

Potential role of bio-stimulants in administering floral characters of Asiatic lily cv.

Indian Summerset

ABSTRACT

An experiment on “Potential role of bio-stimulants in administering floral characters of Asiatic lily cv. Indian Summerset” was conducted at Horticulture Research Farm, Department of Horticulture, Sam Higginbottom University of Agriculture, Technology And Sciences, Prayagraj during 2021-22 and 2022-23. The experiment consisted of thirteen treatments viz T₁-control, T₂- 10 ppm triacontanol, T₃-15 ppm triacontanol, T₄-20 ppm triacontanol, T₅- 25 ppm triacontanol, T₆- 5 ppm brassinolide, T₇-10 ppm brassinolide, T₈-15 ppm brassinolide, T₉-20 ppm brassinolide, T₁₀-100 ppm nitrobenzene, T₁₁- 200 ppm nitrobenzene, T₁₂-300 ppm nitrobenzene, T₁₃- 400 ppm nitrobenzene which were arranged in Randomized block design with three replications. Foliar application of bio-stimulants were sprayed twice at fifteen days interval. Analysis of the data indicated that T₉ -brassinolide @ 20 ppm reported to be best for parameters namely days taken for opening of first flower from bud emergence (38.6 days), pedicel length (8.2 cm), number of flowers per stalk (3.1), flower length (12.4cm), flower diameter (14.7 cm) and flower yield (7.8 flowers/plot and 33.3 flowers/m²).

Keywords: Asiatic lily, flower, bio-stimulants, triacontanol, brassinolide, nitrobenzene.

Commented [1]: Wouldn't the title be better: "The Potential of Bio-stimulants in Administering Floral Characters of Asiatic Lily cv. Indian Summerset" or "The Role ofetc"

Commented [2]: The abstract is good enough, concise, informative, clear, accompanied by keywords with a clear structure, and the language used is straightforward and easy to understand. The aim of the research has not been stated.

Commented [3]: Of of : double??

Commented [4]: What's the control? Is it just water or something like that?

Introduction

Cut flower production has indeed become a lucrative sector in horticulture, and among the various flowers cultivated, lilies stand out as one of the most significant cut flower. Belonging to the family Liliaceae, which comprises over 100 species, lilies are celebrated for their enchanting beauty, delicate fragrance, and longstanding cultural significance. With advancements in cultivation techniques and the development of new varieties, lily production has seen substantial growth. Asiatic lilies, in particular, have gained immense popularity, becoming iconic symbol in floriculture. Lilies encompass several important groups such as Asiatic, longiflorum, Oriental, Trumpet, and their hybrids, originating predominantly from Asia (Tuyl *et al.*, 1996). Their natural habitat spans various regions in the Northern Plains, including North America, Asia, and Europe, with a wide distribution ranging from the Arctic Circle to the Philippine Islands and Southern India (Klasman *et al.*, 2002). The cultivation of Asiatic lily is greatly enhanced by the strategic application of biostimulants, which play a crucial role in optimizing growth, flowering, and overall productivity of this iconic flower. Bio-stimulants are derived from natural or synthetic sources and can be administered to different plant parts. They trigger modifications in essential and structural plant processes, ultimately enhancing plant growth by bolstering resilience to environmental stress and enhancing the yield and quality of horticultural crops. Triacntanol are plant growth-promoting element that enhances the growth of plants when used exogenously at low concentration in most of the plants (Naeem *et al.*, 2012). Brassinolide has effectively helped in improving the production under stressful condition (Lalarukh *et al.*, 2022). Nitrobenzene is a compound characterized by its pale-yellow, oily appearance and almond-like fragrance. Nitrobenzene has been observed to have positive effects on flowering crops, particularly in enhancing their flowering attributes. Application of nitrobenzene has resulted in improved flower production and quality in numerous plant species.

