

# Effect of Plant Growth Regulators on Growth and Flower Yield of *Lisianthus* (*Eustoma grandiflorum*) Under Prayagraj Agro-Climatic Conditions

## ABSTRACT

An experiment was carried out under naturally ventilated polyhouse conditions, Department of Horticulture, SHUATS, Prayagraj, during November, 2023 to April, 2024. The experiment was laid out in Completely Randomized Design (CRD) with three replications. There were ten treatments comprising of CCC, GA<sub>3</sub>, NAA at different concentrations viz., GA<sub>3</sub> (100, 200, 300) ppm and CCC (250, 500, 750) ppm and NAA (30, 45, 60) ppm along with control. The results revealed foliar application of 300 ppm GA<sub>3</sub> for *Lisianthus*, significantly enhanced the treatment T<sub>3</sub> - GA<sub>3</sub> @300 ppm was found in superior among other treatments in terms of Plant height (20.66cm), number of leaves (33.06), plant spread (11.3cm), stem diameter (3.23cm), leaf area(15.43cm<sup>2</sup>), bud length (2.62cm), number of buds per plant (6.43), stalk length(5.70cm), Number of flowers per plant(6.43), number of branches per plant (3.12),chlorophyll content (66.10), minimum number of days for bud initiation(122.50), Flower yield per plant(6.43), flower yield per 250m<sup>2</sup> (40187.50), benefit cost ratio (4.08) per 250m<sup>2</sup>was also observed in T<sub>3</sub> - GA<sub>3</sub> @300 ppm.

*Keywords:* *Lisianthus*, NAA, GA<sub>3</sub>, CCC, Polyhouse.

## 1. INTRODUCTION

*Eustoma grandiflorum* Shinn. Syn. *Lisianthus russelianus* Hook., (2n=36) belongs to family Gentianaceae. *Lisianthus* is also known as 'Texas Blue Bell' and Prairie Gentian (Halevy and Kofranek, [1]). *Eustoma* is named after the two Greek words Eu (beautiful, good, well), and stoma (mouth). Although delicate and soft-hued in style, *Lisianthus* flowers boast powerful symbolism. This floral species represents charisma, appreciation, and gratitude. It mainly used for beautification, bouquets, showy purposes and decorating walls, fences, gates, hedges. It is not only popular as a cut flower but also a bedding plant and a pot plant in the market (Grueber [2] and Harbaugh [3]). *Lisianthus* (*Eustoma grandiflorum* [Raf.] Shinn) is an ornamental species that has acquired great importance due to the attractiveness of its flowers, bright colours and long vase life. As cut flower, its acceptance in the international markets has increased in recent years and currently it is among the 10 most sold flowers in the world. It is a plant native to the northern states of Mexico and southern United States of America

(Domínguez-Ramírez [4]. Lisianthus is an annual or biannual species; the period from sowing to flowering can be divided into two stages. The first one is from seed germination up to the liners are ready for transplant; in this stage, the plants form a rosette with four leaf pairs. The second stage begins with the elongation of the stems and ends with flowering. Lisianthus has been ranked one of the lucrative and expensive cut flowers in the world. It has a great cosmopolitan demand mainly for its large and attractive flowers, long and hard stem, wide range of colors specially have purple, blue, lavender and has a long vase life which are the common traits of an ideal and best quality cut flower. Lisianthus, herbaceous annual growing to 15 to 60 cm tall, with bluish green, slightly succulent leaves. These are having large funnel-shaped flowers growing on long straight stems, sometimes erect single stems, other times growing on branching stems that can rise to be eighteen feet tall. The flowers can grow up to two inches across and can be found in a variety of colors. Lisianthus are long-stemmed flowers in cymes, with often only a few openings at a time. Sepals on Lisianthus are only fused close to the base and are much smaller than petals Namratha et al. [5]. In Tamil Nadu, production of lisianthus is concentrated in limited area in the hill stations namely, Ooty and Kodaikanal. Lisianthus is a crop which has immense potential as cut flower and pot plant (katharai et al. [6]). Research work on the use of Gibberellic acid, CCC, NAA plays a vital role in improving the vegetative growth characters of the plants as they enhance the elongation and cell division by promoting the DNA synthesis in the cell. It reduced the juvenile phase due to increase in photosynthesis and respiration with enhanced CO<sub>2</sub> fixation in the plant Thirumal et al. [7] Gibberellic acid (GA<sub>3</sub>) is endogenous plant hormone, belongs to gibberellins group and takes part in various metabolic processes (Eason [8]) GA<sub>3</sub> is mostly known to enhance the production of hydrolyses ( $\alpha$ -amylase) in the endosperm of cereals (Kolumbina et al., [9])



