

Investigating the Impact of *Senna alata* Extract on Hematology and Histopathology of Juvenile of *Clarias gariepinus*

Abstract

This paper sheds light on the mechanism of action, potential effects, and implication of *Senna alata* on fish health and ecosystem. *Senna alata* is renowned for its medicinal properties and ornamental value but is also recognized for its potential toxicity, particularly to aquatic organisms. Understanding the toxicity of *Senna alata* to *Clarias gariepinus* fingerlings is essential for sustainable aquaculture practices and environmental conservation efforts. 250 juveniles of *C. gariepinus* were procured from E. I farm Limited in Ikot Ekpene, Akwa Ibom State. Water quality tests were carried out to ensure that optimum water conditions were maintained during, and that the results obtained were solely due to the effects of the treatments (toxicants) on the fish samples. Chronic toxicity bioassay was done to determine the toxicity of the extract on the blood parameters and tissues of sensitive organs of the exposed fish. Toxicology dose response (fish mortality) was subjected to detailed analysis using Statistical Package for Social Sciences (SPSS) 22.0. Data were analyzed using descriptive statistics. The *Senna alata* exerted a toxic effect on the fish in the present study and toxicity increased with increased concentration. These findings underscore the potential adverse effects of prolonged exposure to *Senna alata* on the physiological and cellular integrity of the fish. Intriguingly, the absence of pathological changes in the control groups suggests a distinct response attributable to the treatment. This highlights the importance of continued monitoring and assessment of the impacts of *Senna alata* on aquatic organisms and ecosystem, emphasizing the need for further research to elucidate the underlying mechanisms and potential long-term consequences.

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Keywords: *Senna alata*, *Clarias gariepinus*, Toxicity, Hematology, Histopathology

1.0 Introduction

The use of piscicide in treatment of ponds prior to stocking with desired fish species is a common practice in aquaculture. Use of synthetic piscicides for this purpose is not much appreciated by fish farmers as synthetic piscicides are non-biodegradable and persistent in the environment. Hence, the use of organic piscicides is embraced by fish farmers as they are biodegradable and non-persistent in the environment and fish killed are edible. This makes research on more plants of ichthyotoxic potential important to avail the farmers more of such plants for this purpose.

Senna alata, commonly known as candle bush or candlestick senna, is a tropical shrub belonging to the fabaceae family. It is renowned for its medicinal properties and ornamental value but is also recognized for its potential toxicity, particularly to aquatic organisms (Abo *et al.*, 2008; Kumar *et al.*, 2009). *Senna alata* is characterized by its distinctive appearance, featuring bright yellow flowers arranged in cylindrical clusters and a slender stem. The plants leaves are large, compound and ovate with a glossy texture and serrated edges. These leaves are alternately arranged along the stems and emit a strong odour when crushed or bruised, a trait often attributed to the presence of chemical compounds such as anthraquinones and saponins (Kumar *et al.*, 2009).

The chemical composition of *Senna alata* includes various bioactive compounds, some of which include, phenolics (rhein, chrysoaphanol, kaempferol, aloe emodin, and glycosides), anthraquinones (alatinone and alatonal), fatty acids (oleic, palmitic, and linoleic acids), steroids, and terpenoids (sitosterol, stigmasterol, and campesterol) (Liu *et al.*, 2009, Kare, 2007). These secondary metabolites are reported to display numerous biological activities (Aviello *et al.*, 2014).

The flower, root, leaves, seed, and bark displayed diverse biological activities (Makinde *et al.*, 2007; Tarkang *et al.*, 2012). These pharmacological activities include antimicrobial (Villasenor *et al.*, 2002; Khan *et al.*, 2001 and Somchit *et al.*, 2003), antifungal (Donatus *et al.*, 2010), anticryptococcus (Ranganathan *et al.*, 2000), antibacterial (Otto *et al.*, 2014, Gaikward *et al.*, 2014), antitumor (Olater *et al.*, 2010), anti-inflammatory (Manaogaran *et al.*, 2004; Sagnai *et al.*, 2014), antidiabetic (Varghese *et al.*, 2013), antioxidant (Makinde *et al.*, 2007), wound healing (Midawa *et al.*, 2010),

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and antihelmintic activities (Kundu *et al.*, 2012). In recent times, the outbreak of drug-resistant diseases has led to several health issues. In an attempt to resolve these issues, pharmacological research has been tailored towards the discovery of innovative, potent, and safe drugs from natural compounds.

