and anti-cancer properties, but excessive consumption may also pose health risks.

4. **Vitamins and Minerals:** Rapeseed and mustard seeds contain various vitamins and minerals, including vitamin E, vitamin K, calcium, magnesium, and phosphorus. These nutrients play important roles in maintaining overall health and well-being.

5. Phytosterols: Both rapeseed and mustard seeds contain phytosterols, plant-derived compounds that have been shown to help reduce cholesterol levels and improve heart health.

6. Antioxidants: Rapeseed and mustard seeds contain antioxidants such as tocopherols and phenolic compounds, which help protect cells from oxidative damage and may have anti-inflammatory properties.

Fats and oil

Oils are fats that remain liquid at room temperature, such as vegetable oils commonly used in cooking. They are derived from various plants and sometimes from fish. While oils are not classified as a distinct food group, they provide essential nutrients vital for health.

Fats consist of short, medium, and long-chain molecules composed of essential fatty acids linked together, similar to how proteins are composed of amino acids. Certain fatty acids are considered essential because they cannot be synthesized in sufficient quantities by the body and must be obtained through the diet for optimal health and well-being.

Fats and oils contribute to the texture, taste, and palatability of foods, enhancing their overall sensory appeal. They also serve as dense sources of energy, providing 9 kilocalories per gram compared to 4.9 kilocalories per gram from cereals and pulses. Additionally, fats and oils play crucial roles in the absorption and transportation of fat-soluble vitamins (A, E, and K) and fat-soluble antioxidants and nutrients.

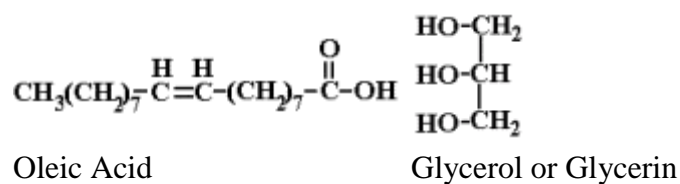
Dietary intake of fats is essential for various nutritional and metabolic processes. Vegetable oils, in particular, serve as the primary source of essential fatty acids for the body. Essential fatty acids serve as precursors to prostaglandins, a group of biologically active compounds that regulate physiological processes such as blood pressure, prevention of blood clotting in arteries, and protection against vascular damage in the brain and heart.

Prostaglandins are increasingly recognized for their anti-proliferative activities, making them valuable in medical contexts as potential "Miracle Drugs." Thus, fats and oils, particularly those derived from vegetable sources, play crucial roles in supporting overall health and well-being beyond their nutritional value alone.

Triglycerides are indeed the primary constituents of both vegetable oils and animal fats. These molecules possess lower densities than water, causing them to float when in contact with water. Triglycerides can exist in solid or liquid states at normal room temperatures. When triglycerides are solid, they are commonly referred to as "fats" or "butters," while in their liquid state, they are termed "oils."

Chemically, a triglyceride, also known as a triacylglycerol (TAG), is composed of one molecule of glycerol linked with three fatty acids. This chemical structure forms the basis of most dietary fats and oils, which are crucial components of human nutrition and metabolism.

Understanding triglycerides is fundamental to comprehending the composition and properties of fats and oils, as well as their roles in human health and nutrition.



Indeed, glycerol, also known as glycerin or glycerine, is a trihydric alcohol, characterized by containing three hydroxyl (-OH) groups in its molecular structure. These hydroxyl groups enable glycerol to combine with fatty acids, leading to the formation of various compounds such as monoglycerides, diglycerides, and triglycerides.

In the process of esterification, the hydroxyl groups of glycerol react with the carboxyl groups of fatty acids, resulting in the formation of ester bonds. Monoglycerides are produced when one fatty acid molecule combines with one hydroxyl group of glycerol, while diglycerides are formed when two fatty acid molecules combine with two hydroxyl groups of glycerol. Triglycerides, on the other hand, are generated when three fatty acid molecules react with all three hydroxyl groups of glycerol.

These compounds, monoglycerides, diglycerides, and triglycerides, are classified as esters due to their chemical structure. Esters are a type of compound created through esterification, a chemical reaction between acids and alcohols. During esterification, water (H₂O) is released as a by-product, and esters are formed as the resultant compounds.