Materials and Methods

The present investigation entitled “Potential role of bio-stimulants in administering floral characters of Asiatic lily cv. Indian summerset” was carried out during 2021-22 and 2022-23 at Horticulture Research Farm, Department of Horticulture, Naini Agricultural Institute, Sam Higginbottom University of Agriculture Technology and Sciences, Naini, Prayagraj. The experiment was laid out with thirteen treatments which were replicated thrice. The treatments were T₁- Control, T₂- 10 ppm Triacantanol, T₃-15 ppm Triacantanol, T₄-20 ppm Triacantanol, T₅- 25 ppm Triacantanol, T₆- 5 ppm Brassinolide, T₇-10 ppm Brassinolide, T₈-15 ppm Brassinolide, T₉-20 ppm Brassinolide, T₁₀-100 ppm Nitrobenzene, T₁₁- 200 ppm Nitrobenzene, T₁₂-300 ppm Nitrobenzene, T₁₃- 400 ppm Nitrobenzene. Application of bio-stimulants was done two times through foliar spray the first spray was done fifteen days after planting.

Results and discussion

Days taken for first flower opening from bud emergence

Significantly lesser number of days taken for first flower opening from bud emergence (33.2 days) which was reported in T₉ (brassinolide at the rate of 20 ppm) followed by T₈ (brassinolide at the rate of 15 ppm, 35.4 days) while T₁ (control) reported significantly more number days (41.9 days). The early onset of flowering, starting from bud emergence, can be attributed to the signalling transduction of brassinolide and the interaction of genes involved in brassinolide biosynthesis. When brassinolide is applied, it accelerates the timing of circadian rhythms, which in turn influences the flowering time of plants. The expression of CPD gene (CONSTITUTIVE PHOTOMORPHOGENESIS AND DWARFISM) is responsible for

exhibiting circadian and diurnal rhythmicity (Bancos *et al.*, 2006). The results are in conformity with the study performed by Amira *et al.*, 2022 in prickly pear and Raja *et al.*, 2020 in *Jatropha*.

Pedicle length

Longer pedicel (9.1cm) were reported in T₉ (brassinolide at the rate of 20 ppm) which was followed by T₈ (brassinolide at the rate of 15ppm, 8.6 cm) while T₁ (control) reported significantly shorter pedicel (5.8 cm). Elongation in pedicel may be attributed to the fact that brassinolide positively influence the cell division along with cell elongation, and splitting of internode, this phenomenon is called brassin activity. The distribution of brassinolide plays an important part in flower organ formation (Li and He, 2013). Brassinolide plays crucial role in regulation of reproductive plant part (Kang *et al.*, 2011). The results are in conformity with the findings of Xia *et al.* (2009) in cucumber and Prakash *et al.* (2008) in sesame.

No. of flowers per stalk

More number of flowers per stalk (4.0) were reported in T₉ (brassinolide at the rate of 20 ppm) which was followed by T₈ (brassinolide at the rate of 15 ppm, 3.5) while T₁ (control) reported significantly lesser number of flowers per stalk (2.8). More number of flowers per stalk may be attributed to the positive effect of brassinolide treated plants empowering production of more photosynthates which resulted in the presence of more assimilates to shoot apex at the time of the commencement of the floral primordial (Lakshmi *et al.*, 2021). Brassinolide is also known to positively enhance the growth of reproductive part which may have eventually lead to positive influence on formation of higher number of buds (Vogler *et al.*, 2014). Brassinolide application might have helped in increasing the number of flowers in rose (Alvarez *et al.*, 2005). Brassinolide

Commented [5]: Number of flowers per stalk, should not be shortened.

spray positively influence the number of flowers/spikes in gladiolus (Mollaei *et al.*, 2018). These results are in conformity with the findings of obtained from the study of Padmalatha *et al.* (2013) in gladiolus Aklade *et al.* (2009) in chrysanthemum and Alvarez *et al.* (2005) in rose.

Flower length

Longer flower (15.0 cm) were reported in T₉ (brassinolide at the rate of 20 ppm) which was followed by T₈ (brassinolide at the rate of 15ppm, 13.4 cm) while T₁ (control) reported significantly shorter flower (10.3 cm). The increased flower length may be due to the effect of brassinolide enhancing cell division and cell elongation in the intercalary meristem. Brassinolide enhances the vegetative development and boost photosynthetic and metabolic activities, which may have resulted in greater translocation and consumption of photosynthetic products resulting in the elongation of the petals (Mollaei *et al.*, 2018). These results are in conformity with the finding done by Babu *et al.* (2009) in jatropha.