department of  
and Sciences,  
ie variety was

planted on polythene bags. The bags were filled with a mixture of soil, vermicompost, FYM and cocopeat in the ratio of 1:1:1:1. Before planting the seedling in the polythene bags, soil was treated with fungicide, i.e. Copper oxychloride @ 2%. During transplanting potting media was pressed firmly around the seedlings so that they do not get disturbed after irrigation. The experiment was conducted in Completely Randomized Design (CRD) with 10 treatments and three replications. Also in this experiment, plant was sprayed with Gibberellic acid (GA<sub>3</sub>) (at 100, 200, 300 ppm), cycocel (CCC) (at 250, 500, 750 ppm) and Naphthalene Acetic Acid (NAA) (at 30, 60, 90 ppm). The treatments were sprayed at 30, 60, 90, 120, 150 days of

intervals, three plants were selected randomly from each treatment per replication and their observation were recorded at 30 days intervals.

**Table 1. Treatment details**

Sl.No.	Treatment	Notation
1	T <sub>0</sub>	Control
2	T <sub>1</sub>	GA3 @100 ppm
3	T <sub>2</sub>	GA3 @200 ppm
4	T <sub>3</sub>	GA3 @300 ppm
5	T <sub>4</sub>	Cycocel @250ppm
6	T <sub>5</sub>	Cycocel @500ppm
7	T <sub>6</sub>	Cycocel @750ppm
8	T <sub>7</sub>	NAA @30 ppm
9	T <sub>8</sub>	NAA @45 ppm
10	T <sub>9</sub>	NAA @60 ppm

### 3. RESULT AND DISSCUSSION

Effect of different concentration of vegetative, flowering, yield and economic parameters are presented in table 2, 3, 4 and 5 respectively.

#### 3.1 Vegetative parameter

##### 3.1.1 Plant height (cm)

The data on vegetative parameters are given in table 2. It is clear from the table that significant difference among the treatments for plant height (cm) of lisianthus. Maximum Plant height (cm) was observed with T<sub>3</sub> - GA<sub>3</sub> @300ppm (20.66 cm) which was at par with T<sub>7</sub>- CCC @750 ppm (18.19 cm) whereas, minimum was reported in T<sub>0</sub> - Control (13.86 cm). These are similar

finding reported by Kathari [6] in lisianthus. and Aparna [10] reported GA<sub>3</sub> induce m-RNA synthesis pertaining to hydrolytic enzymes, which promotes mitotic activity in apical meristem and increase cell division and cell elongation, leading to increased length of internodes, in turn increases the plant height. Providing conducive environment inside the naturally ventilated polyhouse at night time helped in better plant height also.

### **3.1.2 Number of leaves per plant**

The data on vegetative parameters are given in table 2. It is clear from the table that significant difference among the treatments for number of leaves per plant of lisianthus. Maximum number of leaves per plant was observed with T<sub>3</sub> - GA<sub>3</sub>@300ppm (33.06) which was at par with T<sub>9</sub> - NAA @60 ppm (29.40) whereas, minimum was reported in T<sub>4</sub>- CCC @250ppm (25.0). The higher number of side shoots is also associated with higher number of leaves. Similar trend was also observed previously in chrysanthemum Habiba et al. [11] and he reported GA<sub>3</sub> influencing the increased vegetative growth by increasing cell division and cell elongation is the reason behind the production of a greater number of leaves per plant

### **3.1.3 Plant Spread (cm<sup>2</sup>)**

The data on vegetative parameters are given in table 2. It is clear from the table that significant difference among the treatments for plant spread (cm<sup>2</sup>) of lisianthus. Maximum Plant Spread (cm<sup>2</sup>) was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (11.3 cm<sup>2</sup>) which was at par with T<sub>5</sub> - CCC @500 ppm (9.54 cm<sup>2</sup>) whereas, minimum was reported in T<sub>0</sub> - Control (7.96 cm<sup>2</sup>). This might be attributed to the higher concentration of GA<sub>3</sub>, which is more efficient in cell multiplication and elongation of immature tissues, whereas lesser quantities were less favourable. Palei [13]and Doddagoudar [12] comparable results reported that the application of plant growth regulators enhances cell division with rapid internode elongation and is confined in the sub-apical meristem, which increases the plant spread.