Understanding the process of esterification and the formation of esters like monoglycerides, diglycerides, and triglycerides is crucial in comprehending the

chemistry of lipids and their roles in biological systems, including nutrition, metabolism, and cellular structure.

The main biosynthetic pathway of fatty acid synthesis is as follows:

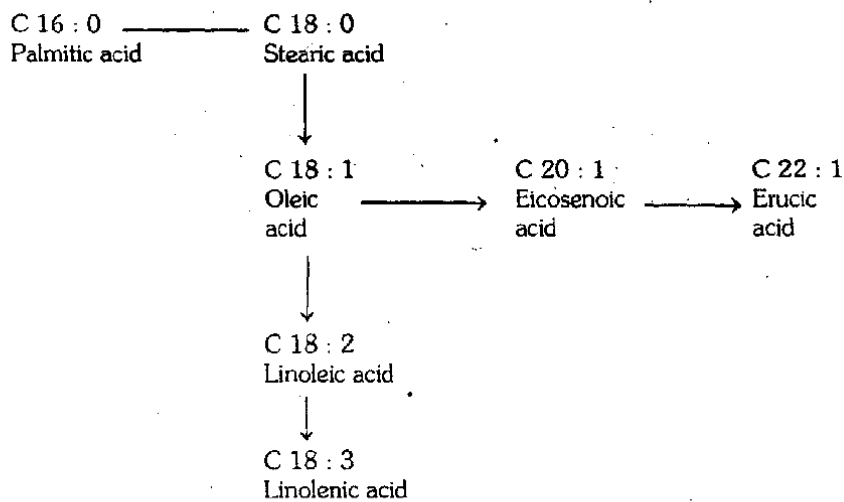


Fig. 1. Biosynthetic pathway of main fatty acids (Jonsson, 1977)

Both erucic acid and linolenic acid are indeed derived from the biosynthetic pathway involving oleic acid, which undergoes either desaturation or further chain elongation processes. In this pathway, oleic acid can undergo decreasing saturation to produce linoleic acid and further elongation to form eicosenoic acid, which eventually leads to the synthesis of erucic acid.

The genetic control of the chain elongation step, particularly in the conversion of oleic acid to erucic acid, is critical in regulating the biosynthesis of erucic acid from oleic, linolenic, and linoleic acids. Canadian breeders have successfully achieved reductions in the levels of erucic acid and linolenic acid through selective breeding methods that target the enzymes responsible for their synthesis.

Interestingly, linoleic acid and linolenic acid are both produced through the same biosynthetic desaturation pathway. Selection for high levels of linoleic acid has been observed to lead to increased levels of linolenic acid as well. Conversely, selection for low levels of linoleic acid has been associated with reductions in linolenic acid levels.

Understanding the genetic regulation and biosynthetic pathways of fatty acids like erucic acid and linolenic acid is crucial for agricultural breeding programs aimed at developing crops with improved oil quality and nutritional profiles. Selective breeding approaches targeting specific enzymes involved in fatty acid synthesis offer promising avenues for achieving desired fatty acid compositions in oilseed crops.

Table 1. Fatty acid composition of rapeseed-mustard seed oil

Fatty acids		Per cent
SFA	Palmitic (16:0)	1-3
	Stearic (18:0)	1-3
MUFA/ PUFA	Oleic (18:1)	8-40
	Linoleic (18:2)	10-29
	Linolenic (18:3)	5-18
	Erucic (22:1)	42-57

- Oil quality in rapeseed and mustard is primarily defined by the types and proportions of fatty acids present in the oil. Linoleic acid (LA) and linolenic acid (18:3) are essential considerations in determining oil quality. LA is a colorless liquid at room temperature with two cis double bonds in its 18-carbon chain. Conversely, linolenic acid's three double bonds make it prone to auto-oxidation, resulting in off-flavors and reduced shelf-life of the oil.
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- Erucic acid, a monounsaturated omega-9 fatty acid, is another key component. Its single double-bonded carbon atom distinguishes it as monounsaturated, and its double carbon bond occurs at the ninth position from the end of its acid chain (n-9 position).
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- Glucosinolates, sulfur and nitrogen-containing compounds derived from glucose and an amino acid, are also crucial in classifying oil quality. Based on fatty acid presence, rapeseed and mustard varieties are categorized into:
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 - 1. Single Zero (Single Low): Varieties with low erucic acid (< 2%) but a high percentage of glucosinolate in seed meal.
 - 2. Double Zero (Double Low or Canola): Varieties with less than 2% erucic acid and less than 30 u moles/g of glucosinolate in seed meal.
 - 3. Triple Zero (Bell 1984): Double low yellow-coated varieties low in fiber, also referred to as triple varieties.
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 - Understanding the composition and classification of fatty acids and glucosinolates is essential for assessing and enhancing the quality of rapeseed and mustard oils, ensuring their suitability for various culinary and industrial applications.