Flower diameter

Larger flower (16.4 cm) were reported in T₉ (brassinolide at the rate of 20 ppm) which was followed by T₈ (brassinolide at the rate of 15ppm, 15.7 cm) while T₁ (control) reported significantly smaller flower (10.2 cm). The use of brassinolide led to an increase in flower diameter as brassinolide contributes to cell elongation, leading to an increase in the bloom's size. Another possibility is translocation of photosynthates leading to increase in the diameter of the flower intensifying the effect of sink. These results are in conformity with the study done by Xia *et al.* (2009) in cucumber.

No. of flower stalks /plot

Significantly more number of flower stalks /plot (9.0) were reported in T₉ (brassinolide at the rate of 20 ppm) which was found to be at par with T₄ (triacontanol at the rate of 20 ppm, 8.6), T₅ (triacontanol at the rate of 25 ppm, 8.6), T₈ (brassinolide at the rate of 15 ppm, 8.6), T₁₁ (nitrobenzene at the rate of 200 ppm, 8.6), T₁₃(nitrobenzene at the rate of 400 ppm, 8.6), T₁₂ (nitrobenzene at the rate of 300 ppm, 8.3), T₆ (brassinolide at the rate of 5 ppm, 8.3), T₇ (brassinolide at the rate of 10 ppm, 8.3) while T₁ (control, reported significantly lesser number of flower stalks /plot (7.1).

No. of flower stalks/m²

Significantly more number of flower stalks/m² (33.3) were reported in T₉ (brassinolide at the rate of 20 ppm) which was found to be at par with T₅ (triacontanol at the rate of 25 ppm, 32.6), T₁₃(nitrobenzene at the rate of 400 ppm, 32.6), T₈ (brassinolide at the rate of 15 ppm, 32.0), T₁₁ (nitrobenzene at the rate of 200 ppm, 32.0), T₇ (brassinolide at the rate of 10 ppm, 31.4), T₁₂ (nitrobenzene at the rate of 300 ppm, 31.4), T₆ (brassinolide at the rate of 5 ppm, 30.8), T₃ (triacontanol at the rate of 15 ppm, 30.2) while T₁ (control) reported significantly lesser number of flowers/m² (26.5). More number of flowers may be attributed to the positive effect of brassinolide treated plants which resulted in the presence of more assimilates to shoot apex at the time of the commencement of the floral primordia (Lakshmi *et al.*, 2021). Brassinolide is also known to positively enhance the growth of reproductive part which may have eventually lead to positive influence on formation of higher number of buds (Vogler *et al.*, 2014). Brassinolide spray positively influence the number of flowers/spikes in gladiolus (Mollaei *et al.*, 2018). These results are in conformity with result obtained from the study of Padmalatha *et al.* (2013) in gladiolus and Alvarez *et al.* (2005) in rose.

Conclusion

The results have shown that T₉ - brassinolide at the rate of 20 ppm had most significant effect in respect to days taken for opening of first flower from bud emergence, pedicel length, number of flowers per stalk, flower length, flower diameter and flower yield.

Table No. 1 Response of bio-stimulants in Days taken for opening of first flower from bud emergence and Pedicel length Asiatic lily plants

	Days taken for opening of first flower from bud emergence			Pedicel length		
	2021-22	2022-2023	Pooled data	2021-22	2022-2023	Pooled data
Treatment	42.0	41.8	41.9	6.2	5.4	5.8
T ₁ - Control	41.5	41.7	41.6	7.2	7.4	7.3
T ₂ - 10 ppm Triacantanol	39.6	42.8	41.2	7.5	7.7	7.6
T ₃ -15 ppm Triacantanol	35.7	40.5	38.1	7.6	7.9	7.7
T ₄ -20 ppm Triacantanol	39.8	36.5	38.1	8.2	8.5	8.4
T ₅ - 25 ppm Triacantanol	41.1	36.3	38.7	8.0	8.4	8.2
T ₆ - 5 ppm Brassinolide	42.3	38.0	40.1	8.1	8.4	8.2
T ₇ -10 ppm Brassinolide	34.5	36.4	35.4	8.4	8.8	8.6
T ₈ -15 ppm Brassinolide	33.7	32.7	33.2	9.1	9.4	9.2
T ₉ -20 ppm Brassinolide	36.3	39.5	38.6	7.7	8.4	8.2
T ₁₀ -100 ppm Nitrobenzene	36.7	37.8	37.8	7.8	7.9	7.9
T ₁₁ - 200 ppm Nitrobenzene	37.7	36.2	37.7	7.9	8.0	7.9
T ₁₂ -300 ppm Nitrobenzene	35.3	38.8	37.9	8.1	8.4	8.4