Despite the traditional use of *Senna alata* for medicinal purposes in humans and livestock, its impacts on aquatic life particularly fish species warrant investigation. In aquaculture, understanding the interactions between *Senna alata* and fish species like *Clarias gariepinus* (African catfish) is crucial due to the plant's widespread presence in aquatic ecosystems and its potential impacts on fish health and growth.

Biologically, the African catfish, *Clarias gariepinus* is the ideal aquaculture species in the world. It is widely distributed, thrives in diverse environments (temperate to tropical), and is highly adoptable and an ecological pioneer species, principally as a major consequence of its air-breathing ability. It feeds on wide array of prey and can adopt its feeding habit depending on the food availability. It is able to withstand adverse environmental conditions highly fecund and spawns under captive conditions (Otoh *et al.*, 2022, Otoh *et al.*, 2023; Otoh *et al.*, 2024 a, b, c, d). It has a wide tolerance of relatively poor water quality and possibly the most exciting features of its species ~~is~~ its potential for highly intensive culture without prerequisite pond aeration or high-water exchange rates and its excellent meat quality (Otoh *et al.*, 2024 a, b, c, d).

Conclusively, understanding the toxicity of *Senna alata* to *Clarias gariepinus* fingerlings is essential for sustainable aquaculture practices and environmental conservation efforts. Also, this paper will shed more light on the mechanism of action, potential effects and implication of *Senna alata* on fish health and ecosystem.

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2.0 Materials and Methods

2.1 Collection of Test organisms (*C. gariepinus*)

250 juveniles of *C. gariepinus* were procured from E. I farm Limited in Ikot Ekpene, Akwa Ibom State. An average weight of 38.58g and length ranging from 20 to 22cm were transported to Vika Farms Limited in plastic containers filled with oxygenated water and drops of palm oil to reduce stress and suffocation before reaching the farm. Collection and transportation were done between 4pm to 6pm to further prevent stressful conditions.

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2.2 Acclimatization of *Clarias gariepinus*

Test organisms were kept in a transparent plastic tank of length 18cm and width 13cm. The test organisms were acclimatized for 14 days (2 weeks) as recommended by APHA (2005). During this period, fishes were fed twice daily according to APHA (2005) at 5% body weight with commercial feeds (blue crown). Water was changed twice daily to remove faecal materials and unconsumed feeds. The tanks were covered with netting material to prevent fish from jumping out from the plastic container and also protected from predators. Feeding was discontinued during the test period.

2.3 Collection and Identification of *S. alata*

Fresh leaves of *S. alata* were gathered from Ndueso's Farm, Ukpap, Ikot Idang, Ikono, Akwa Ibom State southern Nigeria. The plant was identified by Mr. Etefia of Pharmacology Department (Pharmacognosy unit), University of Uyo.

2.4 Preparation of ethanol leaf extract of *Senna alata*

Extraction was done by solvent extraction technique according to Doughari (2012). Collected plant specimens were cleaned and mechanically cut into small pieces using knife and shade-dried to constant weight to obtain a dry matter. The dry matter is pulverized using a mortar and pestle to obtain finer particles of the dry matter.

After pulverizing, a known weight of the pulverized dry matter was macerated in 3L of 70% ethanol concentration for 72 hours, following the methods of Adewoye (2010). The macerated dry matter was filtered into a transparent plastic bucket carefully labeled using a cotton wool and filter paper to obtain the extract. Extract obtained was concentrated in a 1000ml beaker and was evaporated in a water bath at 40°C. Concentrated extracts were transferred using spatula into a beaker and stored with a covered with aluminum foil for bioassay immediately after the evaporation was completed.

2.5 Water Quality Tests

Water quality tests was carried out to ensure that optimum water conditions were maintained during, and that the results obtained were solely due to the effects of the treatments (toxicants) on the fish samples. The physico-chemical parameters of the test water with respect to temperature, dissolved oxygen (DO), pH, Total Dissolved Solids (TDS) and conductivity were determined using (Mercury in glass) thermometer, digital meters designed for the

respective purposes. The measurements were taken before and after acclimatization of organisms, before and after the introduction of toxicant and continued twice daily throughout the 96 ~~hrs~~-hours period of exposure of the bioassay.

