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**Heavy Metal Scavenging Potential of Indigenous Microalgae of  
Bangladesh: A Study on its Application in Textile Effluent Treatment**

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UNDER PEER REVIEW

## 5 ABSTRACT

**Aims:** The aim of study was to identify the physicochemical property improvement and heavy metal scavenging potential of indigenous microalgae (*Spirulina sp.* and *Chlorella sp.*) of Bangladesh for treatment of textile wastewater disposed in the open environment.

**Study design:** The capacity of improving the water quality of the textile effluent by heavy metal absorption was assessed. The quantitative determination included the comparison of physical characters (pH, TDS, EC, DO, COD) and heavy metal profile (Cr, Cd, Pb, Zn, Fe) of the textile effluent before and after bioremediation. Effluent treatment was carried out by individual species separately and in combination of both for a total of 25 days.

**Place and Duration of Study:** Major experiments were carried out at the Applied Botany Laboratory, Dhaka Laboratory, BCSIR, Dhaka, Bangladesh from January 2022 to February 2024. Quantitative estimations were carried out at Soil & Water Laboratory, Dhaka Laboratory, BCSIR, Dhaka, Bangladesh.

**Methodology:** The textile wastewater was characterized by means of physicochemical parameters and heavy metal concentration prior to the experimental procedure. Three treatment plans were designed, two ( $T_{CV}$  and  $T_{SP}$ ) using individual species separately and one treatment ( $T_C$ ) using both the species in combination. The treatment continued for 25 days. The physicochemical parameters and heavy metal concentration of the treated effluent were measured at 5-day interval till the 25<sup>th</sup> day of the experiment. Comparative analysis of the data was utilized to determine useful species for further applied studies in future.

**Results:** The *Chlorella* treatment achieved remarkable pH restoration, with a peak of 7.94 on Day 10 and stabilization at 7.28 by Day 25. Moreover, the same treatment exhibited substantial reductions in TDS, dropping from 7750 mg/L to 455 mg/L by Day 10, and further to 301 mg/L by Day 25. Additionally, it effectively reduced heavy metal concentrations; Cr from 0.783 ppm to 0.462 ppm by Day 25, well below permissible limits.

**Conclusion:** The comparative data suggested the overall improvement of the effluent quality can be achieved by a combined treatment of *C. vulgaris* and *S. platensis*, quickly and cost-efficiently. Further investigation is required for an in-depth understanding of their combined potential.

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7

8 **Keywords:** *Textile effluent, Microalgae, Spirulina, Chlorella, Heavy metal scavenging,*  
9 *Wastewater treatment*

## 10 **1. INTRODUCTION**

11 The increasing global population has presented numerous challenges to the world economy,  
12 particularly regarding environmental preservation and energy security. Discharged textile  
13 wastewater pollutes rivers, lakes and seas worldwide [1]. There is growing concern about the  
14 substantial volume of effluents released from textile processing which consumes large amounts  
15 of water [2]. Untreated effluent from nearby textile factories has been discharged into rivers with  
16 major contaminants located outside the area such as in Narayangonj, Savar and Chattogram in  
17 some industrial areas. The annual global production of dyestuff exceeds 700,000 tonnes [3].The  
18 textile industry uses large amounts of water for various stages of dyeing and cleaning raw  
19 materials [4]. As a result, the wastewater from textile production contains harmful heavy metals  
20 such as cadmium (Cd), chromium (Cr) and lead (Pb). These toxic metals threaten living  
21 organisms, including humans, due to their biotoxic effects which can be acute, chronic, sub-  
22 chronic, neurotoxic, carcinogenic, mutagenic or teratogenic [5,41,42,43]. For example, even low  
23 levels of cadmium can be extremely toxic, leading to bone defects, increased blood pressure,  
24 myocardial dysfunctions, pulmonary oedema and, in severe cases, death [6]. Studies have shown  
25 that lead is the most significant of the toxic heavy metals, being absorbed in inorganic forms  
26 through ingestion of food and water as well as inhalation [7]. Lead poisoning can cause  
27 inhibition of haemoglobin synthesis, kidney and joint dysfunctions, reproductive system issues,  
28 and acute and chronic damage to the central nervous system [8]. Textile wastewater is a major  
29 source of surface water contamination and various technologies are being developed for treating  
30 these effluents. Among these technologies, adsorption is considered one of the most promising

31 methods [9, 10]. Recent studies have focused on the adsorptive removal of heavy metals and  
32 dyes using chitosan-based materials. [11, 12].

33 Four common ways to treat wastewater include physical water treatment, biological water  
34 treatment, chemical treatment and sludge treatment. Agents used for bioremediation are bacteria,  
35 fungi and algae [13]. Microalgae are reported in many studies to alleviate heavy metal toxicity. In  
36 recent years, the use of microalgae and cyanobacteria in the bioremediation of coloured  
37 wastewater has attracted interest due to their central role in carbon dioxide fixation. In addition,  
38 the generated algae biomass has potential as feedstock for biofuel production. Algal ability to  
39 remove dyes from wastewater can be enhanced by stimulating their growth. Living biomass of  
40 microalgae such as *Chlorella* sp. can remove 63.0 – 69.0 % of the colour from the mono-azo dye  
41 tectilon yellow 2G by converting it to aniline [14]. Microalgae such as *Chlorella* sp. and  
42 *Spirulina* sp. grown respectively in CH medium and Zarrouk's medium are demonstrated to be  
43 useful in treating effluent textile wastewater [15]. Harvesting is the crucial step in the production  
44 of algal biomass, as it accounts for 20.0 – 30.0% of production costs. The small size of  
45 microalgae (3.0 – 30.0  $\mu\text{m}$ ) and its low concentration in the culture medium (below 500.0 mg/L)  
46 make cell recovery a very challenging process. Several species of algae with varying  
47 characteristics like shape, size and motility influence their settling.

48 *Chlorella* is a microscopic green alga, spherical or ellipsoidal not much larger than a red blood  
49 cell. The name of this single-celled, non-motile water plant comes from the Greek chloros=green  
50 or yellow-green and ella=small. The cells are usually 2.0 – 12.0  $\mu\text{m}$  in diameter but the size can  
51 vary, even within a single population. The cells are solitary or in irregular clumps. *Chlorella* has  
52 a high growth rate, making it very interesting for research in various fields [15]. There are  
53 various areas where *Chlorella* is used, such as to remove dyes by bio-adsorption, biodegradation

54 and bioconversion. *Chlorella* sp. can degrade dyes by removing nitrogen, phosphorus and carbon  
55 from water [16].

56 *Spirulina* sp. is another organism whose role as an effective material to scavenge metal ions is  
57 exclusively examined [17, 18]. Many studies have aimed to analyze the tolerance and absorption  
58 mechanisms of toxic metals such as Cr, Cd and copper (Cu) in *Spirulina* sp. [19]. It is also  
59 characterized by a higher capacity to remove effluents from textile wastewater [20]. Textile  
60 effluents are the causes that reduce the nutrients of water bodies. It is well known for creating  
61 biofilms on the water's surface so that the lack of sunlight causes aquatic life to suffer [21].  
62 Many studies examined the impact[26-40] of various dyes on water, concluding that higher dye  
63 concentrations inhibit the growth of *Spirulina* sp. and reduce its nutrient levels [22].