### **3.1.4 Number of branches per plant**

The data on vegetative parameters are given in table 2. It is clear from the table that significant difference among the treatments for number of branches per plant of Lisianthus. Maximum number of branches per plant was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (3.12) which was at par with T<sub>2</sub> - GA<sub>3</sub> @200 ppm (2.35) whereas, minimum was reported in T<sub>9</sub>- NAA @60ppm (1.87). The increase in number of branches per plant application of GA<sub>3</sub> may be ascribed to increase cell division and cell enlargement, promotion of protein synthesis in the plant. Stimulation of branching may also be attributed to the breakage of apical dominance.

### **3.1.5 Chlorophyll content**

The data on vegetative parameters are given in table 2. It is clear from the table that significant difference among the treatments for chlorophyll content of Lisianthus. The maximum Chlorophyll content was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (66.10) which was at par with T<sub>7</sub>- NAA @30 ppm (64.61) whereas, minimum was reported in T<sub>1</sub>- GA<sub>3</sub> @100ppm (60.70). Changes in chlorophyll content caused by growth regulators may be attributed to decrease chlorophyll breakdown and enhance the chlorophyll synthesis.

### **3.1.6 Stem diameter (cm)**

The data on vegetative parameters are given in table 2. It is clear from the table that significant difference among the treatments for stem diameter (cm) of lisianthus. The maximum Stem diameter (cm) was observed with T<sub>3</sub> - GA<sub>3</sub> @300ppm (3.23 cm) which was at par with T<sub>8</sub>- NAA @45 ppm (2.53 cm) whereas, minimum was reported in T<sub>0</sub> - Control (1.85 cm). Increase in stem diameter is directly proportional to increase in plant height. The maximum plant height was obtained in T<sub>3</sub> which may have been led to maximum diameter of stem T<sub>3</sub>. With the increase in the concentration of GA<sub>3</sub> diameter increases due to a reflection of the stimulation of cambium and its immediate cell progeny. Similar Doddagoudar [12], reported comparable results in marigold.

### **3.1.7 Maximum Leaf area (cm<sup>2</sup>)**

The data on vegetative parameters are given in table 2. It is clear from the table that significant difference among the treatments for maximum leaf area (cm<sup>2</sup>) of lisianthus. The data on vegetative parameters are given in table 2. It is clear from the table that is significant difference among the treatments for number of leaves per plant of lisianthus. Kumar [14] reported that GA<sub>3</sub> promotes cell division and cell elongation, thereby increase the leaf length and leaf area and enhance sugar translocation.

## **3.2 Floral parameters**

### **3.2.1 Stalk length(cm)**

The data on floral parameters are given in table 3. It is clear from the table that significant difference among the treatments for stalk length (cm) of lisianthus. The maximum Stalk length (cm) was observed with T<sub>3</sub> - GA<sub>3</sub> @300ppm (5.70 cm<sup>2</sup>) which was at par with T<sub>4</sub>- Cycocel @250ppm (5.14 cm<sup>2</sup>) whereas, minimum was reported in T<sub>0</sub> - Control (3.92 cm<sup>2</sup>). GA<sub>3</sub> was the best for obtaining better growth of plants, maximum number of cut blooms with longer stalk as well as bigger flower size obtained. A similar trend was observed previously in China aster by Sailaja and Panchali [15].

### **3.2.2 Bud length (cm)**

The data on floral parameters are given in table 3. It is clear from the table that significant difference among the treatments for bud length (cm) of lisianthus. The maximum Bud length (cm) was observed with T<sub>3</sub> - GA<sub>3</sub> @300ppm (2.62 cm) which was at par with T<sub>2</sub>- GA<sub>3</sub> @200 ppm and T<sub>1</sub> GA<sub>3</sub> @100 ppm (2.45 cm) whereas, minimum was reported in T<sub>7</sub> - NAA @30 ppm (2.08 cm). Justo et al. [16] in carnation reported that the increase in the length of the flower bud in GA<sub>3</sub> treated plants is due to rapid cell elongation, increased cell divisions and cell enlargement. Foliar application of GA<sub>3</sub> also significantly increased flower bud length.