Table 2. Diversified usage of oil with modified fatty acid composition

Type	Use
Zero erucic acid < 2%	Nutritionally superior
High erucic acid (40-55%) (>80%)	Industrial polymers, lubricants, plastic industry Cosmetics, pharmaceuticals.
High stearic acid (20-40%)	Margarines
Epoxy fatty acids	Polymers
Wax esters	Resins
High petroselinic acid	Cosmetics, lubricants
Very low linolenic acids (<3%)	Prolonged shelf life. Margarines
High linoleic acid (40-50%)	Nutritionally superior
Oleic acid (upto 70%)	Nutritionally superior

Breeding for quality, particularly the development of canola or '00' varieties, and their cultivation in India offer numerous advantages:

1. Elevating Nutritional Value: Canola varieties have the potential to enhance the nutritional value of both oil and seed meal, providing healthier options for consumers.

2. Fetching Remunerative Market Price: High-quality canola varieties can command better market prices, benefiting farmers and the agricultural economy.

3. Increasing Market Value and Versatile Usage: Improved quality oils and seed meals from canola varieties can be used in a variety of food, industrial, and pharmaceutical applications, expanding their market value and usage.

4. Enhancing Export Potential: Canola seed meal with reduced erucic acid and glucosinolate content can meet international standards, enhancing its export potential and contributing to foreign exchange earnings.

Quality breeding efforts in India began in the 1970s, initially focusing on evaluating existing genetic variability in indigenous germplasm. Despite efforts, zero erucic acid and low glucosinolate types were not identified initially. Glucosinolate levels in indigenous selections ranged from 63 to 102 μ moles/g on a seed basis.

During this phase, an important achievement was the identification of SM 1, a low erucic acid accession with only 10% erucic acid content. However, this genetic stock had limitations in agronomic performance and could not be utilized either as a commercial variety or as a donor source for quality improvement.

The pace of research gained momentum with the initiation of the Indo-Swedish collaborative project in 1975. Several '0' erucic acid strains were identified during this phase, laying the foundation for further quality improvement efforts in canola breeding in India.

In India, the rapeseed oil and seed meal quality improvement program aims to achieve several objectives:

1. Evaluation of Available Lines: The program involves evaluating low erucic acid and low glucosinolate lines of *Brassica juncea* and *Brassica napus* for both yield and quality parameters.

2. **Development of Varieties:** It focuses on developing low erucic acid and/or low glucosinolate rapeseed-mustard varieties through breeding efforts.
3. **Genetic and Breeding Studies:** Basic studies are conducted to understand the genetics and breeding behavior of erucic acid and glucosinolate content in rapeseed-mustard varieties.
4. **Pest and Disease Resistance:** Information is generated on the reaction of '0/00' types to endemic pests and diseases, aiding in the development of resistant varieties.

Concerted efforts in evaluating indigenous and exotic germplasm have revealed a wide range of variation for different fatty acids in Indian mustard.

Breeding efforts in India, dating back to 1970, have been focused on reducing glucosinolate content in rapeseed-mustard varieties to up to 30 micro moles/g defatted seed meal (low or 0) and erucic acid to up to 2% (low or 0). The goal is to develop double zero or double low varieties that meet internationally acceptable standards for oil and seed meal.

Notably, the first low erucic acid variety, Pusa Karishma of Indian mustard, and the first double low variety, GSC 5 of gobhi sarson, were released in 2004 and 2005, respectively. Currently, five low erucic varieties have been released in Brassica juncea, and five double low varieties have been released in gobhi sarson (*B. napus*).