64 Bioremediation has become the primary choice for contaminated site recovery in America. It is  
65 commonly used globally for situations where previous human activity has left the location  
66 damaged and unusable without remediation [23]. With the country's population rising, the  
67 demand for landfills to relocate polluted material surpasses the available supply [24]. On the  
68 other hand, biological treatment could achieve greater efficiencies in the decolourization and  
69 detoxification of textile wastewater by using native aquatic plants [25]. Using microorganisms to  
70 break down pollutants or waste, such as oil spills, contaminated groundwater or industrial  
71 processes makes bioremediation a very attractive solution [26].

72 This study focused on the growth parameter optimization of *Spirulina* sp. and *Chlorella* sp.  
73 biomass production as one of the key factors influencing heavy metal removal from industrial  
74 wastewater. There were few studies of heavy metal scavenging in the past, under a consortium  
75 condition of *Spirulina* sp. and *Chlorella* sp. We addressed fundamental physicochemical changes

76 of textile effluent under consortium conditions, including its efficiency in scavenging Cr, Cd, Pb,  
77 zinc (Zn), and iron (Fe).

## 78 2. MATERIALS &METHODS

### 79 2.1. Textile effluent collection

80 Textile wastewater was collected from the textile mills in Narayangonj, Bangladesh. The liquid  
81 is collected successively in four 5.0 L gallons that have retention times of 2.0 to 3.0 days  
82 depending on water use, weather and land application practices. The gallons were maintained in  
83 anaerobic conditions and mixed vertically. The samples were not stratified concerning pH,  
84 temperature or electrical conductivity. The water also had a dark-brown colour (Fig. 1B).

85



86

87 **Fig. 1. Textile wastewater samples (A) Collection area (B) Collected wastewater.**

88

### 89 2.2. Preservation of the sample

90 Wastewater preservation techniques were used to prevent retardation of the chemicals and  
91 biological changes that usually continues after the samples have been collected. No preservatives  
92 were used during the transportation of the samples to the laboratory. Around 20.0 L effluent

93 samples were collected (in fresh plastic gallons). 5.0 mL of conc. HNO<sub>3</sub> was added to each  
94 effluent sample bottle to prevent air oxidation and was preserved in a 4.0 °C refrigerator.

95

### 96 2.3. Collection of microalgae

97 The Applied Botany Laboratory, BCSIR, Dhaka Laboratories, from its specialized raceway  
98 culture ponds (Fig. 2), provided the cyanobacteria *Spirulina platensis* and *Chlorella vulgaris*.

99



100

101 **Fig. 2. *Spirulina platensis* raceway pond at BCSIR Laboratories, Dhaka**

### 102 2.4. Culture of microalgae

103 Both *Spirulina platensis* and *Chlorella vulgaris* were cultured in 100 mL culture media in 500  
104 mL Erlenmeyer flasks at 23.0 ± 1.0 °C following aseptic conditions. The cultures were gently  
105 agitated over an orbital shaker (SYC-2102) and exposed to white light for a 24.0 hours  
106 photoperiod by using a cool white fluorescent light. Cell growth was monitored by determination  
107 of optical density (OD<sub>750</sub>) at 750.0 nm. For *Spirulina* culture (Fig. 3A), Zarrouk's medium of pH  
108 9.50 (Table 1) was used and for *Chlorella* culture (Fig. 3B), CH medium was used (Table 2). The  
109 Applied Botany Laboratory, BCSIR Dhaka Laboratories, provided the reagents for media  
110 preparation.

111



112

113 Fig. 3. Flask culture of (A) *Spirulina platensis* in Zarrouk's medium and (B) *Chlorella*  
 114 *vulgaris* in CH medium

115

116 Table 1. Composition of Zarrouk's medium

Chemicals	Amount (g/L)
NaCl	1.00
NaNO <sub>3</sub>	2.50
K <sub>2</sub> SO <sub>4</sub>	1.00
NaHCO <sub>3</sub>	16.8
K <sub>2</sub> HPO <sub>4</sub>	0.50
MgSO <sub>4</sub> .7H <sub>2</sub> O	0.20
FeSO <sub>4</sub> .7H <sub>2</sub> O	0.01
CaCl <sub>2</sub> .2H <sub>2</sub> O	0.04
EDTA-Na <sub>2</sub> .2H <sub>2</sub> O	0.08
A <sub>5</sub> Micro Nutrient (H <sub>3</sub> BO <sub>3</sub> , MnCl <sub>2</sub> .4H <sub>2</sub> O, ZnSO <sub>4</sub> .4H <sub>2</sub> O, Na <sub>2</sub> MoO <sub>4</sub> , CuSO <sub>4</sub> .5H <sub>2</sub> O)	1.00 mL

117

118 **Table 2. Composition of CH medium**

Chemicals	Amount (g/L)
KNO <sub>3</sub>	6.00
K <sub>2</sub> HPO <sub>4</sub>	0.24
MgSO <sub>4</sub>	0.06
FeSO <sub>4</sub>	0.03
CaSO <sub>4</sub> .2H <sub>2</sub> O	0.012

119

120 **2.5. Experimental design for the effluent treatment**

121 The study used twelve glass beakers (1.0 – 2.0 L) representing three treatments and one control  
122 in three replicates. All aquaria were filled with 1.0 L of water from textile wastewater samples.  
123 The control treatment consisted solely of textile wastewater. Treatments were set up in a 1.0 –  
124 2.0 L glass beaker in normal daylight and temperature. Each set up of wastewater and microalgae  
125 amounts of 1.0 L wastewater with 10.0 mL of pure microalgae like *Spirulina* sp. (T<sub>SP</sub>), *Chlorella*  
126 sp. (T<sub>CV</sub>), and Combined (T<sub>C</sub>). The growth of microalgae in the five treatments was quantified in  
127 terms of cell count by using a T80 UV-visible spectrophotometer (OD<sub>600</sub>, PG Instruments,  
128 United Kingdom). No artificial shaker is used in this treatment method. Treatments were carried  
129 out for 25 days and data was collected from all samples at 5 days intervals.

130

131 **2.6. Physicochemical analysis of the samples**

132 For textile wastewater analysis, 100.0 mL samples were taken every five days from each glass  
133 aquarium and were placed in plastic bottles. These samples show some different conditions when  
134 it was treated with microalgae.

135

136 **2.6.1. pH**

137 The pH meter is calibrated using standard buffer solutions of pH 4.0, 7.0, and 9.18 at room  
138 temperature. When testing water samples, the pH is determined at room temperature. Before  
139 each sample measurement, the electrode is thoroughly washed with distilled water and cleaned  
140 with tissue paper. Approximately 100.0 mL of the sample is taken in a glass beaker. The  
141 electrode is then dipped into the sample. The instrument provides a direct measurement of pH.  
142 The pH of the filtered sample is measured using the HachSensION 4-Thermo Fisher.

143

144 **2.6.2. Salinity**

145 The salinity of water samples is determined at room temperature. The electrode is thoroughly  
146 washed with distilled water and cleaned with tissue paper before each measurement of the  
147 sample and buffer solutions. Approximately 100.0 mL of the sample is taken in a glass beaker.  
148 The electrode is then immersed in the sample. The HachSensION 4-Thermo Fisher provides a  
149 direct measurement of salinity.