### **3.2.3 Number of buds per plant**

The data on floral parameters are given in table 3. It is clear from the table that significant difference among the treatments for number of buds per plant of lisianthus. The maximum number of buds per plant was observed with T<sub>3</sub>- GA<sub>3</sub> @300 ppm (6.43) which was at par with T<sub>8</sub> - NAA@45 ppm (5.34) whereas, minimum was reported in T<sub>4</sub>- CCC @250ppm (4.02). The effect of different growth regulators showed significant influence on the number of buds per plant. Similar results were recorded by Kumar et al. [17] in marigold.

### **3.2.4 Number of days for bud initiation**

The data on floral parameters are given in table 3. It is clear from the table that significant difference among the treatments for number of days for bud initiation lisianthus. The minimum Number of days for bud initiation was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (122.50) which was at par with T<sub>5</sub> - CCC @500 ppm (128.7) whereas, maximum was reported in T<sub>7</sub>- NAA@30ppm (131.06). Holkar [18] reported that GA<sub>3</sub> application enhances food translocation for the development of floral primordia, which leads to early flowering. This is due to increase in photosynthesis and respiration along with enhanced fixation by GA<sub>3</sub> that led to flower bud initiation.

### **3.2.5 Number of flowers per plant**

The data on floral parameters are given in table 3. It is clear from the table that significant difference among the treatments for number of flowers per plant of lisianthus. The maximum number of flowers per plant was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (6.43) which was at par with T<sub>8</sub> - NAA@45 ppm (5.34) whereas, minimum was reported in T<sub>4</sub>- CCC @250ppm (4.02). Increased number of flower is attributed to the production of large number of flower buds along with the fact that termination of vertical growth by pinching lead to more laterals/secondary branches at early stage of growth, which then had sufficient time to accumulate carbohydrates for proper flower bud differentiation producing a greater number of flowers per plant. GA<sub>3</sub> helps in breaking bud dormancy and acts as florigen initiating flowering. Similar results were also reported by Mithilesh Kumar [19] in Marigold.

### **3.3 Yield Parameters**

#### **3.3.1 Flower yield per plant**

The data on yield parameters are given in table 4. It is clear from the table that significant difference among the treatments for flower yield per plant of lisianthus. The maximum flower yield per plant was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (6.43) which was at par with T<sub>8</sub>– NAA @45 ppm (5.34) whereas, minimum was reported in T<sub>4</sub>– CCC@250ppm (4.02). Increase in flower yield per plant due to the fact that plant growth regulators stimulate vegetative growth and induced changes in vegetative morphology, and thereby accelerate growth parameters. These findings are also in accordance with those of Rakesh [20] in carnation.

#### **3.3.2 Flower yield per 250 m<sup>2</sup>**

The data on yield parameters are given in table 4. It is clear from the table that significant difference among the treatments for flower yield per 250 m<sup>2</sup> of lisianthus. The maximum flower yield per 250 m<sup>2</sup> was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (40187.50) which was at par with T<sub>8</sub> - NAA @45 ppm (33375.00) whereas, minimum was reported in T<sub>4</sub>– CCC @ 250ppm (25125.00). GA<sub>3</sub> treatment enhance induction of flower bud break i.e., differentiation of floral primordia in the apical region which leads to increase production of flowers per plants and hence increase the flower yield. These results are also in close conformity with the findings of Ryagi [21] in Carnation.

### **3.4 Economic Parameter**

The data on economic parameters are given in table 5. It is clear from the table that significant difference among the treatments of lisianthus. The maximum gross return was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (401875.0) which was at par with T<sub>8</sub> - NAA @45 ppm (3.50) whereas, minimum was reported in T<sub>4</sub>- CCC @ 250 ppm (251250.0). The maximum net return was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (303412.00) which was at par with T<sub>8</sub> - NAA @45 ppm (3.50) whereas, minimum was reported in T<sub>4</sub>- CCC @ 250 ppm (148387.00). The maximum Benefit cost ratio was observed with T<sub>3</sub> - GA<sub>3</sub> @300 ppm (4.08) which was at par with T<sub>8</sub> - NAA @45 ppm (3.50) whereas, minimum was reported in T<sub>4</sub>- CCC @ 250 ppm (2.44).