2.6 Chronic Toxicity Bioassay

Chronic toxicity bioassay was done to determine the toxicity of the extract on the blood parameters and tissues of sensitive organs of the exposed fish. The bioassay was designed to be intermediate termed chronic test with 14 days exposure using sub-lethal concentrations of the treatment (0.0gm/, 0.18mg/L, 0.27mg/L, 0.4mg/L, 0.60mg/L, 0.90mg/L) 10 randomly selected juvenile of *Clarias gariepinus* was exposed for 21 days (Abalaka *et al.*, 2013).

The treatment was renewed every 24 hours and physicochemical parameters of the test media measure before and after every renewal (Ayuba *et al.*, 2012). At the end of the 21 days exposure, blood and tissue samples were collected from randomly selected fish from each treatment for further analysis.

2.7 Collection of Blood and Tissues Samples

Blood samples were collected from randomly selected fish from each treatment. The sample was collected from the fish using syringe and needle and preserved in EDTA treated bottles for hematological analysis. Fish tissues (gill, gut and liver) were obtained by dissecting open the lower part of the operculum to access the internal organs of fish. The organs of interest were carefully removed and preserved in sample bottles containing 40% formalin and properly labelled against the respective organs and treatments. The samples were subsequently taken to the University of Uyo teaching hospital histology laboratory for histology analysis.

2.8 Blood and Tissue Analysis

Collected blood samples were subjected to auto-haem analysis using auto-haem analyser in the haematology laboratory of University of Uyo Teaching Hospital, Uyo, Nigeria.- Blood parameters of interest (Hb, PCV, RBC, WBC and PLT) were analyzed. Histological analysis was carried out on the tissue samples using Haematoxylin and Eosin method.

2.9 Statistical Analysis

Toxicology dose response (fish mortality) was subjected to probit analysis using Statistical Package for Social Sciences (SPSS) 22.0. Data were analyzed using descriptive statistics. Comparison of data and physicochemical properties between the control and other treatments ~~was~~ were carried out using Analysis of Variance (ANOVA).

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3.0 Results

3.1 Initial Physicochemical Parameters of Test Water

Physicochemical parameters of the test water were observed to be within acceptable limit for survival and growth of *C. gariepinus* juveniles prior to commencement of the experiment (IBAR, 2015). Physicochemical parameters of the test water prior to commencement of the bioassay were tested and the values recorded. The values of the basic water quality parameters prior to stocking were, dissolved oxygen (5.60 mg/l), temperature (26.5°C), pH (7.16), TDS (7.1) and EC (12.6) (Table 1).

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Table 1: Initial Physico-chemical parameters of the test water prior to stocking of test organism

Fish Species	Initial physico-chemical parameters prior to stocking				
	DO (mg/l)	Temp (°C)	pH	TDS	EC
<i>Clarias gariepinus</i> juveniles	5.60	26.5	7.16	7.1	12.6

3.2 Variation of in Physico chemical Parameters of Test Media

The effect of *S. alata* leaf extract on the physico-chemical properties of the culture medium was assessed in the study (Table 2). During the 14-day chronic toxicity bioassay physicochemical parameters of the culture medium showed variations in values. No significant change ($p < 0.05$) in temperature was observed. DO and pH were observed to reduce significantly ($p < 0.05$) while TDS and Electric Conductivity were observed to increase significantly ($p < 0.05$). The variations were all observed to be concentration related. Duncan's statistical test revealed that changes in pH of test media were significant ($p < 0.05$) only at higher concentrations (0.60mg/L and 0.90mg/L).