Ongoing efforts focus on recombining low erucic acid with low glucosinolate content in Indian mustard and refining the agronomic base to enhance the yield potential of double low gobhi sarson strains. These efforts underscore the commitment to improving the quality and productivity of rapeseed-mustard varieties in India.

Source of quality traits:

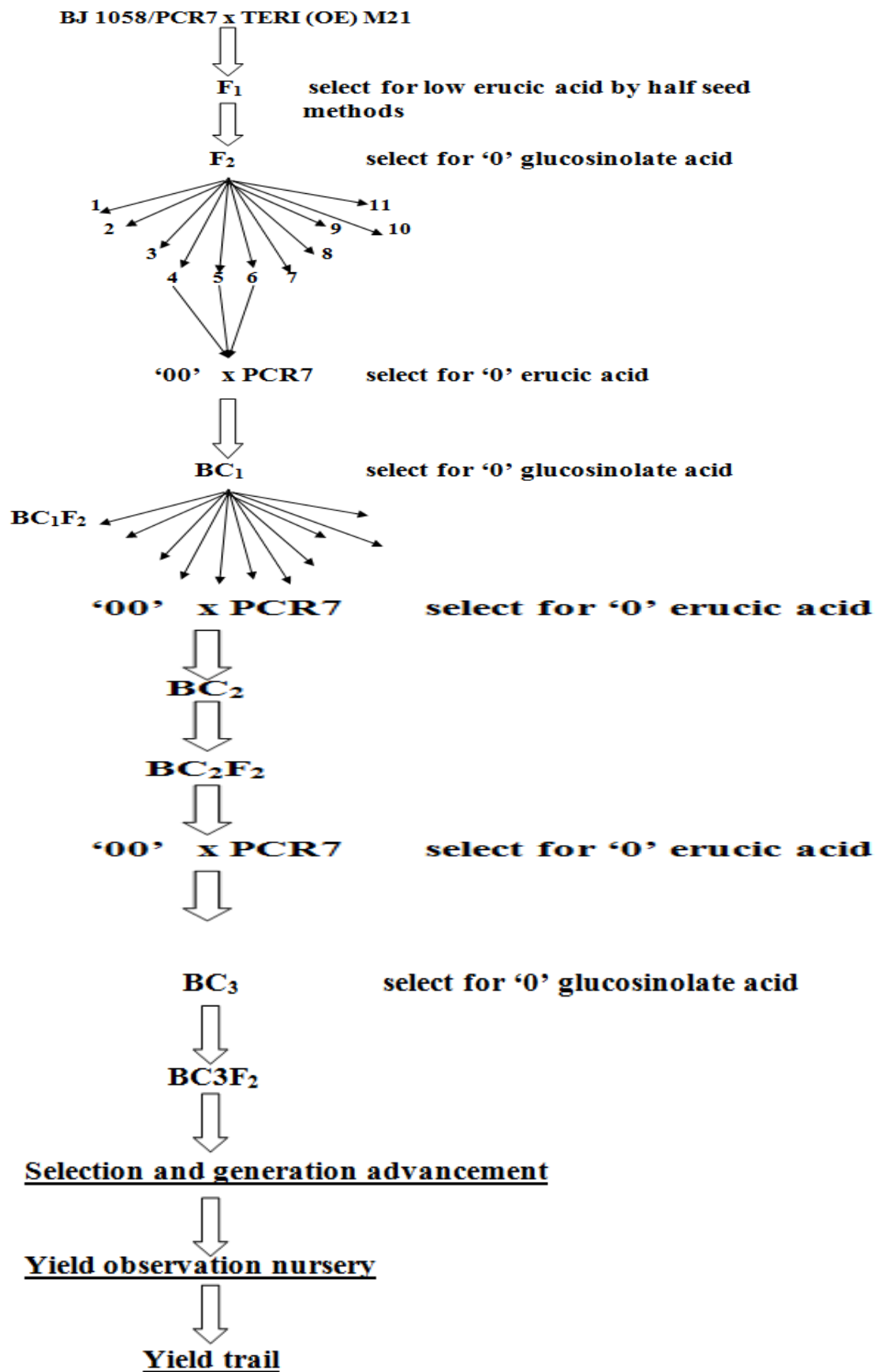


Fig 2. Backcross Method

The backcross method, specifically employing the half-seed technique, is utilized to improve the quality of rapeseed cultivars by eliminating erucic acid. Here's how the process works:

1. Germination Process: Seeds are allowed to germinate on wet filter paper until the radicle (embryonic root) reaches a length of 10 mm.
2. Removal of Outer Cotyledons: Once the radicle reaches the desired length, the outer cotyledons of the seed are removed.
3. Fatty Acid Analysis: The removed outer cotyledons are then analyzed for their fatty acid composition using techniques such as gas-liquid chromatography or paper chromatography.
4. Selection of Embryos: Only those test embryos lacking erucic acid are selected and planted for further growth and development.
5. Growth to Maturity: Selected embryos are grown to maturity under controlled conditions to observe their growth characteristics and performance.

The synthesis of eicosenoic and erucic acids is controlled by two multiple allelic loci, which means that the presence or absence of erucic acid can be influenced by genetic factors at these loci.

Through the backcross method and the half-seed technique, breeders can selectively propagate rapeseed cultivars without erucic acid, thus improving the quality and suitability of the rapeseed oil for various applications, including culinary and industrial uses.

Table 3. Synthesis of eicosenoic and erucic acids is controlled by two multiple allelic loci

S.No.	Breeding methods	Species	Name of varieties/lines	Remarks
01	Selection	<i>B. compestris</i>	Span	Zero Erucic acid

		<i>B. napus</i>	oro	Zero Erucic acid
02	Mutation	<i>B. juncea</i>		3.6% linolenic acid
03	Hybridization	<i>B. juncea</i>	EC287711, Shiva 1, QM 13, QM14, QM39 RW 21-61	Low erucic acid
			Zem-1 Zem-2	Zero & low Erucic acid respectively
Biotechnological Approach				
04	Somaclonal Variation	<i>B. juncea</i>	Pusa jai kisan	Isolated from Varuna
05	Anther culture	<i>B. napus</i>	Winfield	Canola type
06	Transgenic	<i>B. compestris</i>	Laurical	Lauric acid content
07	Transgene(s) Antisense fae 1	<i>B. juncea</i> and <i>B. napus</i>		Low erucic acid

Status of quality improvement programme in India.

In the context of Brassica juncea, several developments have been noted:

- Evaluation of 124 strains with low erucic acid/glucosinolate content occurred under AICRP R&M until the 2010–11 cropping season.
- Pusa Karishma (LES 39), recognized as the first low erucic acid mustard, was released for Delhi state in 2004. Five low erucic varieties were identified and released up to December 2010. Additionally, four strains, including one low erucic and low glucosinolate and two low erucic and low glucosinolate varieties, have been registered with the National Bureau of Plant Genetic Resources (ICAR), New Delhi.

Regarding *B. napus*:

- Evaluation of forty-one strains with low erucic acid and/or low glucosinolate content took place under AICRP R&M until the 2010–11 cropping season.

- Hyola 401, the first double low erucic gobhi sarson hybrid, was released in 1996. By December 2010, five strains with low erucic and low glucosinolate content had been identified and released. Six strains, comprising two low erucic and four low erucic and low glucosinolate varieties, have been registered with the National Bureau of Plant Genetic Resources (ICAR), New Delhi.

In *B. rapa* var. *toria*:

- Two strains of toria, namely PHOP 2-2 (INGR 07033) with high oleic acid (70.1%) and low erucic acid and PHOT 8-2-11 (INGR 07034) with low linolenic acid (3.03%), have been registered with the National Bureau of Plant Genetic Resources (ICAR), New Delhi.

These developments represent significant progress in the development and registration of low erucic acid and low glucosinolate varieties, enhancing the quality and suitability of rapeseed-mustard crops for various agricultural and industrial applications.

Table 7. Varieties recommended for specific conditions/ possessing particular trait.

Crop Species	Traits	Name of variety/varieties /trait
Indian mustard	High oil content	Narendra Swarna Rai 8, NRCDR 02, Rohini
	Quality trait (low erucic acid)	Pusa Karishma, Pusa Mustard 21, (low erucic acid) Pusa Mustard 22, Pusa Mustard 24, ELM 079 (RLC 1)
Gobhi sarson	Quality traits Low erucic acid and glucosinolate	GSC 5, Hyola 401, NUDB 26-11, OCN 3 (GSC 6), TERI Uttam Jawahar

Quality breeding encounters several challenges that impede the development of improved varieties:

1. **Polygenic Nature of Traits:** Many quality traits are polygenic, meaning they are controlled by multiple genes. This complexity makes it challenging to select for desired traits during segregating generations, as the inheritance patterns are intricate and difficult to predict accurately.