150

151 **2.6.3. Total Dissolved Solid (TDS)**

152 The total dissolved solids (TDS) of water samples are determined at room temperature. The  
153 electrode is washed thoroughly with distilled water and cleaned with tissue paper before each

154 measurement of the sample and buffer solutions. About 100.0 mL of the sample is taken in a  
155 glass beaker. The electrode is then dipped in the sample, and the instrument provides a direct  
156 measurement of TDS. The filtered sample was used for measuring the TDS using the  
157 HachSensION 4-Thermo Fisher.

158

#### 159 **2.6.4. Electrical Conductivity (EC)**

160 The conductivity cell should be washed thoroughly with distilled water and cleaned with tissue  
161 paper before each measurement, whether it's for the sample or the KCl standard solution. All  
162 conductance measurements should be taken at a temperature of  $25.0 \pm 0.10$  °C. To calibrate, the  
163 conductivity cell needs to be placed in the standard KCl solution. When taking a sample  
164 measurement, 100.0 mL of the sample should be poured into a 100.0 mL glass beaker. Then, the  
165 conductivity cell should be submerged in the beaker and the EC value noted. The EC of the  
166 filtered sample should be measured using the HachSensION 4-Thermo Fisher.

167

#### 168 **2.6.5. Dissolved Oxygen (DO)**

169 The dissolved oxygen (DO) meter is calibrated using standard solutions at room temperature.  
170 Before each measurement, the electrode is washed thoroughly with distilled water and cleaned  
171 with tissue paper. Approximately 100.0 mL of the sample is taken in a glass beaker and then the  
172 electrode is dipped into the sample. The instrument provides a direct measurement of dissolved  
173 oxygen. For measuring the DO, the filtered sample was used with the HACH Instrument HQ  
174 30D meter.

175

#### 176 **2.6.6. Chemical Oxygen Demand (COD)**

177 The COD of the filtered sample is measured using the reflux digestion method. For each of the  
178 TWW samples collected, 10.0 mL of sample (Sample:Distilled H<sub>2</sub>O = 1:9) was prepared. For  
179 each reaction, 10.0 mL of the prepared sample was mixed with 5.0 mL of K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> solution (0.25  
180 N), 15.0 mL of AgSO<sub>4</sub>-H<sub>2</sub>SO<sub>4</sub> solution (10.0 mg/mL), and 0.02 g HgSO<sub>4</sub> in a digesting tube. The  
181 samples were refluxed for 2.0 hours, then cooled, and the volume was made up to 70.0 mL using  
182 distilled H<sub>2</sub>O. Next, eight (8) drops of Ferroin indicator were added to the mixture and it was  
183 titrated against standard FeSO<sub>4</sub>·(NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>·6H<sub>2</sub>O solution (0.25 N) until the blue-green colour  
184 changes to red wine. The procedure was also carried out for the blank sample.

185

#### 186 **2.7. Heavy metal concentration analysis**

187 To analyze the heavy metals Cr, Cd, Pb, Zn, and Fe, 200 mL of water sample from each setup  
188 was gently evaporated until dried. The residues were dissolved with 5.0 mL of concentrated  
189 HNO<sub>3</sub>, and 5.0 –10.0ml of H<sub>2</sub>O<sub>2</sub> were added to complete the digestion process. Then, 1.0 mL of  
190 this solution was used to determine the concentration of heavy metals using an atomic absorption  
191 spectrophotometer (PerkinElmer "AAAnalyst 700") and the program operated by the software  
192 AAwinlab Analyst-v4.1.

193 On the instrument side, the air and acetylene were manually started, and the instrument was  
194 switched on. After achieving the proper pressure of air and acetylene (75.0 and 30.0 kg/cm<sup>2</sup>,  
195 respectively), the burner was ignited. Calibration involved setting the atomization position for  
196 the required absorbance standard. Standard solutions of 5.0 and 10.0 mg/L of a particular metal  
197 were applied and a linear graph appeared. Once the calibration curve was satisfactory, the  
198 instrument was ready for sample measurement.

199 During sample measurement, the sample blank was aspirated, and distilled water was aspirated  
200 after every measurement. The concentration in mg/L was recorded directly from the screen.

### 201 3. RESULTS AND DISCUSSION

#### 202 3.1. Characteristics of the effluent before treatment

203 The textile effluent sample exhibited the following characteristics before undergoing any  
204 treatment. The pH of the effluent was 7.04, indicating a slightly alkaline nature. The Total  
205 Dissolved Solids (TDS) level was 221.7 mg/L, which is notably high and suggests a significant  
206 presence of dissolved substances that can affect water quality. The Electrical Conductivity (EC)  
207 was measured at 0.245 mS/cm, reflecting the effluent's ability to conduct electricity due to  
208 dissolved inorganic materials. Additionally, the Dissolved Oxygen (DO) content was 2.90 mg/L,  
209 a low value that indicates insufficient oxygen levels to support aerobic life forms effectively.  
210 Heavy metal analysis revealed the presence of several toxic metals in the effluent. The  
211 concentrations were as follows: Chromium (Cr) at less than 0.9 ppm, Cadmium (Cd) at less than  
212 0.8 ppm, Lead (Pb) at less than 1.06 ppm, Zinc (Zn) at less than 0.7 ppm, and Iron (Fe) at less  
213 than 0.9145 ppm. The levels of Lead, Chromium and Cadmium were particularly concerning as  
214 they exceeded acceptable limits, posing serious environmental and health risks.

215

216 **Table 3. Physicochemical Characteristics of The Effluent**

Parameters	Effluent
pH	7.04
TDS	221.7 mg/L
EC	0.245 mS/cm

DO	2.90 mg/L
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217

218 **Table 4. Heavy metal concentration of the effluent**

Heavy Metals	Effluent
Cr	<0.9 ppm
Cd	<0.8 ppm
Pb	<1.06 ppm
Zn	<0.7 ppm
Fe	<0.9145 ppm

219

220 **3.2. Physicochemical characters of the effluent after treatment**

221 The treated samples were analyzed for pH, TDS, EC, DO, and COD after treatments. These  
 222 parameters have shown significant changes after various interventions. Improvement of the  
 223 effluent after different treatments has been summarized in Table 5.