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Table 2: Initial Physico-chemical parameters of the test water prior to stocking of test organism

Parameter						
Conc. (Mmg/L)	0.0	0.18	0.27	0.40	0.60	0.90
DO	5.50 ± 0.58 ^a	5.20 ± 0.28 ^a	4.80 ± 0.64 ^a	4.65 ± 0.67 ^{ab}	4.5 ± 0.85 ^b	4.4 ± 0.78 ^b
Ph	7.16 ± 0.12 ^a	5.25 ± 0.16 ^b	5.10 ± 0.04 ^b	4.93 ± 0.18 ^b	4.89 ± 0.32 ^{bc}	4.86 ± 0.32 ^{bc}
TDS	7.0 ± 0.71 ^a	13.0 ± 0.00 ^b	20.5 ± 0.35 ^{bc}	27.0 ± 2.13 ^{bc}	35.0 ± 2.13 ^c	44.0 ± 1.42 ^d
EC	12.5 ± 0.35 ^a	26.5 ± 0.35 ^b	38.0 ± 2.84 ^{bc}	58.0 ± 7.80 ^{bc}	70.5 ± 3.90 ^c	91.0 ± 4.96 ^d
Temp.	26.2 ± 0.14 ^a	26.2 ± 0.14 ^a	26.2 ± 0.14 ^a	26.2 ± 0.14 ^a	26.3 ± 0.07 ^a	26.2 ± 0.14 ^a

Mean with different superscripts along the same row are significantly different (Duncan's test) p<0.05

3.3 Blood Parameters of Test Media

Results of blood parameters of the respective test media during the period of exposure (14 days) is presented in Table 3. Results from blood analysis show that all the parameters measured, reduced in value when compared with the control. The reductions were observed to be related to the concentration of the extracts. Statistical analysis of data from blood parameters revealed that the observed reduction in the respective parameters (Hb, RCB and PVC) were statistically significant (p<0.05).

Table 3: Effects of *S. alata* on the Blood Parameters of *Clarias gariepinus* Juveniles

Parameter						
Conc. (Mmg/L)	0.0	0.18	0.27	0.40	0.60	0.90
Hb (g/L)	9.0 ± 0.24 ^a	8.8 ± 0.75 ^a	8.65 ± 0.45 ^a	8.45 ± 0.59 ^b	8.45 ± 0.59 ^b	8.1 ± 0.35 ^b
RBC	3.6 ± 0.31 ^a	2.9 ± 0.35 ^a	2.85 ± 0.17 ^a	2.80 ± 0.21 ^a	2.75 ± 0.04 ^{ab}	2.65 ± 0.11 ^b
PCV	34.1 ± 2.05 ^a	26.5 ± 3.19 ^{ab}	26.0 ± 1.42 ^b	25.5 ± 1.77 ^b	25.5 ± 0.35 ^b	24.5 ± 1.03 ^{bc}
PLT	28.5 ± 3.19 ^a	31.0 ± 0.70 ^a	32.5 ± 1.77 ^{ab}	36.0 ± 1.42 ^b	55.0 ± 4.25 ^{bc}	68.0 ± 3.74 ^c
WBC	35.8 ± 0.99 ^a	76.9 ± 0.78 ^b	98.15 ± 11.75 ^b	105.95 ± 5.85 ^{bc}	131.4 ± 0.21 ^c	146.4 ± 1.06 ^c

Mean with different superscripts along the same row are significantly different (Duncan's test) p<0.05.

4.3 Tissue Analysis

The photomicrographs of liver sections from the control group (I) and treatment groups (II–VI) reveal varying degrees of histopathological changes induced by *Senna alata* extract. The control group (I) displayed normal liver parenchyma, characterized by intact hepatocytes, normal-sized blood vessels, and the absence of lesions. In contrast, group II showed diffuse ballooning degeneration and focal areas of macrovesicular fatty changes, indicative of early hepatic distress. Group III mostly maintained normal histology, but with dilated and congested blood vessels, signaling some circulatory disturbance. Group IV exhibited enlarged hepatocytes and mild sinusoidal congestion, suggesting initial hepatic stress. Group V presented significant pathology with diffuse dilated and congested blood vessels and sinusoids, ballooning degeneration, and necrotic cells, indicating severe hepatic damage. Finally, group VI showed extensive damage with widespread congested sinusoids and necrotic hepatocytes, reflecting the most severe hepatic injury (Fig. 1).

The intestinal histology across all groups (I–VI) displayed the lumen (L), mucosa (M) with increased villi (thin arrows), submucosa (SM), and muscularis externa (ME). There were no pathological changes observed in either the control group (I) or the experimental groups (II–VI) (Fig. 2).