UNDER PEER REVIEW

**Table 2. Effect of different concentration of GA3, CCC and NAA on vegetative parameters of Lisianthus.**

<b>Treatment Notation</b>	<b>Treatments</b>	<b>Plant spread (cm<sup>2</sup>)</b>	<b>Plant height (cm)</b>	<b>Number of Leaves</b>	<b>Number of Branches</b>	<b>Chlorophyll content</b>	<b>Stem Diameter (cm)</b>	<b>Leaf area (cm<sup>2</sup>)</b>
<b>T<sub>0</sub></b>	Control	7.96	13.86	26.29	2.18	61.43	1.85	13.34
<b>T<sub>1</sub></b>	GA3@100 ppm	8.06	15.30	25.04	2.30	60.70	2.27	11.29
<b>T<sub>2</sub></b>	GA3@200 ppm	8.09	16.28	26.26	2.35	60.86	2.14	13.35
<b>T<sub>3</sub></b>	GA3@300 ppm	11.3	20.66	33.06	3.12	66.10	3.23	15.43
<b>T<sub>4</sub></b>	Cycocel@250ppm	8.50	15.96	25.0	2.08	62.92	2.37	12.97
<b>T<sub>5</sub></b>	Cycocel@500ppm	9.54	17.61	26.33	2.11	63.12	2.19	13.09
<b>T<sub>6</sub></b>	Cycocel@750ppm	9.38	16.81	25.20	1.90	63.54	2.51	12.20
<b>T<sub>7</sub></b>	NAA@30 ppm	9.5	18.19	27.53	1.94	64.61	2.47	13.38
<b>T<sub>8</sub></b>	NAA@45 ppm	8.99	16.98	28.04	1.92	63.68	2.53	13.60
<b>T<sub>9</sub></b>	NAA@60 ppm	9.1	16.21	29.40	1.87	63.65	2.42	13.93
	F-Test	<b>S</b>	<b>S</b>	<b>S</b>	<b>S</b>	<b>S</b>	<b>S</b>	<b>S</b>
	S.Ed (+)	0.14	0.90	0.30	0.16	0.53	0.25	0.92
	CD (5%)	0.30	0.89	0.64	0.33	1.11	0.54	1.93
	CV	1.95	6.57	1.38	8.97	1.03	13.21	8.52

**Table 3. Effect of different concentration of GA3, CCC and NAA on floral parameters of Lisianthus.**

Treatment Notation	Treatments	Stalk length(cm)	Bud length(cm)	No. of buds per plants	No. of days for bud initiation	No. of flowers per plant
T <sub>0</sub>	Control	3.92	2.13	4.49	4.49	4.49
T <sub>1</sub>	GA3@100 ppm	4.36	2.43	4.99	4.99	4.99
T <sub>2</sub>	GA3@200 ppm	3.77	2.45	5.31	5.31	5.31
T <sub>3</sub>	GA3@300 ppm	5.70	2.62	6.43	6.43	6.43
T <sub>4</sub>	Cycocel@250ppm	5.14	2.09	4.02	4.02	4.02
T <sub>5</sub>	Cycocel@500ppm	4.69	2.30	4.45	4.45	4.45
T <sub>6</sub>	Cycocel@750ppm	4.68	2.11	5.28	5.28	5.28
T <sub>7</sub>	NAA@30 ppm	4.75	2.08	5.13	5.13	5.13
T <sub>8</sub>	NAA@45 ppm	4.88	2.14	5.34	5.34	5.34
T <sub>9</sub>	NAA@60 ppm	4.93	2.11	5.19	5.19	5.19
	F-Test	S	S	S	S	S
	S.Ed (+)	0.12	0.09	0.25	0.25	0.25
	CD (5%)	0.25	0.18	0.54	0.54	0.54
	CV	3.22	4.87	6.25	6.25	6.25