224

225 **Table 5. Analysis of changes in physicochemical parameters of the samples**  
 226 **throughout the treatments**

Parameters	Treatments	Days				
		5	10	15	20	25
pH	T <sub>CV</sub>	7.34	7.94	7.12	7.20	7.28
	T <sub>SP</sub>	7.26	7.38	5.96	6.99	7.38
	T <sub>C</sub>	7.38	7.75	7.22	7.15	7.21
TDS (mg/L)	T <sub>CV</sub>	7750	455	8760	8040	6750
	T <sub>SP</sub>	7250	8120	8320	6960	7310
	T <sub>C</sub>	7070	792	8250	7250	7200
EC (mS/cm)	T <sub>CV</sub>	15.49	0.913	17.43	16.05	13.48
	T <sub>SP</sub>	14.49	16.24	16.62	13.93	15.68

	<b>T<sub>C</sub></b>	14.12	1.786	16.51	14.23	14.43
<b>DO (mg/L)</b>	<b>T<sub>CV</sub></b>	1.36	8.79	1.30	1.37	1.41
	<b>T<sub>SP</sub></b>	1.42	1.44	1.36	1.47	1.35
	<b>T<sub>C</sub></b>	1.39	8.99	1.42	1.44	1.40
<b>COD (mg/L)</b>	<b>T<sub>CV</sub></b>	519	488	413	370	301
	<b>T<sub>SP</sub></b>	511	473	404	355	295
	<b>T<sub>C</sub></b>	490	409	390	302	269

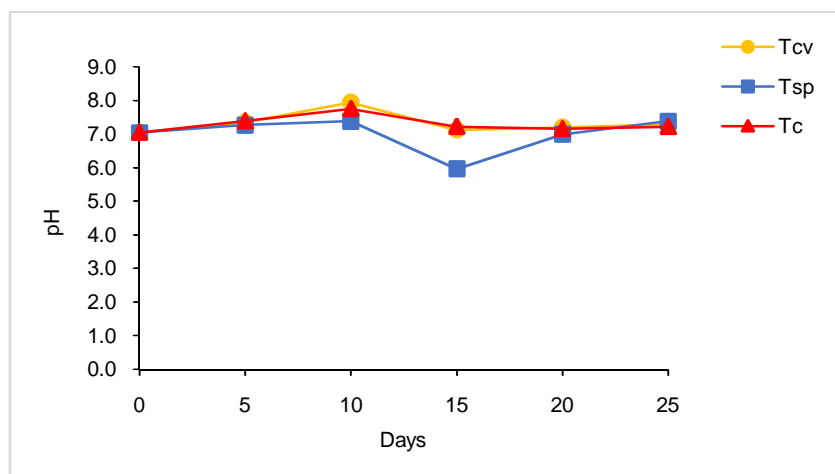
227

### 228 **3.2.1. Changes on pH level**

229 For the T<sub>CV</sub> treatment, pH started at 7.34, peaked at 7.94 on Day 10, then decreased to 7.12 on  
 230 Day 15 before stabilizing at 7.28 by Day 25. This moderate fluctuation suggests the treatment  
 231 maintained a slightly alkaline range overall. In the T<sub>SP</sub> treatment, the pH began at 7.26 and rose  
 232 to 7.38 on Day 10. However, it significantly dropped to an acidic 5.96 on Day 15, indicating a  
 233 drastic change likely due to the treatment process. By Day 20, the pH recovered to 6.99 and  
 234 stabilized at 7.38 by Day 25, returning to a neutral state. The T<sub>C</sub> treatment showed less  
 235 variability, with pH starting at 7.38, increasing to 7.75 on Day 10, then gradually decreasing to  
 236 7.15 by Day 20, and stabilizing around 7.21 by Day 25, as observed in Fig. 4.

237 The T<sub>CV</sub> and T<sub>C</sub> treatments maintained pH The return to near-neutral pH levels by Day 25 across  
 238 all treatments indicates the effluent's buffering capacity, which helps mitigate sudden pH  
 239 changes and reduces environmental risks.

240



241

242 **Fig. 4. Trend of change in pH level in different treatments**

243

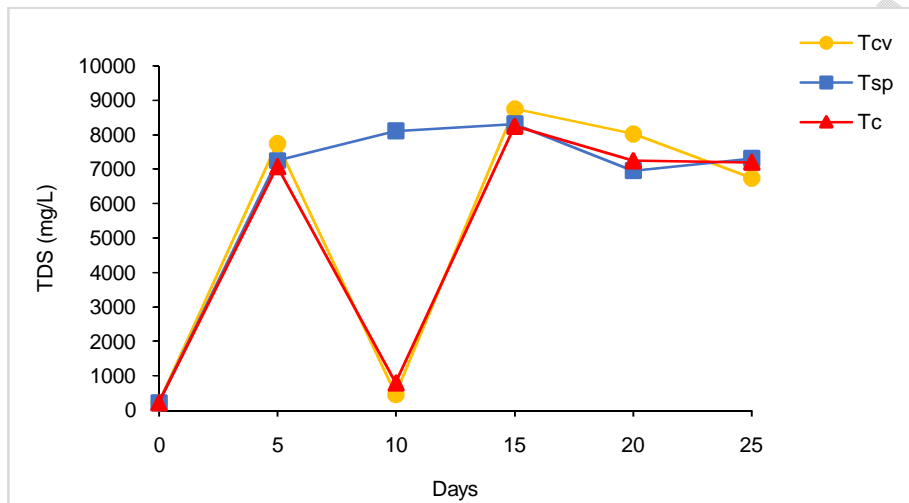
244 **3.2.2. Changes on total dissolved solids (TDS)**

245 For the T<sub>CV</sub> treatment, TDS started at 7750 mg/L, dropped significantly to 455 mg/L by Day 10,  
 246 spiked to 8760 mg/L on Day 15, then decreased to 8040 mg/L on Day 20, and finally reduced to  
 247 6750 mg/L by Day 25. This indicates substantial fluctuations throughout the treatment period. In  
 248 the T<sub>SP</sub> treatment, TDS started at 7250 mg/L, increased to 8120 mg/L on Day 10, and remained  
 249 high, reaching 8320 mg/L on Day 15. It then decreased to 6960 mg/L by Day 20 and slightly  
 250 increased to 7310 mg/L by Day 25. This treatment showed a generally high TDS level with some  
 251 fluctuations. For the T<sub>C</sub> treatment, TDS began at 7070 mg/L, dropped to 792 mg/L on Day 10,  
 252 increased again to 8250 mg/L by Day 15, then decreased to 7250 mg/L on Day 20, and stabilized  
 253 at 7200 mg/L by Day 25, as observed in Fig. 5.

254 T<sub>CV</sub> saw a drop to 455 mg/L by Day 10 followed by a spike to 8760 mg/L on Day 15. T<sub>C</sub> showed  
 255 similar trends, with TDS dropping to 792 mg/L by Day 10 and spiking to 8250 mg/L on Day 15.  
 256 The T<sub>SP</sub> treatment maintained consistently high TDS levels throughout the period, suggesting it  
 257 was less effective in reducing dissolved solids compared to the other treatments. Despite the

258 fluctuations, TDS levels in all treatments showed a trend towards stabilization by Day 25, which  
 259 indicates that the treatment processes were starting to achieve a more balanced state. The  
 260 substantial fluctuations in treatments highlight the need for improved control measures to ensure  
 261 a more consistent reduction in TDS levels.