T ₁₃ - 400 ppm Nitrobenzene	S	S	S	S	S	S
F- test	2.797	2.564	2.139	0.498	0.454	0.311
S. Ed (±)	5.774	5.292	4.416	1.028	0.939	0.643
CD_{0.05}	9.01	8.20	6.87	7.82	6.91	4.81
CV	42.0	41.8	41.9	6.2	5.4	5.8

Table No. 2 Response of bio-stimulants on No. of flowers/stalk and flower length in Asiatic lily plants

	No. of flowers/stalk			Flower length (cm)		
	2021-22	2022-2023	Pooled data	2021-22	2022-2023	Pooled data
Treatment	2.6	2.5	2.5	10.1	10.5	10.3
T ₁ - Control	2.7	2.7	2.7	11.3	11.4	11.3
T ₂ - 10 ppm Triacantanol	3.0	2.7	2.8	11.5	11.6	11.6
T ₃ -15 ppm Triacantanol	3.0	2.7	2.9	11.9	11.7	11.8
T ₄ -20 ppm Triacantanol	3.3	3.3	3.3	12.9	13.3	13.1
T ₅ - 25 ppm Triacantanol	3.1	3.0	3.1	12.5	12.4	12.5
T ₆ - 5 ppm Brassinolide	3.3	3.1	3.2	12.6	12.5	12.6
T ₇ -10 ppm Brassinolide	3.4	3.5	3.4	13.5	13.3	13.4
T ₈ -15 ppm Brassinolide	3.8	3.7	3.8	14.9	15.1	15
T ₉ -20 ppm Brassinolide	3.1	3.1	3.1	12.3	12.5	12.4
T ₁₀ -100 ppm Nitrobenzene	3.2	3.0	3.1	13.2	12.7	12.9
T ₁₁ - 200 ppm Nitrobenzene	3.3	3.3	3.3	13.2	13.0	13.1
T ₁₂ -300 ppm Nitrobenzene	3.4	3.3	3.3	13.3	13.1	13.3
T ₁₃ - 400 ppm Nitrobenzene	S	S	S	S	S	S
F- test	0.131	0.129	0.060	0.767	0.829	0.564

S. Ed (±)	0.272	0.268	0.126	1.585	1.712	1.164
CD_{0.05}	5.09	5.18	2.39	7.55	8.10	5.53
CV	2.6	2.5	2.5	10.1	10.5	10.3

Table No. 3 Response of bio-stimulants on flower diameter and no. of flowers/plot in Asiatic lily plants

	Flower diameter (cm)			No. of flowers/plot		
	2021-22	2022-2023	Pooled data	2021-22	2022-2023	Pooled data
Treatment	10.3	11.7	10.2	7.3	7.0	7.1
T ₁ - Control	13.6	14.0	13.8	8.0	8.0	8.0
T ₂ - 10 ppm Triacontanol	14.2	14.7	14.5	8.3	8.0	8.1
T ₃ -15 ppm Triacontanol	14.3	14.8	14.6	8.3	8.6	8.5
T ₄ -20 ppm Triacontanol	15.3	15.8	15.5	8.6	8.6	8.6
T ₅ - 25 ppm Triacontanol	14.3	14.7	14.5	8.3	8.3	8.3
T ₆ - 5 ppm Brassinolide	14.3	14.8	14.6	8.6	8.3	8.5
T ₇ -10 ppm Brassinolide	15.8	15.9	15.7	8.8	8.6	8.6
T ₈ -15 ppm Brassinolide	16.1	16.7	16.4	9.0	9.0	9.0
T ₉ -20 ppm Brassinolide	15.0	14.4	14.7	8.3	7.3	7.8
T ₁₀ -100 ppm Nitrobenzene	14.6	14.7	14.6	8.6	8.6	8.6
T ₁₁ - 200 ppm Nitrobenzene	14.3	14.7	14.4	8.3	8.3	8.3
T ₁₂ -300 ppm Nitrobenzene	14.7	14.9	14.8	8.6	8.6	8.6
T ₁₃ - 400 ppm Nitrobenzene	S	S	S	S	S	S
F- test	0.752	0.920	0.679	0.408	0.496	0.400
S. Ed (±)	1.553	1.900	1.403	0.843	1.025	0.827
CD_{0.05}	6.43	7.50	5.67	5.93	7.71	6.03
CV	10.3	11.7	10.2	7.3	7.0	7.1