The photomicrograph of the transverse section of the fish gill arch revealed distinct histological features across the control group and treatment groups. In the control group, the gill histology appeared normal, with centrally supporting cartilaginous tissue surrounded by skeletal muscle and basophilic connective tissue. The primary filaments were vascularized and exhibited sporadic fatty changes, indicating normal physiological variation. In treatment group II, the gill sections showed few fatty changes in the

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primary filament, suggesting minimal impact from the extract. However, treatment groups III and IV exhibited more pronounced pathological alterations, including diffused fatty changes across the primary lamella, multiple areas of epithelium degeneration, and congested blood vessels, indicating significant tissue stress and damage. Group V depicted focal epithelium degeneration, while group VI displayed increased fatty lobules in the primary filaments, reflecting extensive fat deposition and potential dysfunction in gill tissue. Group V depicted focal epithelium degeneration, while group VI displayed increased fatty lobules (black arrow) in the primary filaments, reflecting extensive fat deposition and potential dysfunction in gill tissue (Fig. 3).

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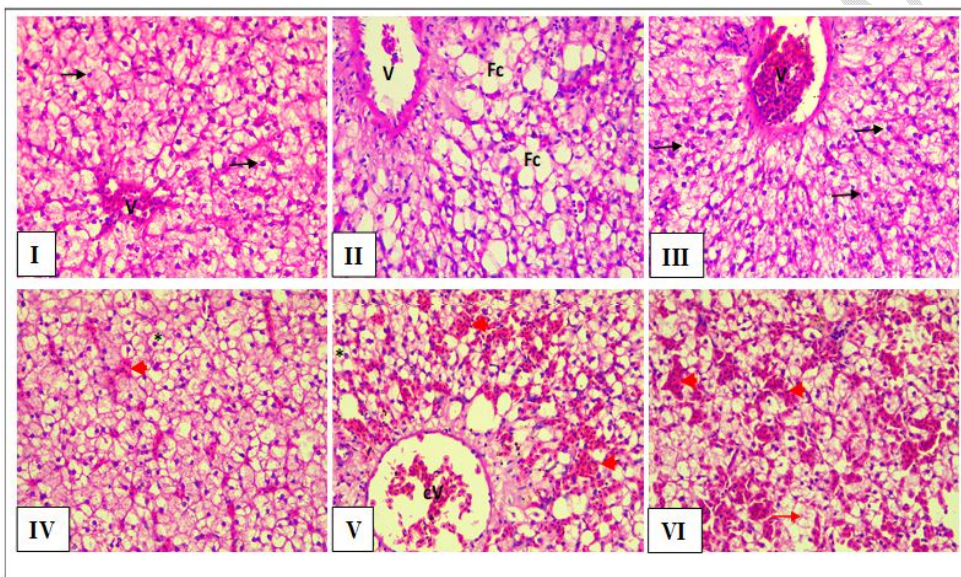


Figure 1: The photomicrograph of the liver section across the groups. **(I)** Normal liver parenchymal showing hepatocytes (**arrow**), normal sized blood vessel (**V**), no lesion seen. **(II)** showed diffused ballooning degeneration and focal area of macrovesicular fatty changes (**Fc**). **(III)** showed normal section with dilated congested blood vessel (**CV**). **(IV)** showed enlarged hepatocytes and mildly congested sinusoid. **(V)** depicts diffused dilated congested blood vessels and sinusoid, ballooning degeneration and necrotic cells. **(VI)** showed diffused congested sinusoid and necrotic hepatocytes. Haematoxylin and Eosin (H&E) stain, x400 magnification