262



263

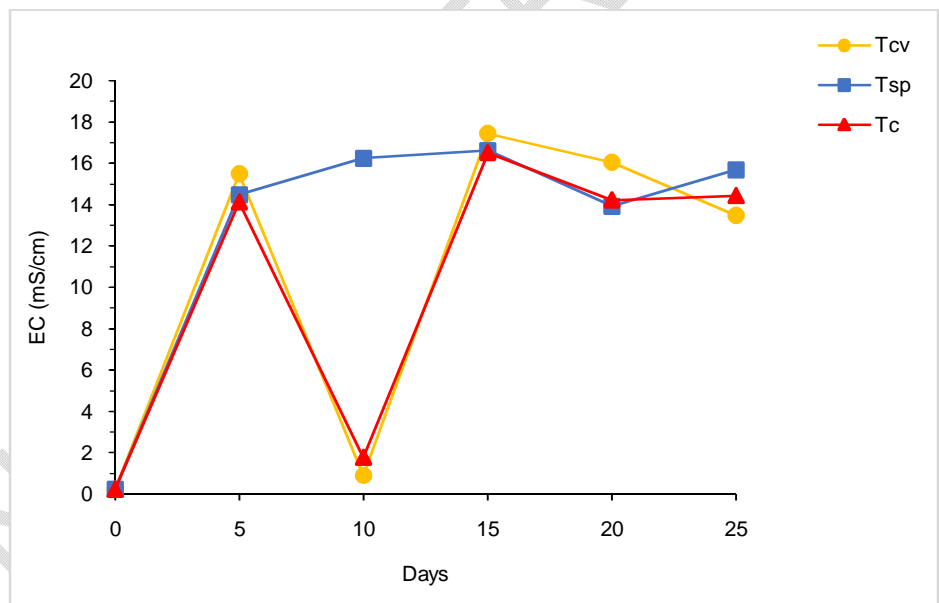
264 **Fig. 5. Trend of change in total dissolved solids in different treatments**

265

266 **3.2.3. Changes on electrical conductivity (EC)**

267 For the T<sub>CV</sub> treatment, EC started at 15.49 mS/cm, dropped significantly to 0.913 mS/cm by Day  
 268 10, spiked to 17.43 mS/cm on Day 15, decreased to 16.05 mS/cm by Day 20, and further reduced  
 269 to 13.48 mS/cm by Day 25. This indicates substantial fluctuations in EC throughout the  
 270 treatment period. In the T<sub>SP</sub> treatment, EC began at 14.49 mS/cm, increased to 16.24 mS/cm by  
 271 Day 10, remained high, reaching 16.62 mS/cm on Day 15, then decreased to 13.93 mS/cm by  
 272 Day 20, and slightly increased to 15.68 mS/cm by Day 25. This treatment maintained generally  
 273 high EC levels with some fluctuations. For the T<sub>C</sub> treatment, EC started at 14.12 mS/cm, dropped  
 274 to 1.786 mS/cm by Day 10, increased again to 16.51 mS/cm by Day 15, then decreased to 14.23

275 mS/cm by Day 20, and stabilized at 14.43 mS/cm by Day 25. This treatment exhibited significant  
276 fluctuations but showed some stabilization towards the end, as observed in Fig. 6.  
277 The T<sub>SP</sub> treatment maintained consistently high EC levels throughout the period, suggesting it  
278 was less effective in reducing dissolved ionic substances compared to the other treatments.  
279 Despite the fluctuations, EC levels in all treatments showed a trend towards stabilization by Day  
280 25, indicating that the treatment processes were starting to achieve a more balanced state. High  
281 EC levels reflect the presence of dissolved salts and inorganic materials in the effluent, which  
282 can affect water quality and treatment processes. The substantial fluctuations in treatments  
283 highlight the need for improved control measures to ensure a more consistent reduction in EC  
284 levels.



286  
287 **Fig. 6. Trend of change in electrical conductivity in different treatments**

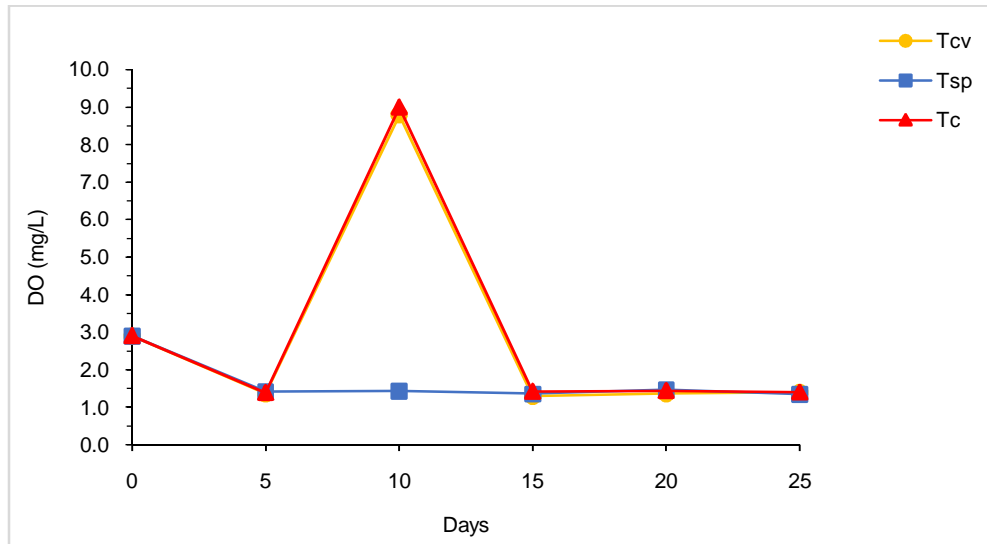
288

289 **3.2.4. Changes on the level of dissolved oxygen (DO)**

290 For the T<sub>CV</sub> treatment, DO started at 1.36 mg/L, increased significantly to 8.79 mg/L by Day 10,  
291 dropped to 1.30 mg/L on Day 15, then slightly increased to 1.37 mg/L by Day 20, and further  
292 stabilized at 1.41 mg/L by Day 25. This indicates substantial fluctuations in DO throughout the  
293 treatment period. In the T<sub>SP</sub> treatment, DO began at 1.42 mg/L, increased marginally to 1.44  
294 mg/L by Day 10, slightly decreased to 1.36 mg/L on Day 15, rose to 1.47 mg/L by Day 20, and  
295 finally dropped slightly to 1.35 mg/L by Day 25. This treatment maintained relatively stable but  
296 low DO levels. For the T<sub>C</sub> treatment, DO started at 1.39 mg/L, increased significantly to 8.99  
297 mg/L by Day 10, dropped to 1.42 mg/L by Day 15, then slightly increased to 1.44 mg/L by Day  
298 20, and stabilized at 1.40 mg/L by Day 25. This treatment also exhibited significant fluctuations  
299 but showed some stabilization towards the end, as observed in Fig. 7.

300 T<sub>CV</sub> saw a spike to 8.79 mg/L by Day 10, followed by a drop to 1.30 mg/L on Day 15. T<sub>C</sub>  
301 showed similar trends, with DO spiking to 8.99 mg/L by Day 10 and dropping to 1.42 mg/L by  
302 Day 15. The T<sub>SP</sub> treatment maintained relatively stable but low DO levels throughout the period,  
303 suggesting it was less effective in increasing oxygen levels in the effluent. Despite the  
304 fluctuations, DO levels in all treatments showed a trend towards stabilization by Day 25,  
305 indicating that the treatment processes were starting to achieve a more balanced state.

306



307

308 **Fig. 7. Trend of change in dissolved O<sub>2</sub> level in different treatments**

309

310 **3.2.5. Changes on the level of chemical oxygen demand (COD)**

311 For the T<sub>CV</sub> treatment, COD started at 519 mg/L, decreased to 488 mg/L by Day 10, continued to  
 312 drop to 413 mg/L on Day 15, further reduced to 370 mg/L by Day 20, and finally lowered to 301  
 313 mg/L by Day 25. This indicates a consistent reduction in COD throughout the treatment period.