**Table 4. Effect of different concentration of GA3, CCC and NAA on yield parameters of Lisianthus.**

Treatment Notation	Treatments	Yield per plant	Yield per (250 m <sup>2</sup> )
T <sub>0</sub>	Control	4.49	28062.50
T <sub>1</sub>	GA3@100 ppm	4.99	31187.50
T <sub>2</sub>	GA3@200 ppm	5.31	33187.50
T <sub>3</sub>	GA3@300 ppm	6.43	40187.50
T <sub>4</sub>	Cycocel@250ppm	4.02	25125.00
T <sub>5</sub>	Cycocel@500ppm	4.45	27812.50
T <sub>6</sub>	Cycocel@750ppm	5.28	33000.00
T <sub>7</sub>	NAA@30 ppm	5.13	32062.50
T <sub>8</sub>	NAA@45 ppm	5.34	33375.00
T <sub>9</sub>	NAA@60 ppm	5.19	32437.50
	F-Test	S	S
	S.Ed (+)	0.25	480.11
	CD (5%)	0.54	1017.78
	CV	6.25	13.05

**Table 5. Effect of different concentration of GA3, CCC and NAA on Economic parameters of Lisianthus.**

Treatment Notation	Treatments combination	Total cost of cultivation	Gross Return	Net Return	Benefit cost ratio
T <sub>0</sub>	Control	94875.00	280625.0	185750.00	2.95
T <sub>1</sub>	GA3@100 ppm	96063.00	311875.0	215812.00	3.24
T <sub>2</sub>	GA3@200 ppm	97263.00	331875.0	214612.00	3.20
T <sub>3</sub>	GA3@300 ppm	98463.00	401875.0	303412.00	4.08
T <sub>4</sub>	Cycocel@250ppm	102863.00	251250.0	148387.00	2.44
T <sub>5</sub>	Cycocel@500ppm	107752.00	278125.0	170373.00	2.58
T <sub>6</sub>	Cycocel@750ppm	113461.00	330000.0	216539.00	2.90
T <sub>7</sub>	NAA@30 ppm	95231.00	320625.0	225394.00	3.36
T <sub>8</sub>	NAA@45 ppm	95355.00	333750.0	238395.00	3.50
T <sub>9</sub>	NAA@60 ppm	95434.00	324375.0	228941.00	3.39

## CONCLUSION

From the present investigation, it is concluded that among the different treatments, the treatment T<sub>3</sub> - GA<sub>3</sub> @300 ppm was found in superior among other treatments in terms of Plant height, number of leaves , plant spread , stem diameter, leaf area, bud length, number of buds per plant, stalk length , Number of flowers per plant, number of branches per plant, chlorophyll content, minimum number of days for bud initiation, Flower yield per plant, flower yield per 250m<sup>2</sup>, benefit cost ratio per250m<sup>2</sup> was also observed in T<sub>3</sub> - GA<sub>3</sub> @300 ppm. GA<sub>3</sub> @300 ppm could be recommended for plant growth and flowering of Lisianthus (*Eustoma grandiflorum*).

## REFERENCES

1. Halevy, A.H., and Kofranek, A.M. (1984). Evaluation of lisianthus as a new flower crop. *HortScience* **19**, 845–847.
2. Grueber KL, Corr BE, Wilkins HF. (1985). Evaluation of *Eustoma grandiflorum* as a bedding plant. *Minn. State Flor. Bull*, **34**(1):16-18.
3. Harbaugh BK. (2007). Lisianthus. *Eustoma grandiflorum*. In: Anderson, N.O. (Ed.), Flower breeding and genetics. *Springer, Netherlands*, 645-663.