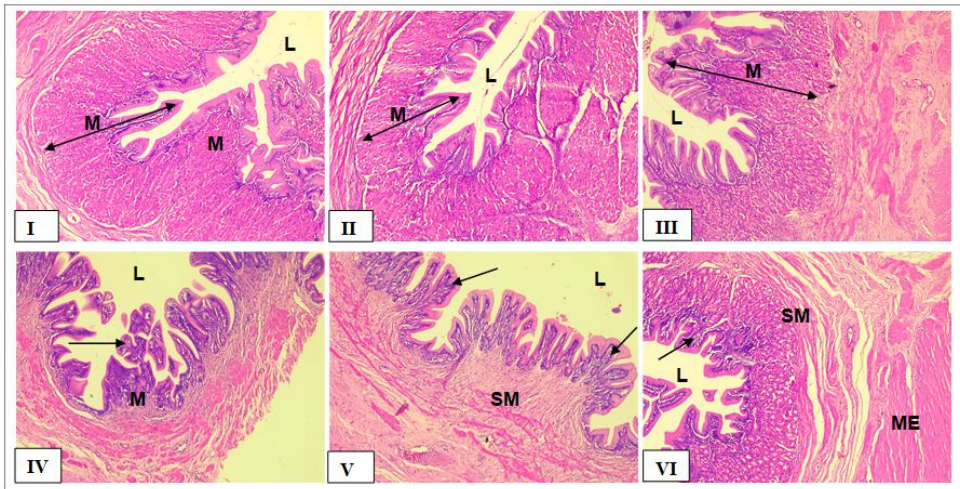


Figure 2: The section of intestinal tissue across the groups (I-VI). The section showed lumen (L), the mucosa (M) showed increased villi (Thin arrows) and the submucosa (SM). Also seen was muscular externa (ME); No pathologic changes seen in the control group (I) and the experimental groups (II-VI). Haematoxylin and Eosin (H&E) stain, x100 magnification.

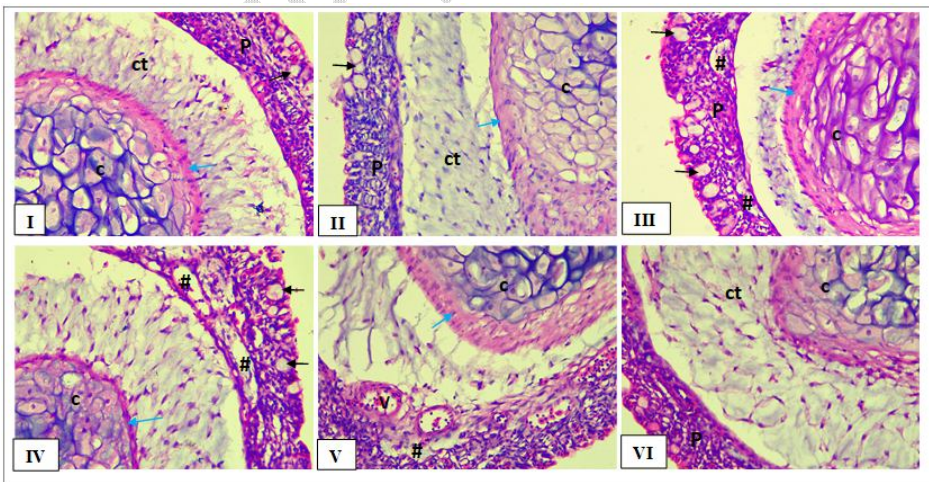


Figure 3: Photomicrograph of the transverse section of the fish gill arch of the Control (I) and the treatment groups (II-VI). (I) showed normal gill histology with centrally supporting cartilaginous tissue (C) surrounded by skeletal muscle (blue arrow) and basophilic connective tissue (Ct). The primary filaments (P) were vascularized and showed sporadic fatty changes (black arrow). (II) showed few fatty changes (black arrow) in the primary filament. (III) & (IV) showed diffused fatty changes across the primary lamella and multiple area of epithelium degeneration (#) and congested blood vessels (red arrowhead). V depict a focal epithelium degeneration, while VI showed increase fatty lobules (black arrow) in the primary filaments.

4.0 Discussion

The *Senna alata* exerted toxic effect on the fish in the present study and toxicity increased with increased concentration. Abnormal behavior such as restlessness, incessant jumping and gulping for air, loss of equilibrium, increase in opercular activities, surface to bottom movement, sudden quick movement and resting at the bottom of the aquaria and death observed in this study were similar to the findings of George *et al.*, 2024 a; b; c; Essien-Ibok *et al.*, 2024).

—The abnormalities observed prior to mortality are an indication of depleted oxygen content due to higher demand for oxygen. Consequently, it was observed in this study that the abnormal behavior and mortality rate of the test organism's increased with increase in concentrations of toxin. This corresponds to the findings of Shobha (2007) that the behaviour and mortality rate of *C. gariepinus* during experiment was found to depend on both duration of exposure and concentration of the toxicant. Similar observations were also, reported by George, *et al.*, 2023 a; b; c; d) in a related study.