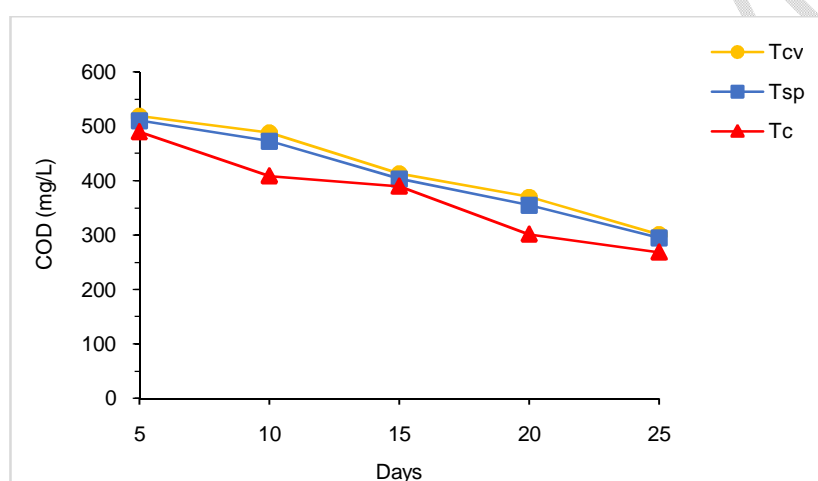
314 In the T<sub>SP</sub> treatment, COD began at 511 mg/L, decreased to 473 mg/L by Day 10, dropped  
 315 further to 404 mg/L on Day 15, continued to decrease to 355 mg/L by Day 20, and finally  
 316 reduced to 295 mg/L by Day 25. This treatment showed a steady decline in COD levels over

317 time. For the T<sub>C</sub> treatment, COD started at 490 mg/L, decreased to 409 mg/L by Day 10,  
 318 continued to drop to 390 mg/L on Day 15, further reduced to 302 mg/L by Day 20, and stabilized  
 319 at 269 mg/L by Day 25. This treatment exhibited a significant and consistent reduction in COD

320 levels, as observed in Fig. 8.

321 All treatments showed a consistent and significant reduction in COD levels over the 25-day  
 322 period. T<sub>CV</sub> saw a decrease from 519 mg/L to 301 mg/L, T<sub>SP</sub> from 511 mg/L to 295 mg/L, and T<sub>C</sub>  
 323 from 490 mg/L to 269 mg/L. This indicates that the treatments were effective in reducing the

324 organic load in the effluent. The steady decline in COD levels suggests that the treatment  
 325 processes were efficient in breaking down organic compounds and reducing the effluent's overall  
 326 oxygen demand. By Day 25, all treatments showed stabilization of COD levels, indicating that  
 327 the treatment processes had reached a more balanced and effective state. The consistent  
 328 reduction and stabilization of COD across all treatments highlight the effectiveness of the  
 329 treatment processes in improving effluent quality, making it safer for discharge into the  
 330 environment.



331  
 332 **Fig. 8. Trend of change in chemical O<sub>2</sub> demand in different treatments**

333  
 334 **3.3. Heavy metal concentration of the effluent after treatment**

335 Heavy metals were analyzed in both treated and untreated effluent by acid-digesting all the  
 336 samples. Collected textile wastewater carries Cr, Cd, Pb, Zn, and Fe. Changes in heavy metal  
 337 quantities after treatments are summarized in Table 6.

338  
 339 **Table 6. Analysis of heavy metal conc. of the effluent throughout the treatment**

Heavy metals (ppm)	Treatments	Days				
		5	10	15	20	25

<b>Cr</b>	<b>T<sub>CV</sub></b>	0.783	0.702	0.651	0.581	0.462
	<b>T<sub>SP</sub></b>	0.774	0.701	0.677	0.602	0.584
	<b>T<sub>C</sub></b>	0.874	0.705	0.633	0.596	0.501
<b>Cd</b>	<b>T<sub>CV</sub></b>	0.721	0.688	0.601	0.552	0.473
	<b>T<sub>SP</sub></b>	0.722	0.702	0.652	0.549	0.533
	<b>T<sub>C</sub></b>	0.779	0.701	0.654	0.593	0.506
<b>Pb</b>	<b>T<sub>CV</sub></b>	0.891	0.782	0.721	0.674	0.553
	<b>T<sub>SP</sub></b>	0.904	0.851	0.750	0.679	0.605
	<b>T<sub>C</sub></b>	1.010	0.901	0.854	0.755	0.673
<b>Zn</b>	<b>T<sub>CV</sub></b>	0.552	0.501	0.441	0.391	0.301
	<b>T<sub>SP</sub></b>	0.691	0.632	0.602	0.599	0.532
	<b>T<sub>C</sub></b>	0.681	0.605	0.567	0.501	0.473
<b>Fe</b>	<b>T<sub>CV</sub></b>	0.788	0.701	0.681	0.601	0.501
	<b>T<sub>SP</sub></b>	0.879	0.805	0.776	0.707	0.632
	<b>T<sub>C</sub></b>	0.867	0.779	0.703	0.679	0.603

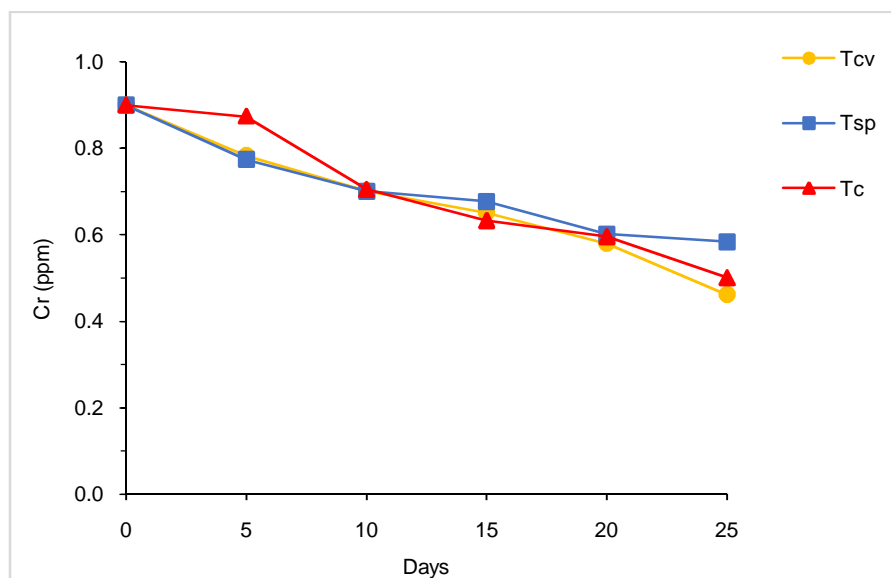
340

### 341 **3.3.1. Impact over Cr Concentration**

342 For the T<sub>CV</sub> treatment, the concentration of Cr started at 0.783 ppm, decreased gradually to 0.462  
343 ppm by Day 25. This treatment showed a consistent reduction in Cr concentration over the  
344 treatment period. In the T<sub>SP</sub> treatment, the concentration of Cr started at 0.774 ppm, decreased to  
345 0.584 ppm by Day 25. Similar to T<sub>CV</sub>, this treatment exhibited a consistent decline in Cr  
346 concentration over time. For the T<sub>C</sub> treatment, the concentration of Cr started at 0.874 ppm,  
347 decreased to 0.501 ppm by Day 25. This treatment also showed a steady reduction in Cr  
348 concentration throughout the treatment period, as observed in Fig. 9.