4. **Domínguez-Ramírez, A. (2002).** El Cultivo del Lisianthus (*Eustoma grandiflorum*). [www.uaaan.mx/postgrado/images/files/hort/simposio2/Ponencia07.pdf](http://www.uaaan.mx/postgrado/images/files/hort/simposio2/Ponencia07.pdf).
5. **Namratha G, Chandrashekar SY, Hemla Naik B, Shivaprasad M and Hanumantharaya L (2021).** Varietal evaluation of Lisianthus (*Eustoma grandiflorum* Shinn.) for morphological parameters under protected cultivation. *The Pharma Innovation Journal*, **10**(12): 2160-2162.
6. **Kathari Lakshmaiah, S Subramanian, M Ganga and P Jeyakumar (2019).** Optimization of pinching and GA3 application to improve growth and flowering of lisianthus (*Eustoma grandiflorum*). *Journal of Pharmacognosy and Phytochemistry* 2019; **8**(6): 614-616.
7. **Thirumal Murugan V, K Manivannan and S Nanthakumar (2021).** Studies on the effect of plant growth regulators on growth, flowering and xanthophyll content of African marigold (*tagetes erecta L.*). *J Pharmacogn Phytochem*, **10**(3):292-294.
8. **Eason, J.R. (2002).** Sandersonia aurantiaca: an evaluation of postharvest pulsingsolutions to maximize cut flower quality. *New Zealand J. Crop Hort. Sci.*, **30**: 273-279.
9. **Kolumbina, M., M. Blesing and D.J. Mares. (2006).**  $\alpha$ -Amylase and programmed cell death in aleurone of ripening wheat grains. *J. Exp. Bot.*, **57**(4): 877-885.
10. **Veluru Aparna, Krishna Prakash, Neema M, Arora Ajay, Naveen Kumar P, Singh MC. (2018).** Effect of gibberellic acid on plant growth and flowering of Chrysanthemum cv. Thai Chen Queen under short day planting conditions. *International Journal of Agriculture Sciences*. ISSN: 0975-3710 & E-ISSN: 0975-9107. **10**(11):6274- 6278.
11. **Habiba SU, Islam MS, Jamal Uddin AFM. (2012).** Influence of terminal bud pinching on growth and yield of chrysanthemum, (*Chrysanthemum indicum L.*). *Journal of Bangladesh Academy of Sciences*, **36**(2):251-255.
12. **Doddagoudar SR, Vyakarnahal BS, Gouda SM. (2004).** Effect of mother plant nutrition and chemical spray on seed germination and seedling vigour of China aster cv. Kamini Karnataka, *Journal of agriculture sciences*, **17**(4):701-704.
13. **Palei S, Das AK, Dash DK. (2016).** Effect of plat growth regulators on growth, flowering and yield attributes of African marigold (*Tagetes erecta L.*), *Research Paper Agriculture*. **2**(1):2454-9916.
14. **Kumar P, Singh A, Laishram N, Pandey RK, Dogra S, Jeelani MI, Sinha BK. (2019)** Effects of plant growth regulators on quality flower and seed production of marigold (*Tagetes erecta L.*)

15. **Sailaja SM, Panchbhai DM.(2014).** Effect of pinching on growth and quality characters of China aster varieties. *Asian J Hort*, **9**(1):36-39.
16. **Benny JC, Devi Singh DS, Fatmi U, Jose DA. (2018).** Effect of plant growth regulators, Gibberellic acid (GA3) and Naphthalene Acetic Acid (NAA) on growth and yield of carnation (*Dianthus caryophyllus*L.) under naturally ventilated polyhouse. *Plant Archives*, **17**. 803-812.
17. **Kumar Suresh P, Bagawathi R, Rajive Kumar, Ronya T.(2008).** Effect of plant growth regulators on vegetative growth, flowering and corm production of gladiolous in Arunachal Pradesh. *J of Orn. Horti*, **10**(2):90-95.
18. **Holkar PS, Chandrashekar SY, Hemanthkumar P, Basavalingaiah, Ganapathi M. (2018).** Effect of gibberellic acid on growth, flowering, flower quality and corm yield of gladiolus, *Trends in Biosciences*, **11**(24):32113216.
19. **Kumar, Mithilesh & Singh, AK & Kumar, Ashok. (2015).** Effect of plant growth regulators on growth and flowering characters of African marigold (*Tagetes erecta* ). *Current Advances in Agricultural Sciences*. **7**. 85. 10.5958/2394-4471.2015.00020.9.
20. **Rakesh., Singhrot, R. S. and Beniwal, B. S. (2003).** Effect of GA3 and pinching on growth and yield in Chrysanthemum. *Haryana J. hort. Sci.* **32**(1-2): 61-63.
21. **Ryagi VY, Mantur SM, Reddy B S.(2007).** Effect of pinching on growth, yield and quality of flowers of Carnation varieties grown under polyhouse. *Karnataka J Agric. Sci*, **20**(4):816-818.