—The introduction of the leaf extract at different concentrations impairs the swimming pattern, skin colouration and general behaviour of fish. Also, the variation in the behavioural responses and mortality in the sublethal test can be attributed to the low level of accumulation of the toxicant. This suggests that fish can tolerate low concentrations of pollutants with reduced mortality.

The reductions in blood cell indices and tissue deformation observed from the chronic bioassay are in line with findings of Okogwu *et al.*, (2015) during their study on behavioural, haematological and histopathological changes in *C. gariepinus* exposed to 2,4-dichlorophenoxyacetic acid. Similar observations have also been reported in a study on exploitation of ethanol extract of *Adenium obesum* stem bark as a potent organic piscicide (Abalaka *et al.*, 2013). In a study conducted on haematological responses, Serum Biochemistry and Histology of *C. gariepinus*, exposed to sublethal concentrations of cold water fresh root bark extract of *Plumbago zeylanica* (Leadwort), it was reported that, exposure of fish to the toxicant for 21 days resulted in remarkable anaemic condition in *C. gariepinus* resulting from reductions in values of haematological parameters (RBC, Hb, PCV) in the juveniles of *C. gariepinus* exposed to *P. zeylanica* extract (Olusegun and Adadayo, 2014). Olusegun and Adadayo (2014) also reported histopathological alterations observed in the brain, gills, liver, intestine and muscle/flesh of the *C. gariepinus* juveniles which align with the results of the present findings.

Stressors evoke non-specific responses in fish which enables the fish to cope with the disturbance and maintain its homeostatic state (Crook, 2012). If severe or long lasting, the response then becomes maladaptive and threatens the fish health and well-being. Therefore, in the presence of stressors (contaminants / pollutants), blood parameters and blood chemistry can be employed as standard laboratory test to determine diseased conditions and metabolic disturbances in fish (Crook, 2012). Blood is a tissue fluid and serves as transport medium whose primary function is to supply oxygen and nutrients as well as constitutional elements to tissues and to remove waste products. Blood also enables hormones and other substances to be transported between tissues and organs. Blood is basically composed of the plasma, red blood cells, white blood cells and platelets, each with sub constituents which collectively contribute to the overall functioning of the blood (Crook, 2012). The WBCs help the immune system and as well remove toxins, waste, foreign bodies and abnormal or damaged cells by phagocytosis. Platelets secrete factors that increase local platelet aggregation e.g., Thromboxane A, enhance vasoconstriction e.g., serotonin and promote blood coagulation e.g., Thromboplastin (Aqra, 2012).

These observations in blood parameters confirm concentration related toxicity of the toxicant to the test organism (Gabriel *et al.*, 2007). The observed reduction in the respective blood parameters results in diseased conditions such as haemolytic anaemia, leukemia and lymphopaenia in exposed fish resulting from interference with blood cells formation process (erythropoiesis) and destruction of blood cells by the toxicants (Crook, 2012).— Haematological changes as recorded in this study are attributed to rapid destruction of the red blood cells by the toxicant and the increase in white blood (WBC) was probably a defense mechanism deployed by the fish to protect itself from assault of *Senna alata*.

Fish gills are the prime target organ of all pollutants due to their extensive surface in contact with the external medium and the reduced distance between the external and internal medium. Gill morphology and morphometric are important biomarkers providing a rapid method for detection of the effects of pollutants (Fafioye, 2012). In

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histopathological studies, gills have also been reported to act as storehouse for bio-accumulating toxicants (Fafioye, 2012). The general morphological changes in the gills recorded in this study have been reported in *Astyanax sp.* The histological examination of gill sections from juvenile *Clarias gariepinus* treated with varying concentrations of *Senna alata* extract demonstrated a clear dose-dependent response. The control group maintained normal gill architecture with minimal fatty changes, while the treatment groups exhibited a progression of pathological features with increasing extract concentration. Group II's minor fatty changes contrasted sharply with the severe alterations in groups III and IV, including widespread fatty changes, epithelial degeneration, and vascular congestion. These changes indicate significant gill tissue stress and compromised function. Group V's focal epithelial degeneration and group VI's extensive fatty lobules further underscore the potential toxicity of higher extract concentrations. These findings suggest that while lower doses of *Senna alata* extract may be relatively safe, higher doses pose a risk of significant gill damage, highlighting the need for careful dose optimization in aquaculture applications.