2. **Difficulty in Estimation and Evaluation:** Evaluating quality traits can be challenging and resource-intensive. Traits such as oil composition, glucosinolate content, and erucic acid levels require specialized techniques such as gas chromatography, which demand substantial resources, including time and money.

3. **Low Heritability and Environmental Influence:** Quality traits often exhibit low heritability, meaning that the proportion of variation due to genetic factors is limited. Additionally, these traits are highly influenced by environmental factors, leading to variations in trait expression across different growing conditions. The interaction between genetics and environment complicates the selection process and slows down progress in improving quality through breeding efforts.

Addressing these challenges requires a multidisciplinary approach, incorporating advances in genetics, genomics, breeding methodologies, and agronomic practices. Collaborative efforts among researchers, breeders, and agricultural stakeholders are crucial for overcoming these obstacles and developing superior varieties with improved quality traits. Additionally, the development of robust phenotyping techniques and the utilization of molecular markers can aid in the accurate evaluation and selection of desired quality traits, contributing to the advancement of quality breeding programs.

Future Strategies

In order to advance quality breeding programs for rapeseed and mustard, future strategies could include:

1. Diversification of Genetic Sources: Expanding the genetic diversity of low glucosinolate varieties and incorporating them into breeding programs can enhance the range of options available to breeders. This diversification helps in developing varieties with improved traits and resilience to various environmental stresses.

2. Development of Rapid Screening Methods: Investing in the development of cost-effective and efficient screening methods for fatty acids and glucosinolates can streamline the early generation selection process. Rapid screening techniques enable breeders to identify promising lines more quickly, accelerating the breeding cycle and reducing resource expenditure.

3. Utilization of Biotechnological Tools: Embracing biotechnological tools, such as dihaploid breeding and marker-assisted selection (MAS), can significantly enhance selection efficiency and precision. Dihaploid breeding facilitates the production of homozygous lines in a single generation, expediting the breeding process. Meanwhile, MAS enables the targeted selection of desirable traits by utilizing molecular markers linked to specific genes of interest, thereby enhancing the accuracy and efficiency of selection.

By implementing these future strategies, breeders can overcome existing challenges and expedite the development of superior rapeseed and mustard varieties with improved quality traits, ultimately contributing to the sustainability and productivity of agriculture.

Conclusion

breeding for oil quality in rapeseed and mustard holds significant promise for improving agricultural productivity and enhancing human nutrition. The review of breeding efforts highlights several key findings and implications:

1. **Progress in Genetic Enhancement:** Over the years, substantial progress has been made in enhancing the oil quality of rapeseed and mustard varieties. Breeding programs have successfully developed varieties with reduced erucic acid and glucosinolate content, thereby improving the nutritional profile and suitability of the oils for various applications.

2. **Challenges and Limitations:** Despite advancements, breeding for oil quality faces challenges such as the polygenic nature of quality traits, difficulty in estimation and evaluation, and low heritability coupled with environmental influences. These challenges underscore the need for innovative breeding strategies and collaborative research efforts to overcome genetic and environmental constraints.

3. **Future Directions:** Future strategies for breeding programs include diversifying genetic sources, developing rapid screening methods, and leveraging biotechnological tools such as dihaploid breeding and marker-assisted selection. These approaches can enhance selection efficiency, accelerate breeding cycles, and facilitate the development of superior varieties with improved oil quality traits.

4. **Implications for Agriculture:** Improving oil quality in rapeseed and mustard varieties not only benefits farmers by increasing market value and versatility but also contributes to global food security and nutrition. High-quality oils derived from rapeseed and mustard play a crucial role in various culinary, industrial, and pharmaceutical applications, underscoring the importance of continued investment in breeding programs.

In summary, breeding for oil quality in rapeseed and mustard represents a dynamic and evolving field with immense potential for addressing agricultural challenges and meeting the growing demand for nutritious and sustainable food sources. By embracing innovative breeding approaches and fostering collaboration across disciplines, researchers and breeders can pave the way for a more resilient and productive agricultural sector.