Table No. 4 Response of bio-stimulants on no. of flowers/m² in Asiatic lily plants

Treatment	No. of flowers/m ²		
	2021-22	2022-23	Pooled
T ₁ - Control	27.1	25.9	26.5
T ₂ - 10 ppm Triacantanol	29.6	29.6	29.6
T ₃ -15 ppm Triacantanol	30.8	29.6	30.2
T ₄ -20 ppm Triacantanol	30.8	32.0	31.4
T ₅ - 25 ppm Triacantanol	32.0	32.0	32.6
T ₆ - 5 ppm Brassinolide	30.8	30.8	30.8
T ₇ -10 ppm Brassinolide	32.0	30.8	31.4
T ₈ -15 ppm Brassinolide	32.0	32.0	32.0
T ₉ -20 ppm Brassinolide	33.3	33.3	33.3
T ₁₀ -100 ppm Nitrobenzene	30.8	27.1	28.9
T ₁₁ - 200 ppm Nitrobenzene	32.0	32.0	32.0
T ₁₂ -300 ppm Nitrobenzene	30.8	32.0	31.4
T ₁₃ - 400 ppm Nitrobenzene	32.0	33.3	32.6
F- test	S	S	S
S. Ed (±)	1.510	1.693	1.420
CD_{0.05}	3.118	3.494	2.932
CV	5.93	7.40	6.00

References

Commented [6]: Please adjust it to the template (if it is appropriate, please ignore it)

Aklade, S.A., Bardhan, K., Singh, P., Kakade, D.K. and Pathan, A.B. (2009). Effect of PGR's on growth, flowering and flower yield of chrysanthemum (*Chrysanthemum indicum*) cv. Local white. *Asian Journal of Horticulture*, **4**(2): 491-493.

Alvarez, R., Farias, Y. and Angarita, M. (2005). Effect of application of brassinosteroid (Biobras-16) on the growth and number of buds of Madame Delbard and Lidia roses. *Proceedings of the Interamerican Society for Tropical Horticulture*, **48**: 189-190.

Amira, K.G., Atteya, M., Rasha, S., Serafy, E. and Esmail, A. E. (2022). Brassinolide- Maximized the Fruit and Oil Yield, Induced the Secondary Metabolites, and Stimulated Linoleic Acid Synthesis of *Opuntia ficus-indica* Oil. *Horticulturae*, **8**: 452-455.

Aziz, M.A. and Miah, M.A. (2009). Effect of flora on the growth and yield of wetland rice. *Journal of Agricultural Research and Development*, **12**(1): 112-120.

Babu, C.R., Vanangamudi, M., Paramathma, M. and Moorthi, K.S. (2009). Effect of Growth Regulators on Flowering and Fruit set in Two *Jatropha* (*Jatropha curcas*) genotypes. *International Journal of Applied Agricultural Research*, **4**(2): 151-154.

Bancos, S., Szatma, A.M., Castle, J., Shibata, K., Yokota, T., Bishop, G.J., Nagy, F. and Szekeres, M. (2006). Diurnal regulation of the brassinosteroid-biosynthetic CPD gene in *Arabidopsis*. *Plant Physiology*, **141**: 299-309.

Jimenez, L., Zhang, Y., Yan, L., Guo, Y. and Niu, L. (2012). Phenolic compounds and antioxidant activity of bulb extracts of six liliium species native to China. *Communications in soil science & plant analysis*, **17**(8): 9361-9378.