Vascular changes in gills of exposed fish could be attempts by the fish to supply more blood to the gills to increase oxygen uptake and supply to the internal organs. The gill of fish is a multipurpose organ that, in addition to providing for aquatic gaseous exchange, plays a dominant role in osmotic and ionic regulation, acid-base regulation and excretion of nitrogenous waste. Thus, despite the fact that all fish groups have functional kidneys, the gill epithelium is the site many processes that are mediated by the renal epithelia in terrestrial vertebrates. Hence, impairment of the gill functions by the overall effect of the pathological changes in the gill of exposed fish will have grave consequences for the fish with respect to the normal functions of the gills (Gabriel, 2007; George *et al.*, 2014 a; b).

The intestine tissue is responsible for storage of food and package of food items in the fish. In the present study, it is evident that the gut tissue without treatment shows the normal mucosa, submucosa and muscular externa layer with no pathologic changes while that of the treatment revealed photomicrographs of the intestine showing the lumen with the mucosa and a dilated congested blood vessel in the submucosa.

The liver is the primary organ for metabolism, detoxification of xenobiotics and excretion of harmful substances. Glucose is stored in the liver as glycogen and is returned to the blood via circulation when needed to maintain stable blood sugar levels. Other than glycogen, the liver stores fat soluble vitamins. The liver has the ability to degrade toxic compounds but its regulatory mechanism can be overwhelmed by elevated concentration of toxic substance which could subsequently result in structural damage. The histopathological findings demonstrate a dose-dependent hepatotoxic effect of *Senna alata* extract on juvenile *Clarias gariepinus*. While the control group maintained normal liver structure, the treatment groups exhibited varying degrees of hepatic alterations with increasing severity correlating with higher extract concentrations. Group II's early signs of hepatotoxicity progressed to moderate hepatic stress in groups III and IV, as evidenced by vascular congestion and cellular hypertrophy. The most severe damage in groups V and VI, marked by extensive congestion, ballooning degeneration, and necrosis, underscores the extract's potential hepatotoxic effects at higher doses. These results highlight the necessity for careful dosage determination of *Senna alata* extract to avoid liver toxicity in aquaculture applications. Further research is essential to understand the mechanisms driving these changes and to establish safe and effective usage guidelines for *Senna alata* in fish health management. The liver of freshwater fish is known to accommodate a high level of toxicants (Musa, 1999 and Ojutiku, 2013).

Physicochemical parameters such as Temperature, Dissolved oxygen, pH, total dissolved solids and electric conductivity are vital aquatic indices which determine the fish health, growth and reproduction. pH decreases because the extract is biodegradable and when it is much in water biological activities degrades it and microbial activities will occur which uses oxygen and more of the oxygen present in the water would be used by the microorganisms to decompose the extract and that is why there is vast depletion of dissolved oxygen, because of the aerobic process. The pH decreases because the extract is acidic pushing the pH of the treatment to acidity and reducing the alkalinity. TDS and electronic conductivity are increasing with increased concentration because of the presence of more particles in the water while the temperature did not get affected because there was nothing to influence the atmospheric temperature in the room.

4.1 Conclusion

In conclusion, the chronic toxicity studies on *Senna alata* conducted on juvenile *Clarias gariepinus* have unveiled notable alterations in both haematological parameters and histopathological features, particularly evident in the gills, liver and intestine of treated groups. These findings underscore the potential adverse effects of prolonged exposure to *Senna alata* on the physiological and cellular integrity of the fish. Intriguingly, the absence of pathological changes in the control groups suggests a distinct response attributable to the treatment. This highlights the importance of continued monitoring and assessment of the impacts of *Senna alata* on aquatic organisms and ecosystem, emphasizing the need for further research to elucidate the underlying mechanisms and potential long-term consequences. Such insights are crucial

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for informed decision-making regarding the management and regulation of *Senna alata* and its interaction with aquatic environments.

5.0 References

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