References:

1. RAHMAN, M. S. (2015). *Comparative study on the chemical composition of different varieties and advanced line of rapeseed and mustard (brassica spp.)* (Doctoral dissertation, Department of Biochemistry, Sher-e-Bangla Agricultural University).
2. Aslan, V. (2023). An overview of biodiesel produced from 2nd generation feedstock: Mustard seed types. *BioEnergy Research*, 16(3), 1380-1400.
3. Thakur, A. K., Singh, K. H., Sharma, D., Parmar, N., & Nanjundan, J. (2019). Breeding and genomics interventions in Ethiopian mustard (*Brassica carinata* A. Braun) improvement—A mini review. *South African Journal of Botany*, 125, 457-465.
4. Banga, S. S. 2008. Breeding for improved quality in Brassica oilseeds. *Sustainable production of oilseeds rapeseed - mustard technology*. Agrotech Publishing Academy Udaipur-313 002:118-130.
5. Panjabi, P., Yadava, S. K., Kumar, N., Bangkim, R., & Ramchiary, N. (2019). Breeding Brassica juncea and B. rapa for sustainable oilseed production in the changing climate: progress and prospects. *Genomic Designing of Climate-Smart Oilseed Crops*, 275-369.
6. Beszterda, M., & Nogala- Kalucka, M. (2019). Current research developments on the processing and improvement of the nutritional quality of rapeseed (*Brassica napus* L.). *European Journal of Lipid Science and Technology*, 121(5), 1800045.
7. Bell, J. M. 1984. Nutrients and toxicants in rapeseed meal: a review. *J. Ani. Sci.* 58: 996– 1010.
8. Chauhan, J. S., Singh, K. H., Singh, V. V. and Kumar, S. 2011. Hundred Years of Rapeseed- Mustard Breeding in India: Accomplishments and Future Strategies. *I. J. Agril. Sci.* 81 (12): 1093–1109.
9. Singh, V. K., Bhoyar, P. I., Anu, & Sharma, V. (2022). Application of Genomics and Breeding Technologies to Increase Yield and Nutritional Qualities of Rapeseed-Mustard and Sunflower. *Technologies in Plant Biotechnology and Breeding of Field Crops*, 103-131.
10. Chauhan, J. S., Tyagi, M. K., Kumar, P. R., Tyagi, P., Singh, M., and Kumar, S. 2002. Breeding for oil and seed meal quality in Rapeseed - mustard in india – A review. *Agric. Rev.* 23(2):71-92.
11. Mehta, N., Tiwari, A., & Nag, S. K. (2015). A review: breeding for oil quality in rapeseed and mustard. *Trends in Biosciences*, 8(22), 6024-6031.

12. Agnihotri, A., Prem, D., & Gupta, K. (2007). The chronicles of oil and meal quality improvement in oilseed rape. *Advances in botanical research*, 45, 49-97.
13. Gurr, M.I. and Harwood, J.L. 1991. Fatty acid structure and metabolism. In: Gurr MI and Harwood J L (eds) *Lipid Biochemistry, An Introduction*. London: Chapman and Hall:21-27.
14. Shekhawat, K., Rathore, S. S., Premi, O. P., Kandpal, B. K., & Chauhan, J. S. (2012). Advances in agronomic management of Indian mustard (*Brassica juncea* (L.) Czernj. Cosson): an overview. *International journal of Agronomy*, 2012.
15. Hemingway, J. S. 1976. Mustard: *Brassica* spp. and *Sinapis alba* (Cruciferae). *Evolution of Crop Plants*. Simmonds (Ed.). Longmans, London: 56–69.
16. Kumar, S. and Chauhan, J. S. 2010. *Rapeseed-mustard: diatary and nutritional aspect*. Directorate of Rapeseed-Mustard Research, Sewar, Rajasthan Bharatpur-321 303:1-3.
17. McVetty, P. B., & Duncan, R. W. (2015). Canola, rapeseed, and mustard: for biofuels and bioproducts. *Industrial Crops: Breeding for BioEnergy and Bioproducts*, 133-156.
18. NBPGR. 2011. *Annual Report 2010–11*. National Bureau of Plant Genetic Resources, Pusa, New Delhi: 109.
19. USDA. 2010. United States Department of Agriculture - Rapeseed area, yield and production Table No.15. <http://www.fas.usda.gov/psdonline/psdreport>. Asps.
20. Willis, J.C. 1973. *A Dictionary of the Flowering Plants and Ferns. Eighth Edition*. Cambridge University Press, Cambridge et alibi. 1245pp.
21. Chauhan, J. S., Singh, K. H., Singh, V. V., & Kumar, S. (2011). Hundred years of rapeseed-mustard breeding in India: accomplishments and future strategies. *Indian J Agric Sci*, 81(12), 1093-1109.
22. Chand, Subhash, Om Prakash Patidar, Rajat Chaudhary, Ranjit Saroj, Kailash Chandra, Vijay Kamal Meena, Omkar M. Limbalkar, Manoj Kumar Patel, Priya P. Pardeshi, and Prashant Vasisth. "Rapeseed-mustard breeding in India: Scenario, achievements and research needs." *Brassica breeding and biotechnology* (2021): 174.
23. Nweze, C. C., & Muhammad, B. Y. (2023). Wandoo Tseaa, Rahima Yunusa, Happy Abimiku Manasseh, Lateefat Bisola Adedipe, Eneh William Nebechukwu, Yakubu Atanyi