Klasman, R., Moreira, D. and Benedetto, A. (2002). Cultivation of Asiatic hybrid of *Lilium sp.* in three different substrates. *Revistala Facultad de Agronomia Buenos Aires*, **22**(1): 79-83.

Klasman, R., Moreira, D. and Benedetto, A. (2002). Cultivation of Asiatic hybrid of *Lilium sp.* in three different substrates. *Revistala Facultad de Agronomia Buenos Aires*, **22**(1): 79-83.

Koley, P., Maitra, S. and Sarkar, I. (2019). Studies on the exogenous application of plant growth regulators on morphological and biochemical changes in gladiolus (*Gladiolus grandiflorus*) Leaf. *International Journal of Current Microbiology and Applied Sciences*, **8**(9): 1869-1877.

Lakshmi, R., Reddy, M.L., Rao, D., Bhagavan, B., Subbaramamma, P. and Krishna, K. (2021). Influence of brassinosteroids, jasmonic acid and chlorocholine chloride on yield and economics of tuberose cv. Prajwal. *Journal of Pharmacognosy and Phytochemistry*, **10**(2): 1141-1145

Lalarukh, I., Syeda, F., Amjad, N. and Mansoor, A.A. (2022). Integral effects of brassinosteroids and timber waste biochar enhance the drought tolerance capacity of wheat plant. **9**(3): 201-208.

Li, Q. and He, J. (2013). Mechanisms of signalling crosstalk between brassinosteroids and gibberellins. *Plant Signal Behaviour*, **8**: 24682 -24686.

- Mollaei, S., Farahmand, H. and Tavassolian, I. (2018).** The effects of 24-epibrassinolide corm priming and foliar spray on morphological, biochemical, and postharvest traits of sword lily. *Horticulture Environment Biotechnology*, **59**: 325 -333.
- Naeem, M. M., Khan, M. A., Moinuddin, M., Idrees, M. and Aftab, T. (2011).** Triacontanol mediated regulation of growth and other physiological attributes, active constituents and yield of *Mentha arvensis*. *Plant Growth Regulation*, **65**(1): 195-206.
- Naeem, M., Ansari, A.A., Aftab, T. and Shabbir, A. (2019).** Application of triacontanol modulates plant growth and physiological activities of *Catharanthus roseus* L. *International Journal of Botany Studies*, **4**(2): 421-426.
- Naeem, M.M., Khan, A. and Moinuddin, A.S. (2012).** Triacontanol: A potent plant growth regulator in agriculture. *Journal of Plant Interactions*, **7**(2): 129-142.
- Padmalatha, G., Reddy, S., Chandrasekhar, R., Shankar, A. and Chaturvedi, A. (2013).** Effect of foliar sprays of bioregulators on growth and flowering in gladiolus. *The Indian Journal of Agriculture Research*, **47**(3): 192-199.
- Prakash, M., Suganthi, S., Gokulakrishnan, J. and Sabesan, T. (2008).** Effect of homo brassinolide on growth, physiology and Biochemical aspects of sesame. *Karnataka Journal of Agriculture Science*, **20**:110-112.
- Raja, A.J.S. and Ezradanam, V. (2002).** Pollination ecology and fruiting behaviour in a monoecious species, *Jatropha curcas* (Euphorbiaceae). *Current Science*, **11**(2): 1395-1398.
- Tuyl, J.M.V. and Holsteijin, H.C.M. (1996).** Lily breeding research in the Netherlands. *Acta Horticulturae*, **41**(4): 35- 45.

Vogler, F., Schmalzl, C., Enghart, M. and Bircheneder, M. (2014). Brassinosteroids promote Arabidopsis pollen germination and growth. *Plant Reproduction*, **27**(3): 111-115.

Xia, X.J., Zhang, Y., Wu, J.X, Wang, J.T., Zhou, Y.H., Shi, K, Y.L. and Yu, J.Q. (2009) Brassinosteroids promote metabolism of pesticides in cucumber. *Journal of Agriculture and Food Chemistry*, **57**:8406-8413.

UNDER PEER REVIEW