349 All treatments showed a consistent and significant decrease in Cr concentration over the 25-day  
350 period. The steady decline in Cr concentration suggests that the treatment processes were  
351 efficient in removing chromium from the effluent. By Day 25, all treatments showed  
352 stabilization of Cr concentration, indicating that the treatment processes had reached a more  
353 balanced and effective state. The consistent reduction in Cr concentration observed across all

354 treatments demonstrates the effectiveness of the treatment processes in improving effluent  
355 quality and reducing the potential risks associated with chromium contamination.



356  
357 **Fig. 9. Differences in the Cr conc. pre and post treatments**

358

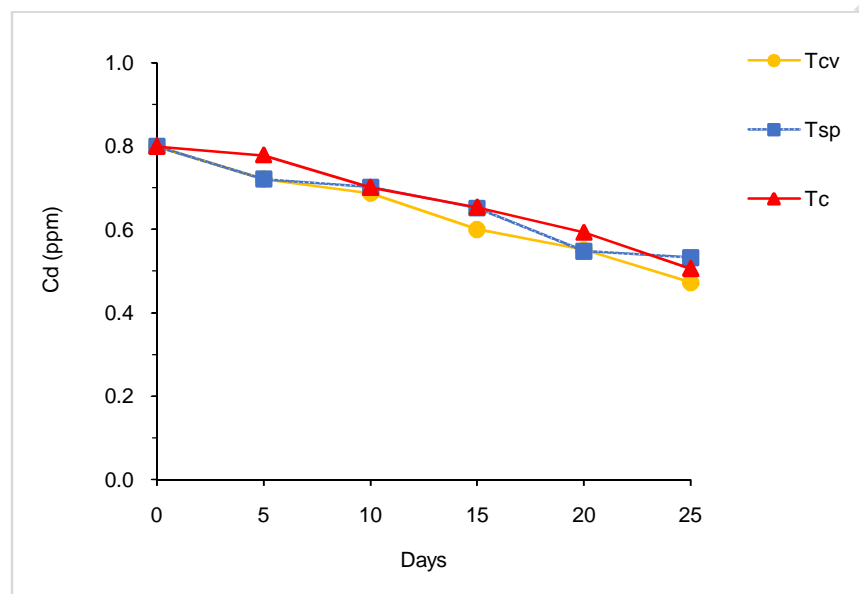
### 359 **3.3.2. Impact over Cd Concentration**

360 For the  $T_{CV}$  treatment, the concentration of Cd started at 0.721 ppm, decreasing gradually to  
361 0.473 ppm by Day 25. This treatment showed a consistent reduction in Cd concentration over the  
362 treatment period. In the  $T_{SP}$  treatment, the concentration of Cd started at 0.722 ppm, decreased to  
363 0.533 ppm by Day 25. Similar to  $T_{CV}$ , this treatment exhibited a consistent decline in Cd  
364 concentration over time. For the  $T_C$  treatment, the concentration of Cd started at 0.779 ppm,  
365 decreased to 0.506 ppm by Day 25. This treatment also showed a steady reduction in Cd  
366 concentration throughout the treatment period, as observed in Fig. 10.

367 All treatments showed a consistent and significant decrease in Cd concentration over the 25-day  
368 period. The steady decline in Cd concentration suggests that the treatment processes were  
369 efficient in removing cadmium from the effluent. By Day 25, all treatments showed stabilization

370 of Cd concentration, indicating that the treatment processes had reached a more balanced and  
371 effective state. The consistent reduction in Cd concentration observed across all treatments  
372 demonstrates the effectiveness of the treatment processes in improving effluent quality and  
373 reducing the potential risks associated with cadmium contamination.

374



375

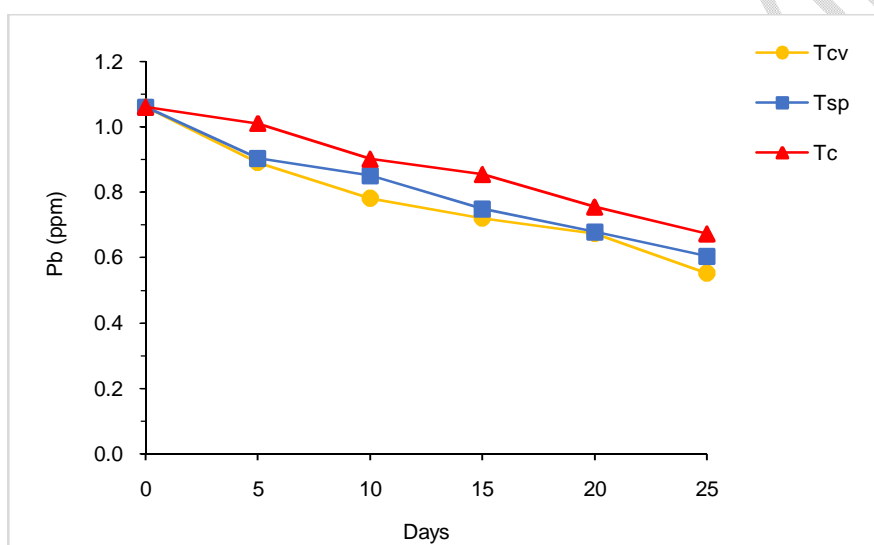
376 **Fig. 10. Differences in the Cd conc. pre and post treatments**

377

### 378 **3.3.3. Impact over Pb Concentration**

379 For the  $T_{CV}$  treatment, the concentration of Pb started at 0.891 ppm, decreased gradually to 0.553  
380 ppm by Day 25. This treatment showed a consistent reduction in Pb concentration over the  
381 treatment period. In the  $T_{SP}$  treatment, the concentration of Pb started at 0.904 ppm, decreased to  
382 0.605 ppm by Day 25. Similar to  $T_{CV}$ , this treatment exhibited a consistent decline in Pb  
383 concentration over time. For the  $T_C$  treatment, the concentration of Pb started at 1.010 ppm,  
384 decreased to 0.673 ppm by Day 25. This treatment also showed a steady reduction in Pb  
385 concentration throughout the treatment period, as observed in Fig. 11.

386 All treatments showed a consistent and significant decrease in Pb concentration over the 25-day  
387 period. The steady decline in Pb concentration suggests that the treatment processes were  
388 efficient in removing lead from the effluent. By Day 25, all treatments showed stabilization of Pb  
389 concentration, indicating that the treatment processes had reached a more balanced and effective  
390 state. The consistent reduction in Pb concentration observed across all treatments demonstrates  
391 the effectiveness of the treatment processes in improving effluent quality and reducing the  
392 potential risks associated with lead contamination.