Emmanuel(2023). Comparative Biochemical Ef-fects of Natural and Synthetic Pesticides on Preserved Phaseolus vulgaris in Male Albino Rats. *Acta Botanica Plantae*, 2, 01-10.

Sabitha, N., Mohan Reddy, D., Lokanadha Reddy, D., Hemanth Kumar, M., Sudhakar, P., Ravindra Reddy, B., & Mallikarjuna, S. J. (2022). Genetic divergence analysis over seasons in single cross hybrids of maize (*Zea mays* L.). *Acta Botanica Plantae*, 1(2), 12-18.

Islam, M. S., Rahman, M. M., & Paul, N. K. (2016). Arsenic-induced morphological variations and the role of phosphorus in alleviating arsenic toxicity in rice (*Oryza sativa* L.). *Plant Science Archives*

Ashokri, H. A. A., & Abuzririq, M. A. K. (2023). The impact of environmental awareness on personal carbon footprint values of biology department students, Faculty of Science, El-Mergib University, Al-Khums, Libya. In *Acta Biology Forum*. V02i02 (Vol. 18, p. 22).

Unnisa, S. A., Rao, B. B., & Vattikoti, P. (2022). Biochemical parameters of selected plants as air pollution indicators. *Acta Botanica Plantae*, 43-50.

Mydeen, A. K. M., Agnihotri, N., Bahadur, R., Lytand, W., Kumar, N., & Hazarika, S. (2023). Microbial Maestros: Unraveling the crucial role of microbes in shaping the Environment. In *Acta Biology Forum* (Vol. 2, pp. 23-28).

Okunlola, A. I., Opeyemi, M. A., Adepoju, A. O., & Adekunle, V. A. J. (2016). Estimation of carbon stock of trees in urban parking lots of the Federal University OF Technology, Akure, Nigeria (Futa). *Plant Science Archives*

Touseef, M. (2023). Exploring the Complex underground social networks between Plants and Mycorrhizal Fungi known as the Wood Wide Web. *Plant Science Archives*. V08i01, 5.

Ogori, A. F., Eke, M. O., Girgih, T. A., & Abu, J. O. (2022). Influence of Aduwa (*Balanites aegyptiaca*. del) Meal Protein Enrichment on the Proximate, Phytochemical, Functional and Sensory Properties of Ogi. *Acta Botanica Plantae*. V01i03, 22-35.

Corpuz, M. C., Balan, H. R., & Panares, N. C. (2016). Biodiversity of benthic macroinvertebrates as bioindicator of water quality in Badiangon Spring, Gingoog City. *Plant Science Archives*

George, U. U., Mbong, E. O., Bolarinwa, K. A., & Abiaobo, N. O. (2023). Ethno-botanical verification and phytochemical profile of ethanolic leaves extract of two medicinal plants (*Phragmites capitata* and *Lantana camara*) used in Nigeria using GC-MS Technique. In *Acta Biology Forum*.

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