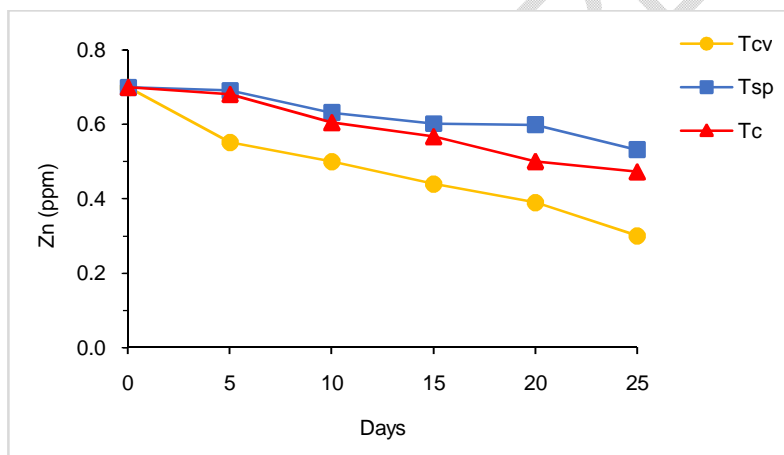
393  
394 **Fig. 11. Differences in the Pb conc. pre and post treatments**

### 396 **3.3.4. Impact over Zn Concentration**

397 For the T<sub>CV</sub> treatment, the concentration of Zn started at 0.552 ppm, decreasing gradually to  
398 0.301 ppm by Day 25. This treatment showed a consistent reduction in Zn concentration over the  
399 treatment period. In the T<sub>SP</sub> treatment, the concentration of Zn started at 0.691 ppm, decreased to  
400 0.532 ppm by Day 25. Similar to T<sub>CV</sub>, this treatment exhibited a consistent decline in Zn  
401 concentration over time. For the T<sub>C</sub> treatment, the concentration of Zn started at 0.681 ppm,

402 decreased to 0.473 ppm by Day 25. This treatment also showed a steady reduction in Zn  
 403 concentration throughout the treatment period, as observed in Fig. 12.

404 All treatments showed a consistent and significant decrease in Zn concentration over the 25-day  
 405 period. The steady decline in Zn concentration suggests that the treatment processes were  
 406 efficient in removing zinc from the effluent. By Day 25, all treatments showed stabilization of  
 407 Zn concentration, indicating that the treatment processes had reached a more balanced and  
 408 effective state. The consistent reduction in Zn concentration observed across all treatments  
 409 demonstrates the effectiveness of the treatment processes in improving effluent quality and  
 410 reducing the potential risks associated with zinc contamination.



411

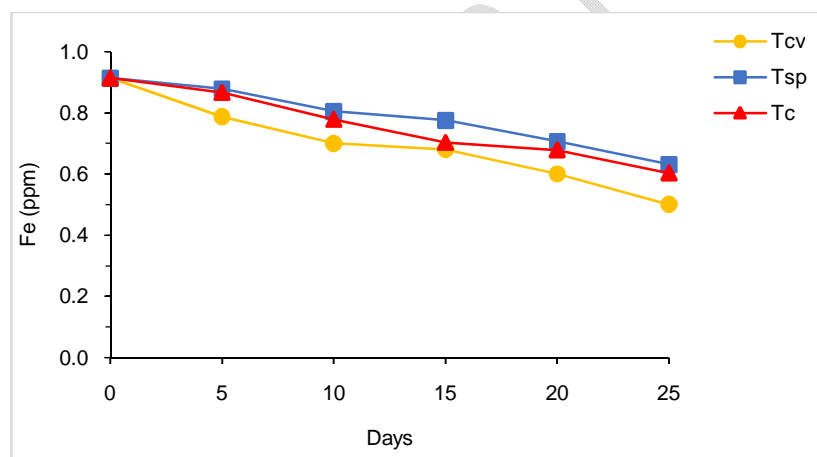
412 **Fig. 12. Differences in the Zn conc. pre and post treatments**

413

414 **3.3.5. Impact over Fe Concentration**

415 For the T<sub>CV</sub> treatment, the concentration of Fe started at 0.788 ppm, decreasing gradually to  
 416 0.501 ppm by Day 25. This treatment showed a consistent reduction in Fe concentration over the  
 417 treatment period. In the T<sub>SP</sub> treatment, the concentration of Fe started at 0.879 ppm, decreased to  
 418 0.632 ppm by Day 25. Similar to T<sub>CV</sub>, this treatment exhibited a consistent decline in Fe  
 419 concentration over time. For the T<sub>C</sub> treatment, the concentration of Fe started at 0.867 ppm,

420 decreased to 0.603 ppm by Day 25. This treatment also showed a steady reduction in Fe  
421 concentration throughout the treatment period, as observed in Fig. 13.  
422 All treatments showed a consistent and significant decrease in Fe concentration over the 25-day  
423 period. The steady decline in Fe concentration suggests that the treatment processes were  
424 efficient in removing iron from the effluent. By Day 25, all treatments showed stabilization of Fe  
425 concentration, indicating that the treatment processes had reached a more balanced and effective  
426 state. Excessive concentrations can lead to promoting the growth of iron-oxidizing bacteria,  
427 which can disrupt aquatic habitats. The consistent reduction in Fe concentration observed across  
428 all treatments demonstrates the effectiveness of the treatment processes in improving effluent  
429 quality.



430  
431 **Fig. 13. Differences in the Fe conc. pre and post treatments**

#### 432 **4. CONCLUSION**

433 Our study aimed to improve textile wastewater effluent by optimizing its physicochemical  
434 properties and reducing heavy metal concentrations through biological treatments using  
435 *Spirulina* sp. and *Chlorella* sp. treatments. The results demonstrated significant improvements in  
436 effluent quality across all treatments over the 25-day period, which effectively restored the

437 effluent's pH to near-neutral levels, mitigating potential environmental risks associated with  
438 acidic or alkaline conditions. Additionally, substantial reductions in TDS, EC, and COD were  
439 observed, indicating the treatments' efficiency in removing dissolved contaminants and organic  
440 matter from the effluent. Moreover, the treatments successfully lowered the concentrations of  
441 toxic heavy metals such as chromium (Cr), cadmium (Cd), lead (Pb), zinc (Zn), and iron (Fe) to  
442 levels below permissible limits, thereby reducing the environmental and health risks associated  
443 with heavy metal contamination.

444 Overall, the results suggest that biological treatments using *Spirulina platensis* and *Chlorella*  
445 *vulgaris* treatments have significant potential for improving textile wastewater effluent quality  
446 and reducing heavy metal pollution. Future research could focus on optimizing treatment  
447 conditions, exploring the mechanisms underlying the remediation processes, and assessing the  
448 long-term environmental impacts of these treatments. Additionally, further investigations into the  
449 potential use of other microalgae species and combinations of treatments could provide valuable  
450 insights into enhancing effluent treatment efficiency and sustainability.

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457 technical support.

458 **COMPETING INTERESTS**

459 The authors declare that they have no known competing interests (or) personal relationships that  
460 could have appeared to influence the work reported in this manuscript.

461 **AUTHORS' CONTRIBUTIONS**

462 NNH: Conceptualization, Methodology, Data curation, Writing- Original Draft, Writing- Review  
463 & Editing, Supervision, Funding Acquisition; MAA: Investigation, Software, ASB: Data  
464 curation, Writing- Review & Editing, Visualization; EAZ: Methodology, Resources; CKR:  
465 Writing- Review & Editing; MKH: Visualization, Formal Analysis and JLM: Supervision,  
466 Project Administration, Funding Acquisition.

467 **DATA AVAILABILITY**

468 All data created for this research are provided within the article/supplementary material; further  
469 enquiries can be directed to the corresponding author(